INSIGHT

CC



BACTERIA

To biofilm or not to biofilm

A new model helps to predict under which conditions a species of bacteria will switch to a static lifestyle.

SHRAVAN PRADEEP AND PAULO E ARRATIA

Related research article Moore-Ott JA, Chiu S, Amchin DB, Bhattacharjee T, Datta SS. 2022. A biophysical threshold for biofilm formation. *eLife* **11**:e76380. doi: 10.7554/eLife.76380

trip to the dentist is seldom fun, but it is often necessary to remove the sticky, slimy deposits (or biofilms) that adhere to our teeth and gums. These structures are formed by bacteria that have adopted a static lifestyle in the moist and warm environment of our mouths. In fact, biofilms are common in a range of natural, clinical, and industrial settings, where they can be dangerous for our health or contaminate equipments (**Davey and O'Toole, 2000; Hall-Stoodley et al., 2004**).

In general, bacteria can either exist in a mobile, 'planktonic' state where they freely disperse and explore their environment for nutrients, or stay statically as 'biofilms', a communal state where the cells share resources and are protected from harmful conditions (*Adler, 1966*). What triggers bacteria to transition from a mobile state to a biofilm lifestyle depends on how each species responds to certain environmental conditions. The factors include nutrient availability, production of certain chemical triggers as well as cellular parameters - such as bacterial concentration, proliferation rate, or diffusing behavior (*Berg, 2018*).

Overall, however, the switch to (immobile) biofilm formation is controlled by bacterial dispersion (which is dependent on nutrient levels), and it occurs when the concentration of bacterial molecules known as autoinducers goes above a certain threshold (**Davies et al., 1998**; **Waters and Bassler, 2005**). These signals, which are produced by bacteria, serve as a proxy for the level of other bacterial cells in the environment and trigger intracellular signals which impact the genes a cell expresses, and the lifestyle it will adopt. Once the biofilm is created, it is maintained by the autoinducer molecules produced by the immobilized bacteria (**Figure 1**).

Yet, how biofilms emerge and the exact conditions that trigger their formation remain a topic of intense research. In general, motile and biofilm lifestyles are studied separately, making it difficult to predict with certainty whether a species of bacteria will form a biofilm under certain conditions. Now, in eLife, Sujit Datta and colleagues at Princeton University – including Jenna Moore-Ott as first author – report having developed a unified framework that can examine both states simultaneously (*Moore-Ott et al., 2022*).

The team developed a series of equations that describe the transition from planktonic state to biofilm under a range of parameters covering all possible conditions. The resulting model, which describes the behavior of the cells, is governed by two main factors: nutrient consumption and bacterial dispersion in the motile state. Both parameters focus on the competition between bacterial dispersion and the production of autoinducer molecules.

Based on the model, Moore-Ott et al. predict two conditions where the concentration of autoinducers remains under the threshold required for biofilm formation. In the first case, nutrients are consumed at such a high rate that the autoinducers are produced (by bacteria) in limited quantities; there is simply not enough autoinducer 'production' time. In the second case, bacteria diffuse and therefore disperse at increased levels

© Copyright Pradeep and Arratia. This article is distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use and redistribution provided that the original author and source are credited.

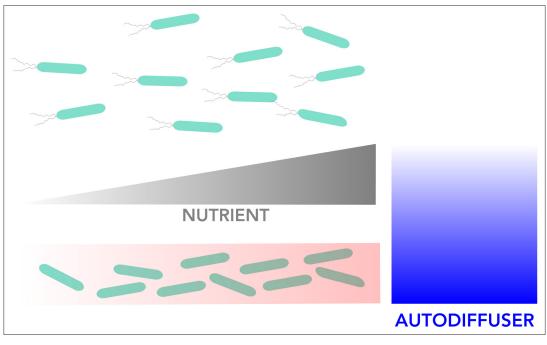


Figure 1. Visual representation of the model for biofilm formation. Bacteria can exist in two different states: a motile state in which they can disperse freely around their environment (top), and an immobile state in which they live together in static as a biofilm (bottom). The red gradient in the biofilm box indicates to which extent bacterial density is increasing in the biofilm from left to right alongside rising nutrient concentrations (grey gradient). The motile bacteria move towards increasing nutrient concentration to the right. The concentration of autodiffusers (molecules produced by bacteria which trigger biofilm formation; blue gradient), is highest close to the biofilm and decreases further away.

(possibly because of environmental conditions), limiting the accumulation of the autoinducers in one location.

In addition, Moore-Ott et al. also pinpointed a third factor the ratio between the time it takes for nutrients to be consumed and for autoinducer to be produced, which affects how fast the biofilm forms and how large they become. For instance, a larger ratio between these two timescales results in the biofilm proliferating, while a smaller ratio slows down the formation of the biofilm. Overall, the combination of these three parameters – nutrient consumption, bacterial dispersion, and ratio of consumption to production time scale – determine which lifestyle a specific species adopts, and at what concentration.

While nutrient consumption and bacterial dispersion vary between different species of bacteria and across environments, they are quantifiable through experiments. This means that the model provides a unique general framework that can be used to predict which state a given bacterial species will adopt under specific circumstances.

Further work should aim to refine the model so it can become closer to real life conditions.

For example, the framework assumes that biofilm formation and the production of autoinducers in a nutrient-dependent fashion are irreversible, two assumptions which can be relaxed for certain species of bacteria (Bridges and Bassler, 2019). In addition, more complex elements could be added to tailor the framework to a specific system, such as incorporating how the biofilm is spatially organized, inputting the role of secondary signaling molecules which fine-tune the impact of autoinducers, or acknowledging how individual cells may respond differently to signals (Bhattacharjee et al., 2022; Jenal et al., 2017; Nadezhdin et al., 2020). Nevertheless, this work represents an important step forward in our quantitative understanding of biofilm formation, which in turn will help us in both fighting and harnessing biofilms, which can be useful in wound healing, bioremediation, or functional materials production.

Shravan Pradeep is in the Department of Earth and Environmental Sciences University of Pennsylvania, Philadelphia, United States bhttp://orcid.org/0000-0001-7483-2385 Paulo E Arratia is in the Department of Mechanical Engineering and Applied Mechanics, University of Pennsylvania, Philadelphia, United States parratia@seas.upenn.edu http://orcid.org/0000-0002-2566-2663

Competing interests: The authors declare that no competing interests exist.

Published 21 July 2022

References

Adler J. 1966. Chemotaxis in bacteria. *Science* 153:708–716. DOI: https://doi.org/10.1126/science. 153.3737.708, PMID: 4957395

Berg HC. 2018. Random Walks in Biology. Princeton University Press. DOI: https://doi.org/10.2307/j. ctv7r40w6

Bhattacharjee T, Amchin DB, Alert R, Ott JA, Datta SS. 2022. Chemotactic smoothing of collective migration. *eLife* **11**:e71226. DOI: https://doi.org/10. 7554/eLife.71226, PMID: 35257660

Bridges AA, Bassler BL. 2019. The intragenus and interspecies quorum-sensing autoinducers exert distinct control over Vibrio cholerae biofilm formation and dispersal. *PLOS Biology* **17**:e3000429. DOI: https://doi.org/10.1371/journal.pbio.3000429, PMID: 31710602

Davey ME, O'Toole GA. 2000. Microbial biofilms: from ecology to molecular genetics. *Microbiology and Molecular Biology Reviews* **64**:847–867. DOI: https://

doi.org/10.1128/MMBR.64.4.847-867.2000, PMID: 11104821

Davies DG, Parsek MR, Pearson JP, Iglewski BH, Costerton JW, Greenberg EP. 1998. The involvement of cell-to-cell signals in the development of a bacterial biofilm. *Science* **280**:295–298. DOI: https://doi.org/10. 1126/science.280.5361.295, PMID: 9535661

Hall-Stoodley L, Costerton JW, Stoodley P. 2004. Bacterial biofilms: from the natural environment to infectious diseases. *Nature Reviews Microbiology* 2:95–108. DOI: https://doi.org/10.1038/nrmicro821, PMID: 15040259

Jenal U, Reinders A, Lori C. 2017. Cyclic di-GMP: second messenger extraordinaire. *Nature Reviews. Microbiology* **15**:271–284. DOI: https://doi.org/10. 1038/nrmicro.2016.190, PMID: 28163311

Moore-Ott JA, Chiu S, Amchin DB, Bhattacharjee T, Datta SS. 2022. A biophysical threshold for biofilm formation. *eLife* **11**:e76380. DOI: https://doi.org/10. 7554/eLife.76380, PMID: 35642782

Nadezhdin E, Murphy N, Dalchau N, Phillips A, Locke JCW. 2020. Stochastic pulsing of gene expression enables the generation of spatial patterns in *Bacillus subtilis* biofilms. *Nature Communications* 11:950. DOI: https://doi.org/10.1038/s41467-020-14431-9, PMID: 32075967

Waters CM, Bassler BL. 2005. Quorum sensing: cell-to-cell communication in bacteria. Annual Review of Cell and Developmental Biology **21**:319–346. DOI: https://doi.org/10.1146/annurev.cellbio.21.012704. 131001, PMID: 16212498