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REFERENCES

- Hamid Q, Boguniewicz M, Leung D. Differential in situ cytokine gene expression in acute versus chronic atopic dermatitis. J Clin Invest. 1994;94(2):870-876.
- Guttman-Yassky E, Nograles KE, Krueger JG. Contrasting pathogenesis of atopic dermatitis and psoriasis Part II: immune cell subsets

- and therapeutic concepts. J Allergy and Clin Immunol. 2011;127(6): 1420-1432
- Kondo S, Yazawa H, Jimbow K. Reduction of serum interleukin-5 levels reflect clinical improvement in patients with atopic dermatitis. J Dermatol. 2001;28:237-243.
- Brunner PM, Leung D, Guttman-Yassky E. Immunologic, microbial, and epithelial interactions in atopic dermatitis. *Ann Allergy Asthma Immunol*, 2018:120:34-41.
- Furue M, Chiba T, Tsuji G, et al. Atopic dermatitis: immune deviation, barrier dysfunction, IgE autoreactivity and new therapies. Allergol Int. 2017:66:398-403.
- Pouliquen IJ, Kornmann O, Barton SV, et al. Characterization of the relationship between dose and blood eosinophil response following subcutaneous administration of mepolizumab. Int J Clin Pharmacol Ther. 2015;53(12):1015-1027.
- Eichenfield LF, Tom WL, Chamlin SL, et al. Guidelines of care of the management of atopic dermatitis: section 1. Diagnosis and assessment of atopic dermatitis. J Am Acad Dermatol. 2014;70:338-351.
- Oldhoff JM, Darsow U, Werfel T, et al. Anti-IL-5 recombinant humanized monoclonal antibody (mepolizumab) for the treatment of atopic dermatitis. Allergy. 2005;60:693-696.
- Ortega HG, Liu MC, Pavord ID, et al. Mepolizumab treatment in patients with severe eosinophilic asthma. N Engl J Med. 2014;371:1198-1207.

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Effect of C1-inhibitor in adults with mild asthma: A randomized controlled trial

To the Editor,

Several newly approved monoclonal antibodies targeting type 2 inflammation have shown remarkable beneficial effects in patients with severe asthma. Biologics directed against IL-5 (mepolizumab, reslizumab), IL-5 receptor (benralizumab), or IL-4 receptor α (dupilumab) have shown to reduce asthma exacerbation rate by about 50%. 1,2 Though promising, these drugs are unable to completely alleviate inflammation-induced asthma symptoms. Moreover, a substantial subset of patients without a pronounced type 2 airway inflammation does not benefit from the currently available biologics. Therefore, novel antiinflammatory treatments targeting other relevant asthma-associated pathways are still warranted. In recent years, the complement system has been implicated in the pathogenesis of type 2 asthma.³ Elevated levels of anaphylatoxins, activation products of the complement system, have been found in the airways of asthma patients following local allergen provocation.⁴ Functional roles for the anaphylatoxins in asthma have been established in experimental studies in mice, showing that these proinflammatory mediators act synergistically and drive allergic inflammation.3 C1-inhibitor (C1-INH) is an endogenous protein with a pivotal regulatory function in the complement system by inhibiting both the classical and lectin pathways. We hypothesized that C1-INH administration inhibits complement activation and attenuates allergen-induced airway eosinophilia in patients with mild asthma.

In this randomized, double-blind, placebo-controlled, parallel study, 24 adults with asthma and house dust mite (HDM) allergy received a continuous intravenous infusion with human plasma-derived C1-INH 100 U kg⁻¹ h⁻¹ or placebo followed after 2 hours by segmental challenge with HDM and lipopolysaccharide (LPS) in one lung and saline in the contralateral lung as control. Bronchoalveolar lavage fluid was obtained seven hours after HDM/LPS or saline challenge. The primary outcome was influx of eosinophils and neutrophils, defined as number of cells/mL, into the bronchoalveolar space. Further details of the study design, subject selection criteria, bronchoalveolar lavage handling, assays, and statistical analysis are described in the supplemental section. Baseline patient characteristics were similar across treatment groups (Table S1).

Two hours after the initiation of C1-INH infusion, median plasma C1-INH antigen concentrations were four times higher in C1-INH-infused patients compared to vehicle-infused controls (Figure S1A). Segmental HDM/LPS challenge resulted in increased C1-INH antigen

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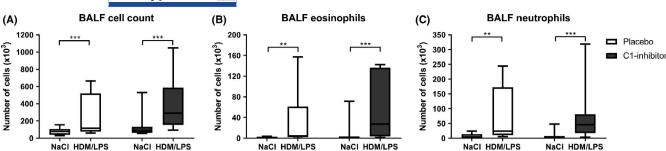


FIGURE 1 Intravenous C1-inhibitor infusion does not modify leukocyte influx after HDM/LPS challenge in the airways of asthma patients. A, Total cell number in BALF, B, eosinophils in BALF, C, neutrophils in BALF. Data represent the median with interquartile range, the smallest and largest observation. **: P < .001, ***: P < .001, BALF: bronchoalveolar lavage

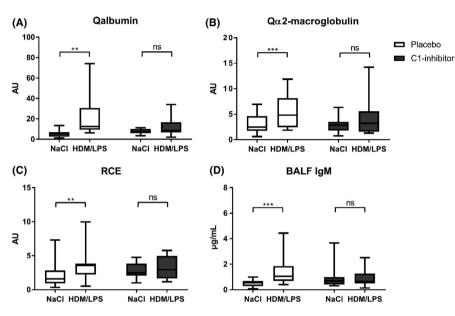


FIGURE 2 Intravenous C1-inhibitor infusion reduces vascular leak following HDM/LPS challenge in the airways of asthma patients. A, Qalbumin: BALF albumin/plasma albumin, B, Q α 2-macroglobulin: BALF α 2-macroglobulin/plasma α 2-macroglobulin, C, Relative coefficient of excretion (RCE): Q α 2M/Qalbumin, (D) BALF IgM levels. Data represent the median with interquartile range, the smallest and largest observation. **: P < .01, ***: P < .001, ns: not significant, BALF: bronchoalveolar lavage

levels in BALF compared to saline instillation in both treatment groups (Figure S1C). C1-INH concentrations were higher in BALF from C1-INH-infused patients compared to the placebo group. C1-INH activity levels in plasma and BALF were similar to C1-INH antigen concentrations (Figure S1B,D), indicating that C1-INH was biologically active.

HDM/LPS challenge induced elevated C4a concentrations compared to saline challenge in the placebo group (Figure S2A). In the C1-INH group, BALF C4a levels were similar between the saline and HDM/LPS-challenged sites. Consistently, using an assay that detects the C4 activation products C4b, C4bi, and C4c (collectively referred to as C4bc), C4 activation in the lung subsegment exposed to HDM/LPS was increased in patients infused with placebo but not in those infused with C1-INH (Figure S2B). We next measured the anaphylatoxin C3a, which is released following C3 cleaved activation. Similar to C4a, HDM/LPS challenge increased BALF C3a in the placebo group, but not in the C1-INH treatment group (Figure S2C). In agreement, C3 activation products were elevated in the HDM/LPS-challenged lung in patients infused with placebo but not in those administered with C1-INH (Figure S2D). These data indicate that C1-INH infusion prevents C4a and C3a generation in the airways upon a bronchial challenge with HDM/LPS.

HDM/LPS instillation augmented total cell counts in BALF compared to saline, partly as consequence of eosinophil and neutrophil

influx (Figure 1A-C). Likewise, HDM/LPS challenge elevated CD4 T cells, but did not alter the number of alveolar macrophages in BALF (Figure S3A,B). C1-INH did not modify this allergen-induced response. HDM/LPS also induced degranulation of eosinophils and neutrophils in the bronchoalveolar space (Figure S4A-D). These responses were not affected by C1-INH with the exception of lactoferrin release, which was inhibited by C1-INH (Figure S4B).

To obtain further insight into the inflammatory response upon HDM/LPS challenge and the effect of C1-INH hereon, we measured a broad spectrum of cytokines and chemokines relevant for allergic inflammation. Of the 35 cytokines and chemokines measured, 15 were detectable in BALF (Table S2, Table S3). HDM/LPS induced increases in neutrophil chemoattractants such as interleukin (IL)-8, IL-1 β , tumor necrosis factor- α , and macrophage inflammatory proteins 1α and 1β (Table S2). Likewise, eosinophil attractants eotaxin-1 and RANTES were increased upon HDM/LPS challenge. These responses were not influenced by C1-INH. HDM/LPS also induced the release of growth-related oncogene- α , stromal cell-derived factor- 1α , and IL-18 in the placebo group. Although statistically insignificant, these rises were also detected in the C1-INH group.

Beside the complement system, C1-INH is an important regulator of the kallikrein-kinin system due to its inhibitory effect on

FXIIa and kallikrein activity.⁶ Elevated levels of kallikrein-kinin system components have been documented in the airways of asthma patients.⁷ We determined C1-INH/FXII and C1-INH/kallikrein complexes as measure for kallikrein-kinin system activation. In BALF, however, these complexes were below detection limit. Hence, the current challenge model is not suitable to study the contribution of the kallikrein-kinin system in allergen-induced inflammation and the effect of C1-INH hereon.

Vascular leak often occurs as consequence of allergen-induced inflammation in the airway of asthma patients and has been associated with the loss of asthma control. We determined the quotients of albumin and α 2-macroglobulin levels in BALF and plasma (QAlb and QA2M, respectively) and the relative coefficient of excretion (QA2M/QAlb) as measures of the permeability of the blood-airway barrier. Intrabronchial HDM/LPS challenge was associated with significant increase in QAlb (Figure 2A), QA2M (Figure 2B), and relative coefficient of excretion (Figure 2C) in the placebo group. These effects were abrogated in the C1-INH group (Figure 2A-C). Likewise, HDM/LPS induced elevated BALF IgM concentrations in placebo-infused patients but not in C1-INH-treated subjects (Figure 2D).

In conclusion, we show that intravenous C1-INH administration prior to intrabronchial HDM/LPS challenge prevents complement activation and vascular leak without attenuating allergic lung inflammation in patients with HDM allergy and asthma. Suppressing vascular leakage could help improve symptoms in patients who do not respond adequately to currently available drugs.

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CONFLICT OF INTEREST

The authors have no conflict of interest in relation to this work.

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REFERENCES

- Israel E, Reddel HK. Severe and difficult-to-treat asthma in adults. N Engl J Med. 2017;377(10):965-976.
- 2. Eger KA, Bel EH. The emergence of new biologics for severe asthma. *Curr Opin Pharmacol.* 2019;46:108-115.
- Zhang X, Kohl J. A complex role for complement in allergic asthma. Expert Rev Clin Immunol. 2010;6(2):269-277.
- de Boer JD, Berger M, Majoor CJ, et al. Activated protein C inhibits neutrophil migration in allergic asthma: a randomised trial. Eur Respir J. 2015;46(6):1636-1644.
- 5. Sarma JV, Ward PA. The complement system. *Cell Tissue Res.* 2011;343(1):227-235.
- 6. Zeerleder S. C1-inhibitor: more than a serine protease inhibitor. *Semin Thromb Hemost*. 2011;37(4):362-374.
- Christiansen SC, Proud D, Sarnoff RB, Juergens U, Cochrane CG, Zuraw BL. Elevation of tissue kallikrein and kinin in the airways of asthmatic subjects after endobronchial allergen challenge. Am Rev Respir Dis. 1992;145(4 Pt 1):900-905.
- 8. Fick Jr RB, Metzger WJ, Richerson HB, et al. Increased bronchovascular permeability after allergen exposure in sensitive asthmatics. *J Appl Physiol* (1985). 1987;63(3):1147-1155.
- 9. Li X, Wilson JW. Increased vascularity of the bronchial mucosa in mild asthma. Am J Respir Crit Care Med. 1997;156(1):229-233.

SUPPORTING INFORMATION

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