

REVIEW ARTICLE OPEN Immunomodulatory effects and mechanisms of distraction osteogenesis

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Distraction osteogenesis (DO) is widely used for bone tissue engineering technology. Immune regulations play important roles in the process of DO like other bone regeneration mechanisms. Compared with others, the immune regulation processes of DO have their distinct features. In this review, we summarized the immune-related events including changes in and effects of immune cells, immune-related cytokines, and signaling pathways at different periods in the process of DO. We aim to elucidated our understanding and unknowns about the immunomodulatory role of DO. The goal of this is to use the known knowledge to further modify existing methods of DO, and to develop novel DO strategies in our unknown areas through more detailed studies of the work we have done.

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INTRODUCTION

Distraction osteogenesis (DO) is an endogenous bone tissue engineering technology that utilizes inherent regenerative capacity to reconstruct or lengthen bone tissues.¹ In DO, osteotomy is performed on the bone that needs to be lengthened initially. After a short latency period, a distractor is used to distract the bone ends at an appropriate speed and frequency to spontaneously promote new natural bone regeneration in the distraction gap. This period is referred to as the distraction period. Next, this process follows a long period of consolidation to achieve new bone mineralization and remodeling.^{2,3} DO is superior to other techniques used for bone defect reconstruction because this osteogenetic process does not require bone tissue transplantation.¹ Dr. Gavriil Ilizarov, a Russian surgeon, formally proposed and popularized the DO technology in the 1950s and 1990s.²⁻ Subsequently, its application has expanded from the limb bones to the axial and craniofacial bones. Recently, DO has been widely used in orthopedic, oral, and craniofacial and plastic surgery for congenital or acquired limb disorders, congenital knee flexion contractures, huge bone defect secondary to bone tumors, infections or trauma, craniosynostosis, Pierre Robin sequence, hemifacial microsomia, and other craniofacial dysplasias or malformations. In some diseases, DO is the preferred treatment.4-16 However, it should not be disregarded that a long consolidation period may lead to undesirable outcomes, such as infection at the e needle site, pain syndrome, nonunion, excessive economic burden, psychological burden, etc., which limit the clinical application of the DO¹

To reduce complications and improve the treatment outcomes of DO, several efforts, such as combining bone tissue engineering elements including exogenous biological scaffolds, seed cells, and growth factors with DO, have been made.^{18–20} Moreover, some studies tried to use novel targetable molecules to develop related

adjuvant treatment methods in DO processes.²¹ The outcomes of these novel treatments have some disadvantages in terms of timing and duration of application.^{21,22} According to recent studies, bone tissue engineering products have immunoregulatory functions for controlling the immune microenvironment of bone regeneration sites and for reconstructing the tissue regeneration process.^{23,24} The roles of immune regulation in bone regeneration have been evaluated and used as a basis for applying bone tissue engineering technology.²⁵ Although the DO is also bone regeneration and has been recognized that the immune regulations are involved in the DO, the immune microenvironment and regulation mechanisms should be explored to successfully perform DO. This review aimed to identify all studies about immunomodulatory effects during DO and summarize the changes in and effects of immune-related cytokines, signaling pathways, and cells at different periods of DO processes. Moreover, it identified some related adjuvant treatments and provided new information about immune regulation mechanisms involved in DO.

LATENCY PERIOD

The latency period is similar to the initial phase of fracture. Immediately after osteotomy, hematoma forms at the osteotomy site, and inflammatory responses occur with the gathering and infiltration of different immune cells.²⁶ Pro-inflammatory cytokines, such as interleukin -1(IL-1) and interleukin-6 (IL-6), are released in large quantities to debride the osteotomy site. This mechanism can provide a favorable environment for angiogenesis and osteogenesis and can promote the initial migration, proliferation, and differentiation of mesenchymal stem cells (MSCs).^{26–28} Further, to promote preliminary soft callus formation, growth factors, such as transforming growth factor- β (TGF- β), bone morphogenetic protein (BMP), insulin growth factor, and

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Immunomodulatory effects and mechanisms of distraction osteogenesis Yang et al.

2



Fig. 1 Immune-related biological processes during the latency period of DO. Hematoma is formed at the osteotomy site immediately after the osteotomy, and inflammation occurs with the aggregation and infiltration of different immune cells. Pro-inflammatory cytokines are released to debride the osteotomy site and provide a favorable environment for angiogenesis and osteogenesis. MSCs initially migrate, proliferate, and differentiate in this environment. Growth factors, such as TGF- β , BMP, etc. are secreted by local inflammatory cells and MSCs. Eventually a soft callus with inflammatory cells, fibroblasts, osteogenic precursor cells, collagen, fibrin matrix, and spreading capillaries is formed at the osteotomy site

vascular endothelial growth factor (VEGF), are secreted locally by inflammatory cells and MSCs.^{27,29,30}

The immunomodulation process in the latency period of DO is similar to the well-studied healing inflammatory cascade reaction process in fractures. However, there are evident differences between them.³¹ In the initial phase of fracture healing, immediate inflammatory response occurs with the higher expression of IL-1, IL-6, and tumor necrosis factor-α (TNF-α), recruitment of inflammatory cells, and promotion of extracellular matrix secretion.^{32,33} After 2–3 days, the IL-1, IL-6, and TNF-α expressions rapidly decrease to an almost undetectable level in the next endochondral phase.²⁸ In contrast, only the expression of IL-1 and IL-6 increases after osteotomy during the latency period of DO. Meanwhile, that of TNF- α does not increase significantly.²⁸ This phenomenon is attributed to the fact that TNF- α mainly responds to severe trauma and that osteotomy is less traumatic than fracture. However, data about chronic ethanol exposure impairing DO in a tibial DO mouse model showed that the TNF signaling pathway axis is involved.³⁴ TNF receptor I-deficient mice can be free from the negative effects of chronic ethanol exposure in DO.^{35} Hence, TNF- α can be caused by other factors to participate in the DO process and affect bone regeneration.

After the inflammatory response and preliminary regeneration processes, a soft callus with inflammatory cells, fibroblasts, osteogenic precursor cells, collagen, fibrin matrix, and spreading capillaries is formed at the osteotomy site (Fig. 1).³⁶ DO requires an appropriate latency period because the formation of these soft calluses is a critical step for subsequent successful bone regenerations.

DISTRACTION PERIOD

The next phase is the distraction period, during which a distractor is applied to the osteotomy site to continuously stretch the osteotomy ends at an appropriate and fixed speed. Under distraction stimulation, the cartilage callus formed during the

latency period is absorbed, and a surprising amount of neovascularization migrates toward the central part of the distraction gap. Moreover, several multipotent stem cells infiltrate, proliferate, and differentiate with intramembranous ossification to produce an immature woven bone, and parts of the bone at the ends of the distraction gap mineralize and remodel.^{37–39} These phenomena are caused by a series of biochemical reactions in the tissue of the gap area with a slow and stable outward traction force. Mechanochemical transduction causes changes in the microenvironment of the distraction gap,^{40,41} including migration, infiltration, and differentiation of various cells, expression of proinflammatory cytokines, growth factors and other biologically active substances with specific temporal and spatial characteristics, and activation of numerous inflammatory and immunerelated signal pathways to regulate inflammatory responses and the immune microenvironment in the gap (Fig. 2).

Immune cells, especially macrophage plays essential roles in the distraction period

Numerous studies have shown that an appropriate early inflammatory response during fracture healing facilitates resistance to infection and recruitment of pluripotent stem cells. Further, excessive inflammatory response interferes with new bone regeneration, revascularization, and extracellular matrix mineralization.^{42–44} The transformation from the inflammatory microenvironment to an anti-inflammatory microenvironment is necessary for successful bone regeneration, which involves phenotype changes in several immune cells, including macrophages, and T and B cells, resulting in alterations in the expression of chemokines and cytokines.⁴⁵

Currently, it is recognized that the immunoregulation of osteogenesis is absolutely indispensable during the DO.⁴⁶ However, whether the transformation of immune cell proinflammatory phenotype to anti-inflammatory phenotype occurs during the DO process remains unclear. When will it happen? What is the effect of distraction stimulation on these processes?

Immunomodulatory effects and mechanisms of distraction osteogenesis Yang et al.



Fig. 2 Immune-related biological processes during the distraction period of DO. Relationship between immune cells, cytokines, and inflammation-related signal pathways in the distraction period of DO. Macrophages are polarized under stretch stimulation, thereby transforming from M1 to M2 phenotype, releasing the anti-inflammatory cytokines IL-10 and TGF-β, plus TGF-β that is secreted by osteoblasts and other precursor cells together regulate immunosuppressive microenvironment and promote the proliferation of osteoblasts. Simultaneously, the activation of YAP in macrophages promotes the secretion of BMP by macrophages. Then, BMP will take over the role of TGF-β to continuallypromote the proliferation of osteogenic precursor cells, leading to an effective differentiation into osteocytes. The role of BMP is achieved via the Smad signaling pathway. Moreover, BMP can activate the MAPK kinase TAK1 to activate the MAPK-related signaling pathways, leading to a higher secretion of IL-6 in osteoblasts. IL-6 will promote the recruitment and osteogenic differentiation of osteoblast-related cells. Osteoclasts are also active during the distraction period. That is, they participate in the absorption of actilage callus and the remodeling of new bones on both sides of the distraction gap. The expression of RANKL secreted by osteoblasts and osteocytes increases during the distraction period, and RANKL combines with RANK on osteoclast-related cells to promote the differentiation and activation of osteoclasts. Simultaneously, OPG expression is also upregulated, thereby inhibiting RANKL and regulating balance in bone resorption

There are only few studies discussed about these questions, and focused on the phenotypic transformation and effect of macrophages in the process of DO. Fan Zhang et al. revealed that the expression of CD68-positive macrophages increased in the distraction gap during the distraction period in a lower limb DO mouse model.⁴⁷ When using Saporin-CD11b to induce macrophage mortality and depletion, it could decrease IL-6, BMP2, and TGF-B secretions leading to lower expressions of osteogenesisrelated proteins. Hence, macrophages play an important role in DO.⁴⁷ They identified macrophages using the CD68-positive signal, and reached the conclusion that inflammatory macrophages accelerate mechanically induced bone regeneration.⁴ Nevertheless, CD68 cannot distinguish inflammatory (M1) from anti-inflammatory (M2) macrophages. Moreover, M2 macrophages are more likely to express BMP2 and TGF- β .⁴⁸ This finding must be further evaluated. Simultaneously, several in vitro studies showed that different mechanical stimuli had different effects on the phenotype and cytokine secretion of macrophages. However, they did not assess the role of these stimuli in the process of osteogenesis.⁴⁹⁻⁵¹ A recent in vitro experiment revealed that macrophages polarize to the M2 phenotype and produce antiinflammatory cytokines IL-10 and TGF-β to regulate the immune microenvironment under the induction of mechanical stretch.⁵² In addition, M2 can promote the osteogenic differentiation of bone marrow mescenchymal stem cells (BMSCs) under mechanical stretch stimulation via the YAP/BMP2 axis.⁵² Under stretch stimulation, a higher Wnt5a expression in the M2 activates the expression of the downstream YAP gene, which promotes the expression of BMP2 in macrophages. Then, BMP2 plays a role in

BMSC osteogenesis.⁵² Moreover, Wei et al. showed the polarization of macrophages toward the M2 phenotype and the interaction between macrophages and osteogenic precursor cells in a suture DO mouse model.⁵³ Results showed that chemokine C-C chemokine ligands (CCL2, CCL7, and CCL21) are produced, and they promote macrophage infiltration under stretch stimulation. Subsequently, macrophages polarize to the M2 phenotype and play an important role in promoting osteoblast differentiation of suture-derived stem cells. In addition, mechanical stretching induces macrophage polarization in vitro, which further validates that mechanical stretching has immunomodulatory effects on macrophage polarization.⁵³

These studies revealed that macrophages are recruited to the distraction gap under the action of CCLs. Subsequently, macrophages undergo M2 phenotypic polarization under a specific microenvironment and stretch stimuli. M2 macrophages interact with osteogenic precursor cells and promote osteogenic differentiation by secreting BMP2, TGF- β , and other cytokines, which is similar to the immune responses of macrophages in other bone regeneration processes. Several studies have revealed that macrophages polarize to the M2 phenotype and secrete TGF-β, IL-4, BMP-2, BMP-4, IL-6, VEGF, and other cytokines to promote bone formation during regeneration. 54,55 TGF- β and IL-4 can stimulate bone extracellular matrix secretion during bone regeneration.⁵⁶ BMP-2 and BMP-4 promote the osteogenic differentiation of MSCs by activating the BMP and Wnt signaling pathways during fracture healing.⁵⁵ The IL-6 family can recruit MSCs to the fracture site and induce osteoblast differentiation during fracture healing.⁵⁷ In DO, the macrophage immune

Immunomodulatory effects and mechanisms of distraction osteogenesis Yang et al.

4

response and its interaction with osteogenic precursor cells in the process of stretch must be assessed via detailed systematic investigations. Current studies showed that M2 macrophages have potential molecular mechanisms in promoting the osteogenic differentiation of osteogenic precursor cells in the distraction gap. Further, stretch stimulation plays an important immunomodulatory effect on macrophages, which may be the major difference in macrophage immune response between DO and bone healing process. The roles of T and B cells and other immune cells in the DO process should be also identified to completely understand the whole immune regulation network in the stretching process.

Regulatory roles of cytokines in the distraction period

IL-6, which is a pro-inflammatory cytokine, plays an important role in the inflammatory response during the latency period. Interestingly, it can be still secreted and form second lower peak by the elliptical cells, osteoblasts, and chondrocytes of the fibrous zone in rat tibial DO model until the end of the distraction period when mechanical stretch is applied.²⁸ Recent studies have shown that stretch stimulation is an independent cause of IL-6 upregulation.⁵⁸ However, IL-1 cannot be induced by stretch.⁵⁹ Therefore, it is particularly sensitive to mechanical stretch stimulation. Function assays confirmed that IL-6 promotes osteogenic differentiation from mature osteoblast cell lines.²⁸ Yuji Ando et al. revealed that IL-6 can also promote the migration and osteogenic differentiation of MSCs in a tibial DO mouse model.⁴⁶ Therefore, it plays an important role in DO. The IL-6 expression is upregulated, thereby inducing the phosphorylation of STAT3 and, ultimately, activating osterix, an osteogenic differentiation-related transcription factor, during the process of inducing osteogenic differentiation of human adipose-derived mesenchymal stem cells.⁶⁰ This signal axis may explain the specific mechanism of IL-6. However, whether this axis and other molecular crosstalk between the immune system and osteogenesis involved in DO was not identified.

Further, the pro-inflammatory effects of IL-6 are important.⁶¹ It can increase the recruitment of mononuclear immune cells in the distraction gap, and its negative effect is counteracted by the immunosuppressive effect of MSC secretions.⁴⁶ Therefore, the immunosuppressive response is important in the distraction period. Based on clinical practice, the plasma leukocyte counts and the CRP levels are significantly elevated during the latency period in patients who received DO treatments of the tibia and fibula. Eventually, they return to normal levels during the distraction period.⁶² This finding revealed the existence of immunosuppression in the distraction phase.

TGF-B1 is another important factor of DO. Several studies showed that the expression of TGF-β1 significantly increased in the distraction period in vitro and in vivo.^{59,63–67} In addition, it is widely distributed in the distraction gap, particularly in the region of active proliferation, and cells express TGF-B receptor type II in the same location.^{63–65} Further analyses of the spatial and temporal expressions of TGF-β1 showed that TGF-β1 expression increases to more than 2-folds than that of normal control rats in a mandibular DO model. Further, TGF-B1 is mainly secreted from osteoblasts, MSCs, and connective tissues surrounding the distraction gap during the distraction period.²⁹ The rabbit tibia, minipig mandibula, Chinese mountain goat tibia, sheep mandibula, and human mandibula DO models showed similar results.^{68–72} That is, stretch stimulation sustains the high expression of TGF-B1 in the distraction gap, particularly cells associated with osteogenesis. By contrast, TGF-β1 expression increased shortly after fracture healing.^{68,69} A higher TGF-β1 expression promotes osteoblast proliferation and extracellular matrix synthesis.^{68,71,72} Moreover, it is accompanied by a persistent trough in osteocalcin expression in the canine tibial distraction model. Therefore, TGF-β1 may inhibit osteoblast differentiation, decrease osteocalcin expression, and delay mineralization when TGF-B1 promotes the proliferation of osteoblasts to fill the distraction gap.65

Whether the number of TNF increases during the distraction phase is still debated.^{27,28} However, the expression of TNF superfamily members, NF-KB receptor-activating factor ligands (RANKL), and osteoprotegerin (OPG), which are important regulators of osteoclast activity, increased in DO.73 RANKL is a RANK ligand that is necessary for the differentiation and activation of osteoclasts, which can promote bone resorption.⁷⁴ Meanwhile, OPG is expressed in osteoblasts and distributed on the bone surface beneath osteoclasts to prevent excessive bone resorp-⁴ The regulation of osteoclast activity is necessary for the tion." bone regeneration and remodeling process of DO.73 In the mandibular DO rat model, the expression of RANKL steadily increased at the beginning of the distraction period.⁷⁵ Although the expression of OPG increased and reached the peak at the end of the distraction, the RANKL-to-OPG ratio was continually increasing.⁷⁵ After the RANKL expression became relatively dominant, the osteoclast activity increases accordingly, and the cartilage formed by natural healing during the latency period is absorbed. Moreover, bone regeneration in the gap mainly involves direct intramembranous ossification.⁷⁶

Signaling pathways involved in the distraction period

The focal adhesion kinase (FAK) signaling pathway is a complex, multi-pathway, multi-crossover signaling network that regulates cell migration, proliferation, and differentiation by activating intracellular PI3K/AKT, Ras/MAPK, and other inflammation-related signaling pathways in response to integrins, growth factors, and mechanical stimuli.⁷⁷ Ransom, R. C. et al. showed that the upregulation of the FAK signaling pathway and the subsequent activation of related gene regulation programs in stem cells during the distraction period resulted in the reversal of stem cells to primitive neural crest cells in the developmental state in a mandibular DO mouse model.⁷⁸ This is the basis of osteogenic differentiation of stem cells.⁷⁸ Silencing the FAK signaling pathway will impair the osteogenic differentiation of BMSCs in in vitro uniaxial stretch BMSC experiments.⁷⁹ FAK can partly achieve osteogenesis via the activation of the MAPK signaling pathway in a tibial DO rat model.⁸⁰ Genes correlated with the FAK-MAPK signaling pathway are significantly expressed in the middistraction period and are maintained up to the early consolidation period. Hence, they play an important role in the process of bone regeneration.⁸

P38 is one of the most important members of the MAPK family as it controls inflammation. Moreover, it is involved in new bone regeneration during the distraction period. P38 not only increases the volume of new bones in the distraction gap by promoting the osteogenic differentiation of MSCs but also promotes the recruitment of MSCs via the stromal cell-derived factor-1/CXCR4 (SDF-1/CXCR4) axis.81,82 The expression of SDF-1 does not significantly increase in the early distraction period. However, it is remarkably upregulated in the late distraction period, and it reaches the peak at the end of the distraction period, which is significantly higher than that in the fracture healing zone. The high level of SDF-1 can accurately recruit more MSCs to regions with a higher BMP2 expression to promote the optimal function of BMP-2, thereby leading to sufficient new bone formation. The directional recruitment of MSCs in the distraction space by SDF-1 is indispensable for successful osteogenic differentiation.⁸³ Moreover, SDF-1 is beneficial for the mineralization and remodeling of new bone by promoting the regeneration of blood vessels in the distraction space, which facilitates optimal blood flow during the consolidation phase. Coincidentally, P38 plays a key role in VEGF secretion and angiogenesis during DO.⁸⁴ The underlying mechanism may be due to the presence of CXCR4 in endothelial progenitor cells (EPCs) as well, in parallel to the recruitment of BMSCs, which recruits a large number of EPCs.^{82,85} In addition to the SDF-1/ CXCR4 axis, P38 promotes BMSC migration by controlling the

secretion of matrix metalloproteinases (MMPs) that degrade the extracellular matrix in an in vitro static strain experiment of rat BMSCs.⁸⁶ Several members of the MMP family are involved in the process of osteogenesis. MMP2 is significantly upregulated under stretch stimulation. It remarkably contributes to the osteogenic differentiation induced by stretch than other MMP members, and may have an essential role.^{58,87} Notably, MMP2 is important in the formation, proliferation, and migration of blood vessels.⁸⁸ Enhancing the migration ability of MSCs may be one of the multiple effects of MMP2 via the P368/MMP2 pathway.⁵⁸ The roles of all MMP family members and their interactions with other signaling molecules must be assessed in detail.

Moreover, ERK, another major member of the MAPK family, can promote the osteogenic differentiation of osteoblast precursor cells.⁸⁹ ERK1/2 is highly expressed in the MSCs in the early distraction period and is expressed by mast chondrocytes in the late distraction period.⁹⁰ In particular, the spatial and temporal expression of ERK1/2 is highly consistent with that of BMP2/4, and BMP can activate ERK and p38 via specific non-Smad pathways to promote osteogenic differentiation. Therefore, there is an association between the integrin-related signaling pathways and osteogenic protein expressions in the role of mechanotransduction and osteogenesis.^{90,91} However, the specific paths and mechanisms between the two remain unknown. Based on a previous study, BMP can activate transforming growth factor- β -activated kinase 1 (TAK1), a MAPK kinase kinase, which can then activate MAPK via cascade phosphorylation.⁹² In addition, the TGF superfamily activates TAK1. Then, TAK1 activates the MAPK signaling pathway to induce IL-6 expression in vitro mechanical stretching experiments on murine pre-osteoblast cell line (MC3T3-E1) and murine primary osteoblasts.⁹³

Thus, the FAK signaling pathway is activated under stretch stimulation. Then, the downstream Ras/MAPK signaling pathway is activated, and it plays a role in the process of MSC recruitment and osteogenic differentiation. Activated P38 upregulates the expressions of SDF-1 and MMPs, which are migration-related signal molecules. This then leads to the recruitment of more MSCs to a specific location to assist cytokines including BMP, which play important roles in bone regeneration. In addition, SDF-1 and MMPs synergistically promote vascular regeneration, proliferation, and migration. The activated ERK is essential in the successful differentiation of osteogenic precursor cells. Its underlying mechanism may be correlated with the members of the TGF superfamily including BMP.

PIK3/AKT, another downstream pathway of FAK, also plays an important role in the DO process. Both in vivo and in vitro experiments have shown that activation of the PI3K/AKT signaling pathway can promote the differentiation of MSCs into EPCs and accelerate bone regeneration by stimulating angiogenesis in DO.⁹⁴ Moreover, recent studies have revealed that AKT can prevent GSK-3β-mediated degradation of β-catenin by regulating the Wnt/β-catenin signaling pathway and can enhance cell viability, osteogenic differentiation, and angiogenesis gene expression of BMSCs via the AKT/GSK-3β/β-catenin axis.⁹⁵

The Wnt signaling pathway is an immune-inflammatory related signaling pathway, and its role in bone regeneration and metabolism under mechanical stress has long been well-known.^{96,97} This pathway is continually activated during the distraction period, and it peaks at the end of this period.^{80,98} The Wnt signaling pathway is more active in the distraction period, compared with the consolidation period, and it is mainly essential in the osteogenic differentiation of osteoblast precursor cells.^{80,98-100} Further, Wnt/ β -catenin participates in the differentiation of MSCs into vascular endothelial cells, which couple angiogenesis and osteogenesis.¹⁰¹ In addition, data about in vitro cyclic mechanical stretching experiments of osteogenic precursor cells showed that the non-canonical Wnt signaling pathway is involved in upregulating OPG expression to inhibit bone destruction.¹⁰²

The expression of sclerostin, a Wnt signaling pathway inhibitor, is downregulated in long bone axial load mouse experiments. Thus, the inhibitory effect of the Wnt signaling pathway is partly relieved to promote new bone regeneration.^{103,104} Data about the use of exogenous sclerostin-antibody revealed that antagonism via the inhibitory effect of the Wnt signaling pathway can maximize the activation of the Wnt signaling pathway during the distraction period in the DO model, and can facilitate the earlier expression of osteogenesis-related proteins in BMSCs. This results in more new bone production in the distraction gap.^{105,106} These gratifying animal experiments support the potential clinical application of sclerostin-antibody in DO. Recently, the Food and Drug Administration has approved the use of Romosozumab, a sclerostin monoclonal antibody, for the treatment of osteoporosis among postmenopausal women at high risk of fractures.^{107,108} The application of sclerostin antibody in DO should be further assessed.

NF-KB, a well-known downstream signaling molecule of AKT, is a classical immune and inflammatory response-related signaling pathway in bone regeneration processes, including fracture healing, which plays a negative role.¹⁰⁹ The NF- κ B signaling pathway can be activated in response to stretch stimulation in mouse pre-osteoblastic cells. Moreover, it interacts with the P38 signaling pathway and jointly upregulates the expression of BMP2/4 to promote osteogenic differentiation.¹¹⁰ However, the NF-kB signaling pathway activity is downregulated during the process of mechanically stimulating the osteogenic differentiation of human jaw bone marrow mesenchymal stem cells (hJBMMSCs).¹¹¹ The activation of the NF-κB signaling pathway via the overexpression of P65 could limit the osteogenic differentiation of these cells. Therefore, the NF-kB signaling pathway has a negative role in the process of mechanically stimulated osteogenic differentiation of MSCs.¹¹¹ However, a recent study showed that the activity of the NF-KB signaling pathway is upregulated, and it inhibits the osteogenic differentiation of osteogenic precursor cells, osteoblast-like MG-63 cells in a mechanical stimulation experiment.¹¹² Mammalian target of rapamycin (mTOR), a downstream molecule of AKT, is important for the proliferation and differentiation of osteoblasts. Simultaneously, it is significantly expressed in response to stretch stimulation. Hence, mTOR and NF-KB can work together to maintain osteogenesis at an appropriate level.¹¹² These in vitro experiments have different or even contradictory results, which may be correlated with different cells, mechanical stretching methods, and selected parameters. To date, there is no study about the in vivo DO models of the NF-KB signaling.

CONSOLIDATION PERIOD

After the required length of distraction has been achieved, the distraction is ceased. The new bone in the distraction gap comprises the central unmineralized zone, adjacent primary mineralized tissue zone, and peripheral remodeling braided zone.^{113,114} The following is a long period of consolidation for fusing the central unmineralized zone and completing the mineralization and subsequent remodeling of the new bone. Finally, the mature lamellar bone, which is similar to the normal structure, is generated in the distraction gap. During this period, the number of osteogenic-related cytokines and signaling pathway-related molecules, which are highly expressed in the distraction period, rapidly declines to achieve microenvironment transformation to promote bone remodeling (Fig. 3).^{27,83}

TGF- β 1 decreases during the consolidation phase but remains at a certain level. Further, it localizes mainly in osteoblasts and involves in the process of bone mineralization and remodeling. Then, it decreases to normal levels during the late consolidation period.^{29,115} Further, studies using several exogenous growth factors confirmed the role of TGF- β 1 during the consolidation period.^{116,117} Moreover, responses to TGF- β from different cells in 5



Fig. 3 Immune-related biological processes during the consolidation period of DO. The number of osteogenic-related cytokines and signaling pathway-related molecules, which highly expressed in the distraction period, rapidly declines to achieve microenvironment transformation to promote bone remodeling. RANKL/OPG plays an important role in bone remodeling. Moreover, TGF-β1 receptors rapidly decline in the fibrous area, leading a decline of cell proliferation capacity to enter differentiation and maturity. After sufficient remodeling and mineralization, new mature lamellar bone similar to the natural bone structure are formed

the distraction gap are modified due to changes in the temporal and spatial distribution of TGF- β receptors with the progression of bone maturation and mineralization.¹¹⁸ Analyses of the temporal and spatial expression of TGF- β R I (receptor for TGF- β I) in the consolidation period revealed that the receptor expression in the osteogenic area slightly decreases in the late consolidation period in a femoral DO rat model. Meanwhile, the rapid decline in the fibrous area indicates that the fibrous area loses its sensitivity to TGF- β I prematurely, and the cell proliferation capacity decreases to enter the stage of differentiation and maturity.¹¹⁸ TGF- β receptors are classified as types I and II.¹¹⁹ TGF- β receptor type I is distributed in the bone matrix, and type II is located in the intracellular space.⁷² The spatial and temporal expressions of type II receptors, which are significantly correlated with cell proliferation, are consistent with TGF- β activation. Excessive type I will inhibit cell proliferation and promote bone mineralization.⁷² Bone volume does not significantly change in the distraction gap when applying topical exogenous TGF- β . However, the mineralization degree and bone strength increase, and they are accompanied by the downregulation of type II receptors.⁷² Due to limitations in the

Table 1. Biological p	rocesses and immune-related events in the different periods of	DO		
Stage of DO	Biological processes	Ref.	Immune-related events and their function	Ref.
Latency period	Formation of hematoma	26,27,29,31,124,125	Various immune cells infiltrate and release pro-inflammatory cytokines such as IL-1 and IL-6 to debride the osteotomy site and promote the initial migration, proliferation, and differentiation of MSCs.	26-29,31,124-126
	Inflammatory response Formation of the outer cartilaginous callus adjacent to the periosteum and the soft callus in the gap		Inflammatory cells and MSCs release growth factors such as TGF- $\beta,$ BMP, IGF, and VEGF to promote preliminary soft callus formation.	
			BMP2 and BMP4 are secreted by immature chondrocytes, and their expression can significantly decline because chondrocytes mature and secretion is ceased.	
Distraction period	Absorption of the cartilage callus	37–39	Immunosuppressive response plays an important role in the distraction period.	46,62
	A surprising amount of neovascularization and spread toward the center of the distraction gap		The expression of IL-6 appears a second lower peak in response to mechanical stretch to stimulate intramembranous osteogenesis by promoting recruitment and osteogenic differentiation of osteogenic precursor cells.	28,46
	Formation of fibrous interzone		TGF- β 1 is continually highly expressed and is consistently distributed with type II receptors, thereby contributing to the proliferation of osteoblast precursor cells and the secretion of extracellular matrix.	29,65,68–72
			The expression of BMP and Smad, a downstream signaling pathway molecule, increases, thereby taking over the role of TGF- β and allowing a large number of osteogenic precursor cells to successfully differentiate into osteocytes, which play a role in both intramembranous and endochondral ossification.	71,91,126–132
	Multipotent stem cells infiltrate, proliferate, and differentiate with intramembranous ossification to produce immature woven bones		The RANK-to-OPG ratio continually increases, regulates the activity of osteoclasts, and promotes the absorption of cartilaginous callus that forms during the latency period.	73-76
	Mineralization and remodeling of parts of the bone at the ends of the distraction gap		Numerous inflammation and immune-related signal pathways, including FAK, MAPK, P38, ERK, Smad, TAK1, PIK3/AKT, Wnt, NF-ĸB, and mTOR respond to mechanical stimulation or cytokine signal transmission, thereby participating in angiogenesis and osteogenesis.	78,80-82,84-86,90,93,94, 98-106,109-112
			Macrophages are widely present in the distraction gap, and M2 phenotype polarization occurs, which promotes the osteogenic differentiation of osteogenic precursor cells.	47,52
Consolidation period	Fusion of the central unmineralized zone	27,83	Osteogenic-related cytokines and signal pathway factors that were highly expressed in the distraction period are rapidly downregulated.	27,83
	Complete mineralization of the new bone		The expression of TGF- β 1 is maintained at a certain level in the early consolidation period, and TGF- β 1 participates in bone maturation and mineralization.	29,115
	Remodeling		The Whrt signaling pathway is maintained at a certain level in the early consolidation period, and it plays an important role in the process of bone mineralization.	72,118
			The RANKL-to-OPG ratio continually increases and peaks in the late consolidation period. Hence, osteoclasts, which are essential for bone remodeling, become extremely active in the mid- and late consolidation periods.	97,98,120
			The expression of IL-1 β and TNF- α increases, and they participate in osteoclastogenesis in conjunction with RANKL.	75,76

specific markers of type I receptor at the time, whether type I receptor is upregulated cannot be confirmed.⁷² Therefore, the temporal and spatial expressions and the effects of both TGF-B receptors in the DO process must be further investigated.

The activity of the Wnt signaling pathway remains at a certain level in the early consolidation period and decreases in the midand late consolidation periods.⁹⁸ The role of Wnt in promoting bone mineralization in the process of bone healing is conclusive. Xuemei Wang et al. applied the gastric instillation of LiCL, which is an activator of the Wnt pathway in a tibia DO rat model. Results showed that the Wnt signaling pathway can increase plasm osteocalcin levels and the average mineral density and hardness of the new bone in the consolidation period. Therefore, the Wnt signaling pathway plays a role in promoting the mineralization of new bones during the consolidation period.¹²

The role of RANKL/OPG cannot be overlooked during the consolidation period. The expression of RANKL continually increases during the consolidation period, reaches its peak in the early consolidation period, and declines in the mid-consolidation period. Meanwhile, the expression of osteoprotegerin is maintained in the early and mid-consolidation period; then, it decreases.⁷⁵ The RANKL-to-OPG ratio continually elevates during the consolidation period, reaches its peak in the late consolidation period, and then decreases at the end of the consolidation period.⁷⁵ Osteoclasts in the bone surface and medullary cavity are active in the mid- and late consolidation periods, which are the main periods of bone resorption.75 Excess extra-structural new bone is resorbed, and it undergoes remodeling to form bone tissues that are similar to the normal structure.⁷⁵ In a mandibular DO rabbit model, the expressions of IL-1 β and TNF- α increase during the early consolidation period, and it is maintained until the mid- to late consolidation periods, which induces osteoclastogenesis and upregulates osteoclast activity in conjunction with RANKL.⁷⁶ A higher RANKL-to-OPG ratio is conducive to a certain degree for bone regeneration and remodeling. However, excessive RANKL expression will likely lead to excessive bone resorption and compromise perfect bone regeneration.¹²¹ In a unilateral tibial DO rabbit model, the application of bovine lactoferrin can increase bone mass in the distraction gap by increasing OPG expression while slightly reducing $RANKL^{122}$ A recent study showed that the RANK/RANKL pathway can be upregulated by applying PTH as a bioactive substance to the distraction gap. Moreover, new bone formation and remodeling are promoted, which decreases the consolidation period in a femoral DO rat model.¹ This may be attributed to the fact that PTH activates osteoclasts via RANKL and stimulates the production of osteoblasts, which facilitates the promotion and balance of osteoclasts and osteogenesis simultaneously.¹²³

CONCLUSION

DO is widely used for bone tissue engineering technology. Similar to other bone regeneration processes, immune regulation plays an important role in the DO process. However, as a unique osteogenesis method, its immune regulation mechanism has several distinct features. This study summarized immune-related events including changes in and effects of immune cells, immune-related cytokines, and signaling pathways at different periods in the DO process (Tables 1 and 2). With the use of the general bone healing process as a reference, the specific immune regulations in the process of DO were discussed. Notably, macrophage polarization, cytokine secretion, immunomodulatory roles, and crosstalk between macrophages and osteogenic precursor cells play important roles in DO. The current knowledge about immunomodulatory roles has been beneficial for DO in the current clinical practice. Nevertheless, based on this publication review, immunomodulatory mechanisms are more complex, as several molecules, different cell types, and multiple levels/steps of

Table 2. Changes in immune-related pro-inflammatory cytokines and growth factors during DO

	Latency period		Distraction period		Consolidation period			Ref.		
Items	Early	Late	Early	Late	Early	Middle	Late			
Pro-inflammatory cytokines										
IL-1	111				$\uparrow\uparrow$	1 1	1	28,76		
IL-6	111		$\uparrow\uparrow$	1 1				28		
TNF-α					$\uparrow\uparrow$	† †	1	28,76		
RANKL			$\uparrow\uparrow$	$\uparrow\uparrow\uparrow$	$\uparrow\uparrow\uparrow$	† †	† †	73,75,76		
OPG			↑	$\uparrow\uparrow$	$\uparrow\uparrow$	† †	1	73,75,76		
RANK-to- OPG ratio			↑	1	$\uparrow\uparrow$	$\uparrow \uparrow \uparrow$	<u> </u>	73,75,76		
Growth factors										
TGF-β1		$\uparrow\uparrow\uparrow$	$\uparrow \uparrow \uparrow$	<u> </u>	<u>†</u> †			29,65, 68– 72,115		
BMP2 and BMP4	↑ ↑		$\uparrow \uparrow \uparrow$	$\uparrow\uparrow\uparrow$	† †			71,91, 126–130		

regulations and balances are involved to achieve optimal/ effective new bone formation. There are still several unknown issues about DO. Therefore, more studies must be performed to develop novel molecular biological techniques and to optimally improve the current methods of DO.

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ADDITIONAL INFORMATION

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REFERENCES

- 1. Hamdy, R. C., Rendon, J. S. & Tabrizian, M. Distraction osteogenesis and its challenges in bone regeneration. Bone Regeneration (ed. Tal, H.) Ch. 8, 177-204 (InTech, 2012).
- 2. Ilizarov, G. A. The tension-stress effect on the genesis and growth of tissues: Part I. The influence of stability of fixation and soft-tissue preservation. Clin. Orthop. Relat. Res. 238, 249-281 (1989).
- 3. Ilizarov, G. A. The tension-stress effect on the genesis and growth of tissues: Part II. The influence of the rate and frequency of distraction. Clin. Orthop. Relat. Res. 239, 263-285 (1989).
- 4. Gubin, A. V., Borzunov, D. Y. & Malkova, T. A. The Ilizarov paradigm: thirty years with the Ilizarov method, current concerns and future research. Int. Orthop. 37, 1533-1539 (2013).
- 5. Birch, J. G. & Samchukov, M. L. Use of the Ilizarov method to correct lower limb deformities in children and adolescents. J. Am. Acad. Orthop. Surg. 12, 144-154 (2004)
- 6. Abula, A. et al. Reconstruction of soft tissue defects and bone loss in the tibia by flap transfer and bone transport by distraction osteogenesis: a case series and our experience. Ann. Plast. Surg. 84, S202-s207 (2020).
- 7. Yushan, M. et al. Bifocal or trifocal (double-level) bone transport using unilateral rail system in the treatment of large tibial defects caused by infection: a retrospective study. Orthop. Surg. 12, 184-193 (2020).
- 8. Rohilla, R., Wadhwani, J., Devgan, A., Singh, R. & Khanna, M. Prospective randomised comparison of ring versus rail fixator in infected gap nonunion of tibia treated with distraction osteogenesis. Bone Jt. J. 98-b, 1399-1405 (2016).
- 9. Ajmera, A., Verma, A., Agrawal, M., Jain, S. & Mukherjee, A. Outcome of limb reconstruction system in open tibial diaphyseal fractures. Indian J. Orthop. 49, 429-435 (2015)
- 10. Makhdoom, A., Qureshi, P. A., Jokhio, M. F. & Siddiqui, K. A. Resistant clubfoot deformities managed by Ilizarov distraction histogenesis. Indian J. Orthop. 46. 326-332 (2012).

- Gubin, A. V., Borzunov, D. Y., Marchenkova, L. O., Malkova, T. A. & Smirnova, I. L. Contribution of G.A. Ilizarov to bone reconstruction: historical achievements and state of the art. *Strategies Trauma Limb Reconstr.* **11**, 145–152 (2016).
- Carlson, A. R. & Taylor, J. A. Posterior vault distraction osteogenesis: indications and expectations. *Child's Nerv. Syst.* 37, 3119–3125 (2021).
- Morrison, K. A., Collares, M. V. & Flores, R. L. Robin sequence: neonatal mandibular distraction. *Clin. Plast. Surg.* 48, 363–373 (2021).
- Shakir, S. & Bartlett, S. P. Modern mandibular distraction applications in hemifacial microsomia. *Clin. Plast. Surg.* 48, 375–389 (2021).
- Hopper, R. A. et al. Thirty years later: what has craniofacial distraction osteogenesis surgery replaced? *Plast. Reconstr. Surg.* 145, 1073e–1088e (2020).
- Hatefi, S. et al. Review of automatic continuous distraction osteogenesis devices for mandibular reconstruction applications. *Biomed. Eng. Online* 19, 17 (2020).
- Farr, S., Mindler, G., Ganger, R. & Girsch, W. Bone lengthening in the pediatric upper extremity. J. Bone Jt. Surg. Am. 98, 1490–1503 (2016).
- Makhdom, A. M., Nayef, L., Tabrizian, M. & Hamdy, R. C. The potential roles of nanobiomaterials in distraction osteogenesis. *Nanomed. Nanotechnol. Biol. Med.* 11, 1–18 (2015).
- Konofaos, P., Kashyap, A., Ver & Halen, J. Biomedical approaches to improve bone healing in distraction osteogenesis: a current update and review. *Biomedizinische Tech. Biomed. Eng.* 59, 177–183 (2014).
- Morcos, M. W., Al-Jallad, H. & Hamdy, R. Comprehensive review of adipose stem cells and their implication in distraction osteogenesis and bone regeneration. *BioMed. Res. Int.* 2015, 842975 (2015).
- Alzahrani, M. M., Anam, E., AlQahtani, S. M., Makhdom, A. M. & Hamdy, R. C. Strategies of enhancing bone regenerate formation in distraction osteogenesis. *Connect. Tissue Res.* 59, 1–11 (2018).
- Yang, Y., Lin, S., Wang, B., Gu, W. & Li, G. Stem cell therapy for enhancement of bone consolidation in distraction osteogenesis: a contemporary review of experimental studies. *Bone Jt. Res.* 6, 385–390 (2017).
- Franz, S., Rammelt, S., Scharnweber, D. & Simon, J. C. Immune responses to implants - a review of the implications for the design of immunomodulatory biomaterials. *Biomaterials* 32, 6692–6709 (2011).
- Veiseh, O. et al. Size- and shape-dependent foreign body immune response to materials implanted in rodents and non-human primates. *Nat. Mater.* 14, 643–651 (2015).
- Ouyang, L., Cao, J., Dai, Q. & Qiu, D. New insight of immuno-engineering in osteoimmunomodulation for bone regeneration. *Regenerative Ther.* 18, 24–29 (2021).
- Alman, B. A., Kelley, S. P. & Nam, D. Heal thyself: using endogenous regeneration to repair bone. *Tissue Eng. B: Rev.* 17, 431–436 (2011).
- Ai-Aql, Z. S., Alagl, A. S., Graves, D. T., Gerstenfeld, L. C. & Einhorn, T. A. Molecular mechanisms controlling bone formation during fracture healing and distraction osteogenesis. J. Dent. Res. 87, 107–118 (2008).
- Cho, T. J. et al. Expression and role of interleukin-6 in distraction osteogenesis. *Calcif. Tissue Int.* 80, 192–200 (2007).
- Mehrara, B. J. et al. Rat mandibular distraction osteogenesis: II. Molecular analysis of transforming growth factor beta-1 and osteocalcin gene expression. *Plast. Reconstr. Surg.* **103**, 536–547 (1999).
- Hvid, I., Horn, J., Huhnstock, S. & Steen, H. The biology of bone lengthening. J. Children's Orthop. 10, 487–492 (2016).
- Runyan, C. M. & Gabrick, K. S. Biology of bone formation, fracture healing, and distraction osteogenesis. J. Craniofacial Surg. 28, 1380–1389 (2017).
- Kon, T. et al. Expression of osteoprotegerin, receptor activator of NF-kappaB ligand (osteoprotegerin ligand) and related proinflammatory cytokines during fracture healing. J. Bone Miner. Res. 16, 1004–1014 (2001).
- Marsell, R. & Einhorn, T. A. The biology of fracture healing. *Injury* 42, 551–555 (2011).
- Wahl, E. C. et al. Chronic ethanol exposure inhibits distraction osteogenesis in a mouse model: role of the TNF signaling axis. *Toxicol. Appl. Pharmacol.* 220, 302–310 (2007).
- Wahl, E. C. et al. Distraction osteogenesis in TNF receptor 1 deficient mice is protected from chronic ethanol exposure. *Alcohol (Fayettev., NY)* 46, 133–138 (2012).
- 36. Alshahrani, I. Biomolecular phases in transverse palatal distraction: a review. *Saudi J. Biol. Sci.* **25**, 1322–1325 (2018).
- Ernst, N. & Adolphs, N. Role of distraction osteogenesis in craniomaxillofacial surgery. *Innovative Surgical Sci.* 1, 97–103 (2016).
- Dhaliwal, K., Kunchur, R. & Farhadieh, R. Review of the cellular and biological principles of distraction osteogenesis: an in vivo bioreactor tissue engineering model. J. Plast. Reconstr. Aesthetic Surg. 69, e19–26 (2016).
- Choi, I. H., Chung, C. Y., Cho, T. J. & Yoo, W. J. Angiogenesis and mineralization during distraction osteogenesis. J. Korean Med. Sci. 17, 435–447 (2002).

- Morgan, E. F., Longaker, M. T. & Carter, D. R. Relationships between tissue dilatation and differentiation in distraction osteogenesis. *Matrix Biol.: J. Int. Soc. Matrix Biol.* 25, 94–103 (2006).
 Matrix Biol. 26, 74–103 (2006).
 - Nomura, S. & Takano-Yamamoto, T. Molecular events caused by mechanical stress in bone. *Matrix Biol.: J. Int. Soc. Matrix Biol.* 19, 91–96 (2000).
 - Liu, H., Li, D., Zhang, Y. & Li, M. Inflammation, mesenchymal stem cells and bone regeneration. *Histochemistry Cell Biol.* 149, 393–404 (2018).
 - Gerstenfeld, L. C. et al. Impaired fracture healing in the absence of TNF-alpha signaling: the role of TNF-alpha in endochondral cartilage resorption. *J. Bone Miner. Res.* 18, 1584–1592 (2003).
 - Schmidt-Bleek, K. et al. Inflammatory phase of bone healing initiates the regenerative healing cascade. *Cell Tissue Res.* 347, 567–573 (2012).
 - Tsukasaki, M. & Takayanagi, H. Osteoimmunology: evolving concepts in boneimmune interactions in health and disease. *Nat. Rev. Immunol.* 19, 626–642 (2019).
 - Ando, Y. et al. Stem cell-conditioned medium accelerates distraction osteogenesis through multiple regenerative mechanisms. *Bone* 61, 82–90 (2014).
 - Zhang, F. et al. Inflammatory macrophages facilitate mechanical stress-induced osteogenesis. *Aging* 12, 3617–3625 (2020).
 - Wang, L. X., Zhang, S. X., Wu, H. J., Rong, X. L. & Guo, J. M2b macrophage polarization and its roles in diseases. J. Leukoc. Biol. 106, 345–358 (2019).
 - Maruyama, K., Nemoto, E. & Yamada, S. Mechanical regulation of macrophage function - cyclic tensile force inhibits NLRP3 inflammasome-dependent IL-1β secretion in murine macrophages. *Inflamm. Regeneration* **39**, 3 (2019).
 - Maruyama, K. et al. Cyclic stretch negatively regulates IL-1β secretion through the inhibition of NLRP3 inflammasome activation by attenuating the AMP kinase pathway. Front. Physiol. 9, 802 (2018).
 - 51. Shan, S. et al. Mechanical stretch promotes tumoricidal M1 polarization via the FAK/NF-κB signaling pathway. *FASEB J.* **33**, 13254–13266 (2019).
 - Dong, L., et al. Mechanical stretch induces osteogenesis through the alternative activation of macrophages. J. Cell. Physiol. 236, 6376–6390 (2021).
 - Liang, W. et al. Polarized M2 macrophages induced by mechanical stretching modulate bone regeneration of the craniofacial suture for midfacial hypoplasia treatment. *Cell Tissue Res.* 386, 585–603 (2021).
 - Schlundt, C. et al. The multifaceted roles of macrophages in bone regeneration: A story of polarization, activation and time. *Acta Biomaterialia* 133, 46–57 (2021).
 - 55. Pajarinen, J. et al. Mesenchymal stem cell-macrophage crosstalk and bone healing. *Biomaterials* **196**, 80–89 (2019).
 - 56. Takayanagi, H. Osteoimmunology: shared mechanisms and crosstalk between the immune and bone systems. *Nat. Rev. Immunol.* **7**, 292–304 (2007).
 - Guihard, P. et al. Oncostatin m, an inflammatory cytokine produced by macrophages, supports intramembranous bone healing in a mouse model of tibia injury. *Am. J. Pathol.* **185**, 765–775 (2015).
 - Chen, J., Hua, J. & Song, W. Screen key genes associated with distractioninduced osteogenesis of stem cells using bioinformatics methods. *Int. J. Mol. Sci.* 22, 6505 (2021).
 - Cillo, J. E. Jr., Gassner, R., Koepsel, R. R. & Buckley, M. J. Growth factor and cytokine gene expression in mechanically strained human osteoblast-like cells: implications for distraction osteogenesis. *Oral. Surg. Oral. Med. Oral. Pathol. Oral. Radiol. Endod.* **90**, 147–154 (2000).
 - Huh, J. E. & Lee, S. Y. IL-6 is produced by adipose-derived stromal cells and promotes osteogenesis. *Biochim. Biophys. Acta* 1833, 2608–2616 (2013).
 - Hunter, C. A. & Jones, S. A. IL-6 as a keystone cytokine in health and disease. *Nat. Immunol.* 16, 448–457 (2015).
 - Lee, D. Y. et al. Distraction osteogenesis induces endothelial progenitor cell mobilization without inflammatory response in man. *Bone* 46, 673–679 (2010).
 - Eingartner, C. et al. Growth factors in distraction osteogenesis. Immunohistological pattern of TGF-beta1 and IGF-I in human callus induced by distraction osteogenesis. *Int. Orthop.* 23, 253–259 (1999).
 - Holbein, O., Neidlinger-Wilke, C., Suger, G., Kinzl, L. & Claes, L. Ilizarov callus distraction produces systemic bone cell mitogens. *J. Orthop. Res.* 13, 629–638 (1995).
 - Lammens, J., Liu, Z., Aerssens, J., Dequeker, J. & Fabry, G. Distraction bone healing versus osteotomy healing: a comparative biochemical analysis. *J. Bone Miner. Res.* 13, 279–286 (1998).
 - Zhuang, H. et al. Mechanical strain-induced proliferation of osteoblastic cells parallels increased TGF-beta 1 mRNA. *Biochem. Biophys. Res. Commun.* 229, 449–453 (1996).
 - Weiss, S., Baumgart, R., Jochum, M., Strasburger, C. J. & Bidlingmaier, M. Systemic regulation of distraction osteogenesis: a cascade of biochemical factors. *J. Bone Miner. Res.* 17, 1280–1289 (2002).
 - Yeung, H. Y., Lee, K. M., Fung, K. P. & Leung, K. S. Sustained expression of transforming growth factor-beta1 by distraction during distraction osteogenesis. *Life Sci.* 71, 67–79 (2002).
 - 69. Çakır-Özkan, N., Eyibilen, A., Özkan, F., Gülbahar, M. Y. & Kabak, Y. B. Immunohistochemical analysis of reconstructed sheep mandibles: transport distraction

osteogenesis versus autogenous bone grafting. J. Oral. Maxillofac. Surg. 69, 1248–1254 (2011).

- Haque, T., Amako, M., Nakada, S., Lauzier, D. & Hamdy, R. C. An immunohistochemical analysis of the temporal and spatial expression of growth factors FGF 1, 2 and 18, IGF 1 and 2, and TGFbeta1 during distraction osteogenesis. *Histol. Histopathol.* 22, 119–128 (2007).
- Yates, K. E., Troulis, M. J., Kaban, L. B. & Glowacki, J. IGF-I, TGF-beta, and BMP-4 are expressed during distraction osteogenesis of the pig mandible. *Int. J. Oral. Maxillofac. Surg.* **31**, 173–178 (2002).
- Ozkan, K. et al. The effect of transforming growth factor beta1 (TGF-beta1) on the regenerate bone in distraction osteogenesis. *Growth Factors (Chur, Switz.)* 25, 101–107 (2007).
- Pérez-Sayáns, M., Somoza-Martín, J. M., Barros-Angueira, F., Rey, J. M. & García-García, A. RANK/RANKL/OPG role in distraction osteogenesis. *Oral. Surg. Oral. Med. Oral. Pathol. Oral. Radiol. Endod.* **109**, 679–686 (2010).
- Boyce, B. F. & Xing, L. Functions of RANKL/RANK/OPG in bone modeling and remodeling. Arch. Biochem. Biophys. 473, 139–146 (2008).
- Zhu, W. Q., Wang, X., Wang, X. X. & Wang, Z. Y. Temporal and spatial expression of osteoprotegerin and receptor activator of nuclear factor -kappaB ligand during mandibular distraction in rats. J. Cranio-Maxillo-Facial Surg. 35, 103–111 (2007).
- Wang, L. C., Takahashi, I., Sasano, Y., Sugawara, J. & Mitani, H. Osteoclastogenic activity during mandibular distraction osteogenesis. *J. Dent. Res.* 84, 1010–1015 (2005).
- 77. Urciuoli, E. & Peruzzi, B. Involvement of the FAK network in pathologies related to altered mechanotransduction. *Int. J. Mol. Sci.* **21**, 9426 (2020).
- Ransom, R. C. et al. Mechanoresponsive stem cells acquire neural crest fate in jaw regeneration. *Nature* 563, 514–521 (2018).
- Hu, P. et al. Fak silencing impairs osteogenic differentiation of bone mesenchymal stem cells induced by uniaxial mechanical stretch. *J. Dent. Sci.* 14, 225–233 (2019).
- Song, J. et al. Fak-Mapk, Hippo and Wnt signalling pathway expression and regulation in distraction osteogenesis. *Cell Prolif.* 51, e12453 (2018).
- Yang, Z. H. et al. Targeting P38 pathway regulates bony formation via MSC Recruitment during mandibular distraction osteogenesis in rats. *Int. J. Med. Sci.* 13, 783–789 (2016).
- Xu, J. et al. Effect of SDF-1/Cxcr4 signaling antagonist AMD3100 on bone mineralization in distraction osteogenesis. *Calcif. Tissue Int.* 100, 641–652 (2017).
- Alzahrani, M. M., Anam, E. A., Makhdom, A. M., Villemure, I. & Hamdy, R. C. The effect of altering the mechanical loading environment on the expression of bone regenerating molecules in cases of distraction osteogenesis. *Front. Endocrinol.* 5, 214 (2014).
- Kim, I. S., Song, Y. M. & Hwang, S. J. Osteogenic responses of human mesenchymal stromal cells to static stretch. J. Dent. Res. 89, 1129–1134 (2010).
- Fujio, M. et al. Stromal cell-derived factor-1 enhances distraction osteogenesismediated skeletal tissue regeneration through the recruitment of endothelial precursors. *Bone* 49, 693–700 (2011).
- Yang, Z. et al. The mechanically activated p38/MMP-2 signaling pathway promotes bone marrow mesenchymal stem cell migration in rats. *Arch. Oral. Biol.* 76, 55–60 (2017).
- Kasper, G. et al. Matrix metalloprotease activity is an essential link between mechanical stimulus and mesenchymal stem cell behavior. *Stem Cells (Dayt., Ohio)* 25, 1985–1994 (2007).
- Almalki, S. G. & Agrawal, D. K. Effects of matrix metalloproteinases on the fate of mesenchymal stem cells. *Stem Cell Res. Ther.* 7, 129 (2016).
- Son, H. E. & Jang, W. G. Cip2A modulates osteogenic differentiation via the ERK-Runx2 pathway in MG63 cells. *BioFactors (Oxford, England)* 47, 658–654 (2021).
- Rhee, S. T., El-Bassiony, L. & Buchman, S. R. Extracellular signal-related kinase and bone morphogenetic protein expression during distraction osteogenesis of the mandible: in vivo evidence of a mechanotransduction mechanism for differentiation and osteogenesis by mesenchymal precursor cells. *Plast. Reconstr. Surg.* 117, 2243–2249 (2006).
- Haque, T. et al. Characterizing the BMP pathway in a wild type mouse model of distraction osteogenesis. *Bone* 42, 1144–1153 (2008).
- Yamaguchi, K. et al. Identification of a member of the MAPKKK family as a potential mediator of TGF-beta signal transduction. *Science* 270, 2008–2011 (1995).
- Fukuno, N. et al. TGF-β-activated kinase 1 mediates mechanical stress-induced IL-6 expression in osteoblasts. *Biochem. Biophys. Res. Commun.* 408, 202–207 (2011).
- Mi, J. et al. Calcitonin gene-related peptide enhances distraction osteogenesis by increasing angiogenesis. *Tissue Eng. A* 27, 87–102 (2021).
- Wang, F. et al. Accelerated bone regeneration by astragaloside IV through stimulating the coupling of osteogenesis and angiogenesis. *Int. J. Biol. Sci.* 17, 1821–1836 (2021).
- Hens, J. R. et al. TOPGAL mice show that the canonical Wnt signaling pathway is active during bone development and growth and is activated by mechanical loading in vitro. J. Bone Miner. Res. 20, 1103–1113 (2005).

- Schupbach, D., Comeau-Gauthier, M., Harvey, E. & Merle, G. Wnt modulation in bone healing. *Bone* 138, 115491 (2020).
- Kasaai, B. et al. Spatial and temporal localization of WNT signaling proteins in a mouse model of distraction osteogenesis. J. Histochem. Cytochem. 60, 219–228 (2012).
- Wang, X. et al. Wnt/β-catenin signaling is required for distraction osteogenesis in rats. Connect. Tissue Res. 59, 45–54 (2018).
- Li, F. F. et al. Alterations in β-catenin/E-cadherin complex formation during the mechanotransduction of Saos-2 osteoblastic cells. *Mol. Med. Rep.* 18, 1495–1503 (2018).
- 101. He, X. et al. Hypoxia improved vasculogenesis in distraction osteogenesis through Mesenchymal-Epithelial transition (MET), Wnt/β-catenin signaling pathway, and autophagy. Acta Histochemica **122**, 151593 (2020).
- Yu, H. C. et al. Mechanical stretching induces osteoprotegerin in differentiating C2C12 precursor cells through noncanonical Wnt pathways. J. Bone Miner. Res. 25, 1128–1137 (2010).
- Robling, A. G. et al. Mechanical stimulation of bone in vivo reduces osteocyte expression of Sost/sclerostin. J. Biol. Chem. 283, 5866–5875 (2008).
- 104. Moustafa, A. et al. Mechanical loading-related changes in osteocyte sclerostin expression in mice are more closely associated with the subsequent osteogenic response than the peak strains engendered. Osteoporos. Int. 23, 1225–1234 (2012).
- 105. Makhdom, A. M., Rauch, F., Lauzier, D. & Hamdy, R. C. The effect of systemic administration of sclerostin antibody in a mouse model of distraction osteogenesis. J. Musculoskelet. Neuronal Interact. 14, 124–130 (2014).
- Alzahrani, M. M. et al. Assessment of the effect of systemic delivery of sclerostin antibodies on Wnt signaling in distraction osteogenesis. *J. Bone Miner. Metab.* 36, 373–382 (2018).
- Morrell, A. E., Robinson, S. T., Ke, H. Z., Holdsworth, G. & Guo, X. E. Osteocyte mechanosensing following short-term and long-term treatment with sclerostin antibody. *Bone* 149, 115967 (2021).
- Rauner, M., Taipaleenmäki, H., Tsourdi, E. & Winter, E. M. Osteoporosis treatment with anti-sclerostin antibodies-mechanisms of action and clinical application. J. Clin. Med. 10, (2021).
- Krum, S. A., Chang, J., Miranda-Carboni, G. & Wang, C. Y. Novel functions for NFκB: inhibition of bone formation. *Nat. Rev. Rheumatol.* 6, 607–611 (2010).
- Wang, L. et al. Involvement of p38MAPK/NF-κB signaling pathways in osteoblasts differentiation in response to mechanical stretch. *Ann. Biomed. Eng.* 40, 1884–1894 (2012).
- 111. Chen, X. et al. Mechanical stretch-induced osteogenic differentiation of human jaw bone marrow mesenchymal stem cells (hJBMMSCs) via inhibition of the NFκB pathway. *Cell Death Dis.* **9**, 207 (2018).
- Wang, D. et al. The interactions between mTOR and NF-κB: a novel mechanism mediating mechanical stretch-stimulated osteoblast differentiation. J. Cell. Physiol. (2020).
- 113. Percival, C. J. & Richtsmeier, J. T. Angiogenesis and intramembranous osteogenesis. *Dev. Dyn.* **242**, 909–922 (2013).
- Rachmiel, A., Rozen, N., Peled, M. & Lewinson, D. Characterization of midface maxillary membranous bone formation during distraction osteogenesis. *Plast. Reconstr. Surg.* 109, 1611–1620 (2002).
- 115. Xie, M. K., Hu, C. B., Zhou, B. & Wu, G. P. Effect of gene transfection timing on TGF-β1 expression in rabbit mandibular distraction gap. *Genet. Mol. Res.* 16, gmr16029330 (2017).
- 116. Bernstein, A., Mayr, H. O. & Hube, R. Can bone healing in distraction osteogenesis be accelerated by local application of IGF-1 and TGF-beta1? J. Biomed. Mater. Res. B: Appl. Biomater. 92, 215–225 (2010).
- Wu, G., Zhou, B., Hu, C. & Li, S. Gene expression of osteogenic factors following gene therapy in mandibular lengthening. *J. Craniofacial Surg.* 26, 378–381 (2015).
- 118. Siwicka, K. A., Kitoh, H., Kawasumi, M. & Ishiguro, N. Spatial and temporal distribution of growth factors receptors in the callus: implications for improvement of distraction osteogenesis. *Nagoya J. Med. Sci.* **73**, 117–127 (2011).
- Moustakas, A., Pardali, K., Gaal, A. & Heldin, C. H. Mechanisms of TGF-beta signaling in regulation of cell growth and differentiation. *Immunol. Lett.* 82, 85–91 (2002).
- 120. Wang, X. et al. Systemic administration of lithium improves distracted bone regeneration in rats. *Calcif. Tissue Int.* **96**, 534–540 (2015).
- 121. Liu, H., Luo, T., Tan, J., Li, M. & Guo, J. 'Osteoimmunology' offers new perspectives for the treatment of pathological bone loss. *Curr. Pharm. Des.* 23, 6272–6278 (2017).
- 122. Li, W., Zhu, S. & Hu, J. Bone regeneration is promoted by orally administered bovine lactoferrin in a rabbit tibial distraction osteogenesis model. *Clin. Orthop. Relat. Res.* **473**, 2383–2393 (2015).
- 123. Leiblein, M. et al. Do antiosteoporotic drugs improve bone regeneration in vivo? *Eur. J. Trauma Emerg. Surg.* **46**, 287–299 (2020).
- Cope, J. B. & Samchukov, M. L. Regenerate bone formation and remodeling during mandibular osteodistraction. *Angle Orthod.* **70**, 99–111 (2000).

- Jazrawi, L. M. et al. Bone and cartilage formation in an experimental model of distraction osteogenesis. J. Orthop. Trauma 12, 111–116 (1998).
- 126. Sato, M. et al. Mechanical tension-stress induces expression of bone morphogenetic protein (BMP)-2 and BMP-4, but not BMP-6, BMP-7, and GDF-5 mRNA, during distraction osteogenesis. J. Bone Miner. Res. 14, 1084–1095 (1999).
- Rauch, F. et al. Temporal and spatial expression of bone morphogenetic protein-2, -4, and -7 during distraction osteogenesis in rabbits. *Bone* 26, 611–617 (2000).
- 128. Campisi, P. et al. Expression of bone morphogenetic proteins during mandibular distraction osteogenesis. *Plast. Reconstr. Surg.* **111**, 201–208 (2003). discussion 209-210.
- 129. Farhadieh, R. D., Gianoutsos, M. P., Yu, Y. & Walsh, W. R. The role of bone morphogenetic proteins BMP-2 and BMP-4 and their related postreceptor signaling system (Smads) in distraction osteogenesis of the mandible. *J. Craniofacial Surg.* **15**, 714–718 (2004).
- Haque, T. et al. Immunohistochemical localization of bone morphogenetic protein-signaling Smads during long-bone distraction osteogenesis. J. Histochemistry cytochemistry: Off. J. Histochemistry Soc. 54, 407–415 (2006).
- Rachmiel, A., Aizenbud, D. & Peled, M. Enhancement of bone formation by bone morphogenetic protein-2 during alveolar distraction: an experimental study in sheep. J. Periodontol. **75**, 1524–1531 (2004).

Ko, F. C. & Sumner, D. R. How faithfully does intramembranous bone regeneration recapitulate embryonic skeletal development? *Dev. Dyn.* 250, 377–392 (2021).

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