

Nrf2 and the Nrf2-Interacting Network in Respiratory Inflammation and Diseases



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Abstract Atmospheric pollutants and cigarette smoke influence the human respiratory system and induce airway inflammation, injury, and pathogenesis. Activation of the NF-E2-related factor 2 (Nrf2) transcription factor and downstream antioxidant response element (ARE)-mediated transcriptions play a central role in protecting respiratory cells against reactive oxidative species (ROS) that are induced by airway toxins and inflammation. Recent studies have revealed that Nrf2 can also target and activate many genes involved in developmental programs such as cell proliferation, cell differentiation, cell death, and metabolism. Nrf2 is closely regulated by the interaction with kelch-like ECH-associated protein 1 (Keap1), while also directly interacts with a number of other proteins, including inflammatory factors, transcription factors, autophagy mediators, kinases, epigenetic modifiers, etc. It is believed that the multiple target genes and the complicated interacting network of Nrf2 account for the roles of Nrf2 in physiologies and pathogenesises. This chapter summarizes the molecular functions and protein interactions of Nrf2, as well as the roles of Nrf2 and the Nrf2-interacting network in respiratory inflammation and diseases, including acute lung injury (ALI), asthma, chronic obstructive pulmonary disease (COPD), pulmonary fibrosis (PF), cystic fibrosis (CF), viral/bacterial infections, and lung cancers. Therapeutic applications that target Nrf2 and its interacting proteins in respiratory diseases are also reviewed.

1 Introduction

Environmental toxins generated by modern industry significantly influence human health. The respiratory system is most susceptible to pollutants as it is directly exposed to atmospheric toxins such as airborne chemicals, O₃, particulate matter (PM), and cigarette smoke (CS). Respiratory diseases and relevant secondary symptoms, including ALI, asthma, COPD, bacterial/viral infections, lung fibrosis, and lung cancers, are among the most common causes of severe illness and death worldwide [1].

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Most of the airway pathogenesises are related to xenobiotic and oxidative stresses occurring in different types of respiratory cells. These stresses can be triggered directly by the inhaled toxins (xenobiotics) or by excess reactive oxygen species (ROS) that are induced by many of these toxins. Intracellular accumulation of ROS may damage macromolecules including DNA, proteins, and lipids, thereby inducing cell defects or death. Imbalanced redox homeostasis and cell damage lead to inflammatory responses, which further enhance oxidative stress. Cellular factors that can respond to and reduce oxidative/xenobiotic stresses are essential protective machinery against airway pathogenesis.

NF-E2-related factor 2 (Nrf2), a cap 'n' collar (CNC)-bZIP family transcription factor, is a central regulator that protects cells against ROS and xenobiotics [2, 3]. Activated by oxidative and xenobiotic stimuli, Nrf2 binds to antioxidant response elements (AREs) and activates a battery of antioxidant and detoxifying genes [4]. Nrf2 also activates many developmental genes, allowing Nrf2 to regulate programs that are related with cell proliferation, cell differentiation, cell death, and metabolism [4]. In addition, Nrf2 interacts with many other proteins and pathways, providing a complicated regulatory network for redox homeostasis and other cellular programs. Nrf2 is identified as the key player and important drug target for a number of human diseases including respiratory diseases [5]. Fully understanding the molecular partners and downstream functions of Nrf2 will significantly aid in the development of Nrf2-targeting therapies. This chapter first reviews the molecular mechanisms of Nrf2, focusing on the multiple interacting partners of Nrf2. Next, the molecular functions and therapeutic applications of Nrf2 and the Nrf2-interacting network in respiratory inflammation and diseases are summarized.

2 Molecular Mechanisms of the Nrf2 Network

2.1 *Keap1-Nrf2 Pathway*

The key regulator of Nrf2 activity in response to oxidative and xenobiotic stimuli is kelch-like ECH-associated protein 1 (Keap1). As an adaptor of the Cul3-based E3 ligase, Keap1 confines Nrf2 to the cytoplasm through mediating Nrf2 ubiquitination and degradation [6]. The Keap1–Nrf2 interaction is mediated by the Kelch repeats of the Keap1 dimer and the N-terminal DLG and ETGE motifs of Nrf2. Cysteine residues (e.g., C151, C273, C288) on Keap1 can be modified by a broad range of oxidative and xenobiotic compounds [7, 8]. According to the established model, cysteine modifications alter the conformation of the Keap1 dimer and interfere with the Keap1–Nrf2 interaction, thereby blocking the proteasomal degradation of Nrf2. The stabilized Nrf2 proteins accumulate in the nucleus and activate transcriptions [9, 10]. After induction, Keap1 can enter the nucleus and shuttle Nrf2 back to the cytosol [11]. The genetic interaction of Keap1 and Nrf2 has been verified in vivo. *Keap1*^{-/-} mice die before weaning due to a severe dysfunction of keratinocytes, while co-knockout of Nrf2 significantly rescues the viability of *Keap1*^{-/-} mice [12].

2.2 *Nrf2-Downstream Genes and Functions*

2.2.1 Antioxidant and Detoxifying Genes

Microarray analyses based on Nrf2 activators and *Nrf2* knockout mice have identified many antioxidant and detoxifying genes that are controlled by Nrf2-ARE [4, 13–15]. Most of these genes code for enzymes that catalyze the metabolism or removal of ROS and other toxins, including: (1) phase I antioxidant enzymes such as superoxide dismutases (SODs), glutathione peroxidase (GPx), and glutathione reductase (GR); (2) phase II detoxifying enzymes such as glutathione-S-transferase (GST), NADP(H):quinone oxidoreductase-1 (NQO1), glutamate-cysteine ligases (GCLM/GCLC); (3) phase III xenobiotic transporters such as multidrug resistance protein 1 (MRP1); and (4) other stress response proteins such as heme oxygenase-1 (HO-1). Protective roles of some of these enzymes against respiratory inflammation and damages have been demonstrated by several studies. For example, the lungs of *Sod2* mutant mice have enhanced sensitivity to hyperoxia [16], while overexpression of *Sod2* in the mouse airway reduces the hyperoxia-induced lung inflammation and injury [17]. Activation of the Nrf2-ARE antioxidant pathway is believed to be an efficient therapeutic strategy for redox-related lung diseases.

2.2.2 Other Nrf2-Target Genes and Functions

Recent studies in different model systems have revealed that Nrf2 can target genes independent of oxidative and xenobiotic responses, demonstrating that Nrf2 can act as more than just a detoxifying factor. For examples, a ChIP-seq study combining microarray assays using mouse embryonic fibroblasts with either enhanced or reduced levels of Nrf2 identified around 1000 Nrf2-target genes, more than half of which were involved in cell proliferation [4]. Nrf2 can control adipogenesis through binding to and activating *peroxisome proliferator-activated receptor γ* (*PPAR γ*), *retinoid X receptor α* (*RXR α*), and *small heterodimer partner* (*SHP*) nuclear receptor genes as well as some lipid metabolism genes [18–21]. The Nrf2-activated *RXR α* can also control the differentiation of acute myeloid leukemia cells [22]. In addition, Nrf2 can regulate neuronal stem cell fate through activating genes that inhibit self-renewal or promote differentiation [23]. Another essential Nrf2-target gene is *p62*, which mediates the role of Nrf2 in the regulation of autophagy [24]. In a human lung cancer cell line, NRF2 activates transcripts of anti-apoptotic factor Bcl-2 and glucose metabolic enzymes [25, 26], indicating the promotive role of Nrf2 in cancer cell proliferation. It is notable that Nrf2 can target and inhibit some proinflammatory cytokine genes such as *IL6* through an ARE-independent manner, revealing a novel mechanism that mediates the anti-inflammation function of Nrf2 [27].

Structural and functional conservation of the Keap1-Nrf2 signaling has been revealed in mammals, zebrafish, and *Drosophila* [8, 28]. Studies using non-mamalian model organisms provide additional insights into the biological functions of Nrf2. In *Drosophila*, CncC and dKeap1 (homologs of Nrf2 and Keap1, respectively) regulate metamorphosis through targeting and activating ecdysteroid biosynthetic genes

and response genes in a tissue-specific manner [29], indicating a potential role of Nrf2 family proteins in the regulation of steroid hormones. CncC and dKeap1 also regulate intestinal stem cell proliferation in *Drosophila* [30]. It is believed that multiple developmental functions of Nrf2 and Keap1 in specific tissues contribute to their complicated roles in diseases [31, 32]. How Nrf2 selectively targets and activates developmental genes remains to be elucidated, but it is likely that it acts through mechanisms that differ from the classic ARE-dependent antioxidant pathway. Notably, recent studies showed that dKeap1 can directly bind to chromatin and function as a transcription coactivator with CncC through interaction with CncC at specific genomic loci [29, 33].

2.3 Other Nrf2-Interacting Proteins

Besides Keap1, a number of proteins that interact with Nrf2 have been identified. These protein partners can regulate the activity, subcellular localization, degradation, and chromatin-binding specificity of Nrf2. This places Nrf2 in the center of a multi-layer regulatory network for redox homeostasis and other cellular programs (Fig. 1). These interactions are believed to mediate the complicated roles of Nrf2 in pathogenesis, including respiratory diseases.

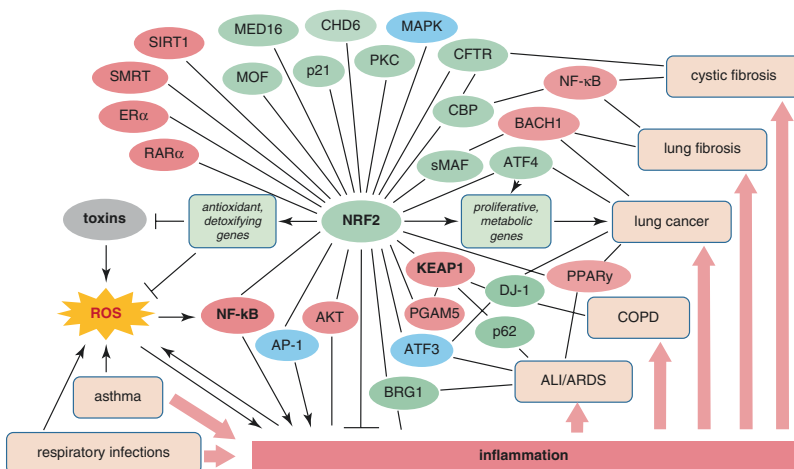


Fig. 1 Nrf2-interacting network and its correlation with respiratory inflammation and diseases. Nrf2 plays a protective role against respiratory inflammation and respiratory diseases through mediating the transcriptional responses to oxidative and xenobiotic stresses. Nrf2 can also promote lung oncogenesis through activating genes that facilitate cell proliferation. In addition, Nrf2 interacts with many proteins and pathways, placing Nrf2 in the center of a network that controls redox homeostasis, inflammation, and pathogenesis. The ovals represent proteins that can interact with Nrf2. The positive and negative regulators of Nrf2 activity are colored in green and red, respectively. Proteins in blue ovals can either activate or inhibit Nrf2 activity. The identified roles of Nrf2 and some of the Nrf2-interacting proteins in respiratory inflammation and diseases are represented with line connections

2.3.1 Inflammatory Factors

Nuclear factor kappa light chain enhancer of activated B cells (NF- κ B) and activator protein 1 (AP-1) are classic transcription factors that induce inflammation in response to oxidative stress through the activation of genes encoding proinflammatory cytokines and chemokines. The essential roles of NF- κ B and AP-1 proteins in inflammation and chronic lung diseases (e.g., asthma, IPF, and COPD) have been identified by many studies [34–36]. It is noted that oxidative stress can co-activate NF- κ B and Nrf2 pathways. For example, PM exposures co-induce NF- κ B and Nrf2-HO-1 in both mouse lungs and human bronchial epithelial cells [37, 38]. NF- κ B can inactivate Nrf2 by competing with the CBP–Nrf2 interaction or by recruiting the transcription repressor HDAC3 to ARE [39]. The AP-1 subunit c-Jun can dimerize with Nrf2 at AREs and co-activate Nrf2-induced transcription, while another AP-1 subunit c-Fos can suppress Nrf2-induced transcription [40, 41]. Crosstalk between Nrf2 and these inflammatory factors establishes a mechanism whereby Nrf2 regulates inflammation.

2.3.2 Small Maf Transcription Factors

Nrf2 can form heterodimers with small Maf (sMaf) transcription factors, including MafF, MafG, and MafK [42]. These interactions are mediated by their leucine zipper domains. It has been well established that the Nrf2-sMaf complexes recognize ARE and the sMaf proteins can support Nrf2-mediated transcriptional activation [43]. Combinatory knockouts of MafF and MafG can rescue the lethality of *Keap1* null mice, suggesting the cooperative roles of sMaf proteins with Nrf2 in vivo [44]. ChIP-Seq analyses of Nrf2-MafG-binding sites indicate that Nrf2-sMaf heterodimers globally regulate antioxidant and metabolic genes [45].

2.3.3 ATF/CREB Transcription Factors and Cofactors

A number of transcription factors, coactivators, and mediators can form complexes with Nrf2 at AREs and control Nrf2-mediated transcription. The mediator MED16 directly interacts with Nrf2 and mediates 75% of Nrf2-dependent transcription [46]. CREB-binding protein (CBP) and its cofactor p300 can bind the Neh4/5 domains of Nrf2. It was observed that p300 and Nrf2 can be co-recruited to ARE in response to oxidative stress [47, 48]. The Nrf2–CBP interaction can be regulated by a Cl⁻/HCO₃⁻ channel protein cystic fibrosis transmembrane conductance regulator (CFTR), which also interacts with Nrf2 and promotes Nrf2-dependent transcription in human bronchial epithelial cells [49]. Activating transcription factor 4 (ATF4) can dimerize with Nrf2 at ARE and co-activate *HO-1* [50]. Formation of the ATF3-Nrf2 dimer can compete with the Nrf2–CBP interaction at ARE and suppress Nrf2-induced transcription [51]. On the other hand, ATF3 depletion in human bronchoalveolar epithelial cells enhances NRF2 degradation through KEAP1 and DJ-1 pathways [52]. Therefore, ATF3 can either positively or negatively control Nrf2 activity.

2.3.4 Nuclear Receptors

Several nuclear receptors can enhance or inhibit Nrf2 activity through interactions with Nrf2. PPAR γ can directly bind to Nrf2 [53], while Nrf2 and PPAR γ also regulate the transcription of each other in the lungs of hyperoxia-induced mice [54, 55]. Estrogen receptor α (ER α) can form a complex with Nrf2 and suppress the Nrf2 pathway in a ligand-dependent manner [56]. Retinoic acid receptor α (RAR α) can inhibit Nrf2 binding to ARE [57]. Direct interactions between DNA-binding domains of RAR α and the Neh7 domain of NRF2 have been reported recently in acute promyelocytic leukemia, in which RAR α fuses with promyelocytic leukemia protein (PML) [58].

2.3.5 p62 and Autophagy

Nrf2 can directly activate gene expression of p62, a selective adaptor that targets ubiquitinated cargos to the autophagosome during autophagy [24, 59]. In addition, phosphorylated p62 can interact with the Kelch repeats domain of Keap1 and compete with the Keap1–Nrf2 interaction, resulting in stabilization of Nrf2 and activation of antioxidant genes [24, 60, 61]. Inhibition of autophagy by arsenic can induce accumulation of autophagosomes and p62, resulting in prolonged activation of Nrf2 [62]. On the other hand, since p62 is an autophagy target, induction of autophagy can enhance the degradation of p62–Keap1 complexes, thereby activating the Nrf2 pathway [63].

2.3.6 Kinases and Nrf2 Phosphorylation

In response to specific compounds, Nrf2-ARE activity can be controlled by protein kinases such as PKC, Akt, ERK, and p38 MAPK in specific cell types, including pulmonary epithelial cells [64]. Nrf2 can be directly phosphorylated by PKC and MAP kinases [65, 66]. How this phosphorylation controls Nrf2 function remains to be elucidated. ERK/MAPK is required for the oxidative-induced NRF2 nuclear localization in HepG2 cells [67]. However, another study indicates that phosphorylation of Nrf2 by MAPK at multiple sites only moderately enhances Nrf2 nuclear accumulation [66]. The p38 MAPK inhibits Nrf2-mediated *HO-1* expression in HepG2 and MEF cells [68, 69]. In *C. elegans*, phosphorylation of the Nrf2 homolog SKN-1 by p38 MAPK or by Akt have opposite effect on SKN-1 nuclear accumulation and transcription activity [70, 71]. In *Drosophila*, CncC functions downstream of the Ras/ERK pathway to activate ecdysone-synthetic genes in the prothoracic gland [29, 72]. In addition, the constitutively activated Ras^{V12} modulates CncC binding at specific chromatic loci, indicating that MAPK may directly control the chromatin binding specificity of Nrf2 [29].

2.3.7 Acetylation of Nrf2

Nrf2 can be acetylated by CBP/p300 [48]. Upon oxidative stimuli, acetylation of Nrf2 at Lys-588/591 by CBP/p300 enhances the binding of Nrf2 to ARE promoters, supporting CBP/p300 as a coactivator of Nrf2 in oxidant responses [48]. Alternative reading frame (ARF), a tumor suppressor that triggers apoptosis, directly interacts with Neh1/3 domains of NRF2 in a human non-small cell lung carcinoma cell line [73]. It was proposed that the ARF–Nrf2 interaction prevents acetylation of NRF2 by CBP and suppresses Nrf2-mediated transcription [73, 74]. Males absent on the first (MOF), a histone acetyltransferase, can acetylate Nrf2 at Lys-588 and is required for Nrf2-induced transcription [75]. Nrf2 can be de-acetylated by sirtuin 1 (SIRT1) at Lys-588/591, which suppresses Nrf2-dependent transcription [76]. These studies indicate acetylation as a positive regulatory mechanism for Nrf2 activity. Alternatively, since both CBP/p300 and MOF are histone acetyltransferases [75, 77], these proteins may cooperate with Nrf2 indirectly through the epigenetic regulation of chromatin structure.

2.3.8 Epigenetic Machinery

Increasing lines of evidences reveal the interactions of Nrf2 with epigenetic modifiers. Brahma-related gene 1 (Brg1), the ATPase of SWI/SNF complex, forms a complex with Nrf2 selectively at the *HO-1* promoter and activates *HO-1* transcription through Brg1-mediated Z-DNA formation [78, 79]. Chromodomain helicase DNA-binding protein-6 (CHD6) interacts with the Neh3 domain of Nrf2 and facilitates Nrf2-induced *NQO1* expression [80]. Silencing mediator for retinoid and thyroid hormone receptor (SMRT), a transcriptional repressor that mediates histone deacetylation, can interact with Nrf2 Neh4/5 domains and inhibit Nrf2-induced *GSTA2* expression [81]. Notably, a recent study in *Drosophila* showed that depletion of CncC or dKeap1 suppresses pericentric silencing and decreases the level of heterochromatin marker H3K9me2, providing direct evidence in support of the epigenetic role of Keap1/Nrf2 family proteins in chromatin remodeling [82].

2.3.9 Other Nrf2-Interacting Proteins

BTB domain and CNC homolog 1 (Bach1) is a CNC family transcription factor that is essential in controlling heme level through the inhibition of *HO-1* gene. Bach1 can dimerize with sMaf and compete with the Nrf2–sMaf interaction at ARE sites and inhibit the activation of *HO-1* [83]. NRF2-induced *HO-1* expression requires inactivation of BACH1 [84]. BACH1 can promote invasion and metastasis by activating metastatic genes [85, 86]. Recent studies showed that Nrf2 activation can inhibit the F-box Protein 22 (Fbxo22)-induced Bach1 degradation, revealing a novel mechanism whereby the Nrf2–Bach1 pathway promotes carcinogenesis [87, 88].

The p21 tumor suppressor can interact with the DLG and ETGE motifs of Nrf2, compete with the Keap1–Nrf2 interaction and stabilize Nrf2 [89]. This discovery adds one more mechanism to the multiple functions of the p53-p21 pathway in cytoprotection and cell survival, indicating a potential role of the Nrf2-p21 network in oncogenesis [90].

DJ-1/PARK7 is a protein that plays multiple antioxidant and cytoprotective functions and is associated with cancer and neurodegeneration. DJ-1 can stabilize Nrf2 by inhibiting the Keap1–Nrf2 interaction and proteasomal degradation of Nrf2 [91]. High levels of both DJ-1 and NRF2 were seen in a large set of lung carcinomas [92].

Phosphoglycerate mutase 5 (PGAM5) is a mitochondrion-binding protein whose ESGE motif can interact with the Kelch domain of Keap1 [93]. It was proposed that a Keap1 dimer can simultaneously interact with one Nrf2 and one PGAM5 molecule. This PGAM5-Keap1-Nrf2 complex can be anchored to mitochondrial surface and likely serve as an antioxidant mechanism specifically targeting ROS leaked from mitochondria [94].

3 The Roles of Nrf2 and Nrf2-Interacting Proteins in Respiratory Diseases

3.1 ALI/ARDS

Acute lung injury (ALI) is a syndrome manifesting as lung edema, inflammation, and alveolar hemorrhaging. If not treated, ALI can progress into its severe form, acute respiratory distress syndrome (ARDS). ALI can be induced by aspiratory injuries caused by various stimuli. For example, mechanical ventilation (MV) along with hyperoxia is used as a therapy for patients with respiratory dysfunction, but can induce lung inflammation and ventilator-induced lung injury (VILI), which directly causes or exacerbates ALI. ROS play a central role in the pathogenesis of lung injury and ALI/ARDS [95]. Protective roles of Nrf2-ARE-activated genes against respiratory injury have been revealed in mouse models with induced lung injury. For examples, overexpression of SOD2 in airways of mice can reduce the hyperoxia-induced lung injury and inflammation [17, 96]. In support of a protective role of Nrf2 in the lungs, enhanced lung injuries and inflammatory responses to MV were found in *Nrf2*^{-/-} mice compared with wild-type mice, and these responses can be restored by supplementing *Nrf2*^{-/-} mice with antioxidant N-acetylcysteine (NAC) [97]. Sulforaphane (SFN), an activator of Nrf2, can reduce pulmonary injury in wild-type mice but not in *Nrf2*^{-/-} mice [98]. CDDO-imidazolide (CDDO-Im), another Nrf2 activator, protects the lungs of mice against CS-induced oxidative stress and alveolar damage through the Nrf2 pathway [99]. These studies support the use of Nrf2-activating antioxidant therapeutics as a way to attenuate pulmonary inflammation and damage in ALI/ARDS patients.

Cooperative roles of Nrf2 with several interacting partners, including ATF3, PPAR γ , Brg1, p62, and Akt, in lung inflammation and injury have been revealed. ATF3-deficient mice have enhanced susceptibility to ALI and VILI, which is associated with the recruitment of inflammatory cells and loss of junctions among resident cells [52, 100]. It was proposed that ATF3 can reduce lung injury through inhibiting Nrf2 degradation via Keap1 and DJ-1 pathways [52]. The protective role of Nrf2 in hypoxia-induced ALI can also be mediated by PPAR γ , which can be transcriptionally induced by Nrf2 and activate several anti-inflammatory and antioxidant genes [55]. In a hepatic ischemia/reperfusion (HIR)-induced ALI mouse model, overexpression of Brg1 increases Nrf2 activity and reduces ROS and inflammatory factors in lung tissues [101]. Autophagy plays both a positive and negative role in ALI, and this dual role is suggested to be mediated by the Keap1-Nrf2-p62 pathway [102]. The protective role of the PI3K/Akt-dependent activation of the Nrf2-HO-1 pathway against lipopolysaccharide (LPS)-induced lung inflammation and injury was revealed in mice treated with desoxyrhapontigenin [103].

3.2 Asthma

Asthma is a lung disorder that is characterized by the swelling of airways, which causes breathing difficulty, coughing, wheezing, and chest tightness. The swelling of the airways can be initiated or exacerbated by environmental allergens (pollen, mold, etc.) and irritants (smoke, dust, gas, etc.). In a mouse model, challenging by the allergen ovalbumin increases inflammation and asthma symptoms by inducing the expression of proinflammatory factors and increasing mucus secretion and airway hyperresponsiveness. In these allergen-challenged mice, Nrf2 knockout resulted in a worsening of asthma symptoms, represented by a higher number of inflammatory cells in bronchoalveolar lavage (BAL) fluid and an increase in mucus-producing cells in the proximal airways [104]. When being exposed to acetylcholine, a molecule involved in asthma pathogenesis, the lungs of *Nrf2*^{-/-} mice are more prone to asthmatic symptoms than the wild-type mice, represented by the less stretch and the greater resistance of air passage through the airways [104]. In humans, a recent study on childhood bronchial asthma in Egyptian children found that children with lower levels of NRF2 in the serum had more severe asthma [105].

Given the role of Nrf2 in protection against asthma, a number of drugs targeting Nrf2 have been tested in rats and mice and been found to reduce asthma symptoms. For example, polydatin treatment of asthmatic mice showed upregulated Nrf2 signaling and reduced oxidative stress in the lungs as well as attenuation of asthma symptoms [106]. Treatment of asthmatic rats with methanolic extract from *Artemisia pallens* increased Nrf2 levels in the lungs and reduced oxidative stress and inflammation [107]. Taken together, these studies suggest that Nrf2 plays an important role in protecting the lungs against asthma and can be a target for asthma treatment.

3.3 COPD

Chronic obstructive pulmonary disease (COPD) is usually caused by exposure to cigarette smoke (CS) or occupational exposure to various chemicals and smoke, and characterized by airway inflammation and emphysema. Emphysema is mainly caused by damage to alveolar walls and enlargement of air sacs, making it difficult to exhale and inhale. Several studies in mice and humans have indicated that Nrf2 appears to play a protective role against emphysema, and the loss of Nrf2 activity correlates with the progression of COPD. Emphysema can be induced in mice by elastase, which is an enzyme that breaks down elastin, a protein that is vital to the elasticity of the lungs. Although both wild-type and *Nrf2*^{-/-} mice have alveolar wall damage and enlarged airspaces upon elastase treatment, these symptoms are exacerbated in the *Nrf2*^{-/-} mice [108]. Elastase-treated mice had a significant increase in expression of Nrf2-regulated antioxidant genes in the alveolar macrophages. Transplantation of wild-type bone marrow to *Nrf2*^{-/-} mice significantly reduced emphysema symptoms, indicating that the Nrf2-induced antioxidant pathway in alveolar macrophages plays a protective role against lung inflammation and emphysema [108]. In *Drosophila*, drug-induced increase in Nrf2/CncC activity partially rescued the flies that had been chronically exposed to CS [109]. Fisetin, a plant flavonoid compound, has been suggested as a potential therapeutic to COPD since it can activate the Nrf2 pathway and reduced the oxidative stress, inflammation, and lung damage caused by CS exposure to rats [110]. In human studies, it has been found that initial exposure to CS results in activation of NRF2 in alveolar macrophages. However, in older smokers with COPD, NRF2 expression was reduced in alveolar macrophages [111]. Genetic polymorphisms in NRF2-regulatory genes or NRF2-target genes have also been found to contribute to COPD [112]. These studies demonstrate the importance of Nrf2 in protecting the lungs against oxidative stress and damage from CS while suggesting Nrf2 as a potential therapeutic target in COPD treatment.

The role of Nrf2 dysfunction in COPD is likely mediated by the loss of DJ-1. In support of this model, DJ-1 overexpression activates Nrf2 and inhibits apoptosis of alveolar type II cells that are treated with CS extract, suggesting the protective role of the DJ-1-Nrf2 pathway against the CS-induced oxidative stress and inflammatory response [113]. Crosstalk of Nrf2 and NF- κ B in inflammation and COPD have been revealed by many studies [35]. In particular, a number of nature products or pharmacological compounds that can simultaneously activate the Nrf2 pathway and inhibit the NF- κ B pathway have shown therapeutic potency against respiratory inflammation and COPD [114–119].

3.4 Lung Fibrosis/IPF

Lung fibrosis is generally caused by the proliferation of fibroblasts from profibroblasts and the subsequent accumulation of extracellular matrix proteins. Idiopathic pulmonary fibrosis (IPF) is a disease that is characterized by progressive scarring

of the lung tissue around the alveoli. This results in thickening and stiffness of the alveolar walls and makes it difficult for oxygen to penetrate into the bloodstream, leading to symptoms that include dry coughing, breathing difficulty, and finger clubbing. Although the causes of IPF remain to be fully understood, oxidative stresses that are induced by stimuli such as CS are thought to be a contributing factor [120]. IPF patients have higher than normal levels of ROS in the BAL fluid and lower than normal levels of antioxidant enzymes (e.g., SOD) and antioxidants in the lungs. Bleomycin-induced pulmonary injury, inflammation, and fibrosis in mice has been used as a model to study lung fibrosis. *Nrf2*^{-/-} mice have significantly higher sensitivity to the bleomycin-induced fibrogenesis than wild-type mice [121], suggesting that Nrf2 plays a protective role against pulmonary fibrosis.

It is well-known that the herbicide paraquat (PQ) can cause pulmonary fibrosis in animals and human [124]. Rapamycin is known to protect against PQ-induced lung injury and has been suggested as a potential treatment for pulmonary fibrosis [122]. Treatment of rats or mice with rapamycin led to an increase in Nrf2 levels and a decrease of PQ-induced ROS and fibrosis-related factors, and the effect of rapamycin can be reversed by an Nrf2 knockdown [123, 124]. In several in vitro assays, the PQ-induced fibroblast-to-myofibroblast transition (FMT) or epithelial-mesenchymal transition (EMT), both important cellular processes in fibrosis, were inhibited by treatment with rapamycin or Nrf2 activator [123, 124]. Rosavin, emodin, and pirfenidone are other drugs that have recently been proposed as treatments for pulmonary fibrosis [125–127]. Treatment of mice or rats with these compounds increases the expression and/or activity of Nrf2 and reduces the bleomycin-induced lung fibrosis. It is also noticed that these drugs can simultaneously inhibit NF- κ B or Bach1, suggesting that the unbalanced Nrf2/NF- κ B or Nrf2/Bach1 equilibrium may contribute to the development of IPF.

3.5 Cystic Fibrosis

Cystic fibrosis (CF) is a genetic disease caused by mutations in the gene that encodes the cystic fibrosis transmembrane conductance regulator (CFTR), a cAMP-dependent and ATP-gated chloride channel that regulates epithelial surface fluid secretion in respiratory and gastrointestinal tracts. In the lung of a CF patient, the mucus blocks the airways and traps bacteria, leading to infections, inflammation, and respiratory dysfunctions. CF is commonly associated with the F508del mutation of CFTR. It was found that CFTR interacts with Nrf2 in human bronchial epithelial (hBE) cells, and that CFTR–NRF2 interaction decreases in CF hBE cells [49]. VX809 and VX661, the approved correctors for CFTR-F508del, restores Nrf2 phosphorylation and its interaction with CBP [49]. Excessive inflammation triggered by the activation of NF- κ B by oxidative stress is highly related with the morbidity and mortality of CF [128]. Roles of CBP–Nrf2–NF- κ B network in CF have been revealed in airway epithelial cells both in vitro and in vivo. Dramatic decrease

of Nrf2 activity is detected in CF cell lines as well as lungs and excised nasal epithelia of CF mice [129]. The defective Nrf2 in CF cells is associated with cAMP signaling, and inhibition of cAMP signaling can interfere in CBP interactions with Nrf2 and NF- κ B, resulting in enhanced Nrf2 activity and reduced NF- κ B activity in CF cells [130]. Combined, these studies implicate that reducing inflammation in CF by strengthening CBP–Nrf2 or weakening CBP–NF- κ B interactions might be potential therapies for CF.

3.6 Respiratory Infections

There are many different viral and bacterial infections that can occur in airways. The most common pathogens are viruses, including respiratory syncytial virus (RSV), influenza A virus (IAV), adenovirus, coronavirus, etc. The less common but very present bacterial pathogens include *Streptococcus pneumoniae*, *Haemophilus influenzae*, and *Mycobacterium tuberculosis* (TB). Both viral and bacterial infections occur in the upper and lower respiratory tract as well as the esophagus. Infection-induced symptoms such as coughing and mucus build up can cause damage to the epithelial tissue and create ROS.

RSV targets the epithelial airway tissues and has been found to activate deacetylation and proteasomal degradation of Nrf2 [131]. *Nrf2*^{-/-} mice that are infected with RSV show more severe lung inflammation and injury as well as attenuated viral clearance than wild-type mice [132]. IAV mainly targets the epithelial tissue and specifically alveolar type I and II epithelial cells. An in vitro study shows that infection of these cells by IAV increases NRF2/HO-1 level. NRF2 knockdown causes these cells to be more sensitive to IAV-induced injury, while overexpression of NRF2 shows a protective effect against IAV [133]. It was found that a NRF2 knockdown is correlated with an increase in viral entry and replication while SFN-induced activation of NRF2 in human nasal epithelial cells significantly decreases IAV entry and replication, directly supporting a protective role of Nrf2 against IAV infection [134]. The effectiveness of entry and replication of a virus can be altered with the cleavage of viral HA surface proteins, which allows the virus to enter the targeted host cell. It has been shown that an increase in NRF2 expression correlates with a lowered expression of the viral HA gene [134]. Several Nrf2 activators have shown protective effect against RSV and IAV. For example, SFN or emodin treatment significantly reduces virus-induced lung inflammation and viral replication [132, 135].

Streptococcus pneumoniae is the most common cause of pneumonia and sepsis. Nrf2 knockout has an increase of proinflammatory mediators and neutrophils present in lung digests from mice treated with *Streptococcus* [136]. Activation of NRF2 by resveratrol significantly reduced oxidative stress in human lung epithelial cells infected by *Streptococcus* [137]. Another common bacterium that infects the lungs is *Tuberculosis* (TB). The Nrf2 activator NAC has been shown

to decrease the bacteria counts and lung injury in guinea pig model infected by TB, suggesting that NAC could be used as an optional aid along with treatment [138].

3.7 Lung Cancers

The double-sided sword roles of Nrf2 in carcinogenesis has been revealed in many studies and discussed in several reviews [10, 139]. On the one hand, Nrf2-deficient mice treated with different carcinogens show increased severities of inflammation-associated carcinogenesis such as the AOM/DSS-induced colonic tumor [140]. On the other hand, enhanced levels of NRF2 or mutations that are predicted to activate NRF2 are found in many human cancers especially lung cancers [10, 141–144]. In support of the promotive role of Nrf2 in lung cancer, targeted deletion of Nrf2 reduces urethane-induced lung tumorigenesis in mice [145]. NRF2 depletion increases chemosensitivity of human lung cancer A549 cells both in vitro and in vivo [146]. Lung tumorigenesis induced by constitutively active K-Ras^{G12D} is suppressed in Nrf2-deficient mice [147]. It is proposed that Nrf2 facilitates cancer cell resistance to chemotherapeutic drugs and radiation through the activation of the antioxidant pathway. Enhanced NRF2-HO-1 pathway is increasingly seen in lung adenocarcinoma [148]. In addition, Nrf2 can promote cancer cell proliferation through activating proliferating and metabolic genes [139]. In human lung cancer cell lines, NRF2 activates transcriptions of anti-apoptotic factor Bcl-2 and glucose metabolic enzymes [25, 26], which to some extent accounts for the role of NRF2 in promoting lung cancers.

Recent studies have revealed that the Nrf2-interacting networks also contribute to the oncogenic role of Nrf2 in lung cancers. For example, single nucleotide polymorphisms (SNPs) in MAF-G have been found to be associated with lung cancers or airway transcription response to CS [149]. Enhanced levels of both MOF and NRF2-target transcriptions were revealed in human non-small cell lung cancer [75]. Co-upregulation of both DJ-1 and NRF2 were observed in a large set of lung carcinomas [92]. Depletion of KEAP1 in human non-small cell lung carcinomas cell lines increases their sensitivity to chemotherapeutic agents, which is likely mediated by enhanced PPAR γ levels and its activation of differentiation genes [150]. Novel mechanisms of the Nrf2-Bach1 pathway in lung cancer metastasis has been recently identified [87, 88]. In normal cells, free heme inhibit Bach1 activity through Fbxo22-mediated ubiquitination. In cells with enhanced Nrf2, activation of HO-1 decreases the cellular level of heme thus stabilizing Bach1. Enhanced Bach1 promotes lung cancer metastasis through the activation of metastatic genes and glycolysis genes. Another protein that cooperates with Nrf2 in the promotion of lung cancer is ATF4. Nrf2-ATF4 dimer can directly target and activate *HO-1* gene [50]. Cooperative activations of *HO-1* by Nrf2 and ATF4 promotes fibrosarcoma lung colonization [151]. Nrf2 and ATF4 can also co-activate biosynthetic enzyme genes involved in serine/glycine metabolism, which plays a promotive role in non-small cell lung cancer [152].

4 Lung Disease Therapies Targeting Nrf2-Interacting Network

Given the protective role of Nrf2 in airway inflammation, injury and pathogenesis, designing drugs that target Nrf2 and its essential regulator Keap1 is valuable for lung disease therapies. Many nature products and pharmaceutical compounds have been identified as activators or inhibitors of Nrf2 and Keap1 [153]. Protective roles of these drugs against lung inflammation, injuries, and diseases have been verified in mouse models, and some drugs are in clinical trials. For example, SFN (an isothiocyanate isolated from cruciferous plants) treatment enhances bacteria removal and phagocytosis in the alveolar macrophages from both mice and COPD patients, supporting the therapeutic effect of SFN in treating COPD [154, 155]. A drug screen in human lung fibroblasts identified a group of synthetic small molecules (HPP-4382) that induced Nrf2-dependent HO-1 expression [156]. Among these compounds, HPP971 has completed phase 1 studies and shown potential therapeutic benefits in diseases, including respiratory injuries (vTv Therapeutics).

The roles of Nrf2 in respiratory inflammation and diseases can be mediated by many other proteins that interact with Nrf2 (Fig. 1). Therefore, based on knowledge of the molecular functions of the Nrf2 network in pathogenesis, drugs targeting the Nrf2-interacting networks can increase the efficiency and selectivity of therapies. Scientists are screening for compounds that directly target Nrf2-associated protein complexes especially the Keap1-Nrf2 complex. In addition, compounds that simultaneously target Nrf2 and its interacting partners have shown therapeutic potency for lung diseases.

4.1 Compounds Targeting Nrf2–Keap1 Interaction

Several groups have successfully identified compounds that directly interfere with the Nrf2–Keap1 interaction. These include ML334 by Hu and colleagues and mono-acidic compounds by GlaxoSmithKline Pharmaceuticals [157, 158]. These compounds have been shown to induce the expression of NRF2-regulated genes in bronchial epithelial cells derived from COPD patients and reverse ozone-induced lung inflammation in rats [158].

4.2 Compounds Co-Targeting Nrf2 and NF- κ B

NF- κ B is an essential factor and therapy target for airway inflammation and chronic lung diseases [159]. Natural products that have both antioxidant and anti-inflammatory effects were seen in recent studies as potential therapies for

inflammation-related lung injuries and diseases. For example, tyrosol (a natural phenolic antioxidant) treatment of LPS-induced ALI mice significantly inhibited NF- κ B and AP-1 and reduced inflammatory cytokines, while at the same time activating Nrf2 and HO-1 [160]. Oridonin (isolated from *Rabdosia rubescens*) and bardoxolone (a synthetic triterpenoid based on oleanolic acid) inhibit the NF- κ B pathway and activate the Nrf2 pathway in LPS-induced ALI mice [161, 162], supporting the antioxidant and anti-inflammatory effects of these compounds for ALI protection and therapy. Investigations of several compounds targeting the Nrf2-NF- κ B network in both mouse and rat models have shown their potential therapeutic effects against CS-induced inflammation, emphysema, and COPD. These include natural products such as forsythiaside (a hydroxycinnamic acid from *Forsythia suspensa*) [114], eucalyptol (a monoterpene oil isolated from *Eucalyptus*) [115], platycodin D (a saponin from the root of *Platycodon grandiflorus*) [116], and isoliquiritigenin (a flavonoid from *Glycyrrhizae* species) [117], as well as pharmacological compounds such as Sul-121 [118] and 15d-PGJ2 [119]. All these drugs can reduce respiratory inflammation and CS-induced emphysema through both the activation of the Nrf2 pathway and the down-regulation of the NF- κ B pathway. In addition, rosavin (found in *Rhodiola rosea*) and emodin (isolated from rhubarb *Rheum palmatum*), nature products that activate Nrf2 and inhibit NF- κ B, have recently been proposed as treatments for pulmonary fibrosis [125, 126]. Dimethyl fumarate (DMF) was approved by the FDA for the treatment of multiple sclerosis. It was found that DMF can both activate Nrf2 and inhibit the NF- κ B pathway and prevent the development of bleomycin-induced lung fibrosis in mice [163]. Notably, DMF can control Nrf2 activity in a dose-dependent manner. In several cancer cells, low concentrations of DMF activate Nrf2, while high concentrations of DMF reduce the nuclear Nrf2 level, likely through the down-regulation of the Nrf2 stabilizer DJ-1 [164].

4.3 Compounds Targeting Nrf2 Phosphorylation Pathways

Desoxyrhapontigenin (isolated from rhubarb plants), a compound that shows significant anti-inflammatory and antioxidant effects, can inhibit MAPK, Akt, and NF- κ B and activate Nrf2 [103, 165]. It was found that desoxyrhapontigenin pretreatment reduces LPS-induced lung inflammation and injury in mice through the PI3K/Akt-dependent activation of the Nrf2-HO-1 pathway [103]. Metformin reduces the risk of lung cancer during anti-diabetic treatment [166]. A recent study showed that metformin suppresses proliferation and induces apoptosis of the human lung carcinoma A549 cells through the inhibition of the Akt and ERK1/2 signaling pathways and activation of Nrf2 [167]. These studies indicate the potency of Nrf2 phosphorylation pathways as therapeutic targets for lung diseases.

4.4 Other Compounds Targeting Nrf2 Network

Interacting with the Keap1-Nrf2 pathway, autophagy can play both positive and negative roles in chronic lung diseases, CS exposure, and ALI [102]. It was found that vitamin D can protect against PM-induced lung injury in mice through the degradation of p62 and Keap1 and the activation of the Nrf2 pathway [168]. Pirfenidone (PFD), an approved drug for IPF treatment, can suppress bleomycin-induced lung fibrosis through both the activation of the Nrf2 pathway and the inhibition of Bach1 [127, 169].

5 Conclusions

As a master antioxidant factor, the protective roles of Nrf2 against respiratory inflammation and pathogenesis, as well as the brilliant potency of Nrf2-targeting therapies for lung diseases, have been verified and supported by numerous studies. However, with the revealing of more and more Nrf2-interacting proteins/pathways and Nrf2-targeting genes/functions, the complexity of the Nrf2 network in physiology and pathology needs to be considered when designing therapies targeting Nrf2. On the bright side, selectively targeting Nrf2-related protein interactions or regulatory networks could increase the efficiency and specificity of therapies. On the negative side, off-target effects that are related to the multiple interacting partners and biological functions of Nrf2 need to be considered when modifying Nrf2 level or activity in different types of cells. Many recent studies have focused on the potential application of natural antioxidant-rich foods that have limited side effects. Although the protective effects of many of the natural products against lung inflammation and pathogenesis have been verified in animal models, their therapeutic efficiency for lung diseases as well as long-term biological effects in humans remain to be evaluated.

Given that activation of the Nrf2 pathway is a strategy for treating most of the respiratory diseases, we should be mindful of the tumorigenic risk of Nrf2 activators. Chemoprevention effects of some Nrf2 inducers, such as SFN and CDDO-Im, have been studied in both animal models and clinical trials [170, 171]. It is necessary to determine the safe thresholds of Nrf2 level in specific cells as well as the safe doses of Nrf2 activators that will not induce oncogenesis. It would also be useful to design therapies that differentially target the oncogenic role and protective role of Nrf2, which may be achieved by targeting specific Nrf2-related protein interactions in the future. Doubtlessly, revealing the complete range of molecular mechanisms and biological functions of the Nrf2 network will contribute to the development of Nrf2-targeting therapies.

Competing Interest Statement The authors declare that they have no competing interest.

References

1. Mortality, G. B. D. & Causes of Death, C. Global, regional, and national life expectancy, all-cause mortality, and cause-specific mortality for 249 causes of death, 1980-2015: a systematic analysis for the global burden of disease study 2015. *Lancet*. 2016;388:1459–544. [https://doi.org/10.1016/S0140-6736\(16\)31012-1](https://doi.org/10.1016/S0140-6736(16)31012-1).
2. Kensler TW, Wakabayashi N, Biswal S. Cell survival responses to environmental stresses via the Keap1-Nrf2-ARE pathway. *Annu Rev Pharmacol Toxicol*. 2007;47:89–116. <https://doi.org/10.1146/annurev.pharmtox.46.120604.141046>.
3. Zhang DD. Mechanistic studies of the Nrf2-Keap1 signaling pathway. *Drug Metab Rev*. 2006;38:769–89. <https://doi.org/10.1080/03602530600971974>.
4. Malhotra D, et al. Global mapping of binding sites for Nrf2 identifies novel targets in cell survival response through ChIP-Seq profiling and network analysis. *Nucleic Acids Res*. 2010;38:5718–34. <https://doi.org/10.1093/nar/gkq212>.
5. Sykietis GP, Bohmann D. Stress-activated cap'n'collar transcription factors in aging and human disease. *Sci Signal*. 2010;3:re3. <https://doi.org/10.1126/scisignal.3112re3>.
6. Kobayashi A, et al. Oxidative stress sensor Keap1 functions as an adaptor for Cul3-based E3 ligase to regulate proteasomal degradation of Nrf2. *Mol Cell Biol*. 2004;24:7130–9. <https://doi.org/10.1128/MCB.24.16.7130-7139.2004>.
7. Eggler AL, Liu G, Pezzuto JM, van Breemen RB, Mesecar AD. Modifying specific cysteine's of the electrophile-sensing human Keap1 protein is insufficient to disrupt binding to the Nrf2 domain Neh2. *Proc Natl Acad Sci U S A*. 2005;102:10070–5. <https://doi.org/10.1073/pnas.0502402102>.
8. Kobayashi M, et al. The antioxidant defense system Keap1-Nrf2 comprises a multiple sensing mechanism for responding to a wide range of chemical compounds. *Mol Cell Biol*. 2009;29:493–502. <https://doi.org/10.1128/MCB.01080-08>.
9. Itoh K, et al. Keap1 represses nuclear activation of antioxidant responsive elements by Nrf2 through binding to the amino-terminal Neh2 domain. *Genes Dev*. 1999;13:76–86. <https://doi.org/10.1101/gad.13.1.76>.
10. Taguchi K, Motohashi H, Yamamoto M. Molecular mechanisms of the Keap1-Nrf2 pathway in stress response and cancer evolution. *Genes Cells*. 2011;16:123–40. <https://doi.org/10.1111/j.1365-2443.2010.01473.x>.
11. Sun Z, et al. KPNA6 (Importin {alpha}7)-mediated nuclear import of Keap1 represses the Nrf2-dependent antioxidant response. *Mol Cell Biol*. 2011;31:1800–11. <https://doi.org/10.1128/MCB.05036-11>.
12. Wakabayashi N, et al. Keap1-null mutation leads to postnatal lethality due to constitutive Nrf2 activation. *Nat Genet*. 2003;35:238–45. <https://doi.org/10.1038/ng1248>.
13. Thimmulappa RK, et al. Identification of Nrf2-regulated genes induced by the chemo preventive agent sulforaphane by oligonucleotide microarray. *Cancer Res*. 2002;62:5196–203.
14. Kwak MK, et al. Modulation of gene expression by cancer chemopreventive dithiolethiones through the Keap1-Nrf2 pathway. Identification of novel gene clusters for cell survival. *J Biol Chem*. 2003;278:8135–45. <https://doi.org/10.1074/jbc.M211898200>.
15. Hu R, et al. Identification of Nrf2-regulated genes induced by chemopreventive isothiocyanate PEITC by oligonucleotide microarray. *Life Sci*. 2006;79:1944–55. <https://doi.org/10.1016/j.lfs.2006.06.019>.
16. Asikainen TM, et al. Increased sensitivity of homozygous Sod2 mutant mice to oxygen toxicity. *Free Radic Biol Med*. 2002;32:175–86. [https://doi.org/10.1016/s0891-5849\(01\)00776-6](https://doi.org/10.1016/s0891-5849(01)00776-6).
17. Folz RJ, Abushamaa AM, Suliman HB. Extracellular superoxide dismutase in the airways of transgenic mice reduces inflammation and attenuates lung toxicity following hyperoxia. *J Clin Invest*. 1999;103:1055–66. <https://doi.org/10.1172/JCI3816>.
18. Pi J, et al. Deficiency in the nuclear factor E2-related factor-2 transcription factor results in impaired adipogenesis and protects against diet-induced obesity. *J Biol Chem*. 2010;285:9292–300. <https://doi.org/10.1074/jbc.M109.093955>.

19. Chorley BN, et al. Identification of novel NRF2-regulated genes by ChIP-Seq: influence on retinoid X receptor alpha. *Nucleic Acids Res.* 2012;40:7416–29. <https://doi.org/10.1093/nar/gks409>.
20. Huang J, Tabbi-Anneni I, Gunda V, Wang L. Transcription factor Nrf2 regulates SHP and lipogenic gene expression in hepatic lipid metabolism. *Am J Physiol Gastrointest Liver Physiol.* 2010;299:G1211–21. <https://doi.org/10.1152/ajpgi.00322.2010>.
21. Kitteringham NR, et al. Proteomic analysis of Nrf2 deficient transgenic mice reveals cellular defence and lipid metabolism as primary Nrf2-dependent pathways in the liver. *J Proteome.* 2010;73:1612–31. <https://doi.org/10.1016/j.jprot.2010.03.018>.
22. Bobilev I, et al. The Nrf2 transcription factor is a positive regulator of myeloid differentiation of acute myeloid leukemia cells. *Cancer Biol Ther.* 2011;11:317–29. <https://doi.org/10.4161/cbt.11.3.14098>.
23. Khacho M, et al. Mitochondrial dynamics impacts stem cell identity and fate decisions by regulating a nuclear transcriptional program. *Cell Stem Cell.* 2016;19:232–47. <https://doi.org/10.1016/j.stem.2016.04.015>.
24. Jain A, et al. p62/SQSTM1 is a target gene for transcription factor NRF2 and creates a positive feedback loop by inducing antioxidant response element-driven gene transcription. *J Biol Chem.* 2010;285:22576–91. <https://doi.org/10.1074/jbc.M110.118976>.
25. Nitire SK, Jaiswal AK. Nrf2 protein up-regulates antiapoptotic protein Bcl-2 and prevents cellular apoptosis. *J Biol Chem.* 2012;287:9873–86. <https://doi.org/10.1074/jbc.M111.312694>.
26. Mitsuishi Y, et al. Nrf2 redirects glucose and glutamine into anabolic pathways in metabolic reprogramming. *Cancer Cell.* 2012;22:66–79. <https://doi.org/10.1016/j.ccr.2012.05.016>.
27. Kobayashi EH, et al. Nrf2 suppresses macrophage inflammatory response by blocking pro-inflammatory cytokine transcription. *Nat Commun.* 2016;7:11624. <https://doi.org/10.1038/ncomms11624>.
28. Sykiotis GP, Bohmann D. Keap1/Nrf2 signaling regulates oxidative stress tolerance and lifespan in *Drosophila*. *Dev Cell.* 2008;14:76–85. <https://doi.org/10.1016/j.devcel.2007.12.002>.
29. Deng H, Kerppola TK. Regulation of *Drosophila* metamorphosis by xenobiotic response regulators. *PLoS Genet.* 2013;9:e1003263. <https://doi.org/10.1371/journal.pgen.1003263>.
30. Hochmuth CE, Biteau B, Bohmann D, Jasper H. Redox regulation by Keap1 and Nrf2 controls intestinal stem cell proliferation in *Drosophila*. *Cell Stem Cell.* 2011;8:188–99. <https://doi.org/10.1016/j.stem.2010.12.006>.
31. Slocum SL, Kensler TW. Nrf2: control of sensitivity to carcinogens. *Arch Toxicol.* 2011;85:273–84. <https://doi.org/10.1007/s00204-011-0675-4>.
32. Deng H. Multiple roles of Nrf2-Keap1 signaling: regulation of development and xenobiotic response using distinct mechanisms. *Fly (Austin).* 2014;8:7–12. <https://doi.org/10.4161/fly.27007>.
33. Deng H, Kerppola TK. Visualization of the *Drosophila* dKeap1-CncC interaction on chromatin illuminates cooperative, xenobiotic-specific gene activation. *Development.* 2014;141:3277–88. <https://doi.org/10.1242/dev.110528>.
34. Rahman I, Adcock IM. Oxidative stress and redox regulation of lung inflammation in COPD. *Eur Respir J.* 2006;28:219–42. <https://doi.org/10.1183/09031936.06.00053805>.
35. Schuliga M. NF-kappaB signaling in chronic inflammatory airway disease. *Biomol Ther.* 2015;5:1266–83. <https://doi.org/10.3390/biom5031266>.
36. Birnhuber A, Biasin V, Schnoegl D, Marsh LM, Kwapiszewska G. Transcription factor Fra-2 and its emerging role in matrix deposition, proliferation and inflammation in chronic lung diseases. *Cell Signal.* 2019;64:109408. <https://doi.org/10.1016/j.cellsig.2019.109408>.
37. Cattani-Cavaliere I, et al. Acute exposure to diesel-biodiesel particulate matter promotes murine lung oxidative stress by Nrf2/HO-1 and inflammation through the NF-kB/TNF-alpha pathways. *Inflammation.* 2019;42:526–37. <https://doi.org/10.1007/s10753-018-0910-8>.
38. Leclercq B, et al. Air pollution-derived PM2.5 impairs mitochondrial function in healthy and chronic obstructive pulmonary diseased human bronchial epithelial cells. *Environ Pollut.* 2018;243:1434–49. <https://doi.org/10.1016/j.envpol.2018.09.062>.

39. Liu GH, Qu J, Shen X. NF-kappaB/p65 antagonizes Nrf2-ARE pathway by depriving CBP from Nrf2 and facilitating recruitment of HDAC3 to MafK. *Biochim Biophys Acta*. 2008;1783:713–27. <https://doi.org/10.1016/j.bbamcr.2008.01.002>.
40. Venugopal R, Jaiswal AK. Nrf1 and Nrf2 positively and c-Fos and Fra1 negatively regulate the human antioxidant response element-mediated expression of NAD(P)H:quinone oxidoreductase1 gene. *Proc Natl Acad Sci U S A*. 1996;93:14960–5. <https://doi.org/10.1073/pnas.93.25.14960>.
41. Venugopal R, Jaiswal AK. Nrf2 and Nrf1 in association with Jun proteins regulate antioxidant response element-mediated expression and coordinated induction of genes encoding detoxifying enzymes. *Oncogene*. 1998;17:3145–56. <https://doi.org/10.1038/sj.onc.1202237>.
42. Katsuoka F, Yamamoto M. Small Maf proteins (MafF, MafG, MafK): history, structure and function. *Gene*. 2016;586:197–205. <https://doi.org/10.1016/j.gene.2016.03.058>.
43. Katsuoka F, et al. Genetic evidence that small maf proteins are essential for the activation of antioxidant response element-dependent genes. *Mol Cell Biol*. 2005;25:8044–51. <https://doi.org/10.1128/MCB.25.18.8044-8051.2005>.
44. Motohashi H, Katsuoka F, Engel JD, Yamamoto M. Small Maf proteins serve as transcriptional cofactors for keratinocyte differentiation in the Keap1-Nrf2 regulatory pathway. *Proc Natl Acad Sci U S A*. 2004;101:6379–84. <https://doi.org/10.1073/pnas.0305902101>.
45. Hirotsu Y, et al. Nrf2-MafG heterodimers contribute globally to antioxidant and metabolic networks. *Nucleic Acids Res*. 2012;40:10228–39. <https://doi.org/10.1093/nar/gks827>.
46. Sekine H, et al. The mediator subunit MED16 transduces NRF2-activating signals into antioxidant gene expression. *Mol Cell Biol*. 2016;36:407–20. <https://doi.org/10.1128/MCB.00785-15>.
47. Katoh Y, et al. Two domains of Nrf2 cooperatively bind CBP, a CREB binding protein, and synergistically activate transcription. *Genes Cells*. 2001;6:857–68.
48. Sun Z, Chin YE, Zhang DD. Acetylation of Nrf2 by p300/CBP augments promoter-specific DNA binding of Nrf2 during the antioxidant response. *Mol Cell Biol*. 2009;29:2658–72. <https://doi.org/10.1128/MCB.01639-08>.
49. Borcherdig DC, et al. Clinically-approved CFTR modulators rescue Nrf2 dysfunction in cystic fibrosis airway epithelia. *J Clin Invest*. 2019;129:3448–63. <https://doi.org/10.1172/JCI96273>.
50. He CH, et al. Identification of activating transcription factor 4 (ATF4) as an Nrf2-interacting protein. Implication for heme oxygenase-1 gene regulation. *J Biol Chem*. 2001;276:20858–65. <https://doi.org/10.1074/jbc.M101198200>.
51. Brown SL, Sekhar KR, Rachakonda G, Sasi S, Freeman ML. Activating transcription factor 3 is a novel repressor of the nuclear factor erythroid-derived 2-related factor 2 (Nrf2)-regulated stress pathway. *Cancer Res*. 2008;68:364–8. <https://doi.org/10.1158/0008-5472.CAN-07-2170>.
52. Shan Y, et al. ATF3 protects pulmonary resident cells from acute and ventilator-induced lung injury by preventing Nrf2 degradation. *Antioxid Redox Signal*. 2015;22:651–68. <https://doi.org/10.1089/ars.2014.5987>.
53. Ikeda Y, et al. Suppression of rat thromboxane synthase gene transcription by peroxisome proliferator-activated receptor gamma in macrophages via an interaction with NRF2. *J Biol Chem*. 2000;275:33142–50. <https://doi.org/10.1074/jbc.M002319200>.
54. Cho HY, Reddy SP, Debiase A, Yamamoto M, Kleiberger SR. Gene expression profiling of NRF2-mediated protection against oxidative injury. *Free Radic Biol Med*. 2005;38:325–43. <https://doi.org/10.1016/j.freeradbiomed.2004.10.013>.
55. Cho HY, et al. Nrf2-regulated PPAR{gamma} expression is critical to protection against acute lung injury in mice. *Am J Respir Crit Care Med*. 2010;182:170–82. <https://doi.org/10.1164/rccm.200907-1047OC>.
56. Ansell PJ, et al. Repression of cancer protective genes by 17beta-estradiol: ligand-dependent interaction between human Nrf2 and estrogen receptor alpha. *Mol Cell Endocrinol*. 2005;243:27–34. <https://doi.org/10.1016/j.mce.2005.08.002>.

57. Wang XJ, Hayes JD, Henderson CJ, Wolf CR. Identification of retinoic acid as an inhibitor of transcription factor Nrf2 through activation of retinoic acid receptor alpha. *Proc Natl Acad Sci U S A*. 2007;104:19589–94. <https://doi.org/10.1073/pnas.0709483104>.
58. Li Y, Wang H, Wang XJ, Tang X. The short isoform of PML-RARalpha activates the NRF2/HO-1 pathway through a direct interaction with NRF2. *FEBS Lett*. 2017;591:2859–68. <https://doi.org/10.1002/1873-3468.12779>.
59. Johansen T, Lamark T. Selective autophagy mediated by autophagic adapter proteins. *Autophagy*. 2011;7:279–96. <https://doi.org/10.4161/autophagy.7.3.14487>.
60. Komatsu M, et al. The selective autophagy substrate p62 activates the stress responsive transcription factor Nrf2 through inactivation of Keap1. *Nat Cell Biol*. 2010;12:213–23. <https://doi.org/10.1038/ncb2021>.
61. Ichimura Y, et al. Phosphorylation of p62 activates the Keap1-Nrf2 pathway during selective autophagy. *Mol Cell*. 2013;51:618–31. <https://doi.org/10.1016/j.molcel.2013.08.003>.
62. Lau A, et al. Arsenic inhibits autophagic flux, activating the Nrf2-Keap1 pathway in a p62-dependent manner. *Mol Cell Biol*. 2013;33:2436–46. <https://doi.org/10.1128/MCB.01748-12>.
63. Taguchi K, et al. Keap1 degradation by autophagy for the maintenance of redox homeostasis. *Proc Natl Acad Sci U S A*. 2012;109:13561–6. <https://doi.org/10.1073/pnas.1121572109>.
64. Xu Y, et al. *Pseudomonas aeruginosa* pyocyanin activates NRF2-ARE-mediated transcriptional response via the ROS-EGFR-PI3K-AKT/MEK-ERK MAP kinase signaling in pulmonary epithelial cells. *PLoS One*. 2013;8:e72528. <https://doi.org/10.1371/journal.pone.0072528>.
65. Huang HC, Nguyen T, Pickett CB. Phosphorylation of Nrf2 at Ser-40 by protein kinase C regulates antioxidant response element-mediated transcription. *J Biol Chem*. 2002;277:42769–74. <https://doi.org/10.1074/jbc.M206911200>.
66. Sun Z, Huang Z, Zhang DD. Phosphorylation of Nrf2 at multiple sites by MAP kinases has a limited contribution in modulating the Nrf2-dependent antioxidant response. *PLoS One*. 2009;4:e6588. <https://doi.org/10.1371/journal.pone.0006588>.
67. Zipper LM, Mulcahy RT. Erk activation is required for Nrf2 nuclear localization during pyrrolidine dithiocarbamate induction of glutamate cysteine ligase modulatory gene expression in HepG2 cells. *Toxicol Sci*. 2003;73:124–34. <https://doi.org/10.1093/toxsci/kfg083>.
68. Keum YS, et al. Mechanism of action of sulfuraphane: inhibition of p38 mitogen-activated protein kinase isoforms contributing to the induction of antioxidant response element-mediated heme oxygenase-1 in human hepatoma HepG2 cells. *Cancer Res*. 2006;66:8804–13. <https://doi.org/10.1158/0008-5472.CAN-05-3513>.
69. Naidu S, Vijayan V, Santoso S, Kietzmann T, Immenschuh S. Inhibition and genetic deficiency of p38 MAPK up-regulates heme oxygenase-1 gene expression via Nrf2. *J Immunol*. 2009;182:7048–57. <https://doi.org/10.4049/jimmunol.0900006>.
70. Inoue H, et al. The *C. elegans* p38 MAPK pathway regulates nuclear localization of the transcription factor SKN-1 in oxidative stress response. *Genes Dev*. 2005;19:2278–83. <https://doi.org/10.1101/gad.1324805>.
71. Tullet JM, et al. Direct inhibition of the longevity-promoting factor SKN-1 by insulin-like signaling in *C. elegans*. *Cell*. 2008;132:1025–38. <https://doi.org/10.1016/j.cell.2008.01.030>.
72. Rewitz KF, Yamanaka N, Gilbert LI, O'Connor MB. The insect neuropeptide PTTH activates receptor tyrosine kinase torso to initiate metamorphosis. *Science*. 2009;326:1403–5. <https://doi.org/10.1126/science.1176450>.
73. Chen D, et al. NRF2 is a major target of ARF in p53-independent tumor suppression. *Mol Cell*. 2017;68:224–232 e224. <https://doi.org/10.1016/j.molcel.2017.09.009>.
74. Hayes JD, Dinkova-Kostova AT. Epigenetic control of NRF2-directed cellular antioxidant status in dictating life-death decisions. *Mol Cell*. 2017;68:5–7. <https://doi.org/10.1016/j.molcel.2017.09.023>.
75. Chen Z, et al. The histone acetyltransferase hMOF acetylates Nrf2 and regulates anti-drug responses in human non-small cell lung cancer. *Br J Pharmacol*. 2014;171:3196–211. <https://doi.org/10.1111/bph.12661>.

76. Kawai Y, Garduno L, Theodore M, Yang J, Arinze IJ. Acetylation-deacetylation of the transcription factor Nrf2 (nuclear factor erythroid 2-related factor 2) regulates its transcriptional activity and nucleocytoplasmic localization. *J Biol Chem.* 2011;286:7629–40. <https://doi.org/10.1074/jbc.M110.208173>.
77. Ogrzyzko VV, Schiltz RL, Russanova V, Howard BH, Nakatani Y. The transcriptional coactivators p300 and CBP are histone acetyltransferases. *Cell.* 1996;87:953–9. [https://doi.org/10.1016/s0092-8674\(00\)82001-2](https://doi.org/10.1016/s0092-8674(00)82001-2).
78. Zhang J, et al. BRG1 interacts with Nrf2 to selectively mediate HO-1 induction in response to oxidative stress. *Mol Cell Biol.* 2006;26:7942–52. <https://doi.org/10.1128/MCB.00700-06>.
79. Ge M, et al. Brg1-mediated Nrf2/HO-1 pathway activation alleviates hepatic ischemia-reperfusion injury. *Cell Death Dis.* 2017;8:e2841. <https://doi.org/10.1038/cddis.2017.236>.
80. Nioi P, Nguyen T, Sherratt PJ, Pickett CB. The carboxy-terminal Neh3 domain of Nrf2 is required for transcriptional activation. *Mol Cell Biol.* 2005;25:10895–906. <https://doi.org/10.1128/MCB.25.24.10895-10906.2005>.
81. Ki SH, Cho IJ, Choi DW, Kim SG. Glucocorticoid receptor (GR)-associated SMRT binding to C/EBPbeta TAD and Nrf2 Neh4/5: role of SMRT recruited to GR in GSTA2 gene repression. *Mol Cell Biol.* 2005;25:4150–65. <https://doi.org/10.1128/MCB.25.10.4150-4165.2005>.
82. Carlson J, Swisse T, Smith C, Deng H. Regulation of position effect variegation at pericentric heterochromatin by Drosophila Keap1-Nrf2 xenobiotic response factors. *Genesis.* 2019;57:e23290. <https://doi.org/10.1002/dvg.23290>.
83. Dhakshinamoorthy S, Jain AK, Bloom DA, Jaiswal AK. Bach1 competes with Nrf2 leading to negative regulation of the antioxidant response element (ARE)-mediated NAD(P)H:quinone oxidoreductase 1 gene expression and induction in response to antioxidants. *J Biol Chem.* 2005;280:16891–900. <https://doi.org/10.1074/jbc.M500166200>.
84. Reichard JF, Motz GT, Puga A. Heme oxygenase-1 induction by NRF2 requires inactivation of the transcriptional repressor BACH1. *Nucleic Acids Res.* 2007;35:7074–86. <https://doi.org/10.1093/nar/gkm638>.
85. Liang Y, et al. Transcriptional network analysis identifies BACH1 as a master regulator of breast cancer bone metastasis. *J Biol Chem.* 2012;287:33533–44. <https://doi.org/10.1074/jbc.M112.392332>.
86. Lee U, et al. A prognostic gene signature for metastasis-free survival of triple negative breast cancer patients. *PLoS One.* 2013;8:e82125. <https://doi.org/10.1371/journal.pone.0082125>.
87. Lignitto L, et al. Nrf2 activation promotes lung cancer metastasis by inhibiting the degradation of bach1. *Cell.* 2019;178:316–329 e318. <https://doi.org/10.1016/j.cell.2019.06.003>.
88. Wiel C, et al. BACH1 stabilization by antioxidants stimulates lung cancer metastasis. *Cell.* 2019;178:330–345 e322. <https://doi.org/10.1016/j.cell.2019.06.005>.
89. Chen W, et al. Direct interaction between Nrf2 and p21(Cip1/WAF1) upregulates the Nrf2-mediated antioxidant response. *Mol Cell.* 2009;34:663–73. <https://doi.org/10.1016/j.molcel.2009.04.029>.
90. Chen W, et al. Does Nrf2 contribute to p53-mediated control of cell survival and death? *Antioxid Redox Signal.* 2012;17:1670–5. <https://doi.org/10.1089/ars.2012.4674>.
91. Clements CM, McNally RS, Conti BJ, Mak TW, Ting JP. DJ-1, a cancer- and Parkinson's disease-associated protein, stabilizes the antioxidant transcriptional master regulator Nrf2. *Proc Natl Acad Sci U S A.* 2006;103:15091–6. <https://doi.org/10.1073/pnas.0607260103>.
92. Merikallio H, Paakko P, Kinnula VL, Harju T, Soini Y. Nuclear factor erythroid-derived 2-like 2 (Nrf2) and DJ1 are prognostic factors in lung cancer. *Hum Pathol.* 2012;43:577–84. <https://doi.org/10.1016/j.humpath.2011.05.024>.
93. Lo SC, Hannink M. PGAM5, a Bcl-XL-interacting protein, is a novel substrate for the redox-regulated Keap1-dependent ubiquitin ligase complex. *J Biol Chem.* 2006;281:37893–903. <https://doi.org/10.1074/jbc.M606539200>.
94. Lo SC, Hannink M. PGAM5 tethers a ternary complex containing Keap1 and Nrf2 to mitochondria. *Exp Cell Res.* 2008;314:1789–803. <https://doi.org/10.1016/j.yexcr.2008.02.014>.

95. Mittal M, Siddiqui MR, Tran K, Reddy SP, Malik AB. Reactive oxygen species in inflammation and tissue injury. *Antioxid Redox Signal*. 2014;20:1126–67. <https://doi.org/10.1089/ars.2012.5149>.
96. Danel C, et al. Gene therapy for oxidant injury-related diseases: adenovirus-mediated transfer of superoxide dismutase and catalase cDNAs protects against hyperoxia but not against ischemia-reperfusion lung injury. *Hum Gene Ther*. 1998;9:1487–96. <https://doi.org/10.1089/hum.1998.9.10-1487>.
97. Papaiahgari S, et al. Genetic and pharmacologic evidence links oxidative stress to ventilator-induced lung injury in mice. *Am J Respir Crit Care Med*. 2007;176:1222–35. <https://doi.org/10.1164/rccm.200701-060OC>.
98. Cho HY, et al. Sulforaphane enriched transcriptome of lung mitochondrial energy metabolism and provided pulmonary injury protection via Nrf2 in mice. *Toxicol Appl Pharmacol*. 2019;364:29–44. <https://doi.org/10.1016/j.taap.2018.12.004>.
99. Sussan TE, et al. Targeting Nrf2 with the triterpenoid CDDO-imidazolide attenuates cigarette smoke-induced emphysema and cardiac dysfunction in mice. *Proc Natl Acad Sci U S A*. 2009;106:250–5. <https://doi.org/10.1073/pnas.0804333106>.
100. Akram A, et al. Activating transcription factor 3 confers protection against ventilator-induced lung injury. *Am J Respir Crit Care Med*. 2010;182:489–500. <https://doi.org/10.1164/rccm.200906-0925OC>.
101. Ge M, et al. Overexpression of Brg1 alleviates hepatic ischemia/reperfusion-induced acute lung injury through antioxidant stress effects. *Oxidative Med Cell Longev*. 2017;2017:8787392. <https://doi.org/10.1155/2017/8787392>.
102. Rojo de la Vega M, et al. Role of Nrf2 and autophagy in acute lung injury. *Curr Pharmacol Rep*. 2016;2:91–101. <https://doi.org/10.1007/s40495-016-0053-2>.
103. Joo Choi R, Cheng MS, Shik Kim Y. Desoxyrhapontigenin up-regulates Nrf2-mediated heme oxygenase-1 expression in macrophages and inflammatory lung injury. *Redox Biol*. 2014;2:504–12. <https://doi.org/10.1016/j.redox.2014.02.001>.
104. Rangasamy T, et al. Disruption of Nrf2 enhances susceptibility to severe airway inflammation and asthma in mice. *J Exp Med*. 2005;202:47–59. <https://doi.org/10.1084/jem.20050538>.
105. Youness ER, Shady M, Nassar MS, Mostafa R, Abuelhamd W. The role of serum nuclear factor erythroid 2-related factor 2 in childhood bronchial asthma. *J Asthma*. 2019;1–6. <https://doi.org/10.1080/02770903.2019.1571081>.
106. Zeng H, et al. Polydatin attenuates reactive oxygen species-induced airway remodeling by promoting Nrf2-mediated antioxidant signaling in asthma mouse model. *Life Sci*. 2019;218:25–30. <https://doi.org/10.1016/j.lfs.2018.08.013>.
107. Mukherjee AA, Kandhare AD, Rojtkar SR, Bodhankar SL. Ameliorative effects of *Artemisia pallens* in a murine model of ovalbumin-induced allergic asthma via modulation of biochemical perturbations. *Biomed Pharmacother*. 2017;94:880–9. <https://doi.org/10.1016/j.biopha.2017.08.017>.
108. Ishii Y, et al. Transcription factor Nrf2 plays a pivotal role in protection against elastase-induced pulmonary inflammation and emphysema. *J Immunol*. 2005;175:6968–75. <https://doi.org/10.4049/jimmunol.175.10.6968>.
109. Prange R, et al. A *Drosophila* model of cigarette smoke induced COPD identifies Nrf2 signaling as an expedient target for intervention. *Aging (Albany NY)*. 2018;10:2122–35. <https://doi.org/10.18632/aging.101536>.
110. Hussain T, et al. The plant flavonoid, fisetin alleviates cigarette smoke-induced oxidative stress, and inflammation in Wistar rat lungs. *J Food Biochem*. 2019;43:e12962. <https://doi.org/10.1111/jfbc.12962>.
111. Suzuki M, et al. Down-regulated NF-E2-related factor 2 in pulmonary macrophages of aged smokers and patients with chronic obstructive pulmonary disease. *Am J Respir Cell Mol Biol*. 2008;39:673–82. <https://doi.org/10.1165/rcmb.2007-0424OC>.
112. Korytina GF, et al. Associations of the NRF2/KEAP1 pathway and antioxidant defense gene polymorphisms with chronic obstructive pulmonary disease. *Gene*. 2019;692:102–12. <https://doi.org/10.1016/j.gene.2018.12.061>.

113. Bahmed K, et al. DJ-1 modulates nuclear Erythroid 2-related Factor-2-mediated protection in human primary alveolar type II cells in smokers. *Am J Respir Cell Mol Biol*. 2016;55:439–49. <https://doi.org/10.1165/rcmb.2015-0304OC>.
114. Cheng L, Li F, Ma R, Hu X. Forsythiaside inhibits cigarette smoke-induced lung inflammation by activation of Nrf2 and inhibition of NF-kappaB. *Int Immunopharmacol*. 2015;28:494–9. <https://doi.org/10.1016/j.intimp.2015.07.011>.
115. Kennedy-Feitosa E, et al. Eucalyptol promotes lung repair in mice following cigarette smoke-induced emphysema. *Phytomedicine*. 2019;55:70–9. <https://doi.org/10.1016/j.phymed.2018.08.012>.
116. Gao W, Guo Y, Yang H. Platycodin D protects against cigarette smoke-induced lung inflammation in mice. *Int Immunopharmacol*. 2017;47:53–8. <https://doi.org/10.1016/j.intimp.2017.03.009>.
117. Yu D, Liu X, Zhang G, Ming Z, Wang T. Isoliquiritigenin inhibits cigarette smoke-induced COPD by attenuating inflammation and oxidative stress via the regulation of the Nrf2 and NF-kappaB signaling pathways. *Front Pharmacol*. 2018;9:1001. <https://doi.org/10.3389/fphar.2018.01001>.
118. Han B, et al. The novel compound Sul-121 inhibits airway inflammation and hyperresponsiveness in experimental models of chronic obstructive pulmonary disease. *Sci Rep*. 2016;6:26928. <https://doi.org/10.1038/srep26928>.
119. Li XY, Luo BL, Wang LJ, Zhang WD, Liu ZG. 15-Deoxy-prostaglandin J2 anti-inflammation in a rat model of chronic obstructive pulmonary disease and human bronchial epithelial cells via Nrf2 activation. *Genet Mol Res*. 2015;14:14037–42. <https://doi.org/10.4238/2015.October.29.22>.
120. Walters DM, Cho HY, Kleeberger SR. Oxidative stress and antioxidants in the pathogenesis of pulmonary fibrosis: a potential role for Nrf2. *Antioxid Redox Signal*. 2008;10:321–32. <https://doi.org/10.1089/ars.2007.1901>.
121. Cho HY, Reddy SP, Yamamoto M, Kleeberger SR. The transcription factor NRF2 protects against pulmonary fibrosis. *FASEB J*. 2004;18:1258–60. <https://doi.org/10.1096/fj.03-1127je>.
122. Chen D, et al. The mechanism of rapamycin in the intervention of paraquat-induced acute lung injury in rats. *Xenobiotica*. 2015;45:538–46. <https://doi.org/10.3109/00498254.2014.995149>.
123. Xu Y, et al. Rapamycin protects against paraquat-induced pulmonary fibrosis: activation of Nrf2 signaling pathway. *Biochem Biophys Res Commun*. 2017;490:535–40. <https://doi.org/10.1016/j.bbrc.2017.06.074>.
124. Tai W, et al. Rapamycin attenuates the paraquat-induced pulmonary fibrosis through activating Nrf2 pathway. *J Cell Physiol*. 2019; <https://doi.org/10.1002/jcp.29094>.
125. Xin X, et al. Protective effects of Rosavin on bleomycin-induced pulmonary fibrosis via suppressing fibrotic and inflammatory signaling pathways in mice. *Biomed Pharmacother*. 2019;115:108870. <https://doi.org/10.1016/j.biopha.2019.108870>.
126. Tian SL, Yang Y, Liu XL, Xu QB. Emodin attenuates bleomycin-induced pulmonary fibrosis via anti-inflammatory and anti-oxidative activities in rats. *Med Sci Monit*. 2018;24:1–10. <https://doi.org/10.12659/msm.905496>.
127. Liu Y, Lu F, Kang L, Wang Z, Wang Y. Pirfenidone attenuates bleomycin-induced pulmonary fibrosis in mice by regulating Nrf2/Bach1 equilibrium. *BMC Pulm Med*. 2017;17:63. <https://doi.org/10.1186/s12890-017-0405-7>.
128. Bodas M, Vij N. The NF-kappaB signaling in cystic fibrosis lung disease: pathophysiology and therapeutic potential. *Discov Med*. 2010;9:346–56.
129. Chen J, et al. Dysfunction of Nrf-2 in CF epithelia leads to excess intracellular H2O2 and inflammatory cytokine production. *PLoS One*. 2008;3:e3367. <https://doi.org/10.1371/journal.pone.0003367>.
130. Ziady AG, et al. Interaction with CREB binding protein modulates the activities of Nrf2 and NF-kappaB in cystic fibrosis airway epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2012;302:L1221–31. <https://doi.org/10.1152/ajplung.00156.2011>.

131. Komaravelli N, et al. Respiratory syncytial virus infection down-regulates antioxidant enzyme expression by triggering deacetylation-proteasomal degradation of Nrf2. *Free Radic Biol Med.* 2015;88:391–403. <https://doi.org/10.1016/j.freeradbiomed.2015.05.043>.
132. Cho HY, et al. Antiviral activity of Nrf2 in a murine model of respiratory syncytial virus disease. *Am J Respir Crit Care Med.* 2009;179:138–50. <https://doi.org/10.1164/rccm.200804-535OC>.
133. Kosmider B, et al. Nrf2 protects human alveolar epithelial cells against injury induced by influenza A virus. *Respir Res.* 2012;13:43. <https://doi.org/10.1186/1465-9921-13-43>.
134. Kesic MJ, Simmons SO, Bauer R, Jaspers I. Nrf2 expression modifies influenza A entry and replication in nasal epithelial cells. *Free Radic Biol Med.* 2011;51:444–53. <https://doi.org/10.1016/j.freeradbiomed.2011.04.027>.
135. Dai JP, et al. Emodin inhibition of influenza A virus replication and influenza viral pneumonia via the Nrf2, TLR4, p38/JNK and NF-kappaB pathways. *Molecules.* 2017;22 <https://doi.org/10.3390/molecules22101754>.
136. Gomez JC, Dang H, Martin JR, Doerschuk CM. Nrf2 modulates host defense during *Streptococcus pneumoniae* pneumonia in mice. *J Immunol.* 2016;197:2864–79. <https://doi.org/10.4049/jimmunol.1600043>.
137. Zahlten J, et al. *Streptococcus pneumoniae*-induced oxidative stress in lung epithelial cells depends on pneumococcal autolysis and is reversible by resveratrol. *J Infect Dis.* 2015;211:1822–30. <https://doi.org/10.1093/infdis/jiu806>.
138. Palanisamy GS, et al. Evidence for oxidative stress and defective antioxidant response in Guinea pigs with tuberculosis. *PLoS One.* 2011;6:e26254. <https://doi.org/10.1371/journal.pone.0026254>.
139. Taguchi K, Yamamoto M. The KEAP1-NRF2 system in Cancer. *Front Oncol.* 2017;7:85. <https://doi.org/10.3389/fonc.2017.00085>.
140. Khor TO, et al. Increased susceptibility of Nrf2 knockout mice to colitis-associated colorectal cancer. *Cancer Prev Res (Phila).* 2008;1:187–91. <https://doi.org/10.1158/1940-6207.CAPR-08-0028>.
141. Singh A, et al. Dysfunctional KEAP1-NRF2 interaction in non-small-cell lung cancer. *PLoS Med.* 2006;3:e420. <https://doi.org/10.1371/journal.pmed.0030420>.
142. Padmanabhan B, et al. Structural basis for defects of Keap1 activity provoked by its point mutations in lung cancer. *Mol Cell.* 2006;21:689–700. <https://doi.org/10.1016/j.molcel.2006.01.013>.
143. Ohta T, et al. Loss of Keap1 function activates Nrf2 and provides advantages for lung cancer cell growth. *Cancer Res.* 2008;68:1303–9. <https://doi.org/10.1158/0008-5472.CAN-07-5003>.
144. Takahashi T, et al. Mutations in Keap1 are a potential prognostic factor in resected non-small cell lung cancer. *J Surg Oncol.* 2010;101:500–6. <https://doi.org/10.1002/jso.21520>.
145. Bauer AK, et al. Targeted deletion of Nrf2 reduces urethane-induced lung tumor development in mice. *PLoS One.* 2011;6:e26590. <https://doi.org/10.1371/journal.pone.0026590>.
146. Bialk P, Wang Y, Banas K, Kmiec EB. Functional gene knockout of NRF2 increases chemosensitivity of human lung cancer A549 cells in vitro and in a xenograft mouse model. *Mol Ther Oncolytics.* 2018;11:75–89. <https://doi.org/10.1016/j.omto.2018.10.002>.
147. DeNicola GM, et al. Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. *Nature.* 2011;475:106–9. <https://doi.org/10.1038/nature10189>.
148. Tsai JR, et al. High expression of heme oxygenase-1 is associated with tumor invasiveness and poor clinical outcome in non-small cell lung cancer patients. *Cell Oncol (Dordr).* 2012;35:461–71. <https://doi.org/10.1007/s13402-012-0105-5>.
149. Wang X, et al. Genetic variation and antioxidant response gene expression in the bronchial airway epithelium of smokers at risk for lung cancer. *PLoS One.* 2010;5:e11934. <https://doi.org/10.1371/journal.pone.0011934>.
150. Zhan L, et al. Regulatory role of KEAP1 and NRF2 in PPARgamma expression and chemoresistance in human non-small-cell lung carcinoma cells. *Free Radic Biol Med.* 2012;53:758–68. <https://doi.org/10.1016/j.freeradbiomed.2012.05.041>.
151. Dey S, et al. ATF4-dependent induction of heme oxygenase 1 prevents anoikis and promotes metastasis. *J Clin Invest.* 2015;125:2592–608. <https://doi.org/10.1172/JCI78031>.

152. DeNicola GM, et al. NRF2 regulates serine biosynthesis in non-small cell lung cancer. *Nat Genet.* 2015;47:1475–81. <https://doi.org/10.1038/ng.3421>.
153. Cuadrado A, et al. Therapeutic targeting of the NRF2 and KEAP1 partnership in chronic diseases. *Nat Rev Drug Discov.* 2019;18:295–317. <https://doi.org/10.1038/s41573-018-0008-x>.
154. Harvey CJ, et al. Targeting Nrf2 signaling improves bacterial clearance by alveolar macrophages in patients with COPD and in a mouse model. *Sci Transl Med.* 2011;3:78ra32. <https://doi.org/10.1126/scitranslmed.3002042>.
155. Bewley MA, et al. Opsonic phagocytosis in chronic obstructive pulmonary disease is enhanced by Nrf2 agonists. *Am J Respir Crit Care Med.* 2018;198:739–50. <https://doi.org/10.1164/rccm.201705-0903OC>.
156. Attucks OC, et al. Induction of heme oxygenase I (HMOX1) by HPP-4382: a novel modulator of Bach1 activity. *PLoS One.* 2014;9:e101044. <https://doi.org/10.1371/journal.pone.0101044>.
157. Hu L, et al. Discovery of a small-molecule inhibitor and cellular probe of Keap1-Nrf2 protein-protein interaction. *Bioorg Med Chem Lett.* 2013;23:3039–43. <https://doi.org/10.1016/j.bmcl.2013.03.013>.
158. Davies TG, et al. Monoacidic inhibitors of the Kelch-like ECH-associated protein 1: nuclear factor erythroid 2-related factor 2 (KEAP1:NRF2) protein-protein interaction with high cell potency identified by fragment-based discovery. *J Med Chem.* 2016;59:3991–4006. <https://doi.org/10.1021/acs.jmedchem.6b00228>.
159. Brasier AR. Therapeutic targets for inflammation-mediated airway remodeling in chronic lung disease. *Expert Rev Respir Med.* 2018;12:931–9. <https://doi.org/10.1080/17476348.2018.1526677>.
160. Wang WC, et al. Protective effects of tyrosol against LPS-induced acute lung injury via inhibiting NF-kappaB and AP-1 activation and activating the HO-1/Nrf2 pathways. *Biol Pharm Bull.* 2017;40:583–93. <https://doi.org/10.1248/bpb.b16-00756>.
161. Yang H, Lv H, Li H, Ci X, Peng L. Oridonin protects LPS-induced acute lung injury by modulating Nrf2-mediated oxidative stress and Nrf2-independent NLRP3 and NF-kappaB pathways. *Cell Commun Signal.* 2019;17:62. <https://doi.org/10.1186/s12964-019-0366-y>.
162. Pei X, Zhang XJ, Chen HM. Bardoxolone treatment alleviates lipopolysaccharide (LPS)-induced acute lung injury through suppressing inflammation and oxidative stress regulated by Nrf2 signaling. *Biochem Biophys Res Commun.* 2019;516:270–7. <https://doi.org/10.1016/j.bbrc.2019.06.006>.
163. Grzegorzewska AP, et al. Dimethyl Fumarate ameliorates pulmonary arterial hypertension and lung fibrosis by targeting multiple pathways. *Sci Rep.* 2017;7:41605. <https://doi.org/10.1038/srep41605>.
164. Saidu NE, et al. Dimethyl fumarate controls the NRF2/DJ-1 axis in cancer cells: therapeutic applications. *Mol Cancer Ther.* 2017;16:529–39. <https://doi.org/10.1158/1535-7163.MCT-16-0405>.
165. Choi RJ, Chun J, Khan S, Kim YS. Desoxyrhapontigenin, a potent anti-inflammatory phytochemical, inhibits LPS-induced inflammatory responses via suppressing NF-kappaB and MAPK pathways in RAW 264.7 cells. *Int Immunopharmacol.* 2014;18:182–90. <https://doi.org/10.1016/j.intimp.2013.11.022>.
166. Lai SW, et al. Antidiabetes drugs correlate with decreased risk of lung cancer: a population-based observation in Taiwan. *Clin Lung Cancer.* 2012;13:143–8. <https://doi.org/10.1016/j.clcc.2011.10.002>.
167. Zhang J, Jiao K, Liu J, Xia Y. Metformin reverses the resistance mechanism of lung adenocarcinoma cells that knocks down the Nrf2 gene. *Oncol Lett.* 2018;16:6071–80. <https://doi.org/10.3892/ol.2018.9382>.
168. Tao S, et al. Vitamin D protects against particles-caused lung injury through induction of autophagy in an Nrf2-dependent manner. *Environ Toxicol.* 2019;34:594–609. <https://doi.org/10.1002/tox.22726>.

169. Margaritopoulos GA, Vasarmidi E, Antoniou KM. Pirfenidone in the treatment of idiopathic pulmonary fibrosis: an evidence-based review of its place in therapy. *Core Evid.* 2016;11:11–22. <https://doi.org/10.2147/CE.S76549>.
170. Cheung KL, Kong AN. Molecular targets of dietary phenethyl isothiocyanate and sulforaphane for cancer chemoprevention. *AAPS J.* 2010;12:87–97. <https://doi.org/10.1208/s12248-009-9162-8>.
171. Liby K, et al. The synthetic triterpenoids CDDO-methyl ester and CDDO-ethyl amide prevent lung cancer induced by vinyl carbamate in a/J mice. *Cancer Res.* 2007;67:2414–9. <https://doi.org/10.1158/0008-5472.CAN-06-4534>.