



Article Mechanisms of Chitosan Nanoparticles in the Regulation of Cold Stress Resistance in Banana Plants

Anbang Wang ¹, Jingyang Li ¹¹, Arwa Abdulkreem AL-Huqail ², Mohammad S. AL-Harbi ³, Esmat F. Ali ³, Jiashui Wang ¹, Zheli Ding ¹, *¹, Saudi A. Rekaby ⁴, Adel M. Ghoneim ⁵ and Mamdouh A. Eissa ⁶, *¹

- ¹ Hainan Banana Healthy Seedling Propagation Engineering Research Center, Haikou Experimental Station, Chinese Academy of Tropical Agricultural Sciences (CATAS), Haikou 571101, China; wanganbang011@126.com (A.W.); jingyanglee@163.com (J.L.); jiashuiwang@126.com (J.W.)
- ² Department of Biology, College of Science, Princess Nourah Bint Abdulrahman University, Riyadh 13324-8824, Saudi Arabia; aaalhuqail@pnu.edu.sa
- ³ Department of Biology, College of Science, Taif University, Taif 21944, Saudi Arabia; mharbi@tu.edu.sa (M.S.A.-H.); a.esmat@tu.edu.sa (E.F.A.)
- ⁴ Department of Soils and Water, Faculty of Agriculture, Al-Azhar University (Assiut Branch), Assiut 71524, Egypt; saudirekaby.4419@azhar.edu.eg
- ⁵ Agricultural Research Center, Field Crops Research Institute, Giza 12112, Egypt; adelrrtc.ghoneim@gmail.com

Abstract: Exposure of banana plants, one of the most important tropical and subtropical plants,

- ⁶ Department of Soils and Water, Faculty of Agriculture, Assiut University, Assiut 71526, Egypt
- Correspondence: dingzheli@zju.edu.cn (Z.D.); mamdouh.eisa@aun.edu.eg (M.A.E.)



Keywords: chitosan nanoparticles; chilling tolerance; antioxidant enzyme activities; membrane damage traits

1. Introduction

Banana plants (*Musa* spp) belong to the *Musaceae* family and are widely cultivated in the tropical and subtropical regions of the world due to their high nutritive value, as well



Citation: Wang, A.; Li, J.; AL-Huqail, A.A.; AL-Harbi, M.S.; Ali, E.F.; Wang, J.; Ding, Z.; Rekaby, S.A.; Ghoneim, A.M.; Eissa, M.A. Mechanisms of Chitosan Nanoparticles in the Regulation of Cold Stress Resistance in Banana Plants. *Nanomaterials* **2021**, *11*, 2670. https://doi.org/10.3390/ nano11102670

Academic Editor: Dalibor Húska

Received: 28 August 2021 Accepted: 8 October 2021 Published: 11 October 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). as their economic importance [1]. Banana fruits are a basic commodity and one of the most important sources of food in both developed and developing countries as they are are rich in carbohydrates, vitamins, and protein in addition to having high nutrient content, notably potassium, phosphorus, and calcium [2]. Climate changes and the continuous increase in the world population have led to an increase in the demand for food [3,4]. Bananas are grown in a wide range of tropical and subtropical regions around the world but their growth is negatively affected by various biotic and abiotic stresses, especially those caused by climate changes [5,6]. Chilling is one of the most important phenomena accompanying the emergence of climate changes, and it is also one of the most important reasons limiting the expansion of banana cultivation to meet the increase in food demand [7–9].

Low temperatures during the growth period of fruit trees lead to many physiological disorders within the plant cells, which have a negative impact on the quality of the fruit and result in a reduction of their marketing values [9,10]. There are many factors that affect the expansion of banana cultivation in the world, the most important of which is cold stress, which may cause losses of up to more than 50% of the yield, and this problem is exacerbated under climatic changes [8]. Banana plants are considered sensitive to cold stress, and when temperatures drop below the critical point (11 °C), physiological disorders occur, and the sensitivity differs depending on the variety [7,11]. The physiological disorders caused by cold stress negatively affect plant growth, [7,11]. Cold stress reduces the photosynthesis rate and causes various forms of damage in the cell membrane [12]. Banana plants tolerate cold stress by increasing the compatible solutes; e.g., proline, soluble carbohydrates, and phenolic compounds [7,8]. Various materials have been used to increase cold stress resistance and reduce damage in economically important plants [8,13]. Li et al. [13] used chitosan to increase the cold stress resistance of tea plants (Camellia sinensis), and found that chitosan increased plant tolerance to chilling. Currently, a technological revolution is taking place through the use of nanomaterials to increase plant resistance in the face of different abiotic stresses; despite this, there is not much information available on the effect of CH-NPs in increasing the resistance of bananas to chilling stress.

Nanotechnology can be used to address many agricultural and environmental challenges, and it can also be used as an effective tool for energy and resource constraints, the sustainable use of resources, urbanization, and fertilizer management [14]. With an everincreasing population, there is a need for higher crop yields and more efficient strategies for improving agricultural practices, and thus the use of nanomaterials in agricultural science is on the rise [15]. Indeed, nanoparticle (NP) compounds may have a significant impact on sustainable agriculture and the development of precision agriculture by maximizing agricultural output (i.e., crop yields) and minimizing inputs (i.e., fertilizers, pesticides, and herbicides) via control of environmental conditions and the application of proper management [16]. NPs have properties distinct from those of bulk materials due to their small size and larger surface area which, as a result, mean that their solubility and surface reactivity are higher [17–20].

Chitosan (CH), produced from the polysaccharide chitin, has many benefits as an ecofriendly polymer in agricultural, biomedical, and fodder production [12,21,22]. CH-NPs are used for many purposes due to their high permeability, eco-friendliness, safety for living organisms, low price, and high solubility and biodegradability [23]. Many recent studies have recommended using CH-NPs to increase the ability of crops and economically important plants to address different stress conditions, such salinity, drought, metal toxicity, and heat [24,25].

The role of chitosan oligosaccharide in increasing the cold stress resistance of tea plants (*Camellia sinensis*) was studied by Li et al. [13]. They found that chitosan increased the accumulation of compatible solutes and the antioxidant defense of tea plants. However, to the best of our knowledge, there are no studies investigating the role of CH-NPs in the fight of banana plants (*Musa* spp) against cold stress. Therefore, the current study investigated the effects of the foliar application of CH-NPs in minimizing the negative

impacts of chilling in banana plants. We assumed that the use of CH-NPs would increase the cold stress in banana plants by upregulating the defense mechanisms.

2. Material and Methods

2.1. Plant Materials and Chitosan Nanoparticles (CH-NPs)

Banana plants of the Baxi variety (*Musa acuminata* var. Baxi) produced with a tissue culture technique were used in the current study [7]. The plants were bought from the National Research Centre. The plants were three months old (40 cm high, 10 leaves) and cultivated in a growing media consisting of sand and compost (3:1 w/w). Twenty uniform, healthy plants were selected for the current experiment. Chitosan was bought from Sigma-Aldrich (St. Louis, MO, USA) and converted into nanoparticles in the Nanotechnology and Advanced Materials Laboratory of the National Research Centre (ARC). The CH-NPs had 99% purity, a specific surface area of 50–80 m² g⁻¹, and a particle size less than 100 nm. Banana plants were sprayed with four different concentrations of nanoparticles, i.e., 0, 100, 200, and 400 mg L⁻¹ of CH-NPs in deionized water.

2.2. Chilling Treatments

Chilling treatment was undertaken in a plant growth chamber (LGP-250E, Shanghai, China). The chamber had a volume of 250 L and the plant growth conditions were automatically controlled. The selected plants were placed in the plant growth chamber under normal growth conditions ($25 \pm 1 \,^{\circ}$ C, $60 \pm 5\%$ relative humidity, 12 h photoperiod with a PPFD of 250 µmol/(m²·s)). The plants were supplied with water and nutrient solution (50% Hoagland nutrient solution) [26]. The NP solutions used were prepared from a 1 g L⁻¹ CH-NP stock solution: 1% (w/v) CH-NP solution was first prepared in 5% acetic acid (v/v) and, after stirring, the solution was diluted to 1:10 with deionized water. The final concentrations of 100, 200, and 400 mg L⁻¹ were achieved by diluting the stock solution in deionized water. The plants in each treatment were sprayed with 250 mL of CH-NP solution containing Tween-80 (40 mL L⁻¹). Seedlings of banana plants were sprayed with four levels of CH-NP solution for three continuous days, then the temperature in the incubator was set at 5 °C for 72 h without changes in the other growth conditions. A group that had not been cold stressed or undergone CH-NP treatment was used as control.

2.3. Plant Samples Analysis

The photosynthetic pigments (chlorophyll a + b and carotenoids) were extracted from the fresh leaves with acetone (80%) and then measured using a modified method from a study by Lichtenthaler and Buschmann [27]. The procedure described by Bates et al. [28] was used to determine the free proline in the fresh leaves of banana plants. Free proline was extracted from the fully expanded fresh leaf (0.1 g) with 3% sulfosalicylic acid (w/v). A method modified from the work of Ainsworth and Gillespie [29] was used in the measurement of the total phenolic compounds. Phenolic compounds were extracted from the banana leaves using methyl alcohol (85%). Soluble carbohydrates were extracted with ethanol (80%) and the total soluble carbohydrates were measured using the anthrone reagent method [7,30]. Two grams of each dried plant sample was digested using a mixture of H_2O_2 , Se, Li₂SO₄, and concentrated H_2SO_4 [31]. The nitrogen in the plant sample extracts was measured using the Kjeldahl distillation method [32]. The phosphorus (P), potassium (K), calcium (Ca), iron (Fe), manganese (Mn), zinc (Zn), and copper (Cu) in the digested plant samples were determined using an inductively coupled plasma emission ICP 6200 (ICP) [32]. Reactive oxygen species (ROS), i.e., hydrogen peroxide (H₂O₂), hydroxyl radicals ($^{\bullet}OH$), and superoxide anions ($O_2^{\bullet-}$), were determined using the method described by Velikova et al. [33] and Kubiś [34]. Hydrogen peroxide (H_2O_2) was extracted from fresh banana leaves with trichloroacetic acid (5%), then the absorbance reading was determined at 390 nm using spectrophotometer [33]. Superoxide anions $(O_2^{\bullet-})$ were extracted from the leaf cuts using the K-phosphate buffer method, then the optical density

was recorded at 580 nm [34]. Malondialdehyde (MDA) was determined based on the method described by Madhava and Sresty [35]. A fresh leaf sample (0.5 g) was extracted with 5 mL of 0.1% trichloroacetic acid (TCA) and then centrifuged, and the supernatant was added to 1 mL of 0.5% thiobarbituric acid in 20% TCA. Malondialdehyde concentration was calculated from the extinction coefficient of 155 mM⁻¹ cm⁻¹ [35]. Hydroxyl radicals (OH–) were extracted from 0.5 g of fresh sample with 10 mM Na phosphate buffer (pH 7.4) and 1% TBA, and then checked using a spectrophotometer at 550 nm [36]. The superoxide dismutase (SOD) activity was measured in 0.1 mM EDTA, sodium carbonate buffer (pH 10.2), epinephrine, and enzymatic extract, as described by Abeed et al. [37]. The peroxidase (POD) activity was measured spectrophotometrically following Abeed et al. [37].

2.4. Data Analysis

One-way ANOVA was used to test the significance of differences between the treatments. Means of treatments were compared using Tukey's test at a 5% level of probability. The results in the figures and tables are shown as means (\pm SD, *n* = 5). All the statistical analyses were run in SPSS 17.0 (SPSS, Chicago, IL, USA).

3. Results

3.1. Effect of CH-NPs on the Growth and Nutrient Content of Banana Plants

Cold stress significantly reduced the growth of banana plants compared to the control treatment (C) (Figure 1). In this study, the foliar application of CH-NPs induced significant increases (p < 0.05) in the fresh and dry weights of banana plants under cold stress when compared to the unsprayed plants. The fresh and dry weights of banana plants treated with CH-NPs (100 mg L⁻¹) were increased by 3 and 18%, respectively, when compared to the unsprayed plants, and these increases amounted to 7 and 30% in the case of the 200 mg L⁻¹ treatment. Spraying banana plants exposed to cold stress with the highest concentration of CH-NPs (400 mg L⁻¹) caused remarkable and significant increases in the fresh and dry weights, with increases of 14 and 41%, respectively, compared to the untreated plants.



Figure 1. Effect of chitosan nanoparticles (CH-NPs) on the fresh and dry weights of banana plants exposed to cold stress (5 °C for 72 h). C = control treatment without cold stress and without CH-NP amendment Different letters refer to significant differences between treatments according to Tukey's test at p < 0.05. Each value is the mean of five replicates (mean \pm SD, n = 5).

NPs (100 mg L⁻¹) induced increases in the concentrations of N, P, K, Ca, and Fe of 13, 12, 28, 27, and 4%, respectively, compared to the unsprayed plants. In the case of the 200 mg L⁻¹ treatment, these increases were 19, 8, 33, 33, and 4%, respectively. Spraying banana plants with the highest concentration of CH-NPs (400 mg L⁻¹) increased the N, P, K, Ca, and Fe by 19, 12, 44, 40, and 5%, respectively, compared to the untreated plants.

Table 1. Effect of chitosan nanoparticles (CH-NPs) on the elemental composition of leaves of banana plants exposed to cold stress (5 °C for 72 h).

CH-NPs mg L ⁻¹	Ν	Р	К	Ca	Fe	Mn	Zn	Cu
	(g kg ⁻¹)	(g kg ⁻¹)	(g kg ⁻¹)	(g kg ⁻¹)	(mg kg $^{-1}$)	(mg kg $^{-1}$)	(mg kg $^{-1}$)	(mg kg $^{-1}$)
С	42 ± 4 a	2.8 ± 0.2 a	30 ± 2 a	23 ± 2 a	215 ± 12 a	152 ± 5 a	133 ± 6 a	57 ± 3 a
0	$32\pm2d$	$2.5\pm0.1b$	$18\pm1~d$	$15\pm1~{ m c}$	$200\pm4~{ m c}$	$150\pm7~\mathrm{a}$	$128\pm5~\mathrm{a}$	$56\pm3~\mathrm{a}$
100	$36\pm3~c$	2.8 ± 0.2 a	$23\pm2c$	$19\pm1\mathrm{b}$	$208\pm5b$	154 ± 6 a	134 ± 7 a	57 ± 4 a
200	$38\pm2b$	2.7 ± 0.3 a	$24\pm3~{ m c}$	$20\pm3b$	$207\pm6b$	152 ± 5 a	125 ± 4 a	$56\pm3~\mathrm{a}$
400	$38\pm3b$	$2.8\pm0.3~\mathrm{a}$	$26\pm3b$	$21\pm2b$	$210\pm8~a$	155 ± 4 a	$135\pm 6~\mathrm{a}$	58 ± 2 a

C = control treatment without cold stress and without CH-NP amendment. Different letters in the same column refer to significant differences between treatments according to Tukey's test at p < 0.05. Each value is the mean of five replicates (mean \pm SD, n = 5).

3.2. Effect of CH-NPs on the Secondary and Osmo-Metabolic Compounds

Cold stress increased the studied biochemical traits compared to the normal growth condaition. The foliar application of CH-NPs clearly increased the biochemical traits in the leaf tissue of banana plants grown under cold stress in comparison to the untreated plants (Table 2). Foliar application of all the studied concentrations of CH-NPs significantly enhanced the accumulation of the total phenolic compunds, soluble carbohydrates, proline, and amino acids in the leaf tissue of the banana plants grown under cold stress. Spraying banana plants with 100 mg L⁻¹ of CH-NPs increased the total phenolic compounds, soluble carbohydrates, proline, and amino acids by 41, 31, 25, and 42%, respectively, compared to the unsprayed plants. Spraying Baxi banana plants with 200 mg L⁻¹ of CH-NPs increased the total phenolic compounds, soluble carbohydrates, proline, and amino acids by 41, 31, 25, and 42%, respectively, compared to the total phenolic compounds, soluble carbohydrates, proline, and amino acids by 41, 31, 25, and 42%, respectively, compared to the unsprayed plants. Spraying Baxi banana plants with 200 mg L⁻¹ of CH-NPs increased the total phenolic compounds, soluble carbohydrates, proline, and amino acids by 69, 33, 37, and 62%, respectively, compared to the control. Spraying banana plants with 400 mg L⁻¹ of CH-NPs increased the phenolic compounds, soluble carbohydrates, proline, and amino acids by 79, 43, 50, and 112%, respectively, compared to the unsprayed plants. CH-NPs at all the studied concentrations increased the secondary and osmo-metabolic compounds in the leaves of the banana plants exposed to cold stress (5 °C for 72 h).

Table 2. Effect of chitosan nanoparticles (CH-NPs) on some biochemical traits in the leaves of banana plants exposed to cold stress (5 $^{\circ}$ C for 72 h).

CH-NPs	Phenolic Compounds	Soluble Carbohydrates	Proline	Amino Acids					
(ling L)	(mg g ⁻¹)								
С	$2.02\pm0.8~\mathrm{d}$	$22.52\pm1.23~\mathrm{c}$	$3.00\pm0.07~\mathrm{e}$	$22\pm2~{ m e}$					
0	$2.45\pm0.05~\mathrm{c}$	$26.57\pm1.53~\mathrm{b}$	$3.12\pm0.09~d$	$26\pm2d$					
100	$3.46\pm0.06~b$	$34.68\pm2.93~\mathrm{a}$	$3.89\pm0.13~\mathrm{c}$	$37\pm3~{ m c}$					
200	$4.15\pm0.09~\mathrm{a}$	$35.23\pm2.48~\mathrm{a}$	$4.27\pm0.10\mathrm{b}$	42 ± 3 b					
400	$4.39\pm0.04~\mathrm{a}$	$38.12\pm3.59~\mathrm{a}$	$4.69\pm0.12~\mathrm{a}$	55 ± 4 a					

C = control treatment without cold stress and without CH-NP amendment. Different letters in the same column refer to significant differences between treatments according to Tukey's test at p < 0.05. Each value is the mean of five replicates (mean \pm SD, n = 5).

3.3. Effect of CH-NPs on the Photosynthesis Pigments and Antioxidant Enzyme Activities

Spraying banana plants with CH-NPs significantly (p < 0.05) increased the photosynthesis pigments and antioxidant enzyme activities (Figure 2). Foliar application of CH-NPs significantly enhanced the carotenoids and total chlorophyll (a + b) in the leaf tissue of banana plants grown under cold stress. Spraying banana plants with 100 mg L⁻¹ of CH-NPs increased the carotenoids and chl (a + b) by 10 and 28%, respectively, compared to the unsprayed plants. Spraying banana plants with 200 mg L⁻¹ of CH-NPs increased the carotenoids and chl (a + b) by 26 and 36%, respectively, compared to the unsprayed plants. Spraying banana plants with 400 mg L⁻¹ of CH-NPs increased the carotenoids and chl (a + b) by 46 and 52%, respectively, compared to the unsprayed plants. Foliar application of chitosan nanoparticles (CH-NPs) significantly enhanced the peroxidase (POD) and superoxide dismutase (SOD) in the leaf tissue of banana plants grown under cold stress.



Figure 2. Effect of chitosan nanoparticles (CH-NPs) on photosynthesis pigments (Chl (a + b) and carotenoids) and antioxidant enzymes superoxide dismutase (SOD) and peroxidase (POD) of banana plants exposed to cold stress (5 °C for 72 h). C = control treatment without cold stress and without CH-NPs amendment. Different letters refer to significant differences between treatments according to Tukey test at p < 0.05. Each value is the mean of five replicates (mean \pm SD, n = 5).

Spraying banana plants with 100 mg L⁻¹ of CH-NPs increased the peroxidase (POD) and superoxide dismutase (SOD) by 8 and 17%, respectively, compared to the unsprayed plants. Spraying banana plants with 200 mg L⁻¹ of CH-NPs increased the peroxidase (POD) and superoxide dismutase (SOD) by 13 and 24%, respectively, compared to the untreated plants. Spraying banana plants with 400 mg L⁻¹ of CH-NPs increased the peroxidase (POD) and superoxide dismutase (SOD) by 38 and 31%, respectively, compared to the control. CH-NPs at all the studied concentrations significantly (p < 0.05) increased photosynthesis pigments and antioxidant enzyme activities in the leaves of banana plants exposed to cold stress (5 °C for 72 h).

3.4. Effects of CH-NPs on the Stress Markers and Membrane Damage Traits

Reactive oxygen species (ROS), i.e., hydrogen peroxide (H₂O₂), hydroxyl radicals ($^{\circ}$ OH), and superoxide anions (O₂ $^{\circ-}$), were determined and the data are shown in Figure 3. Furthermore, the malondialdehyde (MDA) was determined in order to explore the membrane damages, and the data are shown in the same figure.



Figure 3. Effects of chitosan nanoparticles (CH-NPs) on malondialdehyde (MDA) and reactive oxygen species (superoxide anions ($O_2^{\bullet-}$), hydroxyl radicals ($^{\bullet}OH$), and hydrogen peroxide (H_2O_2)). C = control treatment without cold stress and without CH-NP amendment. Different letters refer to significant differences between treatments according to Tukey's test at p < 0.05. Each value is the mean of five replicates (mean \pm SD, n = 5).

The highest significant values (p < 0.05) for MDA and ROS were recorded in banana plants exposed to cold stress (5 °C for 72 h) without the addition of CH-NPs. Foliar application of chitosan nanoparticles (CH-NPs) significantly decreased MDA and ROS in the leaf tissue of banana plants grown under cold stress. Spraying banana plants with 100 mg L⁻¹ of CH-NPs decreased the MDA, H₂O₂, •OH, and O₂•- by 20, 32, 39, and 46%, respectively, compared to the unsprayed plants. Spraying banana plants with 200 mg L⁻¹ of CH-NPs decreased the MDA, H₂O₂, •OH, and O₂•- by 27, 29, 36, and 45%, respectively, compared to the untreated plants. Spraying banana plants with 400 mg L⁻¹ of CH-NPs decreased the MDA, H₂O₂, •OH, and O₂•- by 33, 33, 40, and 48%, respectively, compared to the unsprayed plants. In Figure 3, there is a clear trend indicating decreasing ROS and membrane damage traits in response to foliar application of all CH-NP concentrations.

4. Discussion

Climate change is considered to be a phenomenon that has recently increased in importance due to its negative effects on the growth of field crops and the quality of the crop yield, which directly affect the availability of food [38,39]. Low temperatures are one of the manifestations of these climatic changes that negatively affect the productivity of various crops, especially banana plants, as they are highly susceptible to cold stress [7,40,41]. Chilling harshly limits the physiological and biochemical processes in plant cells, causing leaf chlorosis, wilting, and even necrosis [42]. In the current study, the lowest significant growth values of the banana plants were recorded in the plants exposed to cold stress (5 °C for 72 h) without the addition of chitosan nanoparticles (CH-NPs). Cold stress occurs in banana plants when the temperature drops below the critical point (5–11 $^{\circ}$ C), which leads to various physiological disorders that negatively affect plant growth, and plants' tolerance to cold stress varies according to the variety [7,11]. El-Mahdy et al. [7] studied the responses of two banana plant cultivars (Baxi and Williams) to cold stress (5 $^{\circ}$ C for 48 h) and found that cold stress reduced the growth of the two studied cultivars. They also confirmed that Baxi banana plants have more tolerance to chilling stress than the Williams variety. Banana varieties that have the ability to secrete compatible solutes and have a strong antioxidant defense are more tolerant to cold stress [7]. Furthermore, the protection of the photosynthesis system under cold stress conditions is an important mechanism that determines the degree of success of the plant's resistance to chilling stress [7,8,13]. The protection of the fruit trees against chilling injury is crucial, and this can be done by developing efficient and practical techniques to reduce cold stress, particularly for banana plants. The findings of the current study confirm that chitosan nanoparticles (CH-NPs) are an effective tool in reducing the adverse effects of chilling in banana plants.

Nanoparticles (NPs) are an effective tool for promoting plant growth and enhancing stress tolerance [17–20,43]. NPs have properties distinct from those of bulk materials due to their small size and larger surface area which, as a result, mean that their surface reactivity is higher [17–20]. Spraying plants with nutrients, growth hormones, and important elements is an important agricultural process that stimulates plants to resist abiotic stress [13,18,24]. The application of CH-NPs enhances the growth of banana plants, as evidenced in this study by the increase in the fresh and dry weights. Once CH-NPs enter the leaf tissue, they can move and be transported to reach the centers of vital processes in the plant cell and become more effective in stimulating the plant's resistance to stress [44–47].

In the current study, we hypothesized that CH-NPs would increase the efficiency of banana plants in the face of cold stress by reducing the accumulation of reactive oxygen species and, in consequence, the degree of oxidative stress. The mechanisms behind this response seem to involve the stimulation of the activity of antioxidant enzymes and non-enzymatic antioxidants (phenolic compounds). The accumulation of osmoprotectants (soluble carbohydrates, proline, and amino acids), on the other hand, contributed to enhancing the plants' resistance against chilling stress. In addition, the CH-NPs maintained a balance between nutrients and protection of photosynthesis pigments. The findings of the present study demonstrate that nutrient content, photosynthesis pigments, and antioxidant

enzyme activities were significantly increased by the foliar application of CH-NPs; on the other hand, ROS and MDA were significantly decreased (Figure 4). The positive impacts of CH-NPs in reducing biotic and abiotic stresses have also been mentioned in other studies; e.g., Sen et al. [24], Zhang [25], and Sathiyabama and Manikandan [46].



Figure 4. Schematic diagram exploring the roles of chitosan nanoparticles (CH-NPs) in enhancing the cold stress resistance of banana plants.

The nutrients inside plants are considered one of the most important factors helping plants succeed in overcoming the harsh conditions that occur during cold stress [48,49]. The foliar application of CH-NPs enhanced the minerals status in the leaf tissue, e.g., Fe, Zn, Mn, P, Ca, N, K, and Mg. Chilling stress reduces the uptake of several nutrients, including N, P, and K [48,49]. The use of plant mineral nutrients is a potential option to achieve better crop growth and productivity and to alleviate the detrimental effects of chilling stress in a sustainable way [49,50]. Following the foliar application of CH-NPs, significant increases (p < 0.05) were recorded in most growth parameters and in the nutrient content of the banana plants exposed to cold stress (5 °C for 72 h). In particular, the increased magnesium and total nitrogen content contributed to increased chlorophyll content compared to the control [50–57].

5. Conclusions

The importance of the chilling problem is increasing with the increase in the demand for food in response to climatic changes. The current study clearly demonstrated that cold stress reduces growth, increases cell membrane injuries, and accumulates high levels of reactive oxygen species in leaf tissue. Spraying banana plants with chitosan nanoparticles (CH-NPs) at doses of 100–400 mg L⁻¹ in water reduced the adverse effects of chilling. The positive effects of CH-NPs in enhancing chilling tolerance may be due to their role in reducing oxidative stress by enhancing the activity of antioxidant enzymes and increasing the concentration of osmoprotectant substances, which are used to control cell osmosis. Reducing the negative effects of cold stress with CH-NPs may be a sustainable and economically feasible approach to achieving abiotic stress tolerance. The use of CH-NPs to reduce chilling effects provides the opportunity to develop practical solutions for banana cultivation in places where cold stress occurs, especially following the emergence of various climatic changes in many agricultural production areas around the world.

Author Contributions: Data curation, A.W., J.L., M.S.A.-H., E.F.A., J.W., Z.D., S.A.R., A.A.A.-H. and M.A.E.; Investigation, J.L., E.F.A., Z.D. and M.A.E.; Methodology, J.L., E.F.A., Z.D., S.A.R., A.A.A.-H. and M.A.E.; Resources, A.W. and Z.heli D.; Software, A.W. and E.F.A.; Writing—original draft, M.S.A.-H., E.F.A., J.W., Z.D., S.A.R. and M.A.E.; Writing—review & editing, A.W., J.L., M.S.A.-H., A.M.G., A.A.A.-H., E.F.A., J.W., Z.D. and M.A.E. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by China Agriculture Research System of MOF and MARA (CARS-31-02) Lancang-Mekong Cooperation Special Fund. The authors are thankful to Taif University Researchers Supporting Project (TURSP-2020/64) Taif University, Saudi Arabia, for the financial support and research facilities. This research was also funded by the Deanship of Scientific Research at Princess Nourah bint Abdulrahman University through the Fast-track Research Funding Program.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Robinson, J.C.; Saúco, V.G. Bananas and Plantains; CABI: Wallingford, UK, 2010; Volume 19.
- 2. Manchanda, P.; Gill, M.I.S.; Megha, S.; Gosal, S.S. Production of superelite planting material through in vitro culturing in banana. In *Biotechnologies of Crop Improvement*; Gosal, S., Wani, S., Eds.; Springer: Cham, Switzerland, 2018; Volume 1, pp. 93–129.
- 3. Pozza, L.E.; Field, D.J. The science of soil security and food security. Soil Secur. 2020, 1, 100002. [CrossRef]
- 4. Mukhopadhyay, R.; Sarkar, B.; Jat, H.S.; Sharma, P.C.; Bolan, N.S. Soil salinity under climate change: Challenges for sustainable agriculture and food security. *J. Environ. Manag.* 2021, 280, 111736. [CrossRef] [PubMed]
- Zhang, Q.; Zhang, J.Z.; Chow, W.S.; Sun, L.L.; Chen, J.W.; Chen, Y.J.; Peng, C.L. The influence of low temperature on photosynthesis and antioxidant enzymes in sensitive banana and tolerant plantain (*Musa* sp.) cultivars. *Photosynthetica* 2011, 49, 201–208. [CrossRef]
- He, W.-D.; Gao, J.; Dou, T.-X.; Shao, X.-H.; Bi, F.-C.; Sheng, O.; Deng, G.-M.; Li, C.-Y.; Hu, C.-H.; Liu, J.-H.; et al. Early cold-induced peroxidases and aquaporins are associated with high cold tolerance in Dajiao (*Musa* spp. 'Dajiao'). *Front. Plant Sci.* 2018, *9*, 282. [CrossRef] [PubMed]
- El-Mahdy, M.T.; Youssef, M.; Eissa, M.A. Impact of in vitro cold stress on two banana genotypes based on physio-biochemical Evaluation. S. Afr. J. Bot. 2018, 119, 219–225. [CrossRef]
- Rodríguez-Zapata, L.C.; Gil, F.L.E.Y.; Cruz-Martínez, S.; Talavera-May, C.R.; Contreras-Marin, F.; Fuentes, G.; Sauri-Duch, E.; Santamaría, J.M. Preharvest foliar applications of glycine-betaine protects banana fruits from chilling injury during the postharvest stage. *Chem. Biol. Technol. Agric.* 2015, 2, 43. [CrossRef]
- 9. Chen, L.-L.; Shan, W.; Cai, D.-L.; Chen, J.-Y.; Lu, W.-J.; Su, X.-G.; Kuang, J.-F. Postharvest application of glycine betaine ameliorates chilling injury in cold-stored banana fruit by enhancing antioxidant system. *Sci. Hortic.* **2021**, *287*, 110264. [CrossRef]
- Lima, J.D.; Rozane, D.E.; Gomes, E.N.; Da Silva, S.H.M.G.; Moraes, W.D.S.; Kluge, R.A. Chilling Prevention on Banana 'Nanica' in the Field with Bagging. J. Agric. Sci. 2018, 10, 122. [CrossRef]
- 11. Jones, D.R. (Ed.) Handbook of Diseases of Banana, Abacáand Enset; CAB International: Wallingford, UK, 2019.
- 12. Levitt, J. Responses of plants to environmental stresses. In *Chilling, Partly and High Temperature Stresses*, 2nd ed.; Academic: New York, NY, USA, 1980; Volume 1, p. 49.
- 13. Li, Y.; Zhang, Q.; Ou, L.; Ji, D.; Liu, T.; Lan, R.; Li, X.; Jin, L. Response to the cold stress signaling of the tea plant (*Camellia sinensis*) Elicited by Chitosan Oligosaccharide. *Agronomy* **2020**, *10*, 915. [CrossRef]
- 14. Parisi, C.; Vigani, M.; Rodríguez-Cerezo, E. Agricultural nanotechnologies: What are the current possibilities? *Nano Today* 2015, 10, 124–127. [CrossRef]
- 15. Gogos, A.; Knauer, K.; Bucheli, T.D. Nanomaterials in Plant protection and fertilization: Current state, foreseen applications, and research priorities. *J. Agric. Food Chem.* **2012**, *60*, 9781–9792. [CrossRef]
- 16. Fraceto, L.F.; Grillo, R.; de Medeiros, G.A.; Scognamiglio, V.; Rea, G.; Ebartolucci, C. Nanotechnology in agriculture: Which innovation potential does it have? *Front. Environ. Sci.* **2016**, *4*, 20. [CrossRef]
- Cheng, G.; Chang, T.-H.; Qin, Q.; Huang, H.; Zhu, Y. Mechanical properties of silicon carbide nanowires: Effect of size-dependent defect density. *Nano Lett.* 2014, 14, 754–758. [CrossRef]

- Zahedi, S.M.; Moharrami, F.; Sarikhani, S.; Padervand, M. Selenium and silica nanostructure-based recovery of strawberry plants subjected to drought stress. *Sci. Rep.* 2020, 10, 1–18. [CrossRef] [PubMed]
- 19. Alkharabsheh, H.; Seleiman, M.; Battaglia, M.; Shami, A.; Jalal, R.; Alhammad, B.; Almutairi, K.; Al-Saif, A. Biochar and its broad impacts in soil quality and fertility, nutrient leaching and crop productivity: A review. *Agronomy* **2021**, *11*, 993. [CrossRef]
- 20. Seleiman, M.F.; Almutairi, K.F.; Alotaibi, M.; Shami, A.; Alhammad, B.A.; Battaglia, M.L. Nano-fertilization as an emerging fertilization technique: Why can modern agriculture benefit from its use? *Plants* **2020**, *10*, 2. [CrossRef] [PubMed]
- 21. Rinaudo, M. Chitin and chitosan: Properties and applications. Prog. Polym. Sci. 2006, 31, 603–632. [CrossRef]
- 22. Al-Dhabaan, F.A.; Mostafa, M.; Almoammar, H.; Abd-Elsalam, K.A. Chitosan-based nanostructures in plant protection applications. In *Nanotechnology Applications in Health and Environmental Sciences*; Gabler: Wiesbaden, Germany, 2018; pp. 351–384.
- 23. Shukla, S.K.; Mishra, A.K.; Arotiba, O.; Mamba, B. Chitosan-based nanomaterials: A state-of-the-art review. *Int. J. Biol. Macromol.* **2013**, *59*, 46–58. [CrossRef]
- 24. Sen, S.K.; Chouhan, D.; Das, D.; Ghosh, R.; Mandal, P. Improvisation of salinity stress response in mung bean through solid matrix priming with normal and nano-sized chitosan. *Int. J. Biol. Macromol.* **2020**, *145*, 108–123. [CrossRef]
- Zhang, X.; Hu, C.; Sun, X.; Zang, X.; Zhang, X.; Fang, T.; Xu, N. Comparative transcriptome analysis reveals chitooligosaccharidesinduced stress tolerance of Gracilariopsis lemaneiformis under high temperature stress. *Aquaculture* 2020, 519, 734876. [CrossRef]
- Farghly, K.A.; Gomah, H.H.; Ahmed, M.M.; Abdalla, R.M.; Eissa, M.A. Corn wastes and peanut shell as growing media for production of red radish plants in soilless system. *Commun. Soil Sci. Plant Anal.* 2020, 51, 1799–1810. [CrossRef]
- 27. Lichtenthaler, H.K.; Buschmann, C. Chlorophylls and carotenoids: Measurement and characterization by UV-VIS spectroscopy. *Curr. Protoc. Food Anal. Chem.* **2001**, *1*, F4. 3.1–F4. 3.8. [CrossRef]
- Bates, L.S.; Waldren, R.P.; Teare, I.D. Rapid determination of free proline for water-stress studies. *Plant Soil* 1973, 39, 205–207. [CrossRef]
- 29. Ainsworth, E.A.; Gillespie, K.M. Estimation of total phenolic content and other oxidation substrates in plant tissues using Folin–Ciocalteu reagent. *Nat. Protoc.* 2007, *2*, 875–877. [CrossRef] [PubMed]
- 30. Yemm, E.W.; Willis, A.J. The estimation of carbohydrates in plant extracts by anthrone. Biochem. J. 1954, 57, 508–514. [CrossRef]
- 31. Parkinson, J.A.; Allen, S.E. A wet oxidation procedure suitable for the determination of nitrogen and mineral nutrients in biological material. *Commun. Soil Sci. Plant Anal.* **1975**, *6*, 1–11. [CrossRef]
- 32. Burt, R. Soil Survey Laboratory Methods Manual; Soil Survey Investigations Report No. 42; NRCS: Washington, DC, USA, 2004.
- 33. Velikova, V.; Yordanov, I.; Edreva, A. Oxidative stress and some antioxidant systems in acid rain-treated bean plants: Protective role of exogenous polyamines. *Plant Sci.* **2000**, *151*, 59–66. [CrossRef]
- 34. Kubiś, J. Exogenous spermidine differentially alters activities of some scavenging system enzymes, H2O2 and superoxide radical levels in water-stressed cucumber leaves. *J. Plant Physiol.* **2008**, *165*, 397–406. [CrossRef]
- 35. Rao, K.M.; Sresty, T. Antioxidative parameters in the seedlings of pigeonpea (*Cajanus cajan* (L.) Millspaugh) in response to Zn and Ni stresses. *Plant Sci.* 2000, 157, 113–128. [CrossRef]
- 36. Halliwell, B.; Gutteridge, J.M.C.; Aruoma, O.I. Te deoxyribose method: A simple "test-tube" assay for determination of rate constants for reactions of hydroxyl radicals. *Anal. Biochem.* **1987**, *165*, 215–219. [CrossRef]
- 37. Abeed, A.H.; Eissa, M.A.; Abdel-Wahab, D.A. Effect of Exogenously applied jasmonic acid and kinetin on drought tolerance of wheat cultivars based on morpho-physiological evaluation. *J. Soil Sci. Plant Nutr.* **2021**, 21, 131–144. [CrossRef]
- Eissa, M.A.; Nafady, M.; Ragheb, H.; Attia, K. Effect of soil moisture and forms of phosphorus fertilizers on corn production under sandy calcareous soil. World Appl. Sci. J. 2013, 26, 540–547.
- Hasanuzzaman, M.; Bhuyan, M.; Zulfiqar, F.; Raza, A.; Mohsin, S.; Mahmud, J.; Fujita, M.; Fotopoulos, V. Reactive oxygen species and antioxidant defense in plants under abiotic stress: Revisiting the crucial role of a universal defense regulator. *Antioxidants* 2020, 9, 681. [CrossRef]
- 40. Moreano, J.P.S.; Xu, X.; Criollo, C.B.A.; Chen, X.; Lin, Y.; Munir, N.; Lai, Z. Genome-wide identification and comprehensive analyses of TCP gene family in banana (*Musa* L.). *Trop. Plant Biol.* **2021**, *14*, 180–202. [CrossRef]
- 41. Augspurger, C.K. Reconstructing patterns of temperature, phenology, and frost damage over 124 years: Spring damage risk is increasing. *Ecology* **2013**, *94*, 41–50. [CrossRef]
- 42. Ruelland, E.; Zachowski, A. How plants sense temperature. Environ. Exp. Bot. 2010, 69, 225–232. [CrossRef]
- 43. Mishra, S.K.; Khan, M.H.; Misra, S.; Dixit, V.K.; Khare, P.; Srivastava, S.; Chauhan, P.S. Characterisation of Pseudomonas spp. and *Ochrobactrum* sp. isolated from volcanic soil. *Antonie Van Leeuwenhoek* **2017**, *110*, 253–270. [CrossRef] [PubMed]
- 44. Lv, J.; Christie, P.; Zhang, S. Uptake, translocation, and transformation of metal-based nanoparticles in plants: Recent advances and methodological challenges. *Environ. Sci. Nano* **2019**, *6*, 41–59. [CrossRef]
- 45. Saharan, V.; Pal, A. Current and Future Prospects of Chitosan-Based Nanomaterials in Plant Protection and Growth; Springer: Berlin/Heidelberg, Germany, 2016; pp. 43–48.
- 46. Sathiyabama, M.; Manikandan, A. Foliar application of chitosan nanoparticle improves yield, mineral content and boost innate immunity in finger millet plants. *Carbohydr. Polym.* **2021**, *258*, 117691. [CrossRef] [PubMed]
- 47. Faizan, M.; Rajput, V.; Al-Khuraif, A.; Arshad, M.; Minkina, T.; Sushkova, S.; Yu, F. Effect of foliar fertigation of chitosan nanoparticles on cadmium accumulation and toxicity in solanum lycopersicum. *Biology* **2021**, *10*, 666. [CrossRef] [PubMed]

- 48. Tommasini, L.; Svensson, J.T.; Rodriguez, E.M.; Wahid, A.; Malatrasi, M.; Kato, K.; Wanamaker, S.; Resnik, J.; Close, T.J. Dehydrin gene expression provides an indicator of low temperature and drought stress: Transcriptome-based analysis of Barley (*Hordeum vulgare* L.). *Funct. Integr. Genom.* **2008**, *8*, 387–405. [CrossRef] [PubMed]
- 49. Yan, Q.; Duan, Z.; Mao, J.; Li, X.; Dong, F. Effects of root-zone temperature and N, P, and K supplies on nutrient uptake of cucumber (*Cucumis sativus* L.) seedlings in hydroponics. *Soil Sci. Plant Nutr.* **2012**, *58*, 707–717. [CrossRef]
- 50. Farooq, M.; Aziz, T.; Wahid, A.; Lee, D.-J.; Siddique, K. Chilling tolerance in maize: Agronomic and physiological approaches. *Crop. Pasture Sci.* **2009**, *60*, 501–516. [CrossRef]
- 51. Dzung, N.A.; Khanh, V.T.P.; Dzung, T.T. Research on impact of chitosan oligomers on biophysical characteristics, growth, development and drought resistance of coffee. *Carbohydr. Polym.* **2011**, *84*, 751–755. [CrossRef]
- 52. Al-Huqail, A.A. Changes in antioxidant status, water relations and physiological indices of maize seedlings under drought stress conditions. *J. Biol. Sci.* 2019, *19*, 331–338. [CrossRef]
- 53. Eissa, M.A.; Abeed, A.H. Growth and biochemical changes in quail bush (*Atriplex lentiformis* (Torr.) S.Wats) under Cd stress. *Environ. Sci. Pollut. Res.* **2019**, *26*, 628–635. [CrossRef]
- 54. Abou-Zaid, E.A.; Eissa, M.A. Thompson seedless grapevines growth and quality as affected by glutamic acid, vitamin b, and algae. *J. Soil Sci. Plant Nutr.* **2019**, *19*, 725–733. [CrossRef]
- 55. Ali, A.M.; Awad, M.Y.; Hegab, S.A.; Gawad, A.M.A.E.; Eissa, M.A. Effect of potassium solubilizing bacteria (Bacillus cereus) on growth and yield of potato. *J. Plant Nutr.* **2021**, *44*, 411–420. [CrossRef]
- 56. Al-Sayed, H.; Hegab, S.A.; Youssef, M.; Khalafalla, M.; Almaroai, Y.A.; Ding, Z.; Eissa, M.A. Evaluation of quality and growth of roselle (*Hibiscus sabdariffa* L.) as affected by bio-fertilizers. *J. Plant Nutr.* **2021**, *43*, 1025–1035. [CrossRef]
- 57. Ding, Z.; Zhou, Z.; Lin, X.; Zhao, F.; Wang, B.; Lin, F.; Ge, Y.; Eissa, M.A. Biochar impacts on NH3-volatilization kinetics and growth of sweet basil (Ocimum basilicum L.) under saline conditions. *Ind. Crop. Prod.* **2020**, *157*, 11290. [CrossRef]