

Genome Sequencing in the Parkinson Disease Clinic

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Abstract

Background and Objectives

Genetic variants affect both Parkinson disease (PD) risk and manifestations. Although genetic information is of potential interest to patients and clinicians, genetic testing is rarely performed during routine PD clinical care. The goal of this study was to examine interest in comprehensive genetic testing among patients with PD and document reactions to possible findings from genome sequencing in 2 academic movement disorder clinics.

Methods

In 203 subjects with PD (age = 63 years, 67% male), genome sequencing was performed and filtered using a custom panel, including 49 genes associated with PD, parkinsonism, or related disorders, as well as a 90-variant PD genetic risk score. Based on the results, 231 patients (age = 67 years, 63% male) were surveyed on interest in genetic testing and responses to vignettes covering (1) familial risk of PD (*LRRK2*); (2) risk of PD dementia (*GBA*); (3) PD genetic risk score; and (4) secondary, medically actionable variants (*BRCA1*).

Results

Genome sequencing revealed a *LRRK2* variant in 3% and a *GBA* risk variant in 10% of our clinical sample. The genetic risk score was normally distributed, identifying 41 subjects with a high risk of PD. Medically actionable findings were discovered in 2 subjects (1%). In our survey, the majority (82%) responded that they would share a *LRRK2* variant with relatives. Most registered unchanged or increased interest in testing when confronted with a potential risk for dementia or medically actionable findings, and most (75%) expressed interest in learning their PD genetic risk score.

Discussion

Our results highlight broad interest in comprehensive genetic testing among patients with PD and may facilitate integration of genome sequencing in clinical practice.

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Glossary

ACMG = American College of Medical Genetics and Genomics; HGMD = Human Gene Mutation Database; PD = Parkinson disease; VUS = variants of uncertain significance.

Advances in genetic knowledge have implicated more than 100 genes in Parkinson disease (PD) risk, including both rare and common variants.^{1,2} Along with potential implications for counseling families, identification of these genetic risk factors may also affect clinical decision making. Several clinical trials for targeted therapies in PD are now recruiting patients with *GBA*-PD or *LRRK2*-PD, motivating large-scale efforts to identify eligible subjects.³ In addition, numerous studies highlight links between specific genetic variants and heterogeneous PD symptoms. For example, *GBA*-PD is characterized by more rapid disease progression and an increased risk of dementia.⁴⁻⁷ *APOE* genotype, which has an established role in clinical risk stratification for Alzheimer disease,⁸ is also associated with Lewy body dementia.^{9,10} In *LRRK2*-PD, several studies have suggested a more benign disease course, whereas others reveal a higher likelihood of the postural instability gait difficulty motor phenotype.^{11,12} Finally, although most common PD risk variants (i.e., population allele frequency >1%) have only modest effect sizes in isolation, a genetic risk score incorporating dozens of single nucleotide polymorphisms predicts accelerated PD progression^{13,14} and may also facilitate early diagnosis.¹⁵ Because it is derived from common variant genotypes, the PD genetic risk score may be applicable to many patients with PD. Similar algorithms have been clinically validated for risk assessment in other common and complex genetic disorders, such as cardiovascular disease.^{16,17}

Most current commercial gene panels for PD are largely restricted to testing for monogenic causes of mendelian PD. The available tests also vary widely in the number of genes and variants tested, frequently include loci with uncertain evidence, and do not include common genetic variants or assess a genetic risk score.¹⁸ Compared with targeted panel assays, genome sequencing comprehensively examines both rare and common variants in most genes. In addition, results can be readily filtered and reported based on all available evidence, making possible iterative analytic updates following new discoveries and without requiring an additional blood draw or new data generation. The American College of Medical Genetics and Genomics (ACMG) recommends offering testing for genes conferring a risk of a serious medical condition for which there is an effective intervention or need for surveillance.¹⁹ Such medically actionable genetic findings unrelated to PD can be readily screened using genome sequencing.

Despite the growing potential to affect clinical decision making, genetic testing is rarely used in the routine clinical assessment of PD,²⁰ yet most patients with PD have a strong desire to know their genetic information.²¹⁻²³ Besides potential barriers for reimbursement, many neurologists feel

poorly prepared to provide counseling to patients regarding results.²⁴ In addition, other issues that may arise from comprehensive testing using genome sequencing have not been fully explored, such as disclosure of genetic risk scores or potential discovery of medically actionable findings unrelated to PD diagnosis. To address these gaps, it is essential to document not only the spectrum of findings from genome sequencing that would be expected in clinical practice but also the possible reactions of patients to learning of these results.

Methods

Subject Recruitment

Subjects were recruited from 2 academic movement disorder centers (Baylor College of Medicine and University of Maryland School of Medicine). During routine clinic visits between 2013 and 2019, patients with PD who were diagnosed by a movement disorder specialist were offered participation in a DNA bank repository. Demographics and selected clinical details were collected through chart review and confirmed based on an unstructured interview at the time of sample collection in the clinic, including a family history of PD (defined as any affected blood relative), age at symptom onset, and results from cognitive screening. The DNA bank repository protocol permits genome sequencing for research but does not allow for disclosure of results. Therefore, in a separate study conducted between 2018 and 2019, patients with PD from both clinics were recruited for an approximately 15-minute survey in which responses were recorded to several clinical vignettes including genetic testing results (below). Individuals with dementia and those who had previously undergone clinical genetic testing for PD were excluded from the survey study. There were 203 subjects who contributed DNA samples for genome sequencing and 231 who completed the survey. Twenty-eight subjects participated in both parts of the study.

Standard Protocol Approvals, Registrations, and Patient Consents

All subjects provided informed consent. The institutional review boards at Baylor College of Medicine and University of Maryland approved both the biospecimen collection/sequencing and survey study included in this report.

Custom PD Gene Panel

We developed a custom PD gene panel for filtering sequencing results and creation of personalized PD genome risk profile reports (eTable 1, links.lww.com/NXG/A530). We included 5 categories of genes/variants, including both (1) 5 highly penetrant PD risk genes (*SNCA*, *LRRK2*, *VPS35*, *PARK2/PRKN*, and *PINK1*) and (2) 3 moderately penetrant

risk/modifier genes (*GBA*, *SMPD1*, and *APOE*). *APOE* has an established association with cognitive impairment in PD.^{9,25} Because idiopathic PD clinical manifestations sometimes overlap with other genetic syndromes that cause parkinsonism (e.g., dopamine-responsive dystonias²⁶), we also included an additional category (3) including 41 such genes (eTable 1). All genes selected for the panel were based on careful literature review with the following inclusion criteria: at least 1 pathogenic variant reported in 3 separate families by 2 or more independent groups or at least 1 large sequencing study revealing a significant association with PD or parkinsonism. For the PD genetic risk score (category 4), we considered 90 risk variants identified by PD genome-wide association study (GWAS) meta-analysis, weighted by their individual odds ratios for PD.^{16,27} We divided the genetic risk scores into low, medium, and high risk based on quintiles (first quintile defined as low risk, second–fourth quintile as medium risk, and fifth quintile as high risk).²⁸ Subjects with a high vs low score were compared in terms of age at onset, family history, and the Montreal Cognitive Assessment score using the Student *t* test. Finally (category 5), our panel included 59 genes unrelated to PD, but which were recommended for secondary reporting by the ACMG.²⁹

Genome Sequencing

DNA was extracted from peripheral blood lymphocytes or saliva samples using standard protocols. Whole-genome sequencing data were generated for 203 samples at BCM-HGSC using established library preparation and sequencing methods. Libraries were prepared using KAPA Hyper PCR-free library reagents (KK8505; Kapa Biosystems Inc., Wilmington, MA) on Beckman robotic workstations (Biomek FX and FXp models). In brief, DNA (750 ng) was sheared into fragments of approximately 200–600 bp using the Covaris E220 system (96 well format; Covaris, Inc., Woburn, MA) followed by purification of the fragmented DNA using AMPure XP beads. A double size selection step was used, with different ratios of AMPure XP beads, to select a narrow size band of sheared DNA molecules for library preparation. DNA end-repair and 3'-adenylation were then performed in the same reaction followed by ligation of Illumina unique dual barcode adapters (Cat# 20022370) to create PCR-Free libraries, and the library was run on the Fragment Analyzer (Advanced Analytical Technologies, Inc., Ames, IA) to assess the library size and the presence of remaining adapter dimers. This was followed by a qPCR assay using the KAPA Library Quantification Kit (KK4835) using their SYBR FAST qPCR Master Mix for size estimation and quantification. WGS libraries were sequenced on the Illumina NovaSeq instrument using the S4 flow cells with 24 libraries in each lane. Sequencing reagents used included the NovaSeq 6000 S4 Reagent Kit (Cat# 20012866) and NovaSeq Xp 4-Lane Kit (Cat# 20021665). Libraries were loaded at an average concentration of 365 pM to generate 150-bp paired-end reads. Sequence data were processed using HgV (Mercury V, version 17.5), the HGSC workflow management system, and mapped to human reference build hg19.^{30,31} Unique aligned

sequences in these samples varied between 69.6 and 285.7 Gb, with an average of 129.89 Gb per sample. The average median insert size was 420 bp, and the average mode insert size was 399 bp. For the analysis of copy number variation at the *PRKN* locus, we applied the Dragen calling algorithm with Circular Binary Segmentation caller using the self-normalization method (Illumina, San Diego, CA, version 3.8.4), which integrates both depth-based calling and split read methods to identify potential copy number variation within 500 kb of the gene.

Variant Filtering/Analysis

All variants, including both exonic and intronic variants, were initially annotated using the ClinVar³² and Human Gene Mutation Database (HGMD) in April 2019, supplemented by literature review and discussion by our multidisciplinary team, which included movement disorder neurologists, a clinical geneticist, a genetic counselor, and basic scientists.³³ With the exception of *GBA* (see below), all variants considered pathogenic in this study were annotated in ClinVar as either pathogenic/likely pathogenic or in HGMD as DM, indicating disease causing. In the case of *GBA*, database annotation is largely based on the risk for Gaucher disease; however, we additionally considered 2 *GBA* variants, p.E365K and p.T408M, with strong literature support for an increased risk for PD, but which are nonpathogenic for Gaucher.^{34,35} For all variants, we additionally performed literature review to document consistent reports from multiple independent families and/or large cohort sequencing studies or support from experimental validation in model organisms. This supportive evidence is documented in eTable 2 (links.lww.com/NXG/A530). For category 1 genes, following the exclusion of ClinVar benign or likely benign variants, all other identified findings with MAF <5% were considered variants of uncertain significance (VUS) (eTable 3).

Survey Study

The survey comprised 4 clinical vignettes developed based on empiric results from our genome sequencing study (see eMethods for complete survey, links.lww.com/NXG/A530). The vignettes highlighted the following scenarios: family risk of PD due to a *LRRK2* variant (vignette 1); personal dementia risk in a carrier of a *GBA* variant (vignette 2); negative findings for highly penetrant variants and reporting of the genetic risk score (vignette 3); and discovery of a medically actionable variant in *BRCA1* (vignette 4). These vignettes were designed both as an educational tool and to assess how patient interest in genetic testing may be affected by different results. Vignettes and follow-up questions were read aloud to subjects. The interest level for comprehensive genetic testing was assessed based on a Likert scale (0, “much less interested,” to 5, “much more interested”). Qualitatively, we also recorded comments from subjects on factors influencing their decision making. Comments responding to at least 1 vignette were recorded from 133 of 231 subjects. We also requested feedback on how doctors might better prepare patients for genetic testing in PD.

Table 1 Subject Characteristics

	Sequencing	Survey
Subjects, n	203	231
Age, y, mean (SD)	63 (9)	67 (9.9)
Sex, % male	67	63
Disease duration, y, mean (SD)	7 (6)	9 (5)
Race, % European ancestry	86	87
Family history of PD, ^a %	27	24

Abbreviation: PD = Parkinson disease.

^a Family history was defined as any blood relative with PD.

Data Availability

Complete results from the survey are available in eTable 4 (links.lww.com/NXG/A530). For all subjects who consented for public data sharing, sequencing data are in process for release in relevant genomic databases. The complete sequencing data are also available on request by contacting the corresponding author, Dr. Shulman (joshua.shulman@bcm.edu).

Results

Genome Sequencing

We performed genome sequencing in 203 unselected, unrelated subjects with PD. Subject demographics and clinical characteristics are summarized in Table 1. For filtering and annotation of personal genomes, we developed a custom PD gene and

variant panel including 5 categories of reportable genetic risk (Methods). The gene panel was designed to facilitate generation of individual PD Genome Sequencing Reports (Figure 1). Because genome sequencing was conducted under a protocol that did not allow for disclosure of results, the individual findings instead informed our development of several clinical vignettes that we subsequently presented to a largely nonoverlapping sample of subjects with PD.

Among highly penetrant gene variants (category 1; see also vignette 1, below), our sample included 7 subjects with a *LRRK2* pathogenic variant (3%) and 1 heterozygous carrier of a pathogenic *PRKN* single nucleotide variant. In this individual, sequence data were further interrogated to exclude the possibility of a copy number variant. An additional 28 subjects (14%) were discovered to have a VUS in at least 1 category 1 gene (eTable 3, links.lww.com/NXG/A530). We next examined variants in genes associated with a moderate risk of PD and disease modifiers (category 2; vignette 2). We discovered 21 carriers of *GBA* pathogenic variants (10%). Notably, 1 subject was a carrier of both a *GBA*^{E356K} and a *LRRK2*^{M1646T} variant. For *APOE*, 45 subjects were heterozygous for the *e4* allele. Two individuals were homozygous for *APOE e4*, but based on available clinical documentation, both individuals were not known to have cognitive impairment. Notably, 2 subjects were carriers of *SMPD1* variants previously reported to be pathogenic for Niemann-Pick Type A (p.E517V, c.1264-1G>T); however, these alleles have not been definitively associated with PD. Table 2 shows all reportable pathogenic variants in genes from categories 1 and 2. No individuals in our sample were discovered to have variants linked to other genetic causes of parkinsonism (category 3).

Figure 1 Sample PD Genome Sequencing Report

Parkinson disease (PD) genome sequencing report
<p>1. Highly penetrant PD risk genes Genes analyzed: <i>LRRK2</i>, <i>SNCA</i>, <i>VPS35</i>, <i>PRKN</i>, and <i>PINK1</i> Results: Negative No variants were found in these genes that can cause familial PD or increase risk of PD.</p>
<p>2. Moderately penetrant PD risk genes and disease modifiers Genes analyzed: <i>GBA</i>, <i>SMPD1</i>, and <i>APOE</i> Results: Negative No variants were found in these genes that increase risk for PD or influence risk for dementia.</p>
<p>3. Genes associated with parkinsonism Genes analyzed: <i>DJ-1</i>, <i>SYNJ1</i>, <i>PLA2G6</i>, <i>GCH1</i>, and 37 other genes Results: Negative No variants were found in these genes causing other syndromes that share symptoms with PD.</p>
<p>4. PD genetic risk score Genes analyzed: 90 variants that combine to give a score of low, medium, or high risk of PD Results: Medium risk level A moderate number of common variants were found to increase risk of PD.</p>
<p>5. Medically actionable findings unrelated to PD Genes analyzed: <i>BRCA1</i>, <i>BRCA2</i>, <i>MEN1</i>, <i>RYR2</i>, <i>VHL</i>, and 55 other genes Results: Negative No variants were found in these genes associated with other diseases for which intervention or surveillance is recommended.</p>

This report represents the most common results based on application of our customized PD gene panel (see also eTable 1, links.lww.com/NXG/A530, for all genes included in categories 1–3).

Table 2 Sequencing Results

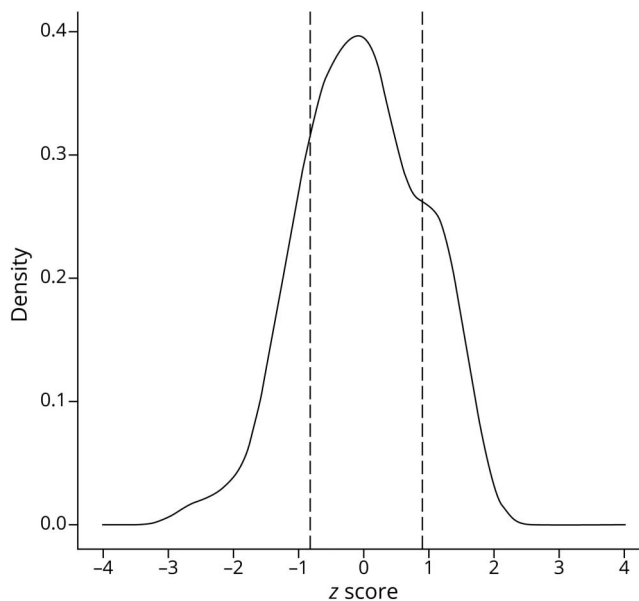
Category	Gene	Variant ^a	Subjects
1: High penetrance	<i>LRRK2</i>	c.1256C>T:p.A419V	1
		c.4937T>C:p.M1646T	5
		c.6055G>A:p.G2019S	1
2: Moderate penetrance	<i>GBA</i>	<i>PRKN</i> c.823C>T:p.R275W	1
		c.1093G>A:p.E365K	12
		c.1448T>C:p.L483P	1
		c.115+1G>A	1
		c.1223C>T:p.T408M	3
3: Medically actionable	<i>APOE</i>	c.1342G>C:p.D448H	1
		c.1226A>G:p.D409S	2
		c.882T>G:p.H294Q	1
		E4 heterozygous	45
		E4 homozygous	2
		<i>DSG2</i> c.523+1G>A	1
		<i>MSH2</i> c.942+3A>T	1

^a Additional details including transcripts, chromosome position, and literature-based evidence supporting all variants can be found in eTable 2 (links. lww.com/NXG/A530).

We next examined the 90-variant PD genetic risk score (category 4), which was normally distributed in our sample (Figure 2). After dividing into quintiles,²⁸ there were 41 subjects each with a high or low genetic risk score. Of note in comparisons of the low- vs high-risk groups, we found no significant difference between positive family history ($p = 0.3$), age at symptom onset ($p = 0.6$), or level of cognitive function ($p = 0.2$), despite reports of such associations in much larger sample sizes.^{36,37} As expected, most subjects ($n = 122$) had a score consistent with a moderate risk, and for most of these ($n = 104$), no other genes were flagged from categories 1 to 3 (representative report, Figure 1; vignette 3). Finally, we examined 59 ACMG medically actionable findings, unrelated to PD risk (category 5; vignette 4). We discovered 2 individuals harboring such alleles, including a *DSG2* variant associated with arrhythmogenic right ventricular cardiomyopathy and a variant in *MSH2*, which causes Lynch syndrome, conferring an increased risk for colon and other cancers (Table 2).²⁹

Survey Study

Because our institutional review board protocol did not permit disclosure of genome sequencing results to participants, we enrolled a largely nonoverlapping sample of 231 subjects with PD from our 2 centers for a survey study, in which representative results were presented as part of clinical vignettes. Subject demographics are summarized in Table 1. A minority of subjects ($n = 28$) completing the survey also contributed samples for genome sequencing. Before hearing

Figure 2 Genetic Risk Score Distribution

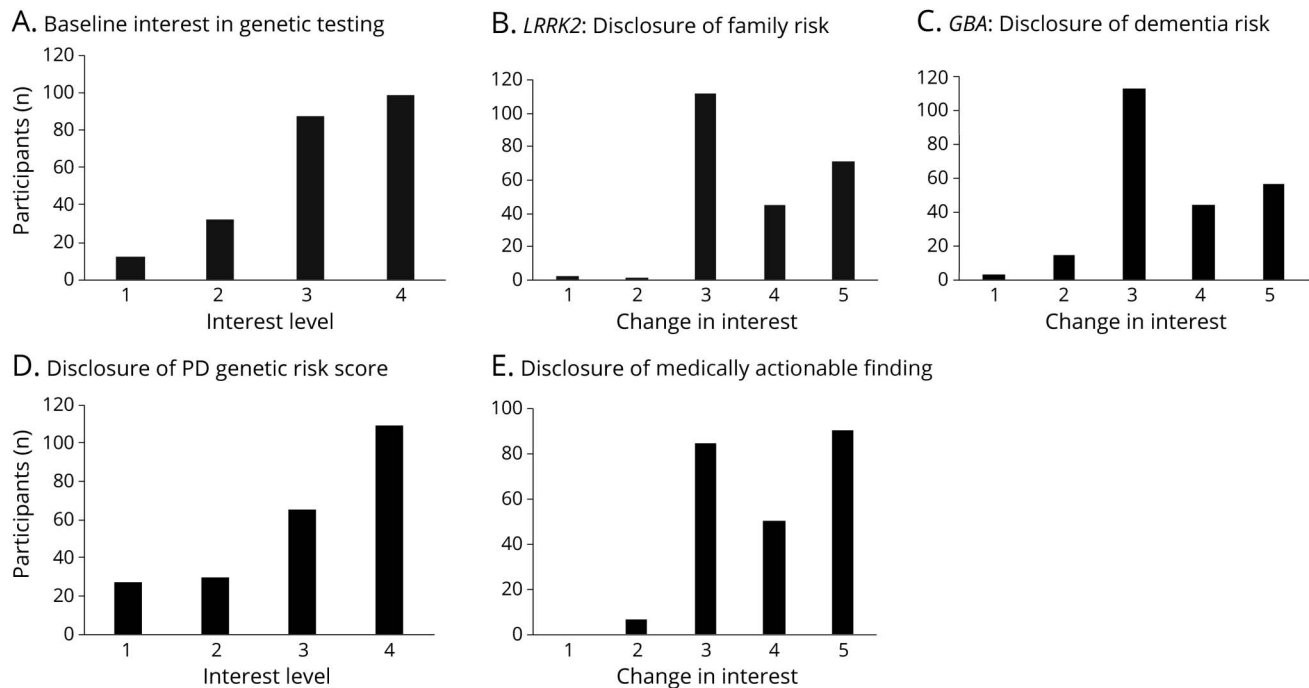
Density plot for the PD genetic risk score. Z scores reflect standard deviations from the mean. The low- and high-risk groups are indicated by dashed lines, highlighting cutoffs for the first and fifth quintiles, respectively. PD = Parkinson disease.

the vignettes, most participants (81%) responded that they were interested in genetic testing for PD (Figure 3). There was no significant difference in interest among respondents with a family history of PD ($p = 0.18$) or those contributing samples for genome sequencing ($p = 0.34$). Most respondents (82%) indicated that they would inform at-risk family members about the potential finding of a high-penetrance PD risk variant, such as *LRRK2*^{G2019S} (vignette 1). Only 2 subjects (1%) reported reduced interest in comprehensive genetic testing based on the potential implications for familial risk. Feedback often reflected concern for children or grandchildren. One subject commented, “I just want to know what I can do to keep my kids from getting this,” and another offered, “I don’t want to do anything that would hurt our grandchildren.” Three subjects expressed concern about potential insurance denial or job discrimination, either for themselves or their relatives.

Clinical vignette 2 focused on the potential for an increased dementia risk following discovery of a *GBA* variant. Only 16 subjects (7%) reported reduced interest in genetic testing, whereas 101 (44%) were, in fact, more interested in genetic testing after learning that genetic variants might indicate a heightened risk for PD dementia. When informed that genetic testing results, such as discovery of a *GBA* variant, would not currently change treatment decisions for the majority of patients, 75% of subjects responded that this did not change their interest in genetic testing.

For most patients, genetic testing is not expected to reveal a highly penetrant PD risk variant. Instead, vignette 3 highlights

Figure 3 Survey Results



Results from survey questions gauging subject interest in comprehensive genetic testing at baseline (A) or following several clinical vignettes, including disclosure of an *LRRK2* variant (B, vignette 1), *GBA* variant (C, vignette 2), genetic risk score (D, vignette 3), or secondary, medically actionable finding unrelated to PD (E, vignette 4). The survey responses for “interest level” (A and C), included 1, “not interested”; 2, “a little interested”; 3, “moderately interested”; and 4, “very interested.” For “change in interest” (B, C, and E), responses included 1, “much less interested”; 2, “a little less interested”; 3, “no change”; 4, “a little more interested”; and 5, “much more interested.” Most subjects reported no change or increased interest in genetic testing following disclosures. For complete survey and all results, see eMethods and eTable 4 (links.lww.com/NXG/A530). PD = Parkinson disease.

that a genetic risk score can be computed based on a large number of genetic polymorphisms. The majority of subjects (75%) expressed an interest in learning their PD genetic risk score. Among the subjects who were not interested in their PD genetic risk score, 4 expressed concerns that PD must be predominantly influenced by environmental factors, making genetic testing less relevant. Finally, most participants (88%) expressed a desire to receive information on medically actionable variants unrelated to PD following vignette 4, which included discussion of a pathogenic *BRCA1* variant. In fact, 61% reported that they were even more interested in genetic testing considering the possibility of learning such information.

Other general comments from subjects praised the potential utility of genetic testing for long-term care planning, improved understanding of PD, and promoting new therapies. Six subjects noted minimal interest in genetic testing due to the lack of availability of disease-modifying treatments. One volunteered, “I don’t think it should be discussed unless there is a cure.” Four subjects raised concerns about the cost of genetic testing. Six reported being unaware that PD is influenced by genetics or that testing was available. Several subjects were surprised to learn that genetic risk in PD is more complicated than in other disorders. One subject expressed that before the survey, he was concerned that genetic testing was clinically useful but was being withheld from patients because of cost. The most frequent comment at the end of the

survey was a request for more information. Fifteen subjects desired more education about genetic testing during doctor visits. Seventeen requested that written material on genetics in PD be made available. Three suggested a didactic class or seminar, and 5 asked for references to websites or online videos. Many expressed an increased interest in genetic testing that was specifically related to new information learned from completing the survey.

Discussion

A substantial majority of patients with PD seen at 2 academic medical centers were interested in information about genetic risk, including the opportunity to inform at-risk family members (72%), discover their possible risk of dementia (75%), obtain their genetic risk score (75%), and learn about potential medically actionable variants (88%). This is largely consistent with other published surveys.²¹⁻²³ Despite rapidly expanding genetic knowledge about PD, including many established variants that affect disease risk and clinical manifestations, testing is rarely pursued in routine PD clinical care. Most commercial genetic testing panels are largely limited to monogenic causes of familial PD.¹⁸ Moreover, such panels often include genes with limited evidence or omit more recently discovered loci. By contrast, genome sequencing offers more comprehensive genetic analysis. We obtained genome

sequencing from 203 patients with PD and developed a custom gene and variant panel for filtering and analysis. In our sample, 29 subjects had reportable, diagnostic findings of either high- or moderate-penetrance PD risk or modifying alleles among 49 genes examined. These results are overall consistent with other published work.³⁸⁻⁴⁰ We also computed a 90-variant PD genetic risk score for all subjects and examined secondary findings for 59 medically actionable genes. It is important to note that compared with a targeted gene panel assay, genome sequencing coupled with focused filtering for genes relevant to PD permits refiltering and updated reporting based on newly published findings.⁴¹ For example, since completing our analyses, the ACMG has already updated its recommendations on medically actionable findings to include 73 genes.¹⁹

Because sequencing was performed under a research protocol that did not allow for disclosure of genetic results to subjects, we designed a survey to assess reactions to clinical vignettes including representative findings. For the majority of subjects, interest in genetic testing remained strong or increased despite potential revelations of familial PD risk, personal risk of PD dementia, or lack of immediate effect on treatment decisions. Interest in clinical genetic testing for PD will also likely rise with increased awareness of clinical trials for disease-modifying treatments in carriers of *LRRK2* and *GBA* variants.^{3,42} If such therapies prove successful, results of this and other larger multicenter studies (e.g., PD GENERation⁴³) will be important to inform roll-out of more routine genetic testing for PD, including guidelines for effective communication with patients and families.

Based on recent studies,²⁴ many neurologists feel unprepared to communicate authoritatively on PD genetics, but some of the issues are commonly encountered in genetic counseling for other neurogenetic syndromes. For example, the discovery of high-penetrance *LRRK2* variants may trigger interest in cascade testing among family members. However, in the case of moderately penetrant variants, such as *GBA* or *APOE*, the implications for relatives may be more difficult to convey and for patients to conceptualize. Despite the potential for clinically meaningful findings, most patients are unlikely to discover responsible high- or moderate-penetrance PD risk variants, at least based on our current genetic knowledge. In our study, many subjects were surprised to learn that such negative results are the most likely outcome (86% of our sequencing sample). Inclusion of the PD genetic risk score expands the potential applicability of genetic testing, and we documented strong interest in such results. Higher genetic risk scores have been shown to be significantly associated with PD progression,^{44,45} cognitive impairment,¹³ and levodopa-induced dyskinesia.⁴⁶ Nevertheless, most neurologists may not be familiar with genetic risk scores. In addition, available risk scores are derived from GWAS conducted almost exclusively in European-ancestry populations, making them poorly generalizable for other groups.

Strengths of this study include genetic sampling from 2 high-volume academic movement disorders clinic populations and

inclusion of more than 200 subjects for both the genome sequencing and survey arms of the study. We also developed a custom panel for filtering and annotation of genome sequencing results based on systematic criteria and literature review. The clinical vignettes developed for our survey may be a valuable tool for engaging and educating patients as part of pretest genetic counseling. We also acknowledge some important limitations. Adoption of genome sequencing as a first-line clinical genetic test for PD may be limited by higher cost for data generation and analysis compared with single gene panels. Moreover, compared with single nucleotide variants, the detection of copy number variants may be less reliable, and our analysis did not systematically consider such alleles, which can be important contributors to PD risk.⁴⁶ In addition, expert consensus annotation of PD pathogenic variants is currently lacking to guide reporting of results for many genes. Notably, in the case of *GBA* and *SMPD1*, current database annotations are focused on lysosomal storage disorders rather than PD. Finally, attitudes about genetic testing from our academic movement disorders clinics may not be generalizable, including for more culturally and ethnically diverse populations. Overall, our findings highlight broad interest in comprehensive genetic testing among patients with PD and can help inform successful integration of genome sequencing in clinical practice.

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Name	Location	Contribution
Emily J. Hill, MD	Department of Neurology, and Parkinson's Disease Center and Movement Disorders Clinic, Baylor College of Medicine, Houston, TX; Department of Neurology and Rehabilitation Medicine, University of Cincinnati, OH	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; study concept or design; and analysis or interpretation of data
Laurie A. Robak, MD, PhD	Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; study concept or design; and analysis or interpretation of data
Rami Al-Ouran, PhD	Department of Pediatrics, Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and analysis or interpretation of data
Jennifer Deger, BSA	Department of Neuroscience, Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; study concept or design; and analysis or interpretation of data
Jamie C. Fong, MS	Department of Molecular and Human Genetics, and Center for Alzheimer's and Neurodegenerative Diseases Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; and analysis or interpretation of data
Paul Jerrod Vandeventer, BS	Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, TX	Analysis or interpretation of data
Emily Schulman, MD	Department of Neurology, University of Maryland School of Medicine, Baltimore	Major role in the acquisition of data
Sindhu Rao, MBBS	Department of Neurology, Baylor College of Medicine, Houston, TX	Major role in the acquisition of data

Appendix (continued)

Name	Location	Contribution
Hiba Saade, MD	Department of Neurology, and Center for Alzheimer's and Neurodegenerative Diseases, Baylor College of Medicine, Houston, TX	Major role in the acquisition of data
Joseph M. Savitt, MD	Department of Neurology, University of Maryland School of Medicine, Baltimore	Drafting/revision of the manuscript for content, including medical writing for content; and study concept or design
Rainer von Coelln, MD	Department of Neurology, University of Maryland School of Medicine, Baltimore	Drafting/revision of the manuscript for content, including medical writing for content; and major role in the acquisition of data
Neeja Desai, MD	Department of Neurology, University of Maryland School of Medicine, Baltimore	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; and study concept or design
Harshavardhan Doddapaneni, PhD	Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX	Major role in the acquisition of data and analysis or interpretation of data
Sejal Salvi, MS	Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX	Major role in the acquisition of data and analysis or interpretation of data
Shannon Dugan-Perez, BA	Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX	Major role in the acquisition of data and analysis or interpretation of data
Donna M. Muzny, MS	Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; major role in the acquisition of data; study concept or design; and analysis or interpretation of data
Amy L. McGuire, JD, PhD	Center for Alzheimer's and Neurodegenerative Diseases, and Center for Medical Ethics and Health Policy, Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; and study concept or design
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Richard A. Gibbs, PhD	Department of Molecular and Human Genetics, and Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX	Major role in the acquisition of data; study concept or design; and analysis or interpretation of data

Appendix (continued)

Name	Location	Contribution
Chad Shaw, PhD	Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, TX	Drafting/revision of the manuscript for content, including medical writing for content; study concept or design; and analysis or interpretation of data
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