

Original Article
Pharmacology,
Drug Therapy & Toxicology



A Meta-Analysis on the Performance of Cystatin C- versus Creatinine-based eGFR Equations in Predicting Vancomycin Clearance

Nor Asyikin Mohd Tahir ,¹ Shamin Mohd Saffian ,¹ Farida Hanim Islahudin ,¹ Abdul Halim Abdul Gafor ,² and Mohd Makmor-Bakry ¹

¹Faculty of Pharmacy, Universiti Kebangsaan Malaysia, Kuala Lumpur, Malaysia

²Nephrology Unit, Department of Medicine, Universiti Kebangsaan Malaysia Medical Centre, Cheras, Malaysia



Received: Apr 5, 2020

Accepted: Jun 25, 2020

Address for Correspondence:

Mohd Makmor-Bakry, PhD

Faculty of Pharmacy, Universiti Kebangsaan Malaysia, Jalan Raja Muda Abdul Aziz, 50300 Kuala Lumpur, Malaysia.

E-mail: mohdclinpharm@ukm.edu.my

© 2020 The Korean Academy of Medical Sciences.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ORCID iDs

Nor Asyikin Mohd Tahir

<https://orcid.org/0000-0002-7330-7148>

Shamin Mohd Saffian

<https://orcid.org/0000-0001-6243-6975>

Farida Hanim Islahudin

<https://orcid.org/0000-0002-4450-4361>

Abdul Halim Abdul Gafor

<https://orcid.org/0000-0003-3950-2844>

Mohd Makmor-Bakry

<https://orcid.org/0000-0002-0592-0645>

Disclosure

The authors have no potential conflicts of interest to disclose.

Author Contributions

Conceptualization: Mohd Tahir NA, Mohd Saffian S, Makmor-Bakry M. Data curation:

ABSTRACT

Background: The objective of this study was to compare the performance of cystatin C- and creatinine-based estimated glomerular filtration rate (eGFR) equations in predicting the clearance of vancomycin.

Methods: MEDLINE and Embase databases were searched from inception up to September 2019 to identify all studies that compared the predictive performance of cystatin C- and/or creatinine-based eGFR in predicting the clearance of vancomycin. The prediction errors (PEs) (the value of eGFR equations minus vancomycin clearance) were quantified for each equation and were pooled using a random-effects model. The root mean squared errors were also quantified to provide a metric for imprecision.

Results: This meta-analysis included evaluations of seven different cystatin C- and creatinine-based eGFR equations in total from 26 studies and 1,234 patients. The mean PE (MPE) for cystatin C-based eGFR was 4.378 mL min⁻¹ (95% confidence interval [CI], -29.425, 38.181), while the creatinine-based eGFR provided an MPE of 27.617 mL min⁻¹ (95% CI, 8.675, 46.560) in predicting clearance of vancomycin. This indicates the presence of unbiased results in vancomycin clearance prediction by the cystatin C-based eGFR equations. Meanwhile, creatinine-based eGFR equations demonstrated a statistically significant positive bias in vancomycin clearance prediction.

Conclusion: Cystatin C-based eGFR equations are better than creatinine-based eGFR equations in predicting the clearance of vancomycin. This suggests that utilising cystatin C-based eGFR equations could result in better accuracy and precision to predict vancomycin pharmacokinetic parameters.

Keywords: Cystatin C; Creatinine; Vancomycin; Clearance; eGFR

INTRODUCTION

Vancomycin is a glycopeptide antibiotic which has been widely used for the treatment of Methicillin-Resistant *Staphylococcus aureus* infection.¹ Concentrations of vancomycin that are too high could result in severe side effects such as nephrotoxicity,² hepatotoxicity,³ and ototoxicity,⁴ especially if it is used together with aminoglycosides.⁵ Subtherapeutic levels

Mohd Tahir NA, Mohd Saffian S. Formal analysis: Mohd Tahir NA, Mohd Saffian S. Investigation: Mohd Tahir NA, Mohd Saffian S. Methodology: Mohd Tahir NA, Mohd Saffian S. Software: Mohd Tahir NA, Mohd Saffian S. Validation: Mohd Tahir NA, Mohd Saffian S. Writing- original draft: Mohd Tahir NA, Mohd Saffian S, Makmor-Bakry M. Writing - review & editing: Mohd Tahir NA, Mohd Saffian S, Islahudin FH, Makmor-Bakry M.

of vancomycin on the other hand, could result in therapeutic failure and development of resistant microorganisms.^{6,7} As such, the narrow therapeutic range of vancomycin and a trend towards higher minimum inhibitory concentration increases the importance of accurate dosing of vancomycin to ensure its safety and efficacy.⁶ Vancomycin is primarily eliminated by the kidneys and hence the maintenance dose is determined by the glomerular filtration rate (GFR).

Accurate estimation of a patient's kidney function is highly essential during vancomycin use. Numerous methods can be used to determine GFR, with estimated creatinine clearance (CL_{cr}) widely accepted as the clinical reference standard.⁸ Renal clearance is usually estimated based on Cockcroft-Gault equation and is assumed to be relative to the renal clearance of vancomycin.⁹⁻¹¹ However, studies conducted previously found that serum creatinine-based estimated GFR (eGFR) was associated with several factors such as age, muscle mass and sex. Overestimation of eGFR often occurs amongst the elderly, highly obese, patients with reduced muscle mass, malnutrition, hepatic dysfunction and bedridden patients.¹²⁻¹⁴ Tubular secretion of creatinine could also increase as chronic kidney disease develop progressively to a later stage, thus, results in an unpredictable overestimation of GFR. Some drugs such as trimethoprim,¹⁵ cimetidine,¹⁶ and antiretroviral drugs,¹⁷ could block the tubular secretion of creatinine, leading to increasing serum creatinine concentration without the presence of GFR modification. Therefore, an alternative biomarker with greater precision is required to accurately measure renal function status for better use of vancomycin.¹⁸

One alternative endogenous biomarker of glomerular filtration that has been suggested is cystatin C. It is a 13 kDa non-glycosylated cysteine protease inhibitor.¹⁹ Cystatin C has a stable production rate because it is produced virtually by all nucleated cells in the body.^{19,20} In a normally functioning kidney, cystatin C is freely filtered by the glomerulus, reabsorbed and catabolised at the renal proximal tubules.¹⁹ Serum cystatin C level also showed less dependence on other factors such as age, sex, weight, and muscle mass compared to serum creatinine.^{21,22} Therefore, the serum concentration of cystatin C will directly reflect the kidney function status. Precise dosing is crucial for drugs with a narrow therapeutic index and/or patient with unpredictable renal function estimates. Thus, Kidney Disease: Improving Global Outcomes (KDIGO) 2012 guidelines outlined a recommendation to use methods based on serum cystatin C for GFR estimation. These recommendations are based on many reports which revealed that cystatin C is better than creatinine in GFR determination because it is not affected by hepatic dysfunction, inflammation, body surface area, age, sex, diet and muscle mass.²³ A meta-analysis conducted previously also found cystatin C showed greater accuracy than creatinine as a biomarker for patients with impaired renal function.²⁴

In spite of these results, the use of serum cystatin C for drug dosage adjustment remains limited, partly because many clinicians do not believe its utility.²⁵ Several studies, often with small sample sizes, have compared the performance of serum cystatin C- and creatinine-based eGFR equations in predicting vancomycin clearance. However, missing from the literature is a comprehensive quantitative assessment on the predictive ability or usefulness of serum cystatin C-based eGFR equations to predict the clearance of vancomycin compared to creatinine. The purpose of this study is to systematically review the literature and compare the effectiveness of pooled cystatin C- and creatinine-based eGFR equations in predicting vancomycin clearance.

METHODS

Identification of relevant studies and eligibility criteria

Relevant studies were located through a literature search in MEDLINE (1946 to April 2019) and Embase (1947 to April 2019) to identify all studies that compared the predictive performance of cystatin C- and/or creatinine-based eGFR in predicting the clearance of vancomycin. The literature search was updated in September 2019 to include any recent publications. The MEDLINE MeSH terms and corresponding keywords used are “cystatin C”, “creatinine”, “eGFR”, “pharmacokinetic”, and “vancomycin”. In the search strategy, subject terms and keywords were applied in combination. The search was limited to human studies and articles published in the English language. The reference lists from the key review publications were further examined for potentially relevant studies as well as citations of the studies identified.

Inclusion and exclusion criteria

Studies that were identified from the database search were screened based on the title and abstract. A full-text assessment was carried out on all studies that compare the performance of cystatin C- and/or creatinine-based eGFR in predicting the clearance of vancomycin. These steps were conducted by two authors. Most of these data come in the form of a scatterplot of the eGFR and vancomycin clearance values. Studies were excluded if they met any of the following criteria: 1) they are review articles, meta-analyses, commentaries, editorial letters, case reports, 2) only summary data of cystatin C- or creatinine-based eGFR values against vancomycin clearance were provided, 3) the clearance values cannot be standardised in mL min⁻¹, 4) no scatter plot or raw data was provided, and 5) the published scatter plot was of poor resolution for the data to be extracted.

Data extraction

Data extracted from the published studies were based on a standardized form, and any discrepancies in data extraction process were resolved by consensus between authors. Study characteristics extracted from the studies were the year of publication, study size, type of patient's group, age, methods to analyse serum concentration of cystatin C, creatinine, vancomycin, type of cystatin C- or creatinine-based eGFR equations used and methods of vancomycin clearance estimation. Data points from each scatter plot were digitized using the WebPlotDigitizer version 4.1 (Austin, TX, USA). Differences in the number of data points on the scatter plot and the number of subjects recruited were found in some studies. Subsequent calculation and analysis were based on the number of data points, instead of the number of subjects being reported. The reproducibility of the data extraction was evaluated by replicating the data extraction process by a researcher who was not involved in this study. The relative difference between the original and replicate of the extracted data points was compared based on the correlation coefficient or coefficient of determination, depending on which information was provided. A difference of < 5% was considered to be acceptable.

The predictive performance of each cystatin C- and creatinine-based eGFR equations was assessed for bias (mean prediction error [MPE]) and imprecision (mean squared error [MSE] and root MSE [RMSE]) according to Sheiner and Beal²⁶ based on the following equation:

$$MPE (mL \min^{-1}) = \frac{\sum_{i=1}^N PE_i}{N}$$

$$MSE = \frac{\sum_{i=1}^N (PE_i)^2}{N}$$

$$RMSE = \sqrt{MSE}$$

where PE_i is the prediction error (the value of eGFR equations minus vancomycin clearance for the i^{th} observation). N is the number of data points extracted from each study. Magnitude and direction of bias was expressed as $MPE \pm 95\%$ confidence interval (CI) of the difference plots. The RMSE defined as average discrepancy of the prediction from the reference values were reported in the original units of the measure. An RMSE of 0 indicates a completely precise predictions.²⁶

Data-analysis and statistical method

The meta-analyses were performed based on the generic inverse variance method using MedCalc for Windows, version 17.9.7 (MedCalc Software, Ostend, Belgium). In this method, each study estimate of the relative treatment is assigned with a weight equivalent to the inverse of the variance of the effect estimate. This method is used to report the difference between the means and the standard error of this difference based on the continuous data. Several cystatin C- and creatinine-based eGFR equations used in more than one study were considered as a repeated measure. Information on the type of eGFR equations used was extracted from each study and similar equations were pooled together. Results on MPE and its standard error mean were compared between pooled cystatin C-based eGFR and creatinine-based eGFR equations through a three-stage hierarchical meta-analytical model. Comparison of each biomarker in predicting the vancomycin clearance will be determined based on the 95% CI of the pooled MPE and RMSE. If the 95% CI of MPE includes zero, it will indicate the presence of unbiased results. The predictions will be considered not to differ significantly from the actual values. Meanwhile, smaller RMSE value will indicate a greater precision of predictive ability by each biomarker.

Heterogeneity analysis for each study was conducted by using the χ^2 test. A P value of less than 0.05 was considered statistically significant. Heterogeneity test based on I^2 statistic were used whereby an I^2 value of 75%, 50%, and 25% is considered as having a high, moderate and low heterogeneity respectively.²⁷ The meta-analysis was based on a random effects model given a priori assumption of significant heterogeneity between studies. Potential causes of heterogeneity were investigated by conducting sensitivity analysis. This was done by excluding one study at a time to determine its influence on the pooled effects.²⁸ If the overall results and conclusions are consistent with the primary analysis, with a reduction in the I^2 value, the study will be considered as an outlier. A pre-specified subgroup analysis among adult patients and methods to estimate vancomycin clearance (e.g., Bayesian, two-stage, Sawchuk-Zaske and population pharmacokinetic analysis) were also conducted. Publication bias was examined using funnel plots and Duval-Tweedie's Trim and Fill method.²⁹

Proportional bias

The data point of eGFR and vancomycin clearance values extracted were further analysed to investigate for the presence of any systematic trend of bias across the different range of vancomycin clearance. A proportional bias, in this case, is defined as a systematic deviation of the PE were plotted against the values of vancomycin clearance on the X-axis. The presence of statistically significant proportional bias is defined when the value of the slope of the difference plot does not equal to zero with a P value less than 0.05.

RESULTS

Literature search results and characteristics of the eligible studies

The literature search retrieved 1,309 articles from MEDLINE and Embase databases. Two additional studies were identified in a manual search of the reference list. Of them, 633 articles were removed due to duplication. After screening by title and abstract, 106 studies investigating the relationship between serum cystatin C and creatinine with vancomycin pharmacokinetics were retrieved for further assessment by studying the full-text articles. Further 80 studies were excluded because the inclusion criteria were not met. Flow diagram for the literature search and study selection process is shown in Fig. 1.

There were 26 studies included in the final analysis.^{9,30-54} Four of the studies provided information on the correlation between both cystatin C- and creatinine-based eGFR with vancomycin clearance.^{33,38,53,54} The study population consisted of patients with various conditions such as burn patients, those who were admitted to the intensive critical unit and patients with unstable renal function. Study characteristics such as assays for serum creatinine, cystatin C and vancomycin level, equations used for each biomarker as well as methods to estimate vancomycin clearance were also presented (Tables 1 and 2, Supplementary Table 1). Data from 1,234 patients were available for analysis.

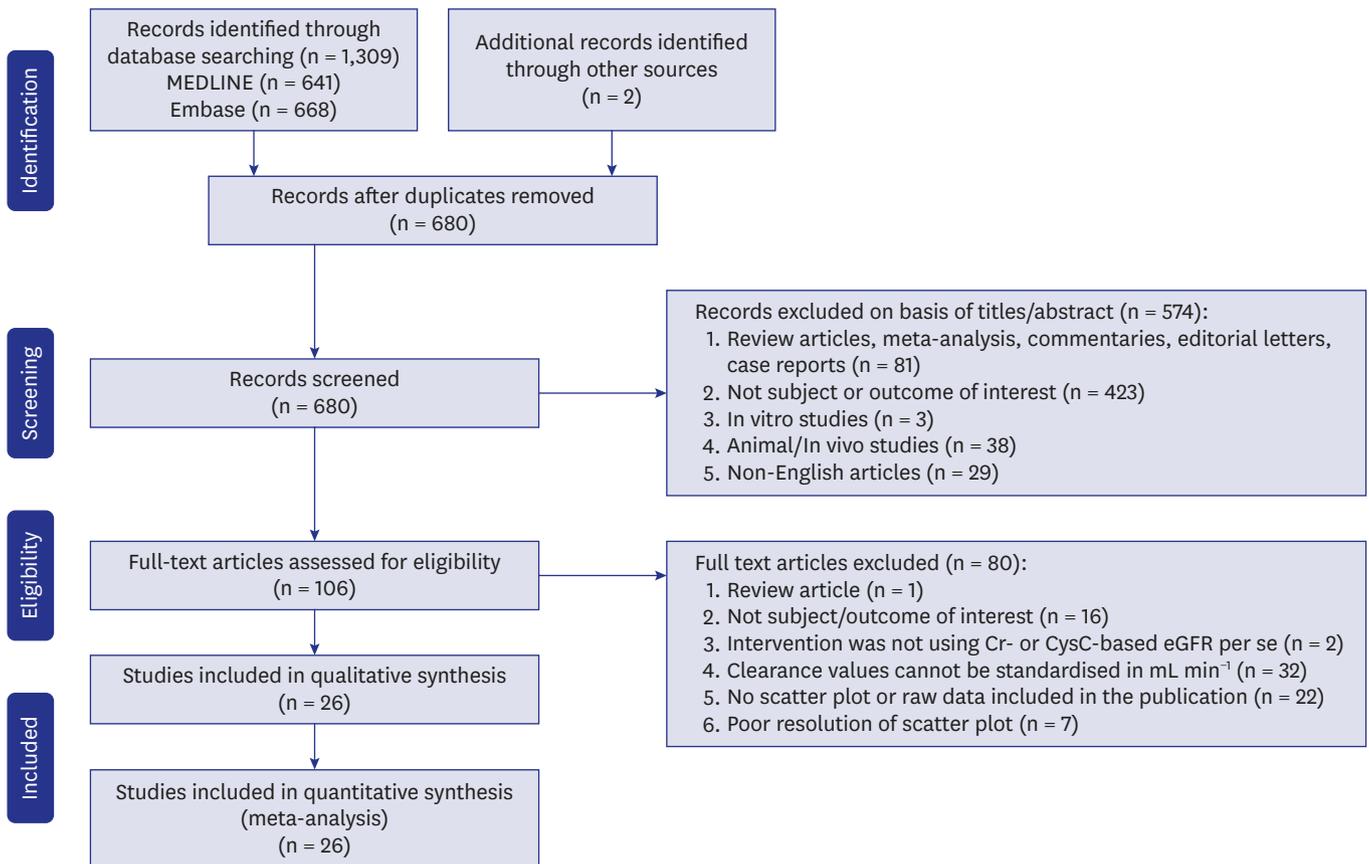


Fig. 1. Study selection process for the meta-analysis.

Cr = creatinine, CysC = cystatin C, eGFR = estimated glomerular filtration rate.

Cystatin C versus Creatinine in Vancomycin Clearance

Table 1. Summary of published studies on CysC-based eGFR equations in predicting vancomycin clearance

CysC-based eGFR equations	Studies	Population			Analytical measure		SCysC, mg/L	eGFR _{CysC} , mL/min ^c	Methods to determine CL _{vanC}	Results from the published studies
		No. of patients	Patients	Age range, yr	CysC	Vancomycin				
Larsson	Okamoto et al. ³³	24	Elderly patients	66–87	PETIA	FPIA	1.28	NA	Bayesian	$\rho = 0.883$ ($P < 0.001$)
	Shin et al. ⁵³	50	Term neonates	19.12 ± 15.55 days ^{a,b}	PETIA	FPIA	1.44	63.63	Bayesian	$r = 0.496$ ($P = 0.001$)
Hoek	Kees et al. ³⁸	25	ICU patients	31–82	PENIA	HPLC	NA	NA	Two-stage approach	$r^2 = 0.70$ ($P < 0.001$)
Flodin	Chen et al. ⁵⁴	65	MRSA-infected patients	19–78	PENIA	FPIA	1.35	71	Two-stage approach	$r^2 = 0.8465$ ($P < 0.001$)

CysC = cystatin C, eGFR = estimated glomerular filtration rate, SCysC = serum cystatin C, CL_{vanC} = clearance of vancomycin, MRSA = methicillin-resistant *Staphylococcus aureus*, ICU = intensive care unit, PENIA = particle-enhanced immunonephelometry assay, PETIA = particle-enhanced immunoturbidimetry assay, HPLC = high performance liquid chromatography, NA = not available, r = correlation coefficient, r^2 = coefficient of determination.

^aAge reported in mean ± standard deviation; ^bAge reported in days; ^ceGFR were variably expressed as means or medians or the measure of central tendency was not reported; eGFR was variably expressed as mL/min or mL/min/1.73 m².

Table 2. Summary of published studies on Cr-based eGFR in predicting vancomycin clearance

Cr-based GFR equations	Studies	Population			Analytical measure		SCr, mg/dL	eGFR _{Cr} , mL/min ^c	Methods to determine CL _{vanC}	Results from the published studies
		No. of patients	Patients	Age-range, yr	Cr	Vancomycin				
CG	Matzke et al. ⁹	56	Patients with various degrees of renal function	17–85	NA	Radio-immunoassay	NA	NA	Sawchuk-Zaske	$r = 0.881$
	Birt et al. ³⁰	22	Adult inpatients	28–79	NA	FPIA	NA	97.4	Sawchuk-Zaske	$r = 0.703$ ($P < 0.001$)
	Polard et al. ³¹	16	Adult ICU patients	19–75	NA	Enzyme-multiplied immunoassay technique	0.7	124	Bayesian	$r^2 = 0.465$ ($P = 0.004$)
	Yoshida et al. ³²	8	Patients with pneumonia	65–88	NA	FPIA	NA	54.92	Bayesian	NA
	Okamoto et al. ³³	24	Elderly patients	66–87	Enzymatic	FPIA	0.63	NA	Bayesian	$\rho = 0.684$ ($P < 0.001$)
	Omote et al. ³⁴	106	Adult Japanese cancer patients with various degrees of renal function	24–92	FPIA	FPIA	0.8	87.05	Bayesian	NA
	Pea et al. ³⁵	70	Critically ill patients	59 ± 18 ^a	NA	FPIA	NA	95.9	Two-stage approach	$r = 0.75$ ($P < 0.001$)
	Yamamoto et al. ³⁶	106	Adult patient & healthy volunteers	20–100	NA	FPIA & bioassay method	0.96	84.45	Bayesian	$r^2 = 0.361$ $r = 0.601$
	Dolton et al. ³⁷	70	Burn patients & control patients	15–95	NA	FPIA	NA	99.7	Bayesian	Burns, $r^2 = 0.572$ Control, $r^2 = 0.758$
	Kees et al. ³⁸	25	ICU patients	31–82	Enzymatic	HPLC	NA	106	Two-stage approach	$r^2 = 0.37$ ($P = 0.001$)
	Jeurissen et al. ³⁹	20	Critically ill patients	Adult (age not specified)	NA	ELISA	NA	95	Two-stage approach	$r = 0.83$ ($P < 0.001$)
	Lee et al. ⁴⁰	109	Spinal cord injury patients	66 ± 11 ^a	NA	NA	0.74	91.24	Sawchuk-Zaske	$r = 0.4$ ($P < 0.001$)
	Shimamoto et al. ⁴¹	101	Adult patients	34–95	NA	FPIA	0.72	82.1	Bayesian	$r^2 = 0.649$
	Shimamoto et al. ⁴²	105	Adult patients	52 ± 17 ^a	NA	FPIA	NA	137.65	Bayesian	$r = 0.70$ ($P < 0.01$)
	Adane et al. ⁴³	29	Extremely obese patients	39–53	NA	PETIA	NA	124.8	Population Pharmacokinetic Analysis	$r = 0.11$

(continued to the next page)

Table 2. (Continued) Summary of published studies on Cr-based eGFR in predicting vancomycin clearance

Cr-based GFR equations	Studies	Population			Analytical measure		SCr, mg/dL	eGFR _{Cr} , mL/min ^c	Methods to determine CL _{Van}	Results from the published studies Correlation of Cr-based eGFR with CL _{Van}
		No. of patients	Patients	Age-range, yr	Cr	Vancomycin				
Measured 24hr-CL _{Cr}	Nielsen et al. ⁴⁴	14	Patients with various degrees of renal function	21–68	Autoanalyzer	Agar diffusion method	NA	62		r = 0.90 (P < 0.001)
	Blouin et al. ⁴⁵	10	Normal and morbidly obese patients	25–37	Jaffe	Radioimmunoassay	NA	163.4	Two-stage approach	r = 0.783 (P < 0.025)
	Rotschafer et al. ⁴⁶	28	Patients with serious staphylococcal infection	18–80	NA	Agar diffusion method & radioimmunoassay	1.4	121.57	Two-stage approach	r = 0.45
	Garaud et al. ⁴⁷	10	Acutely ill patients	Adult (age not specified)	NA	HPLC	NA	55.9	Two-stage approach	r = 0.78 (P < 0.05)
	Brater et al. ⁴⁸	11	Burn patients	23–58	Continuous flow analysis	HPLC	NA	105	Bayesian	r = 0.932 (P < 0.001)
	Taber et al. ⁴⁹	10	Adult liver transplant recipients	24–68	NA	NA	1.1	43	Population Pharmacokinetic Analysis	r ² = 0.828 (P < 0.001)
	Dailly et al. ⁵⁰	70	Burn patients	43 ± 18 ^a	NA	FPIA	NA	173.3	Two-stage approach	r = 0.506 (P < 0.001)
	Chen et al. ⁵⁴	65	MRSA-infected patients	19–78	Enzymatic	FPIA	2.07	75.56	Two-stage approach	r ² = 0.637 (P < 0.001)
Park et al. ⁵¹	20	Patients on extracorporeal membrane oxygenation	50 ± 16 ^a	NA	FPIA	1.02	94.53	Sawchuk-Zaske	NA	
8hr-CL _{Cr}	Baptista et al. ⁵²	79	Critically ill patients	58 ± 16 ^a	NA	NA	0.68	125.1	Two-stage approach	r ² = 0.663 (P < 0.001)
Schwartz	Shin et al. ⁵³	50	Term neonates	19.12 ± 15.55 days ^{a,b}	Enzymatic	FPIA	0.46	70.32	Bayesian	r = 0.527 (P = 0.036)

Cr = creatinine, eGFR = estimated glomerular filtration rate, SCr = serum creatinine, CL_{Van} = clearance of vancomycin, MRSA = methicillin-resistant *Staphylococcus aureus*, ICU = intensive care unit, PETIA = particle-enhanced immunoturbidimetry assay, FPIA = fluorescence polarisation immunoassay, HPLC = high performance liquid chromatography, ELISA = enzyme-linked immunosorbent assay, 24hr-CL_{Cr} = 24-hour creatinine clearance, 8hr-CL_{Cr} = 8-hour creatinine clearance, CG = Cockcroft-Gault, NA = not available, r = correlation coefficient, r² = coefficient of determination.

^aAge reported in mean ± standard deviation; ^bAge reported in days; ^ceGFR or CL_{Cr} were variably expressed as means or medians or the measure of central tendency was not reported; eGFR was variably expressed as mL/min or mL/min/1.73m².

Meta-analysis on the performance of cystatin C- and creatinine-based eGFR equations in predicting vancomycin clearance

The MPE and RMSE per study and per eGFR equation were summarised in Table 3. Fig. 2 presents a forest plot illustrating the predictive performance of cystatin C- and creatinine-based eGFR equations in predicting the clearance of vancomycin. The pooled MPE obtained by cystatin C-based eGFR equations is 4.38 mL min⁻¹, 95% CI, -29.43, 38.18 mL min⁻¹ with significantly high heterogeneity, P < 0.001, I² = 96.92% when compared with the creatinine-based eGFR equations with a pooled MPE of 27.62 mL min⁻¹, 95% CI, 8.68, 46.56 mL min⁻¹ with significantly high heterogeneity, P < 0.001, I² = 87.74%. Cystatin C-based equations with a 95% CI of MPE that include zero indicates an unbiased prediction of vancomycin clearance. Meanwhile, there was statistically significant positive bias demonstrated by the creatinine-based eGFR equations. Greater precision was shown by cystatin C-based eGFR equations in predicting vancomycin clearance due to the smaller RMSE values, when compared with the creatinine-based eGFR equations (28.96 vs. 61.56 mL min⁻¹, respectively), suggesting a better consistency on the ability to predict vancomycin clearance.

Subgroup analysis

Our analysis was further stratified based on age group of patients (i.e., adults only) to further examine the generalisability of the pooled estimate. The study by Shin et al.⁵³ was excluded because the results were based on the neonate population. Re-analysis for the rest of the

Table 3. Summary of bias (MPE) and precision (RMSE) of Cr- and Cys C-based eGFR equations from study

Biomarkers	Equations	Studies	MPE, mL min ⁻¹	RMSE per study	RMSE per eGFR equation	Slope of difference plot (95% CI)
CysC	Larsson	Okamoto et al. ³³	16.21	20.26	26.35	-0.07 (-0.26, 0.12)
		Shin et al. ⁵³	-10.43	30.95		-0.48 (-0.77, -0.18) ^a
	Hoek	Kees et al. ³⁸	26.94	34.88	34.88	0.06 (-0.24, 0.35)
		Flodin	Chen et al. ⁵⁴	-16.90		29.56
Cr	CG	Matzke et al. ⁹	15.24	28.67	54.93	-0.21 (-0.62, 0.20)
		Birt et al. ³⁰	19.87	32.92		-0.26 (-0.61, 0.10)
		Polard et al. ³¹	25.34	39.70		-0.15 (-0.67, 0.38)
		Yoshida et al. ³²	4.17	15.21		-0.53 (-0.70, -0.36) ^a
		Okamoto et al. ³³	37.07	57.12		0.83 (0.21, 1.46) ^a
		Omote et al. ³⁴	26.96	40.46		0.24 (0.01, 0.47) ^a
		Pea et al. ³⁵	30.88	45.87		0.21 (0.03, 0.38) ^a
		Yamamoto et al. ³⁶	32.41	45.13		0.21 (-0.10, 0.52)
		Dolton et al. ³⁷	20.95	37.65		0.05 (-0.12, 0.21)
		Kees et al. ³⁸	66.13	92.02		0.48 (-0.36, 1.32)
		Jeurissen et al. ³⁹	23.66	39.00		0.59 (0.06, 1.12) ^a
		Lee et al. ⁴⁰	21.55	27.88		-0.75 (-0.90, -0.60) ^a
		Shimamoto et al. ⁴¹	30.05	40.20		0.12 (-0.18, 0.43)
		Shimamoto et al. ⁴²	79.37	95.53		0.41 (0.14, 0.69) ^a
		Adane et al. ⁴³	95.72	110.88		0.08 (-0.60, 0.78)
		24hr-CL _{cr}	Nielsen et al. ⁴⁴	29.03		34.11
	Blouin et al. ⁴⁵		18.59	49.61	-0.55 (-0.85, -0.25) ^a	
	Rotschafer et al. ⁴⁶		48.60	80.58	-0.09 (-0.83, 0.66)	
	Garaud et al. ⁴⁷		16.90	25.62	0.02 (-0.68, 0.72)	
	Brater et al. ⁴⁸		19.95	31.78	0.25 (-0.12, 0.62)	
	Taber et al. ⁴⁹		11.31	15.53	0.30 (-0.18, 0.79)	
	Dailly et al. ⁵⁰		58.20	99.96	-0.27 (-0.45, -0.06) ^a	
	Chen et al. ⁵⁴		-21.28	28.27	-0.50 (-0.59, -0.40) ^a	
	8hr-CL _{cr}	Baptista et al. ⁵²	51.94	73.98	73.98	0.93 (0.62, 1.24) ^a
		Schwartz	Shin et al. ⁵³	-2.52		37.85

MPE = mean prediction error, RMSE = root mean squared error, Cr = creatinine, CysC = cystatin C, eGFR = estimated glomerular filtration rate, CI = confidence interval, CG = Cockcroft-Gault, 24hr-CL_{cr} = 24-hour creatinine clearance, 8hr-CL_{cr} = 8-hour creatinine clearance.

^aPresence of statistically significant proportional bias ($P < 0.05$).

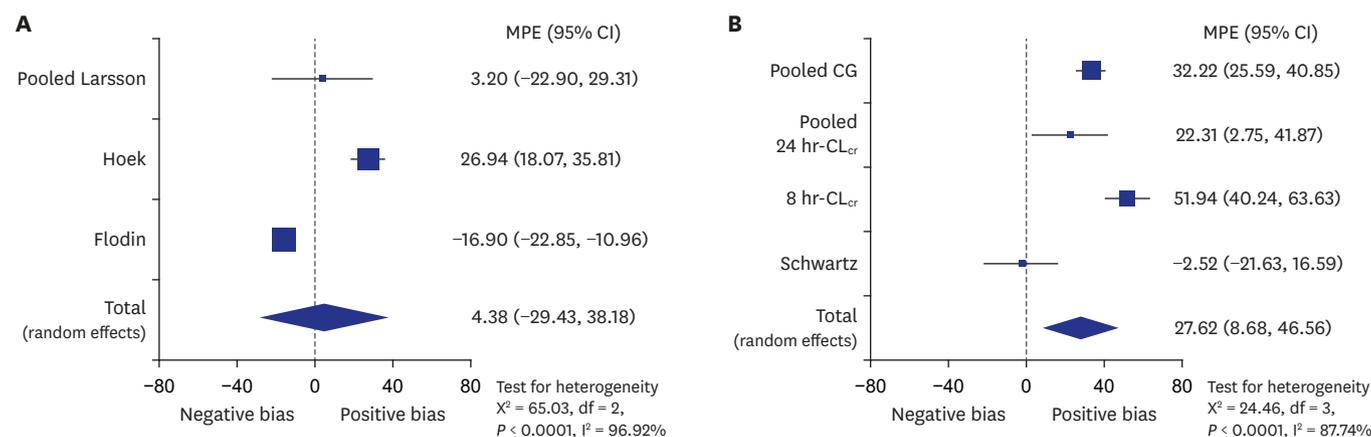


Fig. 2. Forest plot for the primary analysis. (A) On the performance of pooled cystatin C- and (B) creatinine-based estimated glomerular filtration rate equations in predicting vancomycin clearance.

MPE = mean prediction error, CI = confidence interval, CG = Cockcroft-Gault, 24hr-CL_{cr} = 24-hour creatinine clearance, 8hr-CL_{cr} = 8-hour creatinine clearance, df = degrees of freedom.

studies found the pooled MPE obtained by cystatin C-based eGFR equations is 8.63 mL min⁻¹, 95% CI, -15.73, 33.00 mL min⁻¹ with significantly high heterogeneity, $I^2 = 98.04\%$, $P < 0.001$, when compared with the creatinine-based eGFR equations with a pooled MPE of 36.97

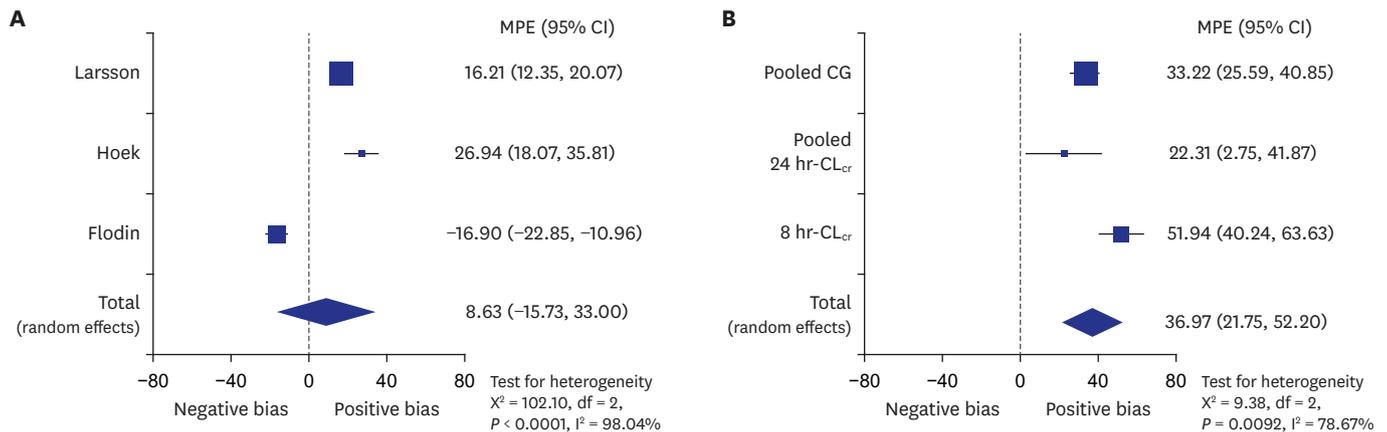


Fig. 3. Forest plot for the subgroup analysis among adult patients only. (A) On the performance of pooled cystatin C- and (B) creatinine-based estimated glomerular filtration rate equations in predicting vancomycin clearance. MPE = mean prediction error, CI = confidence interval, CG = Cockcroft-Gault, 24hr-CL_{cr} = 24-hour creatinine clearance, 8hr-CL_{cr} = 8-hour creatinine clearance, df = degrees of freedom.

mL min^{-1} , 95% CI, 21.75, 52.20 mL min^{-1} with significantly high heterogeneity, $I^2 = 78.67\%$, $P = 0.009$ (Fig. 3). This indicates the presence of unbiased results when cystatin C-based eGFR equations were used to predict the clearance of vancomycin in the adult population, compared to creatinine-based eGFR equations.

Subgroup analysis based on methods to estimate vancomycin clearance (e.g., Bayesian estimates, two-stage approach, Sawchuk-Zaske and population pharmacokinetic method) were also conducted. A similar pattern was observed when the cystatin C-based eGFR equations showed unbiased predictions for vancomycin clearance when it is estimated by using Bayesian (MPE, 3.20 mL min^{-1} ; 95% CI, -22.90, 29.31 mL min^{-1}) and two-stage approach (MPE, 4.89 mL min^{-1} ; 95% CI, -38.07 to 47.85 mL min^{-1}). The details of the subgroup analyses and publication bias assessment are available in **Supplementary Table 2** and **Supplementary Fig. 1**.

Sensitivity analysis

In the sensitivity analysis, one study is excluded at a time to determine its influence on the pooled effects. The dataset is re-analysed after exclusion of a study with negative MPE value, each from cystatin C- and creatinine-based eGFR equations group. There was a statistically significant positive bias by creatinine-based eGFR equations compared to cystatin C-based eGFR equations in vancomycin clearance prediction (Fig. 4). Cystatin C-based equations demonstrated a pooled MPE of 18.38 mL min^{-1} (95% CI, -3.96, 40.71), while creatinine-based equations demonstrated a pooled MPE of 36.97 mL min^{-1} (95% CI, 21.75, 52.20). Homogeneity exists between studies involving cystatin C-based equations (I^2 , 64.87%; $P = 0.092$). Hence, the results of the sensitivity analysis supported the outcomes of the primary analysis which found a better prediction of vancomycin clearance by cystatin C-based eGFR equations compared to creatinine-based eGFR equations.

Systematic trend of the difference plots

Statistically significant proportional bias was defined as differences in prediction against the clearance of vancomycin, with the presence of P value less than 0.05. Presence of significant proportional bias across the range of vancomycin clearance was found in some studies.^{32-35,39,40,42,45,50,52-54} The PE based on both biomarkers were inconsistent across the

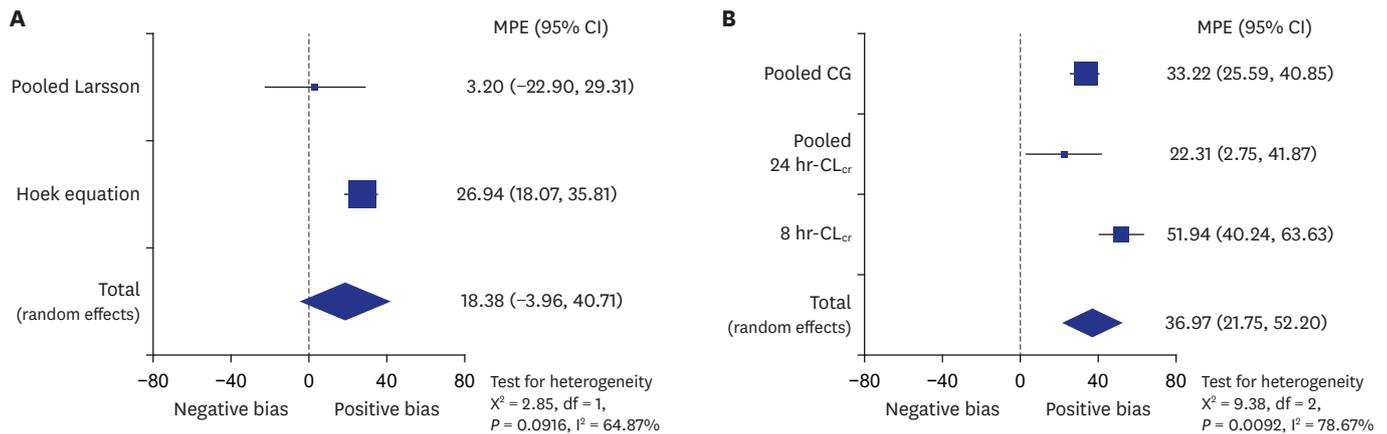


Fig. 4. Forest plot for the sensitivity analysis. **(A)** On the performance of pooled cystatin C- and **(B)** creatinine-based estimated glomerular filtration rate equations in predicting vancomycin clearance.

MPE = mean prediction error, CI = confidence interval, CG = Cockcroft-Gault, 24hr-CL_{cr} = 24-hour creatinine clearance, 8hr-CL_{cr} = 8-hour creatinine clearance, df = degrees of freedom.

range of vancomycin clearance. Two of the studies involving cystatin C-based eGFR equations showed a presence of statistically significant proportional bias.^{53,54} Positive bias was found at lower vancomycin clearance and negative bias was found at higher vancomycin clearance (**Supplementary Fig. 2**). Similar results were also shown in six of the creatinine-based studies (**Supplementary Fig. 3**).^{32,40,45,50,53,54} Majority of these studies uses 24-hour CL_{cr} to obtain the eGFR values. Meanwhile, another six creatinine-based studies with a statistically significant positive slope showed a negative bias at lower vancomycin clearance and positive bias at higher vancomycin clearance. Majority of these studies employed the Cockcroft-Gault equation to estimate patient's GFR. This indicates a different degree of bias as the vancomycin clearance values increases.

DISCUSSION

In this systematic review and meta-analysis, we assessed the performance of serum cystatin C- and creatinine-based eGFR equations in predicting vancomycin clearance. We found that utilising cystatin C-based eGFR equations to predict vancomycin kinetic parameters will be a good alternative to creatinine due to its better accuracy and precision. Cystatin C-based eGFR equations demonstrated better performance than creatinine in estimating vancomycin clearance because the pooled MPE includes zero, indicating the presence of unbiased results (MPE, 4.38 mL min⁻¹; 95% CI, -29.43, 38.18 mL min⁻¹; I², 96.92%; $P < 0.001$). Meanwhile, statistically significant positive bias was demonstrated by the creatinine-based eGFR equations as the 95% CI of the pooled MPE did not include zero (MPE, 27.62 mL min⁻¹; 95% CI, 8.68, 46.46 mL min⁻¹; I², 87.74%; $P < 0.001$). The result is consistent with the findings obtained through a systematic review conducted by Barreto et al.⁵⁵ Better prediction of target levels and elimination of vancomycin were obtained by cystatin C-based eGFR than creatinine-based eGFR. In general, cystatin C-based eGFR showed an equivalent accuracy, if not superior than creatinine-based eGFR in predicting the renal elimination of medications. The conduct of this study is consistent with the recommendations by the KDIGO which encourage more studies on the association between eGFR equations and drug pharmacokinetics to be carried out.⁵⁶

Cystatin C has been proposed as an alternative biomarker because it has a stable production rate and lesser influence by factors known to affect serum creatinine levels such as sex, age, race, and muscle mass.^{19,20} It is expressed in all nucleated cells, freely filtered by the glomerulus and were reabsorbed but not secreted by the proximal tubules.²¹ These characteristics of cystatin C fit the ideal criteria as an endogenous biomarker. Numerous studies conducted previously found cystatin C is better than creatinine as a biomarker for GFR estimation. In a study conducted by Huang et al.,⁵⁷ the performance of creatinine- and cystatin C-based eGFR equations were compared with measured GFR (mGFR). Cystatin C-based Hoek equation showed the best overall performance with low bias, good precision and greatest accuracy. This finding is supported by a meta-analysis which concluded that cystatin C had better diagnostic sensitivity, although lesser specificity in the estimation of GFR.⁵⁸ Hence, there is a promising area for utilisation of cystatin C-based eGFR equations in drug dose adjustment. In a study by Jin et al.,⁵⁹ the coefficient of determination between measured vancomycin concentrations and predicted vancomycin concentrations based on serum cystatin were higher ($r^2 = 0.5436$) when compared with serum creatinine ($r^2 = 0.2776$). A systematic review was later conducted by Brou et al.⁶⁰ to investigate the performance of cystatin C and creatinine in dose prediction of certain drugs such as vancomycin, amikacin, carboplatin and digoxin. Majority of the studies ($n = 13/16$) found a better correlation between serum cystatin C and drug clearance or trough concentrations of drugs being assessed compared to creatinine. This study marked the potential use of cystatin C as a biomarker to obtain more accurate dosing of vancomycin.

Dosage adjustment of renally excreted drugs is crucial among patients with reduced eGFR values. This is important particularly for drugs with a narrow therapeutic index such as vancomycin.⁶¹ This time-dependent antibiotic relied on the value of the area under the curve (AUC) to evaluate its effectiveness. Efforts to predict the AUC based on estimated CL_{cr} without information on drug concentration was suggested.⁶² More than 80% of the vancomycin is excreted by the kidney and its clearance is approximately 50%–80% of the mGFR.¹⁰ Hence, predicting the AUC of vancomycin without having to measure its serum concentration is possible through an equation describing a linear relationship between vancomycin clearance and CL_{cr} .⁶² Although serum creatinine concentrations have been widely used in clinical practice to measure patient's renal function, it often led to an overestimation of eGFR. This is especially true among elderly patients or those with low muscle mass and could possibly result in renal toxicity due to the drug overdose.

Analysis involving cystatin C- and creatinine-based eGFR equations in predicting vancomycin clearance showed the presence of significantly high heterogeneity. Some of the 95% CI of the studies do not overlap, indicating the presence of differences between studies. The various characteristics of these studies are evident from the examination of **Tables 1** and **2**. Subgroup analysis on adult patients did not demonstrate any improvement of homogeneity between studies. However, the MPE of cystatin C-based eGFR equations still showed unbiased predictions of vancomycin clearance (MPE, 8.63 mL min⁻¹; 95% CI, -15.73, 33.00 mL min⁻¹). compared to creatinine-based eGFR equations (MPE, 36.97 mL min⁻¹; 95% CI, 21.75, 52.20 mL min⁻¹). Subgroup analysis on the paediatric population was not conducted as only one study were reported in this category. We also evaluated the influence of different methods of vancomycin clearance estimation on the prediction performance of cystatin C- and creatinine-based eGFR equations. A similar pattern, whereby cystatin C-based eGFR equations showed an unbiased prediction of vancomycin clearance were found. The pooled MPE of both cystatin C-based eGFR equations and creatinine-based eGFR equations was

reduced in subgroup analysis involving vancomycin clearance obtained based on Bayesian estimates (from 4.38 to 3.20 mL min⁻¹ and from 27.62 to 17.72 mL min⁻¹, respectively). These findings could be explained by the fact that Bayesian estimate demonstrated greater accuracy and lower degree of bias compared to other methods of estimation.^{63,64}

Sensitivity analysis was also conducted to examine which study might be an influential outlier to the primary endpoint.²⁸ Outliers are measurements that have a significant deviation from the rest and they are quite common in different kind of datasets.⁶⁵ The amount of residual heterogeneity tends to be inflated by the outliers, and often results in upwardly biased mean estimates.⁶⁶ After exclusion of these outliers from meta-analyses of both biomarkers, the heterogeneity test was further improved, while maintaining the consistency of the pooled MPE results with the primary analysis.

Several limitations in this meta-analysis should be acknowledged. First, some studies had repeated measurements for the same individual (i.e., more than one measurement was available per individual).^{34,35,50} This indicates that the values of eGFR and vancomycin clearance reported are not unique for each individual and that the errors might be correlated. Second, there is a possibility that inaccurate digitisation of the data points to occur during the data extraction process. A good resolution of the scatter plot is very important in ensuring accurate extraction of eGFR and vancomycin clearance values. This is because sometimes overlapping data points could result in imprecise values recorded. However, the error is relatively small overall and less likely to cause a significant effect on the results or conclusion of the present study. A quality control step was conducted by comparing the reproducibility of eGFR and vancomycin clearance data points extracted from the scatter plot by a different researcher. A correlation coefficient or coefficient of determination obtained from the data points extracted were compared to the original values reported in the studies included, with a difference of < 5% were considered acceptable.

Another limitation of this meta-analysis is the inclusion of studies written in the English language only and the clearance values must be reported in units that can be standardised in mL min⁻¹. This had further limited the number of studies that can be included in this meta-analysis. A small number of studies investigating the correlation between cystatin C-based eGFR equations and vancomycin clearance is also one of the limitations in this study. There was a disproportionate focus on creatinine-based eGFR equations (26 studies) compared to cystatin C-based eGFR equations (four studies). It should be noted that although creatinine-based eGFR equations showed a statistically significant bias, it has less variance. This is because a bigger sample size will result in a narrower CI. In this case, creatinine had a smaller 95% CI of the pooled MPE due to the inclusion of a greater number of studies involving creatinine-based eGFR equations. Therefore, more studies investigating the correlation between cystatin C-based eGFR equations and vancomycin clearance should be included to improve the precision of the pooled estimate in future meta-analyses.

Other limitations associated with this study is the high heterogeneity between study designs. Differences in cystatin C, creatinine and vancomycin assays, as well as methods to estimate vancomycin clearance could be among the contributing factors. There is also a plausible effect between different populations of patients on vancomycin (such as renal failure, burns and obese patients) with our study findings. The pharmacokinetic parameters of these patient populations were known to be difficult to predict. This would lead to a very high level of heterogeneity and hence, compromises the interpretation of results. Random-effects

model was used after considering the effects of heterogeneity between studies. We tried to address this limitation by conducting subgroup analysis based on patient age group (i.e., adult population) and methods to estimate clearance of vancomycin. However, subgroup analysis based on the complex patient's condition was not conducted because data were too varied.

In conclusion, this meta-analysis has demonstrated that the cystatin C-based eGFR equations provided the best estimation of vancomycin clearance when compared to the creatinine-based equations. Greater precision of predictive performance was also shown by cystatin C-based eGFR equations due to its smaller RMSE values. This indicates that utilisation of cystatin C could be a good alternative biomarker to creatinine in the maintenance or adjustment of vancomycin doses. Majority of the studies also had a statistically significant proportional bias with a positive mean error at a low vancomycin clearance and vice versa. This finding was similar in both biomarkers. A further study specifically designed to compare the different eGFR equations in predicting vancomycin clearance across different patient population or assay methods for both biomarkers and vancomycin is needed to quantify the effectiveness of these biomarkers.

SUPPLEMENTARY MATERIALS

Supplementary Table 1

Serum cystatin C- and creatinine-based eGFR equations

[Click here to view](#)

Supplementary Table 2

Summary of subgroup analysis

[Click here to view](#)

Supplementary Fig. 1

Funnel plot of observed and imputed studies for publication bias assessment. Presence of an additional nine black circles indicates evidences of publication bias in reported outcomes from inspection of funnel plots and trim and fill analysis.

[Click here to view](#)

Supplementary Fig. 2

Difference plots between cystatin C-based eGFR equations and vancomycin clearance for analysis on the proportional bias.

[Click here to view](#)

Supplementary Fig. 3

Difference plots between creatinine-based eGFR equations and vancomycin clearance for analysis on the proportional bias.

[Click here to view](#)

REFERENCES

1. Mohr JF, Murray BE. Point: vancomycin is not obsolete for the treatment of infection caused by methicillin-resistant *Staphylococcus aureus*. *Clin Infect Dis* 2007;44(12):1536-42.
[PUBMED](#) | [CROSSREF](#)
2. Minejima E, Choi J, Beringer P, Lou M, Tse E, Wong-Beringer A. Applying new diagnostic criteria for acute kidney injury to facilitate early identification of nephrotoxicity in vancomycin-treated patients. *Antimicrob Agents Chemother* 2011;55(7):3278-83.
[PUBMED](#) | [CROSSREF](#)
3. Cadle RM, Mansouri MD, Darouiche RO. Vancomycin-induced elevation of liver enzyme levels. *Ann Pharmacother* 2006;40(6):1186-9.
[PUBMED](#) | [CROSSREF](#)
4. Forouzesh A, Moise PA, Sakoulas G. Vancomycin ototoxicity: a reevaluation in an era of increasing doses. *Antimicrob Agents Chemother* 2009;53(2):483-6.
[PUBMED](#) | [CROSSREF](#)
5. Rello J, Sole-Violan J, Sa-Borges M, Garnacho-Montero J, Muñoz E, Sirgo G, et al. Pneumonia caused by oxacillin-resistant *Staphylococcus aureus* treated with glycopeptides. *Crit Care Med* 2005;33(9):1983-7.
[PUBMED](#) | [CROSSREF](#)
6. Rybak M, Lomaestro B, Rotschafer JC, Moellering R Jr, Craig W, Billeter M, et al. Therapeutic monitoring of vancomycin in adult patients: a consensus review of the American Society of Health-System Pharmacists, the Infectious Diseases Society of America, and the Society of Infectious Diseases Pharmacists. *Am J Health Syst Pharm* 2009;66(1):82-98.
[PUBMED](#) | [CROSSREF](#)
7. Bakke V, Sporse H, Von der Lippe E, Nordøy I, Lao Y, Nyrerød HC, et al. Vancomycin levels are frequently subtherapeutic in critically ill patients: a prospective observational study. *Acta Anaesthesiol Scand* 2017;61(6):627-35.
[PUBMED](#) | [CROSSREF](#)
8. Levey AS, Coresh J, Balk E, Kausz AT, Levin A, Steffes MW, et al. National Kidney Foundation practice guidelines for chronic kidney disease: evaluation, classification, and stratification. *Ann Intern Med* 2003;139(2):137-47.
[PUBMED](#) | [CROSSREF](#)
9. Matzke GR, McGory RW, Halstenson CE, Keane WF. Pharmacokinetics of vancomycin in patients with various degrees of renal function. *Antimicrob Agents Chemother* 1984;25(4):433-7.
[PUBMED](#) | [CROSSREF](#)
10. Matzke GR, Zhanel GG, Guay DR. Clinical pharmacokinetics of vancomycin. *Clin Pharmacokinet* 1986;11(4):257-82.
[PUBMED](#) | [CROSSREF](#)
11. Golper TA, Noonan HM, Elzinga L, Gilbert D, Brummett R, Anderson JL, et al. Vancomycin pharmacokinetics, renal handling, and nonrenal clearances in normal human subjects. *Clin Pharmacol Ther* 1988;43(5):565-70.
[PUBMED](#) | [CROSSREF](#)
12. Lubran MM. Renal function in the elderly. *Ann Clin Lab Sci* 1995;25(2):122-33.
[PUBMED](#)
13. MacDiarmid SA, McIntyre WJ, Anthony A, Bailey RR, Turner JG, Arnold EP. Monitoring of renal function in patients with spinal cord injury. *BJU Int* 2000;85(9):1014-8.
[PUBMED](#) | [CROSSREF](#)
14. Verhave JC, Fesler P, Ribstein J, du Cailar G, Mimran A. Estimation of renal function in subjects with normal serum creatinine levels: influence of age and body mass index. *Am J Kidney Dis* 2005;46(2):233-41.
[PUBMED](#) | [CROSSREF](#)
15. Delanaye P, Mariat C, Cavalier E, Maillard N, Krzesinski JM, White CA. Trimethoprim, creatinine and creatinine-based equations. *Nephron Clin Pract* 2011;119(3):c187-93.
[PUBMED](#) | [CROSSREF](#)
16. Andreev E, Koopman M, Arisz L. A rise in plasma creatinine that is not a sign of renal failure: which drugs can be responsible? *J Intern Med* 1999;246(3):247-52.
[PUBMED](#) | [CROSSREF](#)
17. Maggi P, Montinaro V, Mussini C, Di Biagio A, Bellagamba R, Bonfanti P, et al. Novel antiretroviral drugs and renal function monitoring of HIV patients. *AIDS Rev* 2014;16(3):144-51.
[PUBMED](#)

18. Tanaka A, Aiba T, Otsuka T, Suemaru K, Nishimiya T, Inoue T, et al. Population pharmacokinetic analysis of vancomycin using serum cystatin C as a marker of renal function. *Antimicrob Agents Chemother* 2010;54(2):778-82.
[PUBMED](#) | [CROSSREF](#)
19. Filler G, Bökenkamp A, Hofmann W, Le Bricon T, Martínez-Brú C, Grubb A. Cystatin C as a marker of GFR--history, indications, and future research. *Clin Biochem* 2005;38(1):1-8.
[PUBMED](#) | [CROSSREF](#)
20. Abrahamson M, Olafsson I, Palsdottir A, Ulvsbäck M, Lundwall A, Jensson O, et al. Structure and expression of the human cystatin C gene. *Biochem J* 1990;268(2):287-94.
[PUBMED](#) | [CROSSREF](#)
21. Jacobsson B, Lignelid H, Bergerheim US. Transthyretin and cystatin C are catabolized in proximal tubular epithelial cells and the proteins are not useful as markers for renal cell carcinomas. *Histopathology* 1995;26(6):559-64.
[PUBMED](#) | [CROSSREF](#)
22. Murty MSN, Sharma UK, Pandey VB, Kankare SB. Serum cystatin C as a marker of renal function in detection of early acute kidney injury. *Indian J Nephrol* 2013;23(3):180-3.
[PUBMED](#) | [CROSSREF](#)
23. Shlipak MG, Mattes MD, Peralta CA. Update on cystatin C: incorporation into clinical practice. *Am J Kidney Dis* 2013;62(3):595-603.
[PUBMED](#) | [CROSSREF](#)
24. Roos JF, Doust J, Tett SE, Kirkpatrick CMJ. Diagnostic accuracy of cystatin C compared to serum creatinine for the estimation of renal dysfunction in adults and children--a meta-analysis. *Clin Biochem* 2007;40(5-6):383-91.
[PUBMED](#) | [CROSSREF](#)
25. Rebollo N, Cepeda-Piorno FJ. Cystatin C for therapeutic drug monitoring. *Clin Chem* 2015;61(6):804-7.
[PUBMED](#) | [CROSSREF](#)
26. Sheiner LB, Beal SL. Some suggestions for measuring predictive performance. *J Pharmacokinet Biopharm* 1981;9(4):503-12.
[PUBMED](#) | [CROSSREF](#)
27. Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. *BMJ* 2003;327(7414):557-60.
[PUBMED](#) | [CROSSREF](#)
28. Borenstein M, Hedges L, Higgins J, Rothstein H. *Introduction to Meta-Analysis*. Chichester: John Wiley & Sons, Ltd.; 2009.
29. Duval S, Tweedie R. Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. *Biometrics* 2000;56(2):455-63.
[PUBMED](#) | [CROSSREF](#)
30. Birt JK, Chandler MH. Using clinical data to determine vancomycin dosing parameters. *Ther Drug Monit* 1990;12:206-9.
[PUBMED](#) | [CROSSREF](#)
31. Polard E, Le Bouquin V, Le Corre P, Kérebel C, Trout H, Feuillu A, et al. Non steady state and steady state PKs Bayesian forecasting and vancomycin pharmacokinetics in ICU adult patients. *Ther Drug Monit* 1999;21(4):395-403.
[PUBMED](#) | [CROSSREF](#)
32. Yoshida M, Yasuda N, Nishikata M, Okamoto K, Uchida T, Matsuyama K. New recommendations for vancomycin dosage for patients with MRSA pneumonia with various degrees of renal function impairment. *J Infect Chemother* 2005;11(4):182-8.
[PUBMED](#) | [CROSSREF](#)
33. Okamoto G, Sakamoto T, Kimura M, Ukishima Y, Sonoda A, Mori N, et al. Serum cystatin C as a better marker of vancomycin clearance than serum creatinine in elderly patients. *Clin Biochem* 2007;40(7):485-90.
[PUBMED](#) | [CROSSREF](#)
34. Omote S, Yano Y, Hashida T, Masuda S, Yano I, Katsura T, et al. A retrospective analysis of vancomycin pharmacokinetics in Japanese cancer and non-cancer patients based on routine trough monitoring data. *Biol Pharm Bull* 2009;32(1):99-104.
[PUBMED](#) | [CROSSREF](#)
35. Pea F, Furlanut M, Negri C, Pavan F, Crapis M, Cristini F, et al. Prospectively validated dosing nomograms for maximizing the pharmacodynamics of vancomycin administered by continuous infusion in critically ill patients. *Antimicrob Agents Chemother* 2009;53(5):1863-7.
[PUBMED](#) | [CROSSREF](#)

36. Yamamoto M, Kuzuya T, Baba H, Yamada K, Nabeshima T. Population pharmacokinetic analysis of vancomycin in patients with gram-positive infections and the influence of infectious disease type. *J Clin Pharm Ther* 2009;34(4):473-83.
[PUBMED](#) | [CROSSREF](#)
37. Dolton M, Xu H, Cheong E, Maitz P, Kennedy P, Gottlieb T, et al. Vancomycin pharmacokinetics in patients with severe burn injuries. *Burns* 2010;36(4):469-76.
[PUBMED](#) | [CROSSREF](#)
38. Kees MG, Hilpert JW, Gnewuch C, Kees F, Voegeler S. Clearance of vancomycin during continuous infusion in Intensive Care Unit patients: correlation with measured and estimated creatinine clearance and serum cystatin C. *Int J Antimicrob Agents* 2010;36(6):545-8.
[PUBMED](#) | [CROSSREF](#)
39. Jeurissen A, Sluyts I, Rutsaert R. A higher dose of vancomycin in continuous infusion is needed in critically ill patients. *Int J Antimicrob Agents* 2011;37(1):75-7.
[PUBMED](#) | [CROSSREF](#)
40. Lee JP, Dang AT. Evaluation of methods to estimate glomerular filtration rate versus actual drug clearance in patients with chronic spinal cord injury. *Spinal Cord* 2011;49(12):1158-63.
[PUBMED](#) | [CROSSREF](#)
41. Shimamoto Y, Fukuda T, Tominari S, Fukumoto K, Ueno K, Dong M, et al. Decreased vancomycin clearance in patients with congestive heart failure. *Eur J Clin Pharmacol* 2013;69(3):449-57.
[PUBMED](#) | [CROSSREF](#)
42. Shimamoto Y, Fukuda T, Tanaka K, Komori K, Sadamitsu D. Systemic inflammatory response syndrome criteria and vancomycin dose requirement in patients with sepsis. *Intensive Care Med* 2013;39(7):1247-52.
[PUBMED](#) | [CROSSREF](#)
43. Adane ED, Herald M, Koura F. Pharmacokinetics of vancomycin in extremely obese patients with suspected or confirmed *Staphylococcus aureus* infections. *Pharmacotherapy* 2015;35(2):127-39.
[PUBMED](#) | [CROSSREF](#)
44. Nielsen HE, Hansen HE, Korsager B, Skov PE. Renal excretion of vancomycin in kidney disease. *Acta Med Scand* 1975;197(4):261-4.
[PUBMED](#) | [CROSSREF](#)
45. Blouin RA, Bauer LA, Miller DD, Record KE, Griffen WO Jr. Vancomycin pharmacokinetics in normal and morbidly obese subjects. *Antimicrob Agents Chemother* 1982;21(4):575-80.
[PUBMED](#) | [CROSSREF](#)
46. Rotschafer JC, Crossley K, Zaske DE, Mead K, Sawchuk RJ, Solem LD. Pharmacokinetics of vancomycin: observations in 28 patients and dosage recommendations. *Antimicrob Agents Chemother* 1982;22(3):391-4.
[PUBMED](#) | [CROSSREF](#)
47. Garaud JJ, Regnier B, Inglebert F, Faurisson F, Bauchet J, Vachon F. Vancomycin pharmacokinetics in critically ill patients. *J Antimicrob Chemother* 1984;14 Suppl D:53-7.
[PUBMED](#) | [CROSSREF](#)
48. Brater DC, Bawdon RE, Anderson SA, Purdue GF, Hunt JL. Vancomycin elimination in patients with burn injury. *Clin Pharmacol Ther* 1986;39(6):631-4.
[PUBMED](#) | [CROSSREF](#)
49. Taber DJ, Fann AL, Malat G, Dupuis RE. Evaluation of estimated and measured creatinine clearances for predicting the pharmacokinetics of vancomycin in adult liver transplant recipients. *Ther Drug Monit* 2003;25(1):67-72.
[PUBMED](#) | [CROSSREF](#)
50. Dailly E, Le Floch R, Deslandes G, Pannier M, Jolliet P. Influence of glomerular filtration rate on the clearance of vancomycin administered by continuous infusion in burn patients. *Int J Antimicrob Agents* 2008;31(6):537-9.
[PUBMED](#) | [CROSSREF](#)
51. Park SJ, Yang JH, Park HJ, In YW, Lee YM, Cho YH, et al. Trough concentrations of vancomycin in patients undergoing extracorporeal membrane oxygenation. *PLoS One* 2015;10(11):e0141016.
[PUBMED](#) | [CROSSREF](#)
52. Baptista JP, Roberts JA, Sousa E, Freitas R, Deveza N, Pimentel J. Decreasing the time to achieve therapeutic vancomycin concentrations in critically ill patients: developing and testing of a dosing nomogram. *Crit Care* 2014;18(6):654.
[PUBMED](#) | [CROSSREF](#)
53. Shin JE, Lee SM, Eun HS, Park MS, Park KI, Namgung R. Usefulness of serum cystatin C to determine the dose of vancomycin in neonate. *Korean J Pediatr* 2015;58(11):421-6.
[PUBMED](#) | [CROSSREF](#)

54. Chen YC, Feng JF, Li B, Zhang L, Yang YW. Estimation of safe and effective dose of vancomycin in MRSA-infected patients using serum cystatin C concentrations. *Int J Clin Pharmacol Ther* 2013;51(3):161-9.
[PUBMED](#) | [CROSSREF](#)
55. Barreto EF, Rule AD, Murad MH, Kashani KB, Lieske JC, Erwin PJ, et al. Prediction of the renal elimination of drugs with cystatin C vs creatinine: a systematic review. *Mayo Clin Proc* 2019;94(3):500-14.
[PUBMED](#) | [CROSSREF](#)
56. Matzke GR, Aronoff GR, Atkinson AJ Jr, Bennett WM, Decker BS, Eckardt KU, et al. Drug dosing consideration in patients with acute and chronic kidney disease—a clinical update from Kidney Disease: Improving Global Outcomes (KDIGO). *Kidney Int* 2011;80(11):1122-37.
[PUBMED](#) | [CROSSREF](#)
57. Huang SH, Macnab JJ, Sontrop JM, Filler G, Gallo K, Lindsay RM, et al. Performance of the creatinine-based and the cystatin C-based glomerular filtration rate (GFR) estimating equations in a heterogenous sample of patients referred for nuclear GFR testing. *Transl Res* 2011;157(6):357-67.
[PUBMED](#) | [CROSSREF](#)
58. Zhang Z, Lu B, Sheng X, Jin N. Cystatin C in prediction of acute kidney injury: a systemic review and meta-analysis. *Am J Kidney Dis* 2011;58(3):356-65.
[PUBMED](#) | [CROSSREF](#)
59. Jin SJ, Bae SC, Kim HW, Kim HK, Na EJ, Ahn BS, et al. Evaluation of the effect of initial dose of vancomycin using serum cystatin C as a marker in elderly patients. *Infect Chemother* 2009;41(4):224-9.
[CROSSREF](#)
60. Brou NA, Jacqz-Aigrain E, Zhao W. Cystatin C as a potential biomarker for dosing of renally excreted drugs. *Br J Clin Pharmacol* 2015;80(1):20-7.
[PUBMED](#) | [CROSSREF](#)
61. Tanaka A, Suemaru K, Otsuka T, Ido K, Nishimiya T, Sakai I, et al. Estimation of the initial dose setting of vancomycin therapy with use of cystatin C as a new marker of renal function. *Ther Drug Monit* 2007;29(2):261-4.
[PUBMED](#) | [CROSSREF](#)
62. Yim DS. Overestimation of vancomycin clearance by the linear regression formula in Rodvold's report: why? *Infect Chemother* 2014;46(1):62-3.
[PUBMED](#) | [CROSSREF](#)
63. Garrelts JC, Godley PJ, Horton MW, Karboski JA. Accuracy of Bayesian, Sawchuk-Zaske, and nomogram dosing methods for vancomycin. *Clin Pharm* 1987;6(10):795-9.
[PUBMED](#)
64. Burton ME, Gentle DL, Vasko MR. Evaluation of a Bayesian method for predicting vancomycin dosing. *DICP* 1989;23(4):294-300.
[PUBMED](#) | [CROSSREF](#)
65. Grubbs FE. Procedures for detecting outlying observations in samples. *Technometrics* 1969;11(1):1-21.
[CROSSREF](#)
66. Viechtbauer W, Cheung MW. Outlier and influence diagnostics for meta-analysis. *Res Synth Methods* 2010;1(2):112-25.
[PUBMED](#) | [CROSSREF](#)
67. Flodin M, Jonsson AS, Hansson LO, Danielsson LA, Larsson A. Evaluation of gentian cystatin C reagent on Abbott Ci8200 and calculation of glomerular filtration rate expressed in mL/min/1.73 m² from the cystatin C values in mg/L. *Scand J Clin Lab Invest* 2007;67(5):560-7.
[PUBMED](#) | [CROSSREF](#)
68. Hoek FJ, Kemperman FAW, Krediet RT. A comparison between cystatin C, plasma creatinine and the Cockcroft and Gault formula for the estimation of glomerular filtration rate. *Nephrol Dial Transplant* 2003;18(10):2024-31.
[PUBMED](#) | [CROSSREF](#)
69. Larsson A, Malm J, Grubb A, Hansson LO. Calculation of glomerular filtration rate expressed in mL/min from plasma cystatin C values in mg/L. *Scand J Clin Lab Invest* 2004;64(1):25-30.
[PUBMED](#) | [CROSSREF](#)
70. Cockcroft DW, Gault MH. Prediction of creatinine clearance from serum creatinine. *Nephron* 1976;16(1):31-41.
[PUBMED](#) | [CROSSREF](#)
71. Schwartz GJ, Haycock GB, Edelman CM Jr, Spitzer A. A simple estimate of glomerular filtration rate in children derived from body length and plasma creatinine. *Pediatrics* 1976;58(2):259-63.
[PUBMED](#)