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Anti-PD-1 checkpoint blockade monotherapy in the orthotopic GL261 glioma model: the devil is in the detail

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Abstract

The GL261 cell line, syngeneic on the C57BL/6 background, has, since its establishment half a century ago in 1970, become the most commonly used immunocompetent murine model of glioblastoma. As immunotherapy has entered the mainstream of clinical discourse in the past decade, this model has proved its worth as a formidable opponent against various immunotherapeutic combinations. Although advances in surgical, radiological, and chemotherapeutic interventions have extended mean glioblastoma patient survival by several months, 5-year survival postdiagnosis remains below 5%. Immunotherapeutic interventions, such as the ones explored in the murine GL261 model, may prove beneficial for patients with glioblastoma. However, even common immunotherapeutic interventions in the GL261 model still have unclear efficacy, with wildly discrepant conclusions being made in the literature regarding this topic. Here, we focus on anti-PD-1 checkpoint blockade monotherapy as an example of this pattern. We contend that a fine-grained analysis of how biological variables (age, sex, tumor location, etc.) predict treatment responsiveness in this preclinical model will better enable researchers to identify glioblastoma patients most likely to benefit from checkpoint blockade immunotherapy moving forward.

Key Points

- Anti-PD-1 monotherapy has equivocal efficacy in the GL261 model.
- Known but under-referenced factors impact murine survival in the GL261 model.

Glioblastoma is a devastating malignancy with a median survival of 12–18 months postdiagnosis.^{1–3} Even this brief window of survival is hard-won, requiring a full standard of care upon diagnosis, this includes maximum safe surgical resection, radiotherapy, and chemotherapy by temozolomide.⁴ While this combination of treatments (the "Stupp Protocol") is a significant improvement over previous expectations for glioblastoma patients,^{5,6} the prognosis remains bleak and there exists a clear need for improved therapeutic options.

Increasing numbers of immunotherapy trials are entering the clinic for various cancers,⁷ including multiple different immunotherapy regimens attempted for cancers of the CNS.^{8,9} Immunotherapy is a promising avenue for the treatment of brain tumors, as immune cells can cross into the brain and occupy tumors therein¹⁰ whereas many conventional treatment strategies are confounded by the protective blood–brain barrier.¹¹ The development of these immunotherapies for GBM has benefited enormously from the abundance of immunocompetent murine models of glioblastoma.¹² Among the most commonly used of these is the GL261 model¹³ that shares a number of significant parallels with human glioblastoma.^{14–21} The extensively characterized^{12,13,22} GL261 cell line has been

© The Author(s) 2021. Published by Oxford University Press, the Society for Neuro-Oncology and the European Association of Neuro-Oncology. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (http://creativecommons.org/ licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com repeatedly used in murine survival studies for various immunotherapeutic interventions, but the efficacy of many of these therapies remains controversial. The present work will focus on a single example of this, unpacking the literature to determine why anti-PD-1 checkpoint blockade immunotherapy (CBI) monotherapy, a highly published, flagship immunotherapy agent, has unclear efficacy in the GL261 model in which it has been so repeatedly tested.

We contend that disparate outcomes in publications treating the GL261 model glioma with anti-PD-1 monotherapy are driven by the broad spread of experimental parameters between studies. Furthermore, we will argue that the influence of many of these factors is already known but underappreciated. The systematic study of how preclinical biological variables influence the survival of these animals is a potentially untapped resource that could impact the field's ability to better predict patient outcomes in future immunotherapy trials. Minimally, these variables ought to be duly considered during the experimental design process and, where necessary, controlled for. In summary, this review will lay forth and evaluate the evidence supporting each of a number of experimental parameters as potential drivers of the discord in reported survival times of GL261-bearing mice treated with anti-PD-1 monotherapy.

Rationale

The necessity of this report can be appreciated in comparing these 2 sentences from research publications in 2019:

PD-1 antibody therapy in GL261 resulted in almost half of the animals with long-term survival, which is consistent with previously published findings.—Jahan et al.²³

GL261 tumors are intrinsically unresponsive to anti-PD-1 therapy & All untreated mice succumbed to their disease prior to Day 21, and no animals survived beyond Day 24 in the groups receiving [anti-PD-1 therapy] indicating no significant survival benefit was conferred.—Kim et al.²⁴

Obviously, these absolutist statements on anti-PD-1 antibody's therapeutic efficacy cannot both be true. Some nuance must exist that would permit such discrepant outcomes in the same model system. The jarring dissonance in the literature regarding anti-PD-1 therapy in the GL261 model, which stretches far beyond the 2 publications quoted above, has been previously noted.25 In that review, the differences in outcome were largely attributed to varying frequency and dose of anti-PD-1 administration.²⁵ We view this attribution as insufficiently broad, given the wealth of published knowledge implicating many other experimental variables as well. We aim to expand this conversation and show that anti-PD-1 CBI monotherapy is not simply effective or simply ineffective against the GL261 glioma model but that its efficacy is influenced by myriad factors, far beyond just dosing, which need to be both appropriately accounted for and clearly communicated with the research community. With a solidified understanding of why study outcomes are so disparate, the abundance of preclinical data in the GL261 model can help guide the way to more nuanced personalization of immunotherapies for patients.

Approach

We have observed a wide range of published survival times for anti-PD-1-treated GL261-bearing mice and wanted to obtain an unbiased sample of such studies to examine more deeply. At the time of writing, 29 publications were retrieved by the PubMed "All Fields" search term "(PD-1) AND (GL261)." One of these was excluded for being a Review. Our focus is survival outcomes of anti-PD-1 CBI monotherapy for C57BL/6 mice bearing orthotopic GL261 gliomas, so studies that lacked such trial arms (eg, those only using combination therapies²⁶ or not conducting GL261 survival studies²⁷) were also excluded. The remaining 16 research papers are explored in detail below^{23,24,28–41} (Table 1). This search engine-based sample of the literature does not include all studies that explore outcomes of GL261-bearing mice treated with anti-PD-1 monotherapy⁴² but, rather, provides an approachable number of studies in order to showcase common experimental methods in the field, preserve brevity, and prevent the incorporation of author bias. This scope has prevented us from recognizing the work of many of our talented colleagues and we apologize to those whose relevant studies were unable to include. The variables selected for Table 1 are the ones that have been linked, in a direct or indirect fashion, to the survival of GL261-bearing mice treated with anti-PD-1 monotherapy and each variable will be discussed in greater detail below. In the interest of full transparency, the nature of the control arms of each study has also been included in Table 1. While the sham exposure of the control arm cannot change the absolute survival time or percentage of the anti-PD-1-treated experimental arm, different control set-ups could potentially impact relative statements of treatment efficacy. It is worth noting, however, that variations in the survival of control-treated animals between studies are more likely reflective of the differences in other biological variables like age or tumor inoculum than differences between the types of control interventions. Two of the most common control interventions among our studies (phosphate buffered saline [PBS] and isotype IgG) have been shown to have an indistinguishable effect on murine survival.³³ The strength of evidence linking each variable to the modulation of survival outcomes in this context is color-coded in Figure 1 for easy reference. We hope that assessing the contribution of each variable in a systematic way demystifies some of the enormous spread of murine survival appreciable in Table 1 and in the literature more broadly.

Variables Relating to the Anti-PD-1 Antibody

The dosing strategy of anti-PD-1 antibody has been suggested as a pivotal determinant in the variations in survival

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G4; PIH $6000 GL261; 5 \mu L; 15 mm$ $F, 6-8; Harlan0\%(M; 2.2.5)RMP1-14; BioXcell5000 GL261; 5 \mu L; 15 mmM; 6-8;0\%MaJ43: BioXcell20000 GL261; 2 \mu L; NAM; 6-8;0\%NaJ43: BioXcell20000 GL261; 3 \mu L; 35 mmF; 8; CRL0\%NaRMP1-14; BioLegend30000 GL261; 3 \mu L; 35 mmF; 8; CRL0\%NaRMP1-14; BioLegend30000 GL261; 2 \mu L; 3 mmNa; Na; CRL0\%NaRMP1-14; BioLegend30000 GL261; 2 \mu L; 3 mmNa; T-8; JL0\%NaRMP1-14; BioLegend10000 GL261; 2 \mu L; 3 mmNa; T-8; JL0\%NaRMP1-14; FIH50000 GL261; 2 \mu L; 3 mmNa; P-0; Envigo0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH50000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH10000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%NaRMP1-14; FIH10000 GL261; 10 \mu L; 3 mmF; 6-8; JL0\%$		Mice (sex; weeks old; vendor)		dian Survival am; anti-PD1) ıys)	Ctrl.
RMP1-14: BioXCellB000 GL261; 5 µL; 1.5 mm $W; 6-8;$ moxyu 0% NaJ43: BioXCell20000 GL261; 3 µL; 3.5 mm $; 8; JL$ 0% NaHMP1-14: BioXCell20000 GL261; 3 µL; 3.5 mm $; 6; CRL$ 0% NaRMP1-14: BioXcell20000 GL261; 3 µL; 3.5 mm $Na; Na; CRL$ 0% NaRMP1-14: BioXcell30000 GL261; 3 µL; 3.5 mm $Na; Na; CRL$ 0% Na RMP1-14: BioXcell30000 GL261; 2 µL; 3 mm $Na; Na; CRL$ 0% Na RMP1-30; Bioscience10000 GL261; 2 µL; 3 mm $Na; P-10;$ 0% Na RMP1-30; Bioscience10000 GL261; 2 µL; 3 mm $Na; P-10;$ 0% Na RMP1-30; Bioscience10000 GL261; 1 µL; 3 mm $Na; P-10;$ 0% Na G4; PH130000 GL261; 10 µL; 3 mm $F, e-3; JL$ 0% Na HZ: Bristol-Myers130000 GL261; 10 µL; 3 mm $F, e-3; JL$ 0% Na HZ: Bristol-Myers130000 GL261; 10 µL; 3 mm $Na; N-1L$ 0% Na HZ: Bristol-Myers130000 GL261; 10 µL; 3 mm $F, e-3; JL$ 0% Na HZ: PH130000 GL261; 10 µL; 3 mm $Na; N-1L$ 0% Na G4; PH130000 GL261; 1µL; 3 mm $F, e-3; JL$ 0% Na G4; PH130000 GL261; 1µL; 3 mm $F, e-3; JL$ 0% Na G4; PH130000 GL261; 1µL; 3 mm $F, e-8; JL$ 0% Na MP1-14; BioXell130000 GL261; 1µL; 3 mm $F, e-8; JL$ 0% Na G4; PH130000 G	50 000 GL261; NA; 3 mm	F; 6–8; Harlan	-	A; 22.5)	IgG
J43: BioXCell $20 000 Gl261; 2 \mu; NA$ F; 8; JL0%NARMP1-14; BioXcell $200 000 Gl261; 3 \mu; 3.5 mm$ F; 6; CRL0%(19; 21)RMP1-14; BioLegend $300 000 Gl261; 3 \mu; 3.5 mm$ NA; NA; CRL0%(19; 21)RMP1-14; BioXcell $300 000 Gl261; 2 \mu; 3 mm$ NA; 7-8; JL0%(19; 30.5)RMP1-30; EBioscience $300 000 Gl261; 2 \mu; 3 mm$ NA; 7-8; JL0%(19; 30.5)RMP1-30; EBioscience $300 000 Gl261; 2 \mu; 3 mm$ NA; 7-8; JL0%(19; 30.5)RMP1-30; EBioscience $100 000 Gl261; 2 \mu; 3 mm$ NA; 7-8; JL0%(19; 30.5)RMP1-14; PIH $300 00 Gl261; 1 \mu; 3 mm$ F; 6-8; JL0%(26; 30)RMP1-14; PIH $500 000 Gl261; 1 \mu; 3 mm$ F; 6-8; JL0%(26; 30)RMP1-14; BioXcell $130 000 Gl261; 1 \mu; 3 mm$ F; 6-8; JL20%NARMP1-14; BioXcellNA; NA; HNA; NA; JL50%NARMP1-14; BioXcellNA; NA; JL50%NARMP1-14; BioXcellNA; NA; JL50%NARMP1-14; BioXcellNA; NA; JL50%NAG4; PIH130 000 GL261-Luc; 1 \mu; 3 mmF; 6-8; JL20%NAG4; PIH130 000 GL261-Luc; 1 \mu; 3 mmF; 6-8; JL20%NASFIAT2; PIH100 000 GL261-Luc; 1 \mu; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 1 \mu; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 1 \mu; 3		M; 6–8; CMCYU			Saline
RMP1-14; BioXcell200 000 GL261; $3 \mu l$; $35 \mu m$ F, 6; CRL0%(19; 21)RMP1-14; BioLegend300 000 GL261; $2 \mu l$; $3 m$ Na; Na; CRL0%NaRMP1-14; BioXcell300 000 GL261; $2 \mu l$; $3 m$ Na; Na; CRL0%(19; 30.5)RMP1-30; Blioscience300 000 GL261; $2 \mu l$; $3 m$ Na; $3 - 10$ 0%(19; 30.5)RMP1-30; Blioscience100 000 GL261; $2 \mu l$; $3 m$ Na; $8 - 10$;0%(19; 30.5)G4; PIH130 000 GL261; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 0%(26; 30)RMP1-14; PIH500 00 GL261; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 0%(26; 30)HZ; Bristol-Myers130 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 14.30RMP1-14; BioXCell75 000 GL261; $3 \mu l$; $2.5 m$ Na; Na; $J L$ 0%NaRMP1-14; BioXCell75 000 GL261; $1 \mu l$; $3 m$ Na; Na; $J L$ 20%NaRMP1-14; BioXCell130 000 GL261; $1 \mu l$; $3 m$ Na; Na; $J L$ 20%NaG4; PIH100 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 20%NaG4; PIH100 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 20%NaNA; Bristol-Myers130 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 20%NaNA; Bristol-Myers130 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 20%NaNA; Bristol-Myers130 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 20%NaNA; Bristol-Myers130 000 GL261-Luc; $1 \mu l$; $3 m$ F, 6 - 8; $J L$ 20% <t< td=""><td></td><td>F; 8; JL</td><td></td><td></td><td>PBS</td></t<>		F; 8; JL			PBS
RMP1-14; BioLegend 300 000 GL261; 2 µL; 3 mm N4; N4; CRL 0% N4 RMP1-14; BioXCell 300 000 GL261; 2 µL; 3 mm N4; 7-8; JL 0% (19; 30.5) RMP1-30; Blioscience 100 000 GL261; 2 µL; 3 mm N4; 8-10; 0% (19; 30.5) G4; PIH 130 000 GL261; 1 µL; 3 mm F; 8-8; JL 0% (19; 30.5) RMP1-14; PIH 500 00 GL261; 10 µL; 3 mm F; 8-8; JL 0% (26; 30) RMP1-14; PIH 500 00 GL261; 10 µL; 3 mm F; 8-8; JL 0% (25; 34) 4H2; Bristol-Myers 130 000 GL261; 10 µL; 3 mm F; 6-8; JL 0% NA 4H2; Bristol-Myers 130 000 GL261; 10 µL; 3 mm F; 6-8; JL 0% NA 64; PIH 130 000 GL261; Luc; 1 µL; 3 mm F; 6-8; JL 20% NA 64; PIH 130 000 GL261; Luc; 1 µL; 3 mm F; 6-8; JL 20% NA 64; PIH 130 000 GL261; Luc; 1 µL; 3 mm F; 6-8; JL 20% NA 64; PIH 130 000 GL261; Luc; 1 µL; 3 mm F; 6-8; JL 20% NA 64; PIH 130 000 G		F; 6; CRL		; 21)	Untreated
RMP1-14; BioXCell $30000 G[261; 2 \ \muL; 3 \ mm)Na; 7-8; JL0%(19; 30.6)RMP1-30; eBioscience10000 G[261; 2 \ \muL; 3 \ mm)K; 8-10;0\%0\%0\%G4; PIH130000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL0\%26; 30)RMP1-14; FIH130000 G[261; 10 \ \muL; 3 \ mm)F; 8-8; JL0\%26; 30)G4; PIH130000 G[261; 10 \ \muL; 3 \ mm)F; 8-8; JL0\%26; 30)RMP1-14; FIO-Myers130000 G[261; 11, 13 \ mm)F; 6-8; JL14.3\%25; 34)RMP1-14; BioXCell76000 G[261; 3 \ \muL; 3 \ mm)F; 6-8; JL20\%NaRMP1-14; BioXCellN; NA; Hm)NA; NA; JL20\%NAG4; PIH130000 G[261; 11, 1; 3 \ mm)F; 6-8; JL20\%NAG4; PIH130000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 3)D4; PIH130000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 5)D6; PIH10000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 5)D6; PIH10000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 5)D6; PIH10000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 5)D6; PIH10000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 3)D6; PIH10000 G[261-Luc; 1 \ \muL; 3 \ mm)F; 6-8; JL20\%(16; 25; 3)NA; Bristol-Myers130000 G[261-Luc; 1 \ \muL; 30]F; 6-8; JL$		NA; NA; CRL			Untreated
15 RMP1-30; eBioscience 100 000 CL261; 2 μL; 3 mm W; 8–10; mAS 0% NA C4; PIH 130 000 Cl261-Luc; 1 μL; 3 mm F; 6–8; JL 0% 26; 30) RMP1-14*; PIH 500 000 CL261; 10 μL; 3 mm F; 8–8; JL 0% 26; 30) RMP1-14*; PIH 500 000 CL261; 1 μL; 3 mm F; 8–10; Envigo 55% 24; S; NA) RMP1-14*; BioXCell 75 000 CL261; 3 μL; 2.5 mm F; 6–8; JL 14.2; % 25; 34) L2b RMP1-14; BioXCell No. NA; JL 76, NCl 20% NA L2b RMP1-14; BioXCell NA; NA; JL NA; NL 56-8; JL 70% 70% L2b RMP1-14; BioXCell NA; NA; JL 70% 70% 70% L2b RMP1-14; BioXCell NA; NA; JL 56-8; JL 70% 70% L2b RMP1-14; BioXCell NA; NA; JL 70% 70% 70% 70% L2b G4; PH 130 000 CL261-Luc; 1 μL; 3 mm F; 6–8; JL 20% 70% 70% 70% 70% 70% 70%		NA; 7–8; JL		; 30.5)	IgG
G4; PIHT30 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL0%(26; 30)RMP1-14; PIH600 000 G[261; 10 µL; 3 mmF; 8-10; Envigo55%(245; NA)BMP1-14; PIH500 000 G[261; 10 µL; 3 mmF; 8-10; Envigo55%(25; 34)H2; Britol-Myers130 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL14.3%(25; 34)RMP1-14; BioXCell75 000 G[261; 3 µL; 2.5 mmF; 6-8; JL50%NA22bRMP1-14; BioXCellNA; NA; H mmNA; NA; JL50%NA22bG4; PIH130 000 G[261-Luc; 1 µL; 3 mmNA; NA; JL50%NA1929F1A12; PIH100 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL20%(16; 25.5)1929F1A12; PIH100 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 G[261-Luc; 1 µL; 3 mmF; 6-8; JL30%24; 30)		M; 8–10; RAMS			PBS/IgG
RMP1-14; PIH500 000 GL261; 10 µL; 3m mF; 8-10; Envigo55%(245; Nd)4 H2; Bristol-Myers130 000 GL261-Luc; 1 µL; 3 m mF; 6-8; JL14.3%(25; 34)8quibb75000 GL261; 3 µL; 2.5 m mN3; NA; JL00000NA22bRMP1-14; BioXCell75 000 GL261; 3 µL; 2.5 m mNA; NA; JL60%NA22bRMP1-14; BioXCellNA; NA; JLNA; NA; JL60%NA21bG4; PIH130 000 GL261-Luc; 1 µL; 3 m mF; 6-8; JL20%(16; 25.6)1929F1412; PIH100 000 GL261-Luc; 5 µL; 3 m mF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 5 µL; 3 m mF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 5 µL; 3 m mF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 1 µL; 3 m mF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 1 µL; 3 m mF; 6-8; JL30%23; 33)	130 000 GI261-Luc; 1 µL; 3 mm			; 30)	IgG
4 H2: Bristol-Myers130 000 GL261-Luc; 1 μ L; 3 mmF; 6-8; JL14.3%(25; 34)8 quibbRMP1-14; BioXCell75 000 GL261; 3 μ L; 2.5 mmF; 6-7; NCl20%NA22 bRMP1-14; BioXCellNA; NA; 4 mmNA; NA; JL50%NA22 bRMP1-14; BioXCellNA; NA; 4 mmNA; NA; JL50%NA24 PH130 000 GL261-Luc; 1 μ L; 3 mmF; 6-8; JL278%(22; 33)7929F1A12; PIH130 000 GL261-Luc; 1 μ L; 3 mmF; 6-8; JL20%(16; 25.5)7829F1A12; PIH100 000 GL261-Luc; 5 μ L; 3 mmF; 6-8; JL20%NANA; Bristol-Myers130 000 GL261-Luc; 1 μ L; 3 mmF; 6-8; JL20%NANa; Bristol-Myers130 000 GL261-Luc; 1 μ L; 3 mmF; 6-8; JL20%NA		F; 8–10; Envigo		.5; NA)	lgG
RMP1-14; BioXCell 75 000 GL261; $3 \mu L; 2.5 mm$ F; $6-7; NCl$ 20% NA 22 ^b RMP1-14; BioXCell NA; 4 mm NA; NA; JL 50% NA 22 ^b RMP1-14; BioXCell NA; 4 mm NA; NA; JL 50% NA 22 ^b G4; PIH 130 000 GL261-Luc; 1 $\mu L; 3 mm$ F; 6-8; JLUAF 278% (22; 33) 719 29F1A12; PIH 100 000 GL261-Luc; 5 $\mu L; 3 mm$ F; 6-8; JL 20% NA N3. Bristol-Myers 130 000 GL261-Luc; 1 $\mu L; 3 mm$ F; 6-8; JL 20% NA NA: Bristol-Myers 130 000 GL261-Luc; 1 $\mu L; 3 mm$ F; 6-8; JL 20% NA NA: Bristol-Myers 130 000 GL261-Luc; 1 $\mu L; 3 mm$ F; 6-8; JL 30% (24; 30)				; 34)	Untreated
22 ^b RMP1-14; BioXCell NA; HM NA; HM 50% NA G4; PIH 130 000 GL261-Luc; 1 μL; 3 mM F; 6-8; JHUAF 278% (22; 33) G4; PIH 130 000 GL261-Luc; 1 μL; 3 mM F; 6-8; JL 20% (16; 25.6) (19) 29F1A12; PIH 100 000 GL261-Luc; 5 μL; 3 mM F; 6-8; JL 20% (16; 25.6) NA; Bristol-Myers 130 000 GL261-Luc; 1 μL; 3 mM F; 6-8; Envigo 50% NA NA; Bristol-Myers 130 000 GL261-Luc; 1 μL; 3 mM F; 6-8; Lnvigo 50% NA		F; 6–7; NCI			IgG
G4; PIH 130 000 GL261-Luc; 1 μL; 3 mm F; 6-8; JHUAF 27.8% (22; 33) G4; PIH 130 000 GL261-Luc; 1 μL; 3 mm F; 6-8; JL 20% (16; 25.5) A19 29F1A12; PIH 100 000 GL261-Luc; 5 μL; 3 mm F; 6-8; Hvigo 50% NA NA; Bristol-Myers 130 000 GL261-Luc; 1 μL; 3 mm F; 6-8; JL 30% (24; 30) Squibb Na; Bristol-Myers 130 000 GL261-Luc; 1 μL; 3 mm F; 6-8; JL 30% (24; 30)		NA; NA; JL			Untreated
G4; PIH 130 000 GL261-Luc; 1 μL; 3 mm F; 6–8; JL 20% (16; 25.6) 19 29F1A12; PIH 100 000 GL261-Luc; 5 μL; 3 mm F; 6–8; Envigo 50% NA NA; Bristol-Myers 130 000 GL261-Luc; 1 μL; 3 mm F; 6–8; JL 30% (24; 30) Squibb Squibb 29% U 29% U 24% U 20% 24% U	130 000 GL261-Luc; 1 µL; 3 mm			; 33)	Unspec.
29F1A12; PIH 100 000 GL261-Luc; 5 µL; 3 mm F; 6-8; Envigo 50% NA NA; Bristol-Myers 130 000 GL261-Luc; 1 µL; 3 mm F; 6-8; JL 30% (24; 30) Squibb Squibb 24; 30 24; 30 24; 30 24; 30	130 000 GL261-Luc; 1 µL; 3 mm			; 25.5)	PBS
NA; Bristol-Myers 130 000 GL261-Luc; 1 μL; 3 mm F; 6–8; JL					Unspec.
				; 30)	Untreated

these scaled doses have been standardized assuming a weight of 20 g, which would not be unusual for a young female C57BL/6 mouse. ^bDemarcates a dosing timecourse where significant ambiguity existed in the original manuscript and we were forced to draw an inference regarding the methods. ^cThis manuscript listed the clone used as "RPMI-14," but, as no corroborating evidence as to the existence of this clone could be found, we have made the assumption that the authors intended to write "RMP1-14" which is a commonly used clone of anti-PD-1 antibody.

outcomes observed in the GL261 model²⁵ and so we chose to begin our investigations there (Table 1). More specifically, that review stated based on examining 2 manuscripts^{34,43} that "In the preclinical GL261 model, the success of anti-PD-1 monotherapy is dependent on antibody dosage levels."²⁵WithTable 1 we can assess this statement in light of our broad selection of preclinical GL261 studies and observe that, with few exceptions, 30,38 the anti-PD-1 antibody was delivered intraperitoneally and the doses, after those given in mg/kg,^{30,34,37} are put in the context of an average mouse's weight, all hovered around 200 µg (Table 1). The success of the monotherapeutic intervention clearly varied dramatically between studies but the antibody dosage did not, suggesting that other variables are contributing to differential murine survival. That being said, antibody dose certainly has some role to play. It is known that in the GL261 model, all else held equal, very low doses of anti-PD-1 monotherapy (eg, 100 µg/day) have minimal impact on mouse survival in conditions where a very high dose (eg, 500 µg/day) has therapeutic efficacy.44 Thus, the notion that the success of anti-PD-1 monotherapy is dosage dependent²⁵ appears to be a necessary but not a sufficient relationship to ensure therapeutic success in the GL261 model. Dose alone (µg antibody delivered), which hardly changes between studies, fails to explain the variability in Table 1.

While the amount of anti-PD-1 antibody delivered per dose may vary little across Table 1, significant variation between studies was observed in the schedule with which anti-PD-1 is administered (Table 1). The most prevalent day for initiation of therapy is day 10 after tumor inoculation (6/16; 37.5%), with the range spreading from day 2 to day 10. Other immunotherapeutic interventions in the GL261 model have shown great dependency upon time of therapy initiation⁴⁵⁻⁴⁷; anti-CTLA-4 monotherapy, for instance, had indistinguishable efficacy from a control IgG when given to GL261-bearing mice 12 days postinoculation, while it had a statistically significant impact on murine survival when given 3 days postinoculation.45 To the best of our knowledge, the direct association between earlier treatment and better outcomes in the GL261 model has yet to be demonstrated for anti-PD-1 CBI. What has been demonstrated, however, is the link between GL261 tumor volume at the time of anti-PD-1 administration and the survival outcome.44 Knowing that GL261 tumor volume is a function of time,⁴⁸ it seems prudent to assume that increased time after tumor inoculation could influence survival outcome following anti-PD-1 CBI monotherapy until the contrary has been established. This presumed association is impossible to determine from a retrospective literature review because many other variables beyond the altered date of therapy initiation fluctuate between these studies (Table 1). Accordingly, there is no stark delineation between early treatment (eg, day 2 postinoculation²⁹) and late treatment (eg, day 10 postinoculation³⁴) clearly driving differential survival (Table 1). That being said, the links that have been forged between GL261 tumor volume and time, and between tumor volume and treatment resistance, lead us to suggest that CBI studies, anti-PD-1 therapy included, in the GL261 model attempt both an "early" and a "late" time-point therapy initiation. This would not only be enormously beneficial in determining the extent to which early treatment of this model glioblastoma is a determinant of survival outcome, but also do significant good in moving preclinical practices closer to clinical reality, as many of the presenting symptoms leading to the diagnosis and treatment of glioblastoma are those accompanying late-stage disease.¹

Another antibody-related factor beyond dosing strategy that could contribute to the variable outcomes of anti-PD-1-treated GL261-bearing animals is the potential for non-equivalencies in various anti-PD-1 clones used for preclinical work, a possibility which has remained underexplored. Intriguing new data demonstrates the variable impact of anti-PD-1 clones routinely utilized in murine preclinical studies.⁴⁹ Particularly, this work shows the Armenian Hamster IgG G4 clone to promote depletion of PD-1+T cells in comparison to the Rat IgG2a RMP1-14 clone⁴⁹ (these clones were used in 4/16 and 7/16 of the studies discussed here, respectively; Table 1). Furthermore, there is evidence to suggest that the RMP1-30 clone, used in one of the 16 studies explored here,³³ does not actually block the interaction of the PD-1 molecule with PD-L1 and PD-L2 like the J43 and RMP1-14 clones do.⁵⁰ While we do not know the impact of either of those findings in the context of CBI for the GL261 glioma model, the potential intrinsic differences between the various clones are certainly worth further investigation as a potential contributor to the mixed preclinical data on CBI in GL261, along with dose differences, timing of administration, and route of administration.

Variables Relating to the Intracranial Inoculation of GL261 Cells

When considering disparate outcomes of CBI in an implantable tumor model, there are multitudinous variables that contribute to the actual process of inoculation. Some of these factors related to GL261 inoculation are already known to contribute to varying survival in the context of anti-PD-1 CBI and some have a more indirect connection that merits further investigation. It has been established that higher cell counts for the initial tumor inoculum can shorten the survival of mice bearing GL261 gliomas.¹⁵ Accordingly, comparing the efficacy of anti-PD-1 CBI in the GL261 model when starting tumor load can vary by an order of magnitude or more across studies is potentially confounded. In the 16 papers examined, a range of GL261 inoculum from 20 000 cells³⁰ to 500 000 cells³⁵ was observed, with a plurality (5/16; 31.25%) using 130 000 cells (Table 1). One publication failed to list the number of cells inoculated at all.³⁷ Like with anti-PD-1 dosing, no clear trend exists to separate groups A, B, and C by starting cell count alone (Table 1). It is possible that this variable is either insufficient to change the outcome of a study by itself, or that the impact of increasing tumor cellularity has a nonlinear relationship with anti-PD-1 resistance. The relationship between absolute GL261-tumor volume and the efficacy of anti-PD-1 monotherapy⁴⁴ likely has interplay with this variable as well, as different initial tumor inoculums could-even at equivalent days postinoculation-vary dramatically in size from one another and therefore display different therapeutic outcomes even were all other variables to be held constant.

dvances

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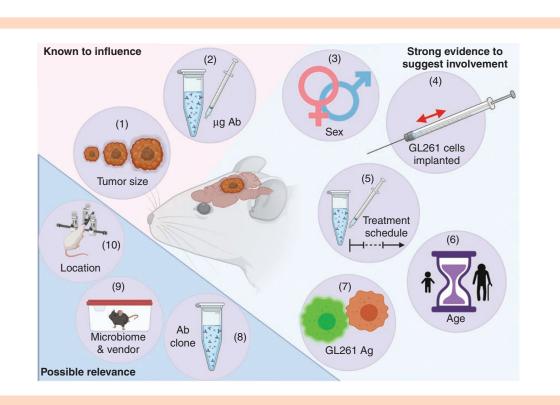


Figure 1. Factors known to, or likely to, impact survival of GL261-bearing mice treated with anti-PD-1 monotherapy categorized by strength of supporting evidence. Many experimental variables that fluctuate across experiments investigating the survival benefit of anti-PD-1 monotherapy in the GL261 model are known to, or suspected of, modulating murine survival time. The factors for which there is evidence of direct modulation of survival time in anti-PD-1-treated GL261-bearing animals include the tumor size at the time of treatment (1) and the dosage of delivered antibody (2). This direct evidential support is indicated by the pink background. The pale blue background includes variables that are known to alter the survival of GL261-bearing animals and could, presumably, also do so in the context of anti-PD-1 monotherapy. These include the sex of the treated animals (3), the number of GL261 cells inoculated (4), the schedule with which the checkpoint blockade agent is administered (5), the age of mice at the time of tumor inoculation (6), and the antigenic potential of the GL261 cell line used (7). Experimental variables for which indirect evidence in related models suggests a possible relation to the survival of anti-PD-1-treated GL261-bearing animals are on a darker blue background. These factors include the clone of anti-PD-1 antibody used (8), the source of the mice and their accompanying microbiome (9), the coordinates at which the tumor is implanted (10).

Other tumor-related factors beyond the starting cell count may interface with murine survival in underappreciated manners, such as tumor antigenicity and location. First, the relative immunogenicity of wild-type GL261 compared to the firefly luciferase-expressing GL261 cell line (GL261-Luc), while somewhat conflicted in the literature, is certainly worth noting. While an older study failed to observe any gross changes in overall growth kinetics or T-cell infiltration between GL261 and GL261-Luc tumors,⁵¹ a more recent study found that wild-type GL261bearing mice had shorter survival than mice bearing either of 2 types of Luciferase-expressing GL261.52 The ability of each of these 3 cell lines to replicate in vitro was comparable and the difference in survival was instead linked to a more inflammatory state within the Luciferase-expressing tumors as shown by a cytokine microarray revealing elevated IFN-y and IL-1 α levels.⁵² Whether the differential cytokine levels in the tumor microenvironment are sufficient to dictate a differential response to anti-PD-1 therapy is yet unknown, but the equivalence of GL261 and GL261-Luciferase cell lines should not be taken for granted without further study. Another minimally explored, but potentially significant, factor in influencing murine survival between studies is the location of tumor inoculation. Some orthotopic glioma models have been shown to have a strongly location-dependent impact on survival.⁵³ This trend was hypothesized to be due to each tumor's relative proximity to the ventricles increasing its odds of growing into the ventricles and causing obstructive hydrocephalus, leading to endpoint-associated symptoms even without significant tumor mass.⁵³ A similar finding with ventriclecontacting tumor growth has been observed to limit survival in glioblastoma patients,⁵⁴ but we have yet to find a study investigating the impact of varying depths or locations of GL261 inoculation on murine survival. If tumor size, antigen composition, and location do systematically alter tumor susceptibility to anti-PD-1 CBI, methodical study of these factors in the GL261 model could help both to clear up existing confusion in the literature and to identify key hallmarks of susceptible tumors in order to focus future CBI clinical trials on patients likely to receive the maximal benefit.

Variables Relating to Mice Used

Glioblastoma is primarily diagnosed in the older adult population and is more common among men than

women.^{55,56} Despite these facts, the vast majority of the 16 studies explored here (11/16; 68.75%) were conducted in 6- to 12-week-old female mice (Table 1). By one approximation, 6-week-old mice, which are used in at least 10 of the 16 studies discussed above, have been compared to 12-year-old human adolescents.⁵⁷ This is far removed from the 64 years of life the average glioblastoma patient has accumulated before their diagnosis.¹This discrepancy matters as increased age is known to diminish expected survival times both in human GBM patients⁵⁸ and in the CT-2A and GL261 murine glioma models.57 This increased age was also a negative prognostic factor for response to an anti-PD-1 inclusive combination immunotherapy in both murine models.⁵⁷The fact that heterochronic bone marrow transplant was unable to reverse this trend suggests that non-hematopoietic factors were involved in the ageassociated immunosuppression preventing therapeutic success.⁵⁷ While we are unaware of any studies specifically examining anti-PD-1 monotherapy in the context of aging in the GL261 model, the weight of evidence suggests that there is likely to be a strong age-associated decline in therapeutic efficacy (Figure 1) which makes the justification of choosing young mice an important task if the preclinical work is meant to have clinical relevance. Furthermore, these facts make the complete omission of mouse age in some of the studies examined in Table 1,^{31,37} a point of concern in regards to the generalizability of their findings.

Pertaining to biological sex, it has been established that males are not only more commonly diagnosed with glioblastoma⁵⁹ but also have shorter survival after diagnosis than do females.^{59,60} This has been shown to hold true with the implantable GL261 and SB28 murine glioma models⁶¹ as well. Interestingly, this survival benefit for female mice has been linked to the hematopoietic compartment, with elegant bone marrow transfers from male donors into female hosts completely abrogating the sex-specific survival benefit in both murine glioma models.⁶¹ This difference was determined to be driven by sex-specific differences in myeloid-derived suppressor cell (MDSC) accumulation at the tumor site,⁶¹ which is of particular interest in the context of anti-PD-1 treatment given the increasingly appreciated effect of this intervention on sculpting myelopoiesis.⁶² Despite the sex-associated immunologic and clinical differences of both glioblastoma and murine glioma models, only 2 of the murine studies explored here^{29,33} use males as experimental animals for their immunotherapy regimen. Three of the studies fail to list the sex of their animals at all^{31,32,37} and not a single paper used both sexes of animals for their survival studies. It has not been established that sex influences the efficacy of anti-PD-1 monotherapy in the GL261 model, but the fact that sex has an impact on survival alone⁶¹ (Figure 1) provides a significant barrier to interstudy comparison and perhaps contributes to why neither of the studies using male mice falls into Group C of Table 1. Vetting a therapeutic strategy that will largely be utilized in older male humans on young/adolescent female mice, while certainly not invalidating the work, raises questions of applicability and generalizability that will be addressed at length below.

Another potential issue related to variations in mice used for these studies is the vendor from which the mice are obtained. While this facet has received significant attention in recent years in the context of gut microbiota differences, particularly between C57BL/6 animals from Taconic versus The Jackson Laboratory,63 this only accounts for part of the variance among C57BL/6 mice. Not only the vendorassociated gut microbiome varies, but also the subspecies of C57BL/6 that have arisen through separate inbreeding at discreet centers. C57BL/6 animals from multiple different vendors have been shown to have differential susceptibility to various pathogens,⁶⁴⁻⁶⁶ to have differential behaviors,^{67,68} and to have various genetic differences that have accumulated over time.⁶⁹ While these vendor-specific effects have been underexplored in the context of the glioma models, the gut microbiome has been implicated both in responsiveness to anti-PD-1 CBI in humans⁷⁰⁻⁷² and in controlling glioma progression⁷³ and anti-PD-1 responsiveness in the GL261 model.⁷⁴ It could certainly be possible for C57BL/6 mice from different vendors to respond differently both to glioma inoculation and to treatment with anti-PD-1 on the grounds of either slight genetic alterations or shifts in gut microbiome, and these potential sources of variability should be explored further.

Discussion

The lack of consensus in the literature pertaining to the therapeutic efficacy of checkpoint blockade strategies in the GL261 model should give us pause. This is especially true in light of the reproducibility crisis observed in other disciplines.75-77 Overcoming, and even benefiting from, the significant deviation that exists within our field will require being methodical in the choosing and reporting of our methods. Even analysis of a single measure of a single experiment type-survival time of GL261-bearing C57BL/6 mice when treated with anti-PD-1 CBI monotherapy-we observed widely discrepant outcomes (Table 1). Were we to broaden our scope to other immunotherapies and other preclinical glioma models, we would observe similar broad variability. For instance, some studies find that anti-CTLA-4 antibody monotherapy leads to 50% long-term survival of GL261-bearing mice,⁷⁸ while others show the intervention to not lead to any survival extension at all.⁷⁹ Relatedly, in the CT-2A glioma model, some researchers have observed up to 60% long-term survival following anti-PD-1 monotherapy⁸⁰ while most observe no survival benefit whatsoever.^{28,81} We do not intend to cast doubt on the scientific practices or the honest work of our colleagues, but rather to contend that common and innocuous variations in the execution of a simple experiment can lead to drastically different conclusions. Obviously, the complete standardization of experimental practices across the scientific community is an unrealistic expectation. Given the heterogeneous nature of any human patient population, such standardization might not even be desirable. Thus, the goal of our article is not to argue for homogeneity in approach but rather to advocate for intentionality in the selection and reporting of experimental parameters.

In Figure 1, we arrange all 10 discussed variables by the degree of published data supporting their involvement in the modulation of animal survival in orthotopic GL261 experiments. These factors, covered at length in the sections above and enumerated in the figure legend, all have direct

or circumstantial evidence tying them to murine survival in glioma studies. The effort put into addressing these individual variables should be commensurate with the degree to which they are likely to play an important role in affecting the study outcome of murine survival. By that reasoning, the least supported variables of anti-PD-1 clone, mouse origin, and coordinates of tumor inoculation could all be the subjects of interesting studies in the future but likely do not need to be widely addressed by individual research programs. Without more supportive evidence that these variables are sufficient to drive survival study outcomes, the effort for individual research programs to test their immunotherapies against tumors inoculated at varying coordinates or with varying anti-PD-1 clones may not be warranted.

The variables which have a greater deal of evidence supporting their modulation of GL261-bearing mouse survival should, minimally, be considered during the process of experimental design. Just as increased knowledge about the mutational burden of GL261 tumors has led increasingly to them being viewed as a model of hypermutated glioblastoma,^{26,82} increased knowledge of other experimental variables should change our practices moving forward. For instance, knowing that modification of the GL261 cell line with bioluminescent reporters enhances its antigenicity⁵² should temper the enthusiasm of successful immunotherapeutic strategies vetted in that model and perhaps encourage the utilization of a second model system like CT-2A, SB28, or wild-type GL261 for verification. Other sets of experimental factors should inform our practices as well. Human glioblastoma patients are often first diagnosed on account of symptoms associated with late-stage disease¹ and a number of experimental variables in the murine GL261 setting go into recapitulating that scenario. This interweaving of variables, including cell count of tumor inoculum, the time allowed for tumor engraftment prior to therapy, and the stochastic differences in tumor size and growth all make it difficult to determine exactly what qualifies as "late stage." One way to encapsulate all these variables and to minimize the difficulties in interstudy comparisons is by encouraging the reporting of measurements of tumor size at the time of treatment, for example, by the use of T2-weighted MRI imaging. Knowing this absolute value would make the comparison of a study inoculating 20 000 cells and treating 8 days later³⁰ versus a study inoculating 500 000 cells and treating 5 days later³⁵ more easily comprehensible. This may not be within every research program's ability to perform. Therefore, an alternative approach could be to initiate therapy at the first sign of murine behavioral changes under that laboratory's particular inoculation protocol. A move toward either of these practices would increase the interpretability of future immunotherapy regimens in preclinical glioma models and facilitate an easier recapitulation of the observed clinical trend of glioblastoma diagnosis at the time of advanced symptoms.

In considering how choices in experimental design change the nature of what is being modeled, it is also worth considering the experimental variables of murine sex and age. Increased age negatively impacts the survival of both CT-2A and GL261-bearing mice.⁵⁷ Meanwhile, male sex is a negative prognostic factor for the survival of both SB28 and GL261-bearing mice.⁶¹ Both these trends have been observed in human glioblastoma patients as well.⁵⁸⁻⁶⁰ If preclinical immunotherapy studies properly report these variables, or even more rigorously assess the effect of murine sex and age groups in their studies, we could maximize the utility of our preclinical data and gain a better understanding of human GBM patients most likely to benefit from the tested intervention. Being more cognizant of which comparable human population is being modeled will ensure the most accurate interpretation and translation of preclinical study results.

Reflecting on the broadly discrepant outcomes of anti-PD-1 antibody monotherapy in the GL261 model gives us an opportunity to reassess our common assumptions regarding our experimental practices. Acknowledging interstudy variability will allow therapeutic success achieved in the GL261 model to be considered more rigorously against competing therapeutic approaches. Furthermore, systematically assessing the contribution of initial conditions like age, sex, and tumor size on ultimate survival outcomes will help researchers both to better understand the limits of their model and add additional nuance to our preclinical understanding before presumptive therapies are translated to the clinic. This degree of heightened experimental rigor will give the best chance of capitalizing on the enormous potential that immunotherapeutic interventions carry to revolutionize the life expectancy of patients diagnosed with the grim malignancy of glioblastoma.

Keywords

Anti-PD-1 | GBM | GL261

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