

Article

Genome Size and Chromosome Number Evolution in Korean *Iris* L. Species (Iridaceae Juss.)

Bokyung Choi¹, Hanna Weiss-Schneeweiss², Eva M. Temsch², Soonku So³, Hyeon-Ho Myeong³ and Tae-Soo Jang^{1,*}

- ¹ Department of Biological Science, College of Bioscience and Biotechnology, Chungnam National University, Daejeon 34134, Korea; cbokyung@cnu.ac.kr
- ² Department of Botany and Biodiversity Research, University of Vienna, Rennweg 14, A-1030 Vienna, Austria; hanna.schneeweiss@univie.ac.at (H.W.-S.); eva.temsch@univie.ac.at (E.M.T.)
- ³ Korea National Park Research Institute, 171, Dangu-ro, Wonju-si 26441, Gangwon-do, Korea; ssk822@knps.or.kr (S.S.); ecomyung@knps.or.kr (H.-H.M.)
- * Correspondence: jangts@cnu.ac.kr; Tel.: +82-42-821-6555

Received: 13 September 2020; Accepted: 25 September 2020; Published: 28 September 2020



Abstract: Chromosome numbers, karyotypes, and genome sizes of 14 *Iris* L. (Iridaceae Juss.) species in Korea and their closely related taxon, *Sisyrinchium rosulatum*, are presented and analyzed in a phylogenetic framework. To date, understanding the chromosomal evolution of Korean irises has been hampered by their high chromosome numbers. Here, we report analyses of chromosome numbers and karyotypes obtained via classic Feulgen staining and genome sizes measured using flow cytometry in Korean irises. More than a two-fold variation in chromosome numbers (2n = 22 to 2n = 50) and over a three-fold genome size variation (2.39 pg to 7.86 pg/1 C) suggest the putative polyploid and/or dysploid origin of some taxa. Our study demonstrates that the patterns of genome size variation and chromosome number changes in Korean irises do not correlate with the phylogenetic relationships and could have been affected by different evolutionary processes involving polyploidy or dysploidy. This study presents the first comprehensive chromosomal and genome size data for Korean *Iris* species. Further studies involving molecular cytogenetic and phylogenomic analyses are needed to interpret the mechanisms involved in the origin of chromosomal variation in the *Iris*.

Keywords: chromosome number; genome size; Iris; karyotype; systematics

1. Introduction

Chromosomal changes play a major role in plant evolution and are thus important in diversification and speciation in angiosperms [1–3]. Variation of chromosome numbers, ploidy levels, and genome sizes have been frequently analyzed for a better understanding of evolutionary patterns and species relationships in plants [4–7]. The genome size values, in combination with data on chromosome numbers, allow ploidy levels to be inferred and can also provide insights into evolutionary relationships between closely related taxa in wild plant groups [7–12]. While the monocots are known to have the widest range of genome sizes (0.2–152.2 pg/1 C) among angiosperms [13–15], the known genome sizes in the family Iridaceae range from 0.5 pg/1 C in *Hesperantha* and *Sisyrinchium* species to 31.4 pg/1 C in *Moraea* taxa [15,16].

Among monocots, *Iris* is an excellent system in which to study the evolutionary patterns of chromosome number and genome size evolution because this group has an exceptionally high diversity of chromosome numbers (n = 7, 8, 9, 10, 11, 12, 13, 15, 16, 17, 18, 19, 20, 21, 22, 24, 25, 27, 36, 54) [17] and genome sizes (1.05–28.2 pg/1 C) [15]. The genus *Iris* L. is perennial and comprises approximately 300 species worldwide, with the greatest number of endemic species occurring in the

Mediterranean and Asia [18–21]. Generic and sub-generic classification of the genus *Iris* has been mainly based on morphological characters of flower organs and roots [22,23]. Subsequently, Wilson [24] recognized nine subgenera in the genus based on phylogenetic analysis of chloroplast sequence data: *Iris, Crossiris* Spach, *Limniris, Lophiris* (Tausch) C. A. Wilson, *Nepalensis, Pardanthopsis* (Hance) Baker, *Siphonostylis* (W. Schulze) C. A. Wilson, *Xiphium*, and *Xyridon* (Tausch) Spach.

Currently, 14 species from *Iris* (subgenus *Limniris* and *Pardanthopsis*) are recognized in Korea: *Iris ensata* Thunb., *I. dichotoma* Pall., *I. domestica* (L.) Goldblatt & Mabb., *I. koreana* Nakai, *I. oxypetala* Bunge, *I. pseudacorus* L., *I. uniflora* Pall. ex Link, *I. laevigata* Fisch., *I. sanguinea* Donn ex Hornem., *I. setosa* Pall. ex Link, *I. odaesanensis* Y.N. Lee, *I. minutoaurea* Makino, *I. rossii* Baker, *I. ruthenica* Ker Gawl. [24–27]. Species of the Korean irises usually have a narrow geographical distribution, and some are endemic (*Iris koreana*) or subendemic to Korea (*Iris odaesanensis* with a small/disjunct population in Jilin Province, China) [19,25]. While several species of Korean irises (*I. dichotoma*, *I. laevigata*, *I. setosa*, and *I. ruthenica*) are either endangered or threatened due to habitat loss caused by anthropogenic activities [25,28], *Iris pseudacorus* and *Sisyrinchium rosulatum* were introduced as horticultural elements and are now naturalized [25]. Despite an increasing importance of conservation status of the Korean irises, comprehensive systematic studies including morphological, phylogenetic, and cytological analyzes have not been performed in the group. Molecular phylogenetic analyses based on plastid and nuclear ribosomal DNA data have allowed for the inferring established relationship patterns of major lineages of among the Korean irises [27,29,30], although some sectional and species level relationships still remain unresolved.

Worldwide, chromosome numbers in the genus *Iris* have been reported for 168 taxa in validly published references (Table S1; Chromosome Counts Database, CCDB, ver. 145, http://ccdb.tau.ac. il/Angiosperms/Iridaceae/Iris/) [17]. Chromosome numbers and karyotypes have been reported for eight species of the Korean irises (*I. ensata, I. dichotoma, I. koreana, I. oxypetala, I. pseudacorus, I. uniflora, I. setosa, I. sanguinea*), but none of the studied plants were collected from natural populations [31]. Thus, detailed cytogenetic studies of Korean irises from natural populations are required. Analyses of chromosome numbers accompanied by genome size measurements allow for inferences of the incidence of polyploidy [5,7]. Although the genome size data of 44 *Iris* species have been conducted so far [15], the nuclear DNA amounts using flow cytometry for Korean irises are not available to date [15].

Thus, this study examines the patterns of chromosome number evolution, karyotypes, and genome size variation in a phylogenetic context in the Korean irises. The specific aims of this study are (1) to determine chromosome numbers and size variation as well as karyotypes structure all 14 Korean *Iris* species from multiple populations; (2) to examine patterns of genome size change in this group of taxa to address to the lack of the C-value data in the genus *Iris*; and (3) to interpret the patterns of evolution of cytological data within the available phylogenetic framework to better understand the genome evolution in Korean irises.

2. Materials and Methods

2.1. Plant Materials

A total of 76 individuals representing 36 populations of 14 species of *Iris* and one outgroup species *Sisyrinchium rosulatum* were collected from natural populations with few individuals used from other sources (cultivation; Table 1). The plants used in this study were transplanted in a glasshouse in Chungnam National University, Daejeon, Republic of Korea. All individuals were used for analyses of chromosome numbers, karyotypes and measurements of genome size (Table 1). Voucher specimens of the examined plants were deposited in CNUK (Chungnam National University Herbarium). Four Korean *Iris* species (*I. dichotoma, I. laevigata, I. setosa, I. ruthenica*) that are endangered and threatened were collected with special permissions from the Korean Government (collected under permits no. 2018-18, 2019-13, 2019-14, 2019-20, and 2019-30; Table 1). In order to avoid illegal removal

to the endangered or vulnerable species, we refrain from providing details of the locality information (GPS coordinates) in this study.

2.2. Chromosome Numbers and Karyotype Analysis

Fresh root tips were pretreated in 0.05% colchicine (Sigma-Aldrich Co., St. Louis, USA) solution at room temperature in the dark for 4.5 h. The sampled roots were fixed in ethanol:acetic acid (3:1) for two hours at room temperature and then stored at -20 °C. For karyotype and chromosome number analyses, meristematic root cells were used (Table 1). Prior to staining, the fixed roots were washed with tap water, and hydrolyzed in 5 N HCL at room temperature for 30 min. Subsequently, to perform Feulgen staining, the root tips were treated in Schiff's reagent (Merck KGaA, Darmstadt, Germany) in darkness for 1 h. Microscopic slides were prepared by squashing the meristematic tissue in 60% acetic acid. A minimum of three well spread chromosome plates were selected for the analyses [32]. The chromosomes were cut using Corel Photo-Paint 12.0 (Corel Company, Ottawa, Canada). For chromosomal measurements, MicroMeasure ver.3.3 program (https://micromeasure.software.informer.com/3.3/) [33] was used, and a minimum of three chromosomal spreads were measured with a comparable medium degree of chromosomal condensation (Table 2).

2.3. Genome Size Measurement by Flow Cytometry (FCM)

Genome sizes of 76 individuals of 14 *Iris* species and *Sisyrinchium rosulatum* were measured using flow cytometry with *Solanum pseudocapsicum* (1 C = 1.29 pg) [34] or *Pisum sativum* "Kleine Rheinländerin" (1 C = 4.42 pg) [35] as an internal standard. Approximately 20–30 mg of fresh leaves of each sample were chopped with an internal reference standard using Otto's buffer I [36]. Subsequently, each sample was filtered using a nylon mesh, and incubated with RNase A in a water-bath at 37 °C for 30 min. The nuclei staining was conducted using propidium iodide, which contains Otto's buffer II, and the sample was stored at 4 °C. A total of three measurements of DNA content were estimated for each sample using a flow cytometer (Partec, Münster, Germany). 1 C-values were calculated using estimation of linear fluorescence intensity of nuclei of the examined taxon and internal standard. The coefficient of variation (CV) of all estimations were mostly below 3% on average, and never exceeded 7% [7].

Taxon; Collection Number	Locality; GPS Coordinate; Collector	Chromosome Number (2 <i>n</i>)	Genome Size 1 C ± S.D. (pg)				
	Iris						
Subgenus Lin	nniris section Limniris						
Iris ensata Thunb.			7.76 ⁴				
IMHAE103	Yang-Yang, Kangwon; N38°05′44″, E128°39′15″, 154 m; TS, SK, CM	24	7.86 ± 0.022				
IMHAE117	Pyeong-Chang, Kangwon; N37°41'34.03'', E128°45'25.79'', 889 m; TS, SK, CM	24	7.66 ± 0.011				
I. koreana Nakai ^{1, 2}			7.29 ⁴				
sck00042	Byeonsanbando National Park Endangered Species Botanical Garden (Cult.): SS_RL_CB_SK	50	7.22 ± 0.068				
	Byeonsanbando National Park						
sck00043	Endangered Species Botanical Garden (Cult.); SS, RI, CB, SK	50	7.35 ± 0.029				
<i>I. laevigata</i> Fisch. ¹			7.12 ⁴				
Jebi-2	Gangwon; TS, CB	32	7.06 ± 0.010				
Jebi-3	Gangwon; TS, CB	32	7.18 ± 0.046				
I. minutoaurea Makino	-		3.85 ⁴				
Gasan_8	Gasan, Kyungsang; N36°02'41'', E128°34'12'', 733 m; TS, CB, HJ	22	3.92 ± 0.008				
JCKCK193269	Gasan, Kyungsang; N36°02'35'', E128°34'36'', 675 m; TS, CB, SK, YM	22	3.71 ± 0.171				
JCKCK193273-1	Gasan, Kyungsang; N36°02'26'', E128°34'26'', 779 m; TS, CB, SK, YM	22	3.71 ± 0.024				
JCKCK193273-2	Gasan, Kyungsang; N36°02'26'', E128°34'26'', 779 m; TS, CB, SK, YM	22	3.72 ± 0.006				
JC041913	Gasan, Kyungsang; N36°02'44'', E128°33'51'', 629 m; TS, YM	22	4.09 ± 0.042				
JCKC1932211-2	Gasan, Kyungsang; N36°02'28'', E128°34'18'', 806 m; TS, CB, SK, YM	22	3.79 ± 0.019				
JCKC1932211-3	Gasan, Kyungsang; N36°02'28'', E128°34'18'', 806 m; TS, CB, SK, YM	22	3.70 ± 0.003				
I. odaesanensis Y.N.Lee ^{1,3}			3.72 ⁴				
YSG18-66_1	Gangwon; YS	28	3.67 ± 0.012				
YSG18-66_2	Gangwon; YS	28	3.68 ± 0.011				
YSG18-66_4	Gangwon; YS	28	3.70 ± 0.019				
BKC928	Kyungsang; CB, YC	28	3.79 ± 0.007				

Table 1. Information on the plant material used for the karyotype and genome size analyses of *Iris* L. in Korea.

Table 1. Cont.

Taxon; Collection Number	Locality; GPS Coordinate; Collector	Chromosome Number (2 <i>n</i>)	Genome Size 1 C ± S.D. (pg)					
JCKC1932271	Kyungsang; TS, CB, KS, YM	28	3.76 ± 0.021					
JC041904	Kyungsang; TS, YM	28	3.74 ± 0.042					
JC041903	Kyungsang; TS, YM	28	3.67 ± 0.015					
JC041940	Kyungsang; TS, YM	28	3.73 ± 0.018					
I. oxypetala Bunge			6.08 ⁴					
CC0621-50	Ga-Gokri, Chungnam; N37°01'21'', E126°34'30'', 4 m; CB, CM	40	6.38 ± 0.014					
CC20190621-58	Ga-Gokri, Chungnam; N37°01'17", E126°34'32", 3 m; CB, CM	40	6.06 ± 0.010					
CC20190621-64	Ga-Gokri, Chungnam; N37°01'24'', E126°34'26'', 4 m; CB, CM	40	6.00 ± 0.017					
JCCMH025	Gang-Wha, Incheon; N37°45'37'', E126°14'29'', 80 m; TS, CB, CM	40	5.90 ± 0.005					
I. pseudacorus L.			5.89 ⁴					
Keumjeong-34	Mt. Keumjeong, Busan; N35°14' 5.5", E129°03'24.6", 455 m; CB, YK, YC	34	5.82 ± 0.012					
Chungdae_2	Sinseong-dong, Daejeon; N36°22'44.8", E127°20'35.5", 83 m; CB	34	5.87 ± 0.017					
CC20190621-1	Is. Hwangdo, Chungnam; N36°35′58.2″, E126°22′42.5″, 14 m; CB	34	5.87 ± 0.024					
CC20190621-48	Dangjin, Chungnam; N37°01'21", E126°34'30"E, 4 m; CB, CM	34	5.98 ± 0.011					
J20190616-1	Nogokjae, Kimchoen; N36°04'04'', E128°12'50'', 242 m; TS, KS, CB	34	6.02 ± 0.026					
BKC910	Mt. Keumjeong, Busan; N35°14'06", E129°03'28", 459 m; CB, YK, YC	34	5.77 ± 0.009					
CC20190621-57	Ga-Gokri, Chungnam; N37°01'21'', E126°34'30'', 4 m; CB, CM	34	5.91 ± 0.009					
I. rossii Baker			3.73 ⁴					
SJH1830	Mt. Cheon-Gwan, Jeon-Ra; N34°32'39.4'', E126°55'40.8'', 225 m; SJH	32	3.69 ± 0.014					
Pusanuni_2	Mt. Keumjeong, Busan; N35°14'23.8", E129°4'27.23", 169 m; CB, YK	32	3.72 ± 0.005					
J513	Mun-San, Kyung-Gi; N37°57′55″, E126°56′13″, 58 m; TS	32	3.85 ± 0.010					
J514	Mun-San, Kyung-Gi; N37°57′55″, E126°56′11″, 74 m; TS	32	3.67 ± 0.013					
Jeokseong_2	Mun-San, Kyung-Gi; N37°57′55″, E126°56′13″, 58 m; TS	32	3.66 ± 0.005					
Keumjeong_8	Mt. Keumjeong, Busan; N35°14'23.8'' E129°4'27.23'', 169 m; CB, CM	32	3.79 ± 0.018					

Table 1.	Cont.
----------	-------

Taxon; Collection Number	Locality; GPS Coordinate; Collector	Chromosome Number (2 <i>n</i>)	Genome Size 1 C ± S.D. (pg)					
JCK05_7	Mt. Saeng-Aeng; N36°19'21.9", E126°30'45.7", 110 m; TS, CB, SK	32	3.74 ± 0.033					
JCKC190512	Mojioreum, Jeju; N33°23'42'', E126°45'58'', 247 m; TS, CB, SK, CM	32	3.75 ± 0.003					
SCK00021	Mt. Bong-Wha, Chungnam; N36°47′08″, E126°26′20″, 79 m; CB, SK	32	3.71 ± 0.003					
I. sanguinea Hornem.			5.23 ⁴					
K02-41	Mt. Hwa-Ak, Kyung-Gi; N38°00'16.60'', E127°52'51.93'', 1097 m; SK	28	5.22 ± 0.012					
Hwaya_1	Mt. Hwa-Ya, Kyung-Gi; N36°56'24.5", E127°16'46.8", 162 m; TS	28	5.30 ± 0.009					
K025-04-01	Yeon-Cheon, Gangwon; N38°09'97.95", E127°11'28.64", 129 m; SK	28	5.42 ± 0.021					
K025-04-04	Yeon-Cheon, Gangwon; N38°09'97.95", E127°11'28.64", 129 m; SK	28	5.18 ± 0.022					
Munsan_1	Mun-San, Kyung-Gi; N37°57′55″, E126°56′13″, 58 m; TS	28	5.12 ± 0.009					
Cheonma-3	Mt. Cheon-Ma, Kyung-Gi; N37°41'24", E127°24'36", 157 m; YS, SJH	28	5.11 ± 0.011					
JC041939	Dae-Gu, Kyungsang; N35°54'44", E128°38'46", 629 m; TS, YM	28	5.25 ± 0.020					
<i>I. setosa</i> Pall. ex Link ¹			5.63 ⁴					
Ga-3	Gangwon; TS, CB	38	5.55 ± 0.012					
Ga_4-1	Gangwon; TS, CB	38	5.64 ± 0.007					
Ga-7	Gangwon; TS, CB	38	5.57 ± 0.004					
Seo-2	Gangwon; TS, CB	38	5.78 ± 0.016					
Ga-5	Gangwon; TS, CB	38	5.59 ± 0.035					
Subgenus L	imniris section Ioniris							
<i>I. ruthenica</i> Ker Gawl. ¹			2.42 ⁴					
Mo 2	Jeju; YS, SJH	42	2.45 ± 0.004					
	Incheon: SS	42	2.42 ± 0.002					
BKC939	Busan; CB, YM	42	2.45 ± 0.006					
JC532	Daegu; TS, CM	42	2.39 ± 0.001					
JC352-1	Daegu; TS, CM	42	2.43 ± 0.006					
IC534-1	Daegu; TS, CM	42	2.42 ± 0.017					
SCK00032	Chungnam; SS, CB, SK	42	2.42 ± 0.001					
BKC937	Busan; CB	42	2.40 ± 0.001					
ICCMH004	Incheon; TS, CB, CM	42	2.43 ± 0.002					
I. uniflora Pall. ex Link	· · ·		2.46 ⁴					
JCK2019-77	Mt. Sorak, Gangwon; N38°09'27'', E128°29'19'', 790 m; TS, CB, SK	42	2.48 ± 0.002					

Taxon; Collection Number	Locality; GPS Coordinate; Collector	Chromosome Number (2 <i>n</i>)	Genome Size 1 C ± S.D. (pg)				
JCK2019-78	Mt. Sorak, Gangwon; N38°09'27", E128°29'19", 790 m; TS, CB, SK	42	2.44 ± 0.003				
JCK2019-79	Mt. Sorak, Gangwon; N38°09'27'', E128°29'19'', 790 m; TS, CB, SK	Mt. Sorak, Gangwon; N38°09'27", E128°29'19", 790 m; TS, CB, SK 42					
Subgenus Parda	anthopsis (Hance) Baker						
Iris dichotoma Pall. ¹			3.34 ⁴				
SS03-1	Cheonripo Arboretum (regenerated individual from a wild population); SS	32	3.35 ± 0.004				
SS20-1	Cheonripo Arboretum (regenerated individual from a wild population); SS	32	3.35 ± 0.008				
SS20-2	Cheonripo Arboretum (regenerated individual from a wild population); SS	32	3.32 ± 0.011				
SS20-3	Cheonripo Arboretum (regenerated individual from a wild population); SS	32	3.34 ± 0.006				
I. domestica (L.) Goldblatt & Mabb.	• •						
Beom_1	Busan (cult.); CB, YK	32	4.75 ± 0.010				
Sisyrinchium rosulatum E.P.Bicknell.			0.91 ⁴				
JCKC190515	Hyomyeong, Jeju; N33°19'25'', E126°35'39'', 327 m; TS, CB, SK, CM	32	0.91 ± 0.001				
JCKC190451	Gujwa, Jeju; N33°27'05'', E126°48'06'', 244 m; TS, CB, SK, CM	32	0.90 ± 0.001				

Table 1. Cont.

¹ GPS coordinates, latitude and longitude of the collected sites of endangered, threatened and endemic species are not indicated for protection purpose (collected under permits no. 2018-18, 2019-13, 2019-14, 2019-20, and 2019-30 from the Ko-Seong, Dae-Gu, Dae-Jeon, Busan, and Jeon-Nam populations, respectively); ² Taxa endemic to Korea; ³ Taxa subendemic to Korea; Collectors: TS = Tae-Soo Jang; CB = Bokyung Choi; KS = Sun-Yu Kim; YK = Yusun Kweon; RI = II Roh; SK = Seong-Yeon Kang; SS = Soonku So; CM = Young-Min Choi; YC = Yeongmun Choi; YS = Sungyu Yang; SJH = Jun-Ho Song; ⁴ Results in bold indicates mean genome size values for each species.

Tayon (Collection Number)		Chromosome Length (µr	$A = 1^{2} (0/)$	DI 3	Figure	
Taxon (Conection Number)	Largest (Mean ± S.D.)	Smallest (Mean ± S.D.)	Total HKL 1 (Mean \pm S.D.)	ASI - (%)	KI ^o	Tiguie
Iris						
Subgenus Limniris section Limniris						
Iris ensata (IMHAE117)	4.8 ± 0.2	2.8 ± 0.1	82.5 ± 1.7	66.7	1.69	1A, 2A
I. koreana (sck00043)	5.3 ± 0.8	1.5 ± 0.1	141.7 ± 9.9	61.9	3.51	1B, 2B
I. laevigata (Jebi-2)	3.7 ± 0.1	2.0 ± 0.1	81.8 ± 1.7	61.9	1.85	1 C, 2C
I. minutoaurea (Cheonma-2)	5.1 ± 0.2	1.9 ± 0.1	75.8 ± 2.4	58.3	2.58	1D, 2D
I. odaesanensis (YSG18-66_2)	4.3 ± 0.1	1.9 ± 0.1	86.3 ± 2.4	59.5	2.19	1E, 2E
I. oxypetala (CC0621-50)	2.9 ± 0.1	1.7 ± 0.1	86.4 ± 3.8	59.1	1.72	1F, 2F
I. pseudacorus (CC20190621-57)	4.9 ± 0.3 4.5 ± 0.2	2.1 ± 0.1	107.1 ± 4.2	62.0	2.34	1G, 2G
I. rossii (Keumjeong_8)		2.1 ± 0.1	100.5 ± 3.0	60.9	2.08	1H, 2H
I. sanguinea (Hwaya_1)	4.5 ± 0.2	2.1 ± 0.2	87.2 ± 2.8	62.5	2.09	1I, 2I
I. setosa (Seo-2)	4.6 ± 0.4	2.1 ± 0.1	122.9 ± 6.9	61.6	2.11	1J, 2J
Subgenus Limniris section Ioniris						
I. ruthenica (JC534-1)	2.9 ± 0.2	1.1 ± 0.1	78.3 ± 4.3	58.0	2.52	1K, 2K
I. uniflora (JCK2019-79)	2.3 ± 0.2	1.1 ± 0.1	61.9 ± 1.9	58.3	1.95	1L, 2L
Subgenus Pardanthopsis						
I. dichotoma (SS20-3)	4.6 ± 0.3	2.0 ± 0.1	97.0 ± 4.4	61.9	2.19	1M, 2M
I. domestica (Beom_1)	3.9 ± 0.3	2.1 ± 0.1	92.8 ± 4.2	59.7	1.76	1N, 2N
Sisyrinchium rosulatum (JCKC190515)	1.8 ± 0.1	1.0 ± 0.1	43.4 ± 1.9	59.9	1.84	10, 20

Table 2. Karyotype analyses in the genus *Iris* L. and its related taxon *Sisyrinchium rosulatum*.

¹ HKL: total haploid karyotype length; ² AsI (asymmetry index): the proportion of all long arms to the total haploid karyotype length; ³ RI (Ratio Index): the ratio of the longest to the shortest chromosome.

3. Results and Discussion

3.1. Chromosome Numbers and Karyotypes of Iris L.

Chromosome numbers for 74 individuals of 14 currently recognized the Korean *Iris* species and two individuals of *Sisyrinchium rosulatum* are provided in Table 1 and Figures 1 and 2. The karyotype analyses of each species with detailed chromosome size measurements are presented in Table 2 and Figure 2. The previously reported chromosome number for the genus *Iris* varied from 2n = 14 (*I. vorobievii*) to 2n = 108 (*I. versicolor*) representing approximately a 7.7-fold difference (Table S1). To date, the chromosome numbers have been determined for approximately 168 taxa of ca. 300 currently recognized species in the genus *Iris* worldwide (c. 56%; Table S1) [17].



Figure 1. Mitotic metaphase chromosomes of Korean irises. (A) *Iris ensata* (2n = 24). (B) *I. koreana* (2n = 50). (C) *I. laevigata* (2n = 32). (D) *I. minutoaurea* (2n = 22). (E) *I. odaesanensis* (2n = 28). (F) *I. oxypetala* (2n = 40). (G) *I. pseudacorus* (2n = 34). (H) *I. rossii* (2n = 32). (I) *I. sanguinea* (2n = 28). (J) *I. setosa* (2n = 38). (K) *I. ruthenica* (2n = 42). (L) *I. uniflora* (2n = 42). (M) *I. dichotoma* (2n = 32). (N) *I. domestica* (2n = 32). (O) *Sisyrinchium rosulatum* (2n = 32). Inset in (K) shows same chromosome group of a single cell which were lying at some distance using high magnification objectives. Scale bar = 5 µm.

A		3	8 (4	5	6 68	7 80	8 86	9 60	10	11 \$8	12 0 0	13	14	15	16	17	18	19	20	21	22	23	24	25
В	11 11	6	1	1	81	24	8.8	81	89	88	81		80	88	80	81	84	5.6	61	48		80	81	*6	**
С	88 8	8 8	1	2	Ba	60	08	88	8 A	81	61	æ ()	44	8 18	86	88									
D	111	1	()	g	11	13	66	63	8.8	8.6	88														
Ε	18 31	2	11	1	И	11	11	38	и	81	88	8.6	88	84											
F	8,11 at		5 5	15	88	84	88	88	88	88	#8	88	8.8	sa	88		68	-	66	**					
G	111	1 8	1	8	80	88	60	68	86	88	88	8.8	60	88	60	**	88								
н	RE	1.1	11	8	81	11	81	0.1	15	11	6.4	51	8.8	14	8.8	0.8									
I	11 1	1	1 1	8	15	11	14	u	51	=	61	8.0	11	8.8											
J	11 6	1 8	5 1	18	51	55	86	38	85	6.5	24	84	15	88	85	2.8	яs	45	**						
К	83 X			61	A1	81	11	=	0	14	81	**		4.8		*5	**	**	**	**					
L	16 41	8	1	4	28		65	#1	#1	46	65	84	Ĥ	**	45	81	85	**	**	2	<u></u>				
М	1) 1	16	\$))	33	н	11	55	Ð	"	u	88	15	63	**	4									
Ν		1 1	1		83	68	10		88			18	88	84											
0	55 55	6					44	8.8	84		.,				**										

Figure 2. Karyotypes of the Korean irises. (A) *Iris ensata* (2n = 24). (B) *I. koreana* (2n = 50). (C) *I. laevigata* (2n = 32). (D) *I. minutoaurea* (2n = 22). (E) *I. odaesanensis* (2n = 28). (F) *I. oxypetala* (2n = 40). (G) *I. pseudacorus* (2n = 34). (H) *I. rossii* (2n = 32). (I) *I. sanguinea* (2n = 28). (J) *I. setosa* (2n = 38). (K) *I. ruthenica* (2n = 42). (L) *I. uniflora* (2n = 42). (M) *I. dichotoma* (2n = 32). (N) *I. domestica* (2n = 32). (O) *Sisyrinchium rosulatum* (2n = 32). Blue and red bars indicate metacentric and submetacentric chromosomes, respectively. Scale bar: 5 µm.

3.1.1. Iris L. Subgenus Limniris Spach Section Limniris Spach

The chromosome numbers in sect. Limniris ranged from 2n = 22 in I. minutoaurea to 2n = 50in *I. koreana* (Table 1) confirming earlier studies [17]. *I. odaesanensis* (2n = 28) is reported here for the first time. Both analyzed plants of *I. ensata* had 2n = 24 (Figure 1A), AsI (asymmetry index) of 66.7% and karyotype formula of 2n = 4 m + 20 sm (Figure 2A) with Haploid Karyotype length (HKL) of $82.5 \pm 1.7 \,\mu\text{m}$. The two individuals of *I. koreana* had 2n = 50 (Figure 1B), with its AsI of 61.9% and the karyotype formula of 34 m + 16 sm (Figure 2B) and HKL of 141.7 \pm 9.9 μ m. The four plants of *I. laevigata* had 2n = 32 (Figure 1C). The species had AsI of 61.9%, karyotype formula of 2n = 22 m + 10 sm (Figure 2C) and HKL of $81.8 \pm 1.7 \mu \text{m}$. All 11 analyzed plants of *I. minutoaurea* had 2n = 22 (Figure 1D) and AsI of 58.3%. The karyotype formula was 18 m + 4 sm (Figure 2D) with HKL of 75.8 \pm 2.4 μ m. All eight plants of *I. odaesanensis* had 2n = 28 (Figure 1E) with AsI of 59.5%, karyotype formula of 2n = 24 m + 4 sm (Figure 2E) and HKL of 86.3 ± 2.4 µm. All four plants of *I. oxypetala* had 2n = 40 (Figure 2F), AsI of 59.1% and karyotype formula of 2n = 36 m + 4 sm (Figure 2F). HKL of this species was $86.4 \pm 3.8 \mu m$. All seven plants of *I. pseudacorus* had 2n = 34 (Figure 1G), AsI was 62.0%and the karyotype formula was 26 m + 8 sm (Figure 2G) with HKL of $107.1 \pm 4.2 \mu \text{m}$. All nine plants of *I. rossii* had 2n = 32 (Figure 1H). This species had AsI of 60.9%, karyotype formula of 2n = 24 m + 8 sm (Figure 2H) and HKL of $100.5 \pm 3.0 \,\mu\text{m}$. All seven plants of *I. sanguinea* possessed 2n = 28 (Figure 1I), AsI of 62.5%, and the karyotype formula of 18 m + 10 sm (Figure 2I) with HKL of 87.2 \pm 2.8 μ m. All five plants of *I. setosa* had 2n = 38 (Figure 1J), in agreement with previous reports [17] except for the one study reporting 2n = 40 for this species [31]. This deviating count might have potentially referred to another taxon (either a species or a variety), but it is difficult to verify due to unknown origin of the examined sample (e.g., cultivated material) [31]. Most previous studies used cultivated material. Thus, any discrepancies that might exist between different reports might have been affected by domestication and associated potential hybridization [31], as also suggested for Iberian irises [37]. *Iris setosa* had AsI of 61.6% and karyotype formula of 2n = 20 m + 18 sm (Figure 2J) with HKL of 122.9 ± 6.9 µm.

3.1.2. Iris L. Subgenus Limniris Spach Section Ioniris Spach

All nine plants of *I. ruthenica* had 2n = 42 (Figure 1K), in agreement with previous reports (Table S1) [17]. This species had AsI of 58.0% and karyotype formula of 2n = 40 m + 2 sm (Figure 2K) with HKL of 78.3 ± 4.3 µm. The three analyzed plants of *I. uniflora* had 2n = 42 (Figure 1L) confirming previous reports (Table S1) [17]. Its AsI was 58.3% and karyotype formula of 2n = 34 m + 8 sm (Figure 2L) with HKL of 61.9 ± 1.9 µm.

3.1.3. Iris L. Subgenus Pardanthopis (Hance) Baker

All four examined individuals of *I. dichotoma* had 2n = 32 (Figure 1M). This number was consistent with previous records for this species from Russian and Chinese populations [17], except for the study of Kim et al. [31], in which analyzed cultivated material was reported to have 2n = 34. This species had AsI of 61.9% and karyotype formula of 2n = 18 m + 14 sm (Figure 2M) with HKL of 97.0 ± 4.4 µm. *Iris domestica* had 2n = 32 (Figure 1N), which was in agreement with previous reports (Table S1) [17]. Its AsI was 59.7% and karyotype formula was 2n = 22 m + 10 sm (Figure 2N) with HKL of 92.8 ± 4.2 µm.

3.1.4. Sisyrinchium rosulatum E.P.Bicknell.

Both of the *S. rosulatum* individuals analyzed had the chromosome number of 2n = 32 (Figure 1O), and the results were in agreement with previous reports [17]. The AsI of this species was 59.9% and karyotype formula was 2n = 24 m + 8 sm (Figure 2O) with HKL of 43.4 ± 1.9 µm.

3.2. Genome Size Variation in Iris L.

DNA content analyses using an internal standard revealed clear and well-defined peaks for all samples with the coefficients of variation (CV) for the internal standard and sample peaks ranging from 1.03% to 5.40% (mean \pm S.D. = 2.24 \pm 0.80) and from 1.23% to 6.50% (mean \pm S.D. = 2.87 \pm 1.06), respectively.

Genome sizes for all Korean irises, except for *Iris pseudacorus*, are reported here for the first time (Table 1; Figure 3). The analysis revealed a 3.29-fold variation in nuclear DNA content among the analyzed taxa, with genome sizes ranging from 2.39 pg/1 C in *Iris ruthenica* to 7.86 pg/1 C in *I. ensata*. To date, the 1 C-values of 44 *Iris* species have been reported ranging from 1.05 pg/1 C in *I. versicolor* to 28.20 pg/1 C in *I. histrio* [15]. Genome size of *Iris pseudacorus* reported earlier (1 C = 5.67 pg) [38] is very similar with the current data (5.77–6.02 pg/1 C; Table 1). The outgroup species *Sisyrinchium rosulatum* had 0.90 pg/1 C (Table 1; Figure 3). The 1 C-values in Korean irises vary significantly among species, but are constant within populations and species, as previously observed in closely related species within a genus [10,39]. This suggests that ploidy levels in the populations of Korean irises examined in this study are at least partially stable within species. The new data presented here will allow for better understanding of the dynamics and patterns of chromosomal evolutionary changes in the genus *Iris* [15].



Figure 3. Chromosome numbers (2*n*; solid black squares) and genome sizes (GS) of Korean irises mapped on the phylogenetic tree based on nrITS sequences modified and simplified from Sim et al. [27]. The mean 1 C-values per species are indicated with solid dots.

3.3. Evolution of Chromosome Numbers and Genome Sizes

The newly obtained chromosome numbers and genome sizes were mapped on nrITS phylogeny of Korean irises (modified from [27]). The data revealed that there is no correlation between chromosome numbers and genome sizes among the phylogenetic lineages of the analyzed Korean *Iris* species. As an example, *Iris ensata* with low chromosome number of 2n = 24 has the highest 1 C-value (7.76 pg/1 C) whereas its close relative *I. setosa* has 2n = 38, but 1.4-fold lower genome size (Figure 4). It suggests that dysploidy and polyploidy are not solely responsible for the observed variation of genome sizes and chromosome numbers, but also other processes, like independent accumulation or reduction of repetitive DNAs amount (e.g., satellite DNAs and/or transposable elements) play an important role, as previously reported in other plant groups [7,40–43].

Other lineages within the genus *Iris* might be more stable evolutionarily. Both *Iris ruthenica* and *I. uniflora* (subgenus *Limniris* section *Ioniris*) have the same chromosome numbers of 2n = 42 and similar genome sizes (ca. 2.4 pg/1 C). Thus, the current cytogenetic data support the monophyly of this section, as proposed based on phylogenetic analyses of plastid markers [44–46], but also suggest low levels of genome dynamics accompanying their evolution. *Iris dichotoma* and *I. domestica* of the subgenus *Pardanthopsis* have also been recovered in monophyletic clade in ITS- and plastid-based phylogenetic analyses [46,47]. However, although they share the same chromosome numbers of 2n = 32, they differ nearly a 1.5-fold in genome sizes. Such variation of genome sizes suggests independent accumulation/loss of repetitive DNAs in the absence of numerical chromosome changes as reported also in other monocot genera [7,40].

In contrast, allopolyploidy can be hypothesized for the origin of *Iris koreana* (2n = 50) from closely related *I. minutoaurea* (2n = 22) and *I. odaesanensis* (2n = 28) [30]. It is also supported from the additive genome size of this taxon. Allopolyploidy has been implied to be an important process in the early diversification of the whole family of Iridaceae [48,49]. Thus, further data employing GISH (genomic *in situ* hybridization) will allow testing the allopolyploid origin of this and other species in the section *Limniris*.



Figure 4. Distribution of genome size data [1 C (pg)] in relation to chromosome number (2*n*). Colors represent subgenus *Limniris* section *Limniris* (red), section *Ioniris* (green), subgenus *Pardanthopsis* (black), and *Sisyrinchium rosulatum* (blue).

The subgenus *Limniris* section *Limniris* has previously been shown to be polyphyletic [27,30,44–46]. This study shows that there is also quite wide variation of chromosome numbers and genome sizes within the newly identified polyphyletic clades of this subgenus (Figures 3 and 4). It suggests that the genome size and chromosome number evolved rather dynamically in those evolutionary lineages, as indeed also observed in other monocot genera [11,50,51].

4. Conclusions

The present study provides the first comprehensive analyses of chromosome numbers, karyotype features and genome size variation of Korean irises from natural populations. Despite the large variation of chromosome numbers (n = 11, 12, 14, 16, 17, 19, 20, 21, 25), some variation of genome sizes (2.42–7.76 pg/1 C) has been demonstrated. This variation is only partially phylogenetically structured. Numerical chromosomal changes, both polyploidy and dysploidy, seem to be accompanied to a different degree by the likely independent changes in the amounts of repetitive DNAs in different phylogenetic lineages. Various combinations of these processes acting on the genomes of closely related species create complex patterns of genome size and chromosome number variation observed in Korean irises. Further analyses are needed to elucidate how these processes affect the evolution of the genomes of individual species, not only of the *Iris* species in Korea but ideally in a broader phylogenetic context of whole subgenera/sections.

Supplementary Materials: The following are available online at http://www.mdpi.com/2223-7747/9/10/1284/s1, Table S1. Chromosome counts from the published data according to Rice et al. [17].

Author Contributions: Conceptualization, B.C. and T.-S.J.; Collection and identification of plant material, B.C., S.S., H.-H.M., and T.-S.J.; Methodology, B.C., E.M.T., and T.-S.J.; Formal analysis, B.C., H.W.-S., E.M.T., and T.-S.J.; Writing—original draft preparation, B.C., H.W.-S., E.M.T., and T.-S.J.; Writing—review and editing, B.C., H.W.-S., and T.-S.J.; Visualization, B.C. and T.-S.J.; Supervision, T.-S.J.; Project administration, T.-S.J.; Funding acquisition, T.-S.J. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Research Foundation of Korea (NRF), grant number NRF-2018R1C1B6003170 to T.-S. Jang. Open access funding provided by grants from the Korea National Park Research Institute (NPRI 2019-18).

Acknowledgments: We thank Sun-Yu Kim (Nakdonggang National Institute of Biological Resources), Sungyu Yang and Jun-Ho Song (Korea Institute of Oriental Medicine), Chang-Ho Choi (Chollipo Arboretum), Il Noh (National Park Services), Seon-Wook Lee (National Park Services), Yusun Kweon, Yeongmun Choi, and all the lab members at CNUK (Herbarium of Chungnam National University) for their help in collecting materials for this study.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Lysak, M.A.; Berr, A.; Pecinka, A.; Schmidt, R.; McBreen, K.; Schuber, I. Mechanisms of chromosome number reduction in *Arabidopsis thaliana* and related Brassicaceae species. *Proc. Natl. Acad. Sci. USA* 2006, 103, 5224–5229. [CrossRef] [PubMed]
- 2. Schubert, I. Chromosome evolution. Curr. Opin. Plant Biol. 2007, 10, 109–115. [CrossRef] [PubMed]
- 3. Weiss-Schneeweiss, H.; Schneeweiss, G.M. Karyotype diversity and evolutionary trends in angiosperms. In *Plant Genome Diversity 2: Physical Structure, Behaviour and Evolution of Plant Genomes;* Springer: Vienna, Austria, 2013; Volume 2, pp. 209–230.
- Bennett, M.D.; Leitch, I.J. Nuclear DNA amounts in angiosperms: Targets, trends and tomorrow. *Ann. Bot.* 2011, 107, 467–590. [CrossRef] [PubMed]
- 5. Weiss-Schneeweiss, H.; Blöch, C.; Turner, B.; Villaseñor, J.L.; Stuessy, T.F.; Schneeweiss, G.M. The promiscuous and the chaste: Frequent allopolyploid speciation and its genomic consequences in American Daisies (*Melampodium* sect. *Melampodium*; Asteraceae). *Evolution* **2012**, *66*, 211–228. [CrossRef]
- 6. Jang, T.S.; McCann, J.; Parker, J.S.; Takayama, K.; Hong, S.P.; Schneeweiss, G.M.; Weiss-Schneeweiss, H. rDNA loci evolution in the genus *Glechoma* (Lamiaceae). *PLoS ONE* **2016**, *11*, e0167177. [CrossRef] [PubMed]
- Jang, T.S.; Parker, J.S.; Emadzade, K.; Temsch, E.M.; Leitch, A.R.; Weiss-Schneeweiss, H. Multiple origins and nested cycles of hybridization result in high tetraploid diversity in the monocot *Prospero. Front. Plant Sci.* 2018, 9, 433. [CrossRef]
- 8. Róis, A.S.; Castro, S.; Loureiro, J.; Sádio, F.; Rhazi, L.; Guara-Requena, M.; Caperta, A.D. Genome sizes and phylogenetic relationships suggest recent divergence of closely related species of the *Limonium vulgare* complex (Plumbaginaceae). *Plant Syst. Evol.* **2018**, *304*, 955–967. [CrossRef]
- 9. Sassone, A.B.; López, A.; Hojsgaard, D.H.; Giussani, L.M. A novel indicator of karyotype evolution in the tribe Leucocoryneae (Allioideae, Amaryllidaceae). *J. Plant Res.* **2018**, *131*, 211–223. [CrossRef]
- 10. Choi, B.; Yang, S.; Song, J.H.; Jang, T.S. Karyotype and genome size variation in *Ajuga* L. (Ajugoideae-Lamiaceae). *Nord. J. Bot.* **2019**, *37*, e02337. [CrossRef]
- Cruz, G.A.S.; Filho, J.R.M.; Vasconcelos, S.; Gitaí, J.; Campos, J.S.; Viccini, L.F.; Zizka, G.; Leme, E.M.C.; Brasileiro-Vidal, A.C.; Benko-Iseppon, A.M.B. Genome size evolution and chromosome numbers of species of the cryptanthoid complex (Bromelioideae, Bromeliaceae) in a phylogenetic framework. *Bot. J. Linn. Soc.* 2020, 192, 887–899. [CrossRef]
- Tlili, A.; Gouja, H.; Vallés, J.; Garnatje, T.; Buhagiar, J.; Neffati, M. Chromosome number and genome size in *Atriplex mollis* from southern Tunisia and *Atriplex lanfrancoi* from Malta (Amaranthaceae). *Plant Syst. Evol.* **2020**, 306, 11. [CrossRef]
- Leitch, I.J.; Leitch, A.R. Genome size diversity and evolution in land plants. In *Plant Genome Diversity* 2: *Physical Structure, Behaviour and Evolution of Plant Genomes*; Springer: Vienna, Austria, 2013; Volume 2, pp. 307–322.
- 14. Schubert, I.; Vu, G.T.H. Genome stability and evolution: Attempting a holistic view. *Trends Plant Sci.* **2016**, 21, 749–757. [CrossRef] [PubMed]
- 15. Pellicer, J.; Leitch, I.J. The plant DNA C-values database (release 7.1): An updated online repository of plant genome size data for comparative studies. *New Phytol.* **2020**, *226*, 301–305. [CrossRef] [PubMed]
- 16. Goldblatt, P.; Walbot, V.; Zimmer, E. Estimation of genome size (C-value) in Iridaceae by cytophotometry. *Ann. Missouri Bot. Gard.* **1984**, *71*, 176–180. [CrossRef]
- 17. Rice, A.; Glick, L.; Abadi, S.; Einhorn, M.; Kopelman, N.M.; Salman-Minkov, A.; Mayzel, J.; Chay, O.; Mayrose, I. The chromosome counts database (CCDB): A community resource of plant chromosome numbers. *New Phytol.* **2015**, *206*, 19–26. [CrossRef]

- Goldblatt, P.; Manning, J.C.; Rudall, P.J. Iridaceae. In *The Families and Genera of Vascular Plants. Vol. 3,* Flowering Plants. Monocotyledons: Lilianae (Except Orchidaceae); Springer: Berlin, Germany, 1998; pp. 295–333.
- Zhao, Y.T.; Noltie, H.J.; Mathew, B. Iridaceae. In *Flora of China*; Missouri Botanical Garden Press: St. Louis, MA, USA, 2000; Volume 24, pp. 297–313.
- 20. Wilson, C.A. Patterns of evolution in characters that define *Iris* subgenera and sections. *Aliso* **2006**, *22*, 425–433. [CrossRef]
- Guo, J.; Wilson, C.A. Molecular phylogeny of crested *Iris* based on five plastid markers (Iridaceae). *Syst. Bot.* 2013, *38*, 987–995. [CrossRef]
- 22. Rodionenko, G.I. The Genus Iris; Academy of Science USSR: Moscow, Russia, 1961.
- 23. Mathew, B. The Iris; Batsford: London, UK, 1989.
- 24. Wilson, C.A. Subgeneric classification in *Iris* re-examined using chloroplast sequence data. *Taxon* **2011**, *60*, 27–35. [CrossRef]
- Sim, J.K. Iridaceae Juss. In *The Genera of Vascular Plants of Korea*; Academy Publ. Co.: Seoul, Korea, 2007; pp. 1326–1331.
- 26. Lee, T.B. Colored Flora of Korea; Hyangmunsa: Seoul, Korea, 2003; Volume II.
- 27. Sim, J.K.; Park, H.D.; Park, S.J. Phylogenetic study of Korean *Iris* (Iridaceae) based on nrDNA ITS sequences. *Korean J. Plant Taxon.* **2002**, *32*, 33–53. [CrossRef]
- Shin, H.; Kim, Y.D.; Lim, Y.S.; Kim, Y.I. Endangered vascular plants in the republic of Korea. In Korean Red List of Threatened Species, 2nd ed.; National Institute of Biological Resources: In-Cheon, Korea, 2014; pp. 137–194.
- 29. Lee, H.J.; Park, S.J. A phylogenetic study of Korean *Iris* L. based on plastid DNA (*psbA-trnH, trnL-F*) sequences. *Korean J. Plant Taxon.* **2013**, *43*, 227–235. [CrossRef]
- 30. Son, O.; Son, S.W.; Suh, G.U.; Park, S.J. Natural hybridization of *Iris* species in Mt. Palgong-san, Korea. *Korean J. Plant Taxon.* **2015**, *45*, 243–253. [CrossRef]
- 31. Kim, H.H.; Park, Y.W.; Yoon, P.S.; Choi, H.W.; Bang, J.W. Karyotype analysis of eight Korean native species in the genus *Iris. Korean J. Medicinal Crop Sci.* **2004**, *12*, 401–405. [CrossRef] [PubMed]
- 32. Choi, B.; Kim, S.Y.; Jang, T.S. Micromorphological and cytological comparisons between *Youngia japonica* and *Y. longiflora* using light and scanning electron microscopy. *Microsc. Res. Techniq.* **2020**, in press. [CrossRef] [PubMed]
- 33. MicroMeasure 3.3 Program. Available online: https://micromeasure.software.informer.com/3.3 (accessed on 1 August 2020).
- 34. Temsch, E.M.; Greilhuber, J.; Krisai, R. Genome size in liverworts. Preslia 2010, 82, 63-80.
- 35. Greilhuber, J.; Ebert, I. Genome size variation in Pisum sativum. Genome 1994, 37, 646–655. [CrossRef]
- 36. Otto, F.; Oldiges, H.; Gohde, W.; Jain, V.K. Flow cytometric measurement of nuclear DNA content variations as a potential in vivo mutagenicity test. *Cytometry* **1981**, *2*, 189–191. [CrossRef]
- Martínez, J.; Vargas, P.; Luceño, M.; Cuadrado, Á. Evolution of *Iris* subgenus *Xiphium* based on chromosome numbers, FISH of nrDNA (5S, 45S) and *trnL-trnF* sequence analysis. *Plant Syst. Evol.* 2010, 289, 223–235. [CrossRef]
- Siljak-Yakovlev, S.; Pustahija, F.; Šolić, E.M.; Bogunić, F.; Muratović, E.; Bašić, N.; Catrice, O.; Brown, S.C. Towards a genome size and chromosome number database of Balkan Flora: C-values in 343 taxa with novel values for 242. *Adv. Sci. Lett.* 2010, *3*, 190–213. [CrossRef]
- 39. Samad, N.A.; Dagher-Kharrat, M.B.; Hidalgo, O.; Zein, R.E.; Douaihy, B.; Siljak-Yakovlev, S. Unlocking the karyological and cytogenetic diversity of *Iris* from Lebanon: *Oncocyclus* section shows a distinctive profile and relative stasis during its continental radiation. *PLoS ONE* **2016**, *11*, e0160816.
- 40. Emadzade, K.; Jang, T.S.; Macas, J.; Kovařík, A.; Novák, P.; Parker, J.; Weiss-Schneeweiss, H. Differential amplification of satellite *PaB6* in chromosomally hypervariable *Prospero autumnale* complex (Hyacinthaceae). *Ann. Bot.* **2014**, *114*, 1597–1608. [CrossRef]
- Weiss-Schneeweiss, H.; Leitch, A.R.; McCann, J.; Jang, T.S.; Macas, J. Employing next generation sequencing to explore the repeat landscape of the plant genome. In *Next Generation Sequencing in Plant Systematics*. *Regnum Vegetabile* 157; Koeltz Scientific Books: Königstein, Germnay, 2015; Volume 158, pp. 155–179.
- 42. Macas, J.; Novák, P.; Pellicer, J.; Čížková, J.; Koblížková, A.; Neumann, P.; Fuková, I.; Doležel, J.; Kelly, L.J.; Leitch, I.J. In depth characterization of repetitive DNA in 23 plant genomes reveals sources of genome size variation in the legume tribe Fabeae. *PLoS ONE* 2015, *10*, e0143424. [CrossRef] [PubMed]

- 43. McCann, J.; Jang, T.S.; Macas, J.; Schneeweiss, G.M.; Matzke, N.J.; Novák, P.; Stuessy, T.F.; Villaseñor, J.L.; Weiss-Schneeweiss, H. Dating the species network: Allopolyploidy and repetitive DNA evolution in American Daisies (*Melampodium* sect. *Melampodium*, Asteraceae). *Syst. Biol.* **2018**, *67*, 1010–1024. [CrossRef] [PubMed]
- 44. Makarevitch, I.; Golovnina, K.; Scherbik, S.; Blinov, A. Phylogenetic relationships of the Siberian *Iris* species inferred from noncoding chloroplast DNA sequences. *Int. J. Plant Sci.* **2003**, *164*, 229–237. [CrossRef]
- 45. Wilson, C.A. Phylogenetic relationships among the recognized series in *Iris* section *Limniris*. *Syst. Bot.* **2009**, 34, 277–284. [CrossRef]
- 46. Jiang, Y.L.; Huang, Z.; Liao, J.Q.; Song, H.X.; Luo, X.M.; Gao, S.P.; Lei, T.; Jiang, M.Y.; Jia, Y.; Chen, Q.B.; et al. Phylogenetic analysis of *Iris* L. from China on chloroplast *trn*L-F sequences. *Biologia* **2018**, *73*, 459–466. [CrossRef]
- 47. Wilson, C.A. Phylogeny of *Iris* based on chloroplast *mat*K gene and *trn*K intron sequence data. *Mol. Phylogenet. Evol.* **2004**, *33*, 402–412. [CrossRef]
- 48. Goldblatt, P.; Takei, M. Chromosome cytology of Iridaceae—Patterns of variation, determination of ancestral base numbers, and modes of karyotype change. *Ann. Missouri Bot. Gard.* **1997**, *84*, 285–304. [CrossRef]
- 49. Lim, K.Y.; Matyasek, R.; Kovařík, A.; Leitch, A.R. Parental origin and genome evolution in the allopolyploids *Iris versicolor. Ann. Bot.* **2007**, *100*, 219–224. [CrossRef]
- 50. Jersáková, J.; Trávníček, P.; Kubátová, B.; Krejčíková, J.; Urfus, T.; Liu, Z.J.; Lamb, A.; Ponert, J.; Schulte, K.; Curn, V.; et al. Genome size variation in Orchidaceae subfamily Apostasioideae: Filling the phylogenetic gap. *Bot. J. Linn. Soc.* **2013**, *172*, 95–105. [CrossRef]
- Šmarda, P.; Bureš, P.; Horová, L.; Leitch, I.J.; Mucina, L.; Pacini, E.; Tichý, L.; Grulich, V.; Rotreklová, O. Ecological and evolutionary significance of genomic GC content diversity in monocots. *Proc. Natl. Acad. Sci. USA* 2014, *111*, 4096–4102. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).