# Antigen Recognition Properties of Mutant $V\beta 3^+$ T Cell Receptors Are Consistent with an Immunoglobulin-like Structure for the Receptor

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#### Summary

We examined the effect of mutations in the V $\beta$  portion of a pigeon cytochrome *c* (cyto *c*)-specific V $\beta$ 3<sup>+</sup>/V $\alpha$ 11<sup>+</sup> T cell receptor on its ability to recognize cyto *c*/IE<sup>k</sup> and various superantigens. The results were consistent with an immunoglobulin-like structure for the receptor V $\beta$  domain and with separate interaction sites on V $\beta$  for conventional antigen and superantigens. An amino acid predicted to lie in CDR1 was critical for cyto *c*/IE<sup>k</sup> but not superantigen recognition, while several amino acids predicted to lie in the hypervariable region 4 loop were critical for superantigen but not cyto *c*/IE<sup>k</sup> recognition.

The V domains of the TCR  $\alpha/\beta$  are similar to those of Igs both in primary amino acid sequence (1-3) and in the organization of their rearranging gene elements (4-7). A great deal is known from x-ray crystallography about the molecular structure of Ig V domains (8-13). Several authors have proposed that the conservation of critical amino acids among Ig and TCR V domains predicts that the structure of the TCR V domains will be very similar to those of Ig with Ig light chain corresponding to the TCR  $\alpha$  chain, and heavy chain corresponding to the TCR  $\beta$  chain (1-3). The TCR V domains are, therefore, predicted to be formed from a series of antiparallel  $\beta$  strands with the loops corresponding to the three Ig CDR regions brought to one face of the domain to form the binding site for antigen/MHC complexes. Some of the  $\beta$  strands (C, C', F, G) are predicted to be involved in the interaction between the  $\alpha$  and  $\beta$  V domains and therefore sequestered from the solvent. The other  $\beta$  strands (A, B, D, E) in combination with a fourth nonantigen binding loop (hypervariable region 4 [HV4])<sup>1</sup> between  $\beta$  strands D and E are predicted to form a lateral solvent-exposed face.

There are some data directly supporting this Ig model for the TCR V domains. Several laboratories have shown that amino acids important for antigen/MHC recognition in regions of the  $\alpha$  and  $\beta$  chain V domains correspond to the CDRs of Ig, especially CDR3 (14–18). We (18–20) and others (21) have demonstrated amino acids in HV4 and in the predicted solvent-exposed  $\beta$  strand leading up to CDR1 ( $\beta$  strand B) are involved in superantigen recognition.

In the current study we examined a TCR specific both for a cytochrome c (cyto c) peptide complexed with IE<sup>k</sup> and for the bacterial superantigen toxin SEC3. We tested several amino acids for their importance in the recognition of both types of antigen. An allelic residue in CDR1 was shown to be required for cyto  $c/IE^k$  but not staphylococcal enterotoxin (SE)C3 recognition, while exchange of amino acids in the predicted HV4 loop with those from the closely related V $\beta$ 17 element eliminated SEC3 recognition without affecting cyto  $c/IE^k$  reactivity. These results provide additional evidence for an Ig model for TCR V domains and further indicate that conventional antigens and superantigens interact with different sites on the TCR V $\beta$  domain.

# Materials and Methods

T Cell Hybridomas. The two T cell hybridomas used in these studies were produced as previously described (22, 23). The first, 5KC-73, was produced from T cells from B10.BR mice immunized with pigeon cyto c in CFA. The hybrid was selected on the basis of strong reactivity to both pigeon and moth cyto c peptide 88–104 presented by IE<sup>k</sup>-expressing APC and the presence of a V $\beta$ 3/ V $\alpha$ 11<sup>+</sup> TCR  $\alpha/\beta$  as judged by reactivity with the V $\beta$ 3-specific mAb KJ25 (24), and with the V $\alpha$ 11-specific mAb RR-8 (25). A  $\beta$  chain loss variant of this hybridoma was produced by first screening random subclones for the absence of surface TCR  $\alpha/\beta$  and then screening RNA from the TCR  $\alpha/\beta^-$  subclones by the PCR for the absence of V $\beta$ 3<sup>+</sup>  $\beta$  chain, but not V $\alpha$ 11<sup>+</sup>  $\alpha$  chain, mRNA

<sup>&</sup>lt;sup>1</sup>Abbreviations used in this paper: cyto c, cytochrome c; HV4, hypervariable region 4; SE, staphylococcal enterotoxin.

(26, 27). The phenotype was confirmed by the reexpression on the surface of TCR  $\alpha/\beta$  upon transfection of the variant with constructs encoding the complete  $\beta$  chain of 5KC-73 as described below. This transfectant, instead of the original hybridoma, was used as the wild-type control in the experiments presented here.

The second T cell hybridoma, 3QO23-24, was produced from T cells taken from SWR/J mice immunized with chicken (c)OVA and subsequently panned on the V $\beta$ 17a-specific mAb KJ23a (28, 29). This hybridoma was selected for high reactivity to cOVA presented by IA<sup>q</sup> APC and for reactivity with KJ23a (28).

APC. A number of APC lines were used in these experiments. An IE<sup>k</sup>-expressing fibroblast line, DCEK (30), was used for the presentation of cyto c peptides. This line and the mutant  $IE^{k-}/$ IAd+ B cell lymphoma line, M12.B5 (31), were used for presentation of staphylococcal toxin superantigens. These lines had the advantage that neither presented endogenous mammary tumor virus (MTV) superantigens (SAgs), DCEK because of lack of expression of the MTV SAg genes (32) and M12.B5 because of the lack of a functional IE molecule (31). MTV SAgs were presented by two other cell lines. CH12.1, a B cell lymphoma of C57BL origin (33), expresses the SAg of MTV9 (34, 35), which in combination with IE<sup>k</sup> stimulates V $\beta$ 17<sup>+</sup> T cells (29, 36), but not V $\beta$ 3<sup>+</sup> T cells. LBB.1, a class II<sup>+</sup> B cell lymphoma hybridoma produced by fusion of the BALB/c lymphoma, M12, and RF/J spleen cells (37), carries SAgs from a large array of MTVs in addition to MTV9, including MTV6 and MTV1, which are identical in sequence (38) and stimulate both  $V\beta 17^+$  (21) and  $V\beta 3^+$  (38) T cells.

Antigens. Pigeon cyto c was purchased from Sigma Chemical Co. (St. Louis, MO). Peptide 88–104 of pigeon and moth cyto c were prepared by the Molecular Resource Center, National Jewish Center for Immunology and Respiratory Medicine. SEA and SEC3 were purchased from Toxin Technology (Madison WI). Recombinant SEB was produced and purified as previously described (39).

IL-2 Production by T Cell Hybridomas. The relative responses of T cell hybridomas and transfectants to antigenic stimuli were assessed by the amount of IL-2 they produced as previously described using the IL-2-dependent cell line HT-2 (22, 40). Introduction of  $\beta$  Chains into  $\beta^{-}$  5KC-73. Reintroduction of the

TCR  $\alpha/\beta$   $\beta$  chain into the  $\beta$  chain loss variant of 5KC-73 was performed as previously described using the expression vector pBDWC\$ (18). Briefly, cDNA from mRNA prepared from 5KC-73 was used as template in a PCR. The 5' primer 5'-AGGTCG-ACCACCATGGCTACAAGGCTC-3' matched the V $\beta$ 3 leader and contained a Sall site. The 3'-primer 5'-GTCACATTTCTCAGA-TCTTC-3' matched the NH<sub>2</sub>-terminal region of C $\beta$  and introduced a BglII site. The V $\beta$ 3 domain-containing PCR product was cloned into the Sall/BGlII site of a variant of pTZ18R in which the BamHI site in the multiple cloning site was replaced with a BglII site. The insert from a clone established to have the correct sequence was subcloned into the Sall/BglII site of pBDWC $\beta$  in frame with the C $\beta$ 2 gene and transfected by electroporation into the  $\beta$  loss variant of 5KC-73. Transfectants were selected in G418 and one was chosen on the basis of the reexpression of the V $\beta$ 3+/ Val1<sup>+</sup> TCR  $\alpha/\beta$ .

Introduction of Mutations into the  $V\beta3$  of the TCR  $\alpha/\beta$  of 5KC-73. Mutations were introduced into the 5KC-73 V $\beta3$  as previously described (18). Briefly, an oligonucleotide primer containing the mutation was synthesized for one strand. A primer overlapping the first primer past the mutation was synthesized for the other strand. Using each of these with the primers described above, the 5' and 3' portions of the V $\beta3$  domain were synthesized by the PCR. The two fragments were fused by using them as a mixed template in a second PCR again using the leader and C $\beta$  primers described above. The fused fragment containing the mutations in a complete V $\beta$  domain was cloned, sequenced, subcloned into pBDWC $\beta$ , and transfected into the 5KC-73  $\beta$  loss variant as above. Transfectant clones were screened for approximately equal levels of TCR  $\alpha/\beta$  measured with KJ25 (anti-V $\beta$ 3) and HAM-597 (anti-C $\beta$ ) (41) mAbs.

# Results

Amino Acids in V $\beta$ 3 Important for SEC3 Recognition. In the mouse the V $\beta$  element most closely related to V $\beta$ 3 is V $\beta$ 17. These V $\beta$ s are identical in 65% of their amino acids with the differences scattered throughout their primary sequences (Fig. 1). T cells bearing these V $\beta$  elements have similar, but not identical, specificities for the SE superantigens (45, 46). Likewise, these elements have similar but not identical specificities for the mouse MTV-encoded SAgs (21, 24, 29, 36, 38).

These points are illustrated in Fig. 2. Two T cell hybridomas, 5KC-73, bearing V $\beta$ 3 and specific for pigeon cyto c (88–104) plus IE<sup>k</sup>, and 3QO23-24, bearing V $\beta$ 17 and specific for cOVA plus IA<sup>9</sup>, were compared for their response to various *Staphylococcus aureus* and MTV SAgs presented by various class II MHC antigen-presenting lines. Neither hybridoma responded to SEB, but both responded well to SEA presented by either an IE<sup>k</sup>-transfected fibroblast line or an IA<sup>d</sup>-expressing B cell lymphoma. However, the hybridoma differed in their response to SEC3. The V $\beta$ 3<sup>+</sup> hybridoma responded well to SEC3 presented by either cell line, while the response of the V $\beta$ 17<sup>+</sup> hybridoma was reduced ~50-fold with the IA<sup>d</sup>-presenting cells and absent

	10 20 3	0
VB3b (5KC-73)	KVIQTPRYLVKGQGQKAKMRCIPEKGHPV	√ F
VB3a (C57BR)		
VB17a (SWR)	<u> </u>	FΥ
	$\leftarrow \text{ B Strand B } \rightarrow \leftarrow \text{ CDR1} \rightarrow \leftarrow \rightarrow \leftarrow \text{ CDR1} \rightarrow \leftarrow \rightarrow $	
	40 50 60	
VB3b (5KC-73)	WYQQNKNNEFKFLINFQNQEVLQQIDMTE	(R
VB3a (CS7BR)		• •
VB17a (SWR)	· · · · QKQ · LT · · · · R · E · IME · T · LVK ·	• •
	$\leftarrow CDR2 \rightarrow$	
	70 80 90	
VR26 (5KC-73)		5
V830 (7578R)	I SKECFSKSFCSEEIQSSERGUSKEFECK.	
VR17a (SLP)		
ADTLY (2444)		
	← HV4→	
VIS310 (SKC-73)	SENNANSDYTFGSGTREEVI	

Figure 1. Sequences of V $\beta$ 3b, V $\beta$ 3a, and V $\beta$ 17a. The protein sequence of the V $\beta$ 3b (42) containing V $\beta$  domain of the 5KC-73 TCR  $\alpha/\beta$  is shown aligned with that of V $\beta$ 3a (43) and V $\beta$ 17a (29). The 5KC-73 DNA and protein sequence is identical to that of T cell hybridoma, 5C.C7, previously reported by Fink et al. (44). The V $\alpha$ 11.1/J $\alpha$ AC25-containing V $\alpha$ domain of the 5KC-73 receptor was cloned and sequenced as well (data not shown). The sequence is identical to that of hybridoma, 116, reported by Jorgensen et al. (16). For V $\beta$ 3a and V $\beta$ 17a only those amino acids differing from V $\beta$ 3b are shown. Others are designated with a period. Regions corresponding to CDR1, CDR2, CDR3,  $\beta$  strand B, and HV4 based on homology to Ig are indicated (1-3). Boxed amino acids were substituted into the V $\beta$ 3b element of 5KC-73 in these studies.



Figure 2.  $V\beta$ 3b and  $V\beta$ 17a have similar but not identical SAg specificities. Shown are the IL-2 responses of the  $V\beta$ 3b<sup>+</sup>  $\beta$  chain-restored  $\beta$  chain loss variant of 5KC-73 and the  $V\beta$ 17<sup>+</sup> hybridoma, 3QO23-24, to a challenge with either pigeon cyto c peptide 88-104 or various SAgs. Results are averaged from at least three experiments in each case.

with the IE<sup>k</sup>-presenting cells. The hybridomas differed in their responses to MTV SAgs as well. Whereas only the V $\beta$ 17<sup>+</sup> hybridoma responded to the IE-dependent MTV9 SAg of the B cell lymphoma CH12.1, both hybridomas responded to the B cell lymphoma hybridoma, LBB.1, which carries a number of MTV SAgs, including those of MTV9, MTV1, and MTV6.

In general the specificity of a TCR  $\alpha/\beta$  for a SAg/MHC complex is determined by the V $\beta$  element of its receptor. Our previous experiments identified amino acids important in SAg recognition in the region of V $\beta$  predicted to form solvent-exposed  $\beta$  strand B and the adjacent HV4 loop (18-20). V $\beta$ 3 and V $\beta$ 17 differ in seven amino acids in these regions



Figure 3. SEC3 recognition is controlled by amino acids in the predicted  $\beta$  strand B and/or HV4 of V $\beta$ 3b. Antigen recognition by 5KC-73 was compared with that of a transfectant in which the amino acids at positions 18, 20, 22, 66, 68, 72, and 74 of the  $\beta$  chain had been replaced with the corresponding amino acids of V $\beta$ 17a. Both were challenged with either pigeon cyto *c* peptide 88-104 or various SAgs, and the IL-2 response was measured. Results are averaged from at least three experiments in each case.

(Fig. 1). Therefore, we transfected the  $\beta$  chain deletion variant of the V $\beta$ 3<sup>+</sup> hybridoma with a construct carrying a mutant V $\beta$ 3<sup>+</sup>  $\beta$  chain in which these seven amino acids were exchanged for those of V $\beta$ 17. The transfectants were tested for their ability to respond to various toxins and MTV SAgs as well as cyto  $c/IE^k$ . The results are shown in Fig. 3.

The transfectant with the wild-type  $V\beta3^+$   $\beta$  chain responded to cyto *c* peptides plus IE<sup>k</sup>, as well as to SEA, SEC3, and LBB.1, but not to CH12.1. Replacement of the amino acids in  $\beta$  strand B and in HV4 with those from V $\beta$ 17 had no effect on the response of the transfectant to cyto  $c/IE^k$ , SEA, or LBB.1. However, the response to SEC3 was reduced by these substitutions even more dramatically than seen with V $\beta$ 17 itself (Fig. 2). These results pinpointed amino acids in putative  $\beta$  strand B and/or HV4 as important in SEC3 recognition and confirmed that extensive nonconservative mutations in these regions have no effect on the recognition of conventional peptide antigen bound to MHC class II.

Perhaps unexpectedly, conversion of these seven amino acids of V $\beta$ 3 to those of V $\beta$ 17 did not transfer the V $\beta$ 17 reactivity to the MTV9 SAg of CH12.1, indicating that the amino acids in these positions may be necessary, but are not sufficient, for recognition on this MTV SAg.

To pinpoint the amino acids essential for SEC3 recognition we exchanged the three amino acids in  $\beta$  strand B and the four amino acids in HV4 separately and in addition exchanged pairs of amino acids at positions 66 and 74 and at positions 68 and 72 of HV4 separately. Again the transfectants were tested for their response to cyto c peptides presented with IE<sup>k</sup> and to SEA or SEC3 presented by the two antigenpresenting lines. The results are shown in Fig. 4. Mutation of the three amino acids in  $\beta$  strand B alone had no effect on the response to the cyto c peptide, SEA, or SEC3. However, exchange of the four amino acids in HV4 eliminated the response to SEC3 without affecting the response to SEA or to cyto c/IE<sup>k</sup>. Partial reduction of the response to SEC3 was seen when amino acid pair 66/74 was exchanged, and virtually a complete loss of response was seen with exchange of the 68/72 pair. These results identify HV4 as an essential part of the V $\beta$ 3 recognition site for SEC3.

An Amino Acid in VB3 Important for Cyto c/IE<sup>k</sup> Recognition. Pigeon cyto c peptide 88-104 is presented in  $H2^k$  mice by the IE<sup>k</sup> molecule and the response is dominated by T cells bearing V $\beta$ 3/V $\alpha$ 11 (47-49). Two allelic forms of the V $\beta$ 3 element have been identified (Fig. 1). Most mouse strains, such as B10.BR, carry the V $\beta$ 3b allele, which has a valine at position 29 predicted to lie in CDR1. A few strains (e.g., C57BR, SWR) carry the V $\beta$ 3a allele, which has a phenylalanine at this position. Gahm et al. and Fry and Matis (43, 49) have reported that H2<sup>k</sup> mice carrying V $\beta$ 3a fail to respond to cyto c with the usual dominant V $\beta$ 3/V $\alpha$ 11 clonotypes, suggesting that the valine at position 29 is critical for cyto c/IE<sup>k</sup> recognition by these clones. To test this idea directly we replaced this amino acid in the  $\beta$  chain of 5KC-73 with the phenylalanine of V $\beta$ 3a and tested the ability of the resulting hybridoma to respond to cyto c/IE<sup>k</sup> and to several superantigens. The results are shown in Fig. 5. The hybridoma bearing the wild-type V $\beta$ 3b<sup>+</sup>  $\beta$  chain responded



well to pigeon cyto c presented with IE<sup>k</sup>, to SEA and SEC3 presented by IE<sup>k</sup> or IA<sup>d</sup>, and to the MTV1/6 SAgs of LBB.1. Introduction of the V $\beta$ 3a phenylalanine at position 29 eliminated the response to pigeon cyto  $c/IE^k$ . Similar results were seen with the corresponding peptide from moth cyto c (data not shown). There were no effects of this mutation on the response to the bacterial or viral SAgs. These results are consistent with an important role for amino acid 29 of CDR1 in recognition of peptide/MHC, but not a number of SAg/MHC complexes.

Mapping Amino Acids on a Ig Vh Model for V $\beta$ . Fig. 6 shows a model of V $\beta$ 3 derived from the known structures of a number of Ig heavy chain V domains using the alignment of Chothia et al. (2). The four amino acids (66E, 68P, 72P, 74S) important for SEC3, but not cyto  $c/IE^k$ , recognition are predicted to lie on the side of the V $\beta$  domain in



Figure 5. Cyto  $c/IE^k$  recognition is controlled by an amino acid in predicted CDR1 of V $\beta$ 3b. Antigen recognition by 5KC-73 was compared with that of a transfectant in which the value at position 29 of V $\beta$ 3b had been replaced with the phenylalanine of V $\beta$ 3a. Both were challenged with either pigeon cyto c peptide 88–104 or various superantigens, and the IL-2 response was measured. Results are averaged from at least three experiments in each case.

**Figure 4.** SEC3 recognition is controlled by amino acids in predicted HV4 of V $\beta$ 3b. Antigen recognition was compared among a series of 5KC-73 transfectants in which various combinations of the amino acids at positions 18, 20, 22, 66, 68, 72, and 74 of the  $\beta$  chain had been replaced with the corresponding amino acids of V $\beta$ 17a. The transfectants were challenged with either pigeon cyto c peptide 88–104 or various SAgs, and the IL-2 response was measured. Results are averaged from the same experiments as those described in Figure 3.

HV4. The amino acids at positions 67 and 73 are both cysteines and predicted to be immediately adjacent to each other on opposite strands so that there is a high probability that they form a short disulfide loop. The formation of this loop would cause the side chains of amino acids 66, 68, 72, and 74 to point outwards towards the solvent, available for inter-



Figure 6. The structure of the 5KC-73  $\beta$  chain V domain based on Ig. The  $\alpha$ -carbon backbone of the 5KC-73  $\beta$  chain V domain was predicted using the coordinates of a set of Ig Vh domains (8–13) obtained from the Brookhaven Data Base (Upton, NY), the alignment of Chothia et al. (2), and software from the Biosym Corp. (San Diego, CA) running on a Silicon Graphics (Mountain View, CA) IRIS Indigo workstation. The view is of the solvent-exposed face composed of  $\beta$  strands B, D, and E as well as HV4 (colored medium is blue except for amino acids 66, 68, 72, and 74, which are dark blue). A disulfide bond was forced between the cysteines at positions 67 and 73. CDR1 is green (except the valine at position 29, which is red); CDR2 is yellow and CDR3 is orange. All other amino acids are light blue.

action with the SAg. The valine at position 29 essential for cyto  $c/IE^k$ , but not SAg, recognition is predicted to lie at the COOH-terminal end of CDR1 with its side chain pointed out toward the potential antigen/MHC ligand. Therefore, the model for the V $\beta$  domain based on Ig is consistent with the functional properties of the V $\beta$ 3 mutants and places the SAg and peptide/MHC interaction sites on different solventexposed faces of the protein.

# Discussion

The antigenic ligands for Ig and the TCR  $\alpha/\beta$  are very different. While almost all chemical structures are possible ligands for Ig, only peptide or protein antigens that combine with MHC molecules are potential ligands for TCR  $\alpha/\beta$ . Therefore, despite the similarities in gene organization and primary sequences between Ig and the TCR  $\alpha/\beta$  (1-7), it would not be surprising if the V domains of these two molecules had quite different structures. However, a number of authors have argued for an Ig-like structure for the TCR  $\alpha/\beta$ V domains based on the conservation of amino acids in positions critical for the backbone structure of Ig V domains (1-3). In addition, there is mounting direct evidence, including that presented here, supporting an Ig-like structure for the TCR  $\alpha/\beta$  with regions corresponding to the Ig CDR loops involved in peptide antigen/MHC recognition, some  $\beta$  strands buried between the V $\alpha$  and V $\beta$  and others together with HV4 forming another solvent-exposed face involved in SAg recognition.

For example, sites for potential N-linked glycosylation are frequent in mouse and human V $\alpha$  and V $\beta$  elements, and a number of these have been shown to be glycosylated in vivo (19, 50, 51). Glycosylated asparagines of course must be solvent exposed and should be rare on antigen-binding loops. The Ig model of the TCR  $\alpha/\beta$  predicts that these sites will lie almost exclusively on solvent-exposed  $\beta$  strands or loops, rather than CDR loops or buried  $\beta$  strands (C,C',F,G). Likewise, the few amino acids known to be involved in binding of anti-TCR  $\alpha/\beta$  mAbs are predicted by the Ig model to be on solvent-exposed loops (18, 19).

There are a number of examples now of amino acids that affect peptide antigen/MHC interaction, which lie in the predicted CDR regions. This is particularly true for CDR3 where alterations in the predicted CDR3 of the  $\alpha$  or  $\beta$  chain have been associated with changes in peptide antigen specificity (14-16). Recently, the predicted CDR1 region of V $\alpha$  has been implicated in the allo-MHC specificity of a TCR  $\alpha/\beta$  (17). We have identified residues at either end of V $\beta$  CDR1 that are important for antigen/MHC interaction. In a previous study (18) a threonine at position 24 of V $\beta$ 8.2 was found to be critical for recognition of a cOVA peptide with IA<sup>b</sup>, and in the current study a value at position 29 of V $\beta$ 3 was essential for response to cyto c plus IE<sup>k</sup>. This latter finding confirms the prediction of Gahm et al. (43).

There is also accumulating evidence that the predicted solvent-exposed HV4 of V $\beta$  plays an essential role in the recognition of various types of SAgs complexed to MHC class II. Previously, we have found that residues in this loop are critical for mouse V $\beta$ 8.2 interaction with Mls-1<sup>a</sup>, the SAg of MTV7 (18, 19). In this case residues on the adjacent  $\beta$ strand B were also found to be important. Casenave et al. (21) have identified an amino acid in the HV4 loop of V $\beta$ 17a (72Q) that controls in vivo deletion of V $\beta$ 17<sup>+</sup> T cells by an MTV SAg carried by C57BL/6 mice (mostly likely that of MTV9). We previously found HV4 to be important in the recognition of SEC2 and SEC3 by human V $\beta$ 13.2 (20), and now in the current study in the recognition of SEC3 by mouse V $\beta$ 3.

The data have not yet defined the role of MHC class II in SAg recognition. For conventional peptide antigens MHC restriction of antigen recognition is a function of the allelic amino acids involved both in antigen binding to MHC and in TCR  $\alpha/\beta$  interaction with the MHC portion of the ligand. The picture is not so clear for SAgs. Although SAg binding to MHC appears to be a prerequisite for TCR  $\alpha/\beta$  recognition (52-54), the recognition is not MHC restricted in the usual sense. A given TCR  $\alpha/\beta$  may recognize a particular SAg in association with many different allelic (or in some cases, isotypic or even xenogenic) forms of MHC class II (52–55), and upwardly pointing mutations in the  $\alpha$ -helical barrels of MHC, which interfere with TCR  $\alpha/\beta$  recognition of bound peptide antigen, often have no effect on recognition of bound SAg (56, 57). These findings in combination with the overriding importance of HV4, rather than the CDRs, in determining SAg interaction raise the possibility that there are few or no essential interactions between TCR  $\alpha/\beta$  and MHC during SAg recognition and that the main function of the MHC is to capture, orient, and perhaps conformationally alter the SAg, thus promoting interaction with the HV4 site.

Several observations could be used to argue against this view. Occasionally amino acid mutations in the TCR  $\alpha/\beta$ CDRs (18) and the MHC  $\alpha$  helices (57) eliminate T cell recognition of a particular SAg/MHC combination. Also sometimes TCR  $\alpha/\beta$  with the correct V $\beta$  element fails to recognize the appropriate SAg presented by a particular MHC molecule (29, 52), indicating involvement of  $\beta$  chain CDR3 and/or  $\alpha$  chain CDRs. These results indicate a close proximity between TCR  $\alpha/\beta$  CDRs and MHC during SAg recognition and could mean that interactions between the side chains of certain amino acids in the MHC and TCR  $\alpha/\beta$ contribute to binding. However, they are just as easily interpreted to mean that these side chain interactions are irrelevant except when they sterically interfere with docking of the HV4-binding site on V $\beta$  with the SAg bound to MHC. We thank Drs. R. Germain, L. Gilcher, and O. Kanagawa for cell lines and mAbs.

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