

Systematic Analysis of Spleen Tyrosine Kinase Expression and its Clinical Outcomes in Various Cancers

Akram I. Alwithenani, Mohammad A. Althubiti¹

Department of Laboratory Medicine, Faculty of Applied Medical Sciences, Umm Al-Qura University, ¹Department of Biochemistry, Faculty of Medicine, Umm Al-Qura University, Makkah, Saudi Arabia

Abstract

Background: Spleen tyrosine kinase (SYK) is an important enzyme in the proliferation and differentiation of all hematopoietic tissues. Its role as a cancer driver is well documented in liquid tumors; however, cumulative evidence has suggested an opposite role in other tumor types.

Objectives: To systematically assess the expression of SYK, its prognostic value and epigenetic status in different cancers using bioinformatics tools.

Methods: In this bioinformatics study, OncoPrint database and cBioPortal were used to study the SYK gene expression, Kaplan–Meier plotter to study its prognostic value and MethHC to assess the SYK gene methylation in various cancers.

Results: From 542 unique analyses of the SYK gene, it was found to be overexpressed in bladder, breast and colon cancers but downregulated in leukemia, lymphoma and myeloma. Compared with normal tissues, breast and brain tumors showed an overexpression of the SYK gene, whereas lymphoma and leukemia had lower expression. The Kaplan–Meier survival analysis revealed that SYK expression in pancreatic, gastric, liver and lung patients were correlated with better overall survival. Using cBioPortal, prostate cancer was found to have the highest SYK gene mutation frequency, and the mean expression was highest in diffuse large B-cell lymphoma, acute myeloid leukemia and thymoma. Using the MethHC database, SYK promoter hypermethylation was found to be significantly higher in breast, renal, liver, lung, pancreatic, prostatic, skin and stomach cancers compared with the normal tissue ($P < 0.005$).

Conclusion: The results of this study indicate the potential use of SYK as a diagnostic and therapeutic target for different type of cancers. However, further experimental data are required to validate these results before use of SYK in clinical settings.

Keywords: Cancer, expression, methylation, mutation, spleen tyrosine kinase, survival rate

Address for correspondence: Dr. Mohammad A. Althubiti, Department of Biochemistry, Faculty of Medicine, Umm Al-Qura University, Makkah, Saudi Arabia.

E-mail: mathubiti@uqu.edu.sa

Submitted: 29-Aug-2019 **Revised:** 28-Oct-2019 **Accepted:** 06-Feb-2020 **Published:** 17-Apr-2020

INTRODUCTION

Spleen tyrosine kinase (SYK) is a cytosolic nonreceptor protein tyrosine kinase that is highly expressed in all

hematopoietic tissues and plays a role in the development and growth of B-cells.^[1] Stimulation of B-cell receptors activates downstream pathways that involve SYK

Access this article online	
Quick Response Code:	Website: www.sjmms.net
	DOI: 10.4103/sjmms.sjmms_300_19

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

How to cite this article: Alwithenani AI, Althubiti MA. Systematic analysis of spleen tyrosine kinase expression and its clinical outcomes in various cancers. Saudi J Med Med Sci 2020;8:95-104.

stimulation, and consequently, B-cell activation, resulting in B-cell development and growth. SYK activation has been shown to play a role in allergy and autoimmunity responses, in B-cell transformation^[2] and as a viral oncogene.^[3]

Kinase inhibitors have been developed to target and inhibit the abnormal stimulation of SYK. For example, R112, a selective inhibitor of SYK, has been found to have therapeutic effects on allergic rhinitis patients,^[4] while fostamatinib, also a SYK inhibitor, has been shown to reduce the onset and severity of arthritis.^[5] SYK targeting has also been shown to be a therapeutic candidate for lymphoma and leukemia. Entospletinib, for example, is a selective inhibitor of SYK that has shown to have effects on patients with relapsed chronic lymphoblastic leukemia.^[6]

Although SYK is considered as an oncogene in some hematological malignancies,^[7] cumulative data have suggested that it plays a tumor suppressive role in various cancers.^[8] This role has been documented in the epithelial cells of solid cancers such as breast cancer. Low expression of SYK in breast cancer is correlated with low survival rates and a more invasive cancer.^[9] In addition, reactivation of SYK has been shown to inhibit tumor growth of breast cancer cells, whereas inhibition of SYK activation decreases the p53-dependent apoptosis of epithelial cancer cells.^[8]

Based on the current evidence, SYK has been demonstrated to have a dual role in cancer progression and suppression. Therefore, there is a need to better understand the role SYK plays in different cancers. Accordingly, in this study, a systematic analysis of SYK was conducted using online databases to assess its expression in different cancer types, prognostic value and epigenetic status in different cancers.

METHODS

This systematic bioinformatics study used various databases for analyzing SYK gene expression and methylation as well as its prognostic value in various cancers.

Spleen tyrosine kinase expression levels in different cancers

OncomineTM database (<https://www.oncomine.org/resource/login.html>) is an open-access, online tool containing data from numerous published cancer microarray studies. In this study, the level of SYK gene expression in the normal and tumor tissues was identified in different cancer types using the Oncomine database in July 2019.^[10] The threshold of SYK mRNA fold was determined according to the following parameters: $P <$

1×10^{-4} , fold change >2 and gene ranking in the top 10%.

Relationship between spleen tyrosine kinase expression and overall survival

Kaplan–Meier plotter (<http://kmplot.com/analysis/>) is an online database of published microarray datasets that assess the effect of 54,675 genes on survival using 18,674 samples from different cancers. In the current study, this tool was used in July 2019 to assess the prognosis of SYK expression, which was only found in patients with pancreatic, gastric, liver and lung cancer. The hazard ratio with 95% confidence intervals and log-rank P value were computed.^[11]

Spleen tyrosine kinase expression and mutations frequencies

The cBioPortal for cancer genomics is an open-access tool (<http://www.cbioportal.org/>), wherein about 55,833 tumor samples from >210 cancer studies in The Cancer Genome Atlas (TCGA) can be visualized and analyzed. The search page of the tool helps extract customized data that allow exploring genetic mutations in different samples of the gene of interest. In this study, data from TCGA^[12] were retrieved using cBioPortal in July 2019 for analyzing the expression and mutations frequency in SYK gene, and Kaplan–Meier (log rank) test was used to calculate the P value for the differences between patients with/or without alteration in SYK.^[13]

Spleen tyrosine kinase promoter methylation in different cancers

To illustrate other possible mechanisms by which SYK gene is compromised, the DNA methylation status of SYK promoter in different cancer types was assessed using the MethHC database in July 2019,^[14] an online tool that provides data on DNA methylation of different human cancers. MethHC integrates data such as DNA methylation, gene expression and correlation of methylation and gene expression from TCGA. SYK promoter methylation was analyzed in the following tumor types: bladder urothelial carcinoma; breast invasive carcinoma; cervical squamous cell carcinoma and endocervical adenocarcinoma; colon adenocarcinoma; head-and-neck squamous cell carcinoma; kidney renal clear cell carcinoma; kidney renal papillary cell carcinoma; liver hepatocellular carcinoma; lung adenocarcinoma; lung squamous cell carcinoma; pancreatic adenocarcinoma; prostate adenocarcinoma; rectum adenocarcinoma; sarcoma; skin cutaneous melanoma; stomach adenocarcinoma; thyroid carcinoma; and uterine corpus endometrial carcinoma.

In the boxplot, *t*-test was used to test the difference between two groups, i.e., tumor and normal samples, and *P* value is the probability of obtaining a statistically significant result.

RESULTS

Oncomine database analyses for gene expression

From the Oncomine data analysis of *SYK* expression in normal and tumor tissues of various cancers, it was found that the *SYK* gene was overexpressed in bladder, breast and colon cancers, whereas it was underexpressed in leukemia, lymphoma and myeloma [Table 1]. Figure 1 shows the results of 542 unique analyses of *SYK* gene. In total, 25 studies of various cancers showed

a statistical difference in the *SYK* expression between normal and tumor tissues, while 19 studies each found overexpression of *SYK* gene in normal and tumor tissues. Remarkably, breast and brain tumors showed an overexpression of *SYK* gene in relation to normal tissues. On the contrary, hematological malignancies such as lymphoma and leukemia showed that *SYK* expression was significantly higher in normal cells than in tumor cells.

Data also showed that *SYK* expression was significantly downregulated in lung carcinoid tumors in comparison with normal tissues [Figure 2a and f]. However, in cancer types such as invasive breast and ductal breast carcinoma, *SYK* was overexpressed in tumor cells by 2.5- and 4.8-fold, respectively, in relation to normal tissue [Figure 2b and c]. Furthermore, in colorectal carcinoma and brain glioblastoma, *SYK* gene was upregulated by 2.4- and 2.1-fold, respectively [Figure 2d and e].

Kaplan–Meier survival analysis

The Kaplan–Meier plotter data analyses revealed that overexpression of *SYK* gene was significantly associated with better overall survival in pancreatic, gastric and lung cancer patients, while this association was nonsignificant in liver cancer patients [Figure 3].

cBioPortal mutation site and frequency analyses

From the 55,833 samples of 210 studies that were retrieved from TCGA pipeline using cBioPortal, a total of 562 *SYK* gene mutation sites were detected and found to be located between amino acids 0 and 635 [Figure 4]. Prostate cancer had the highest *SYK* gene mutation frequency, occurring at 14 different sites, and was mostly found in SH2 and kinase domains.

After showing the mutation sites and the frequency of *SYK* mutation in different cancers, the mean expression of mutated- and wild-type *SYK* in different cancer types was compared. As shown in Figure 5a, the mean *SYK* expression was high in diffuse large B-cell lymphoma, acute myeloid leukemia and thymoma compared with other cancer types. Nonmutated form of *SYK* represented the majority of the expressed gene. In addition, there is no prognostic value between mutated- and wild-type *SYK* in different cancer types, as shown in Figure 5b.

MethHC promoter methylation analysis

The *SYK* promoter hypermethylated in tumor samples of different cancer types compared with the normal

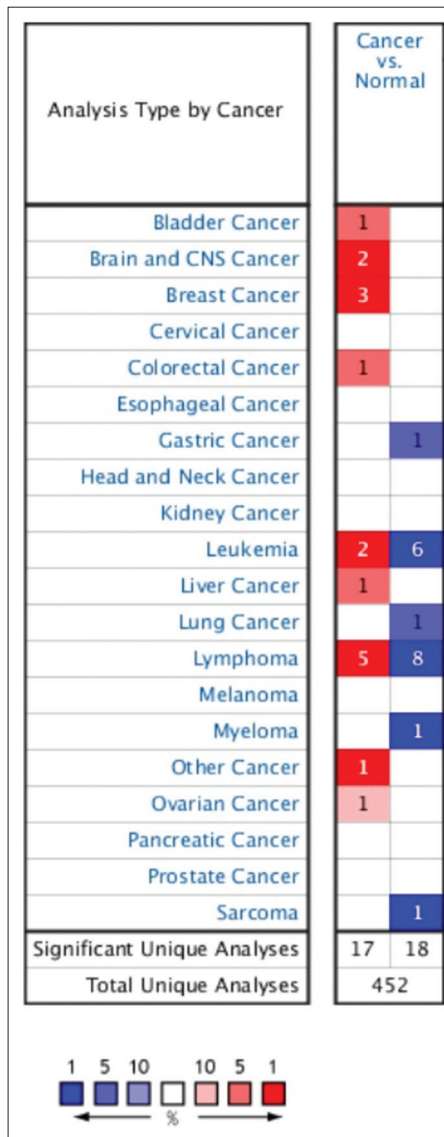


Figure 1: Overexpression (red) or underexpression (blue) of spleen tyrosine kinase mRNA in tumor tissues versus normal tissues

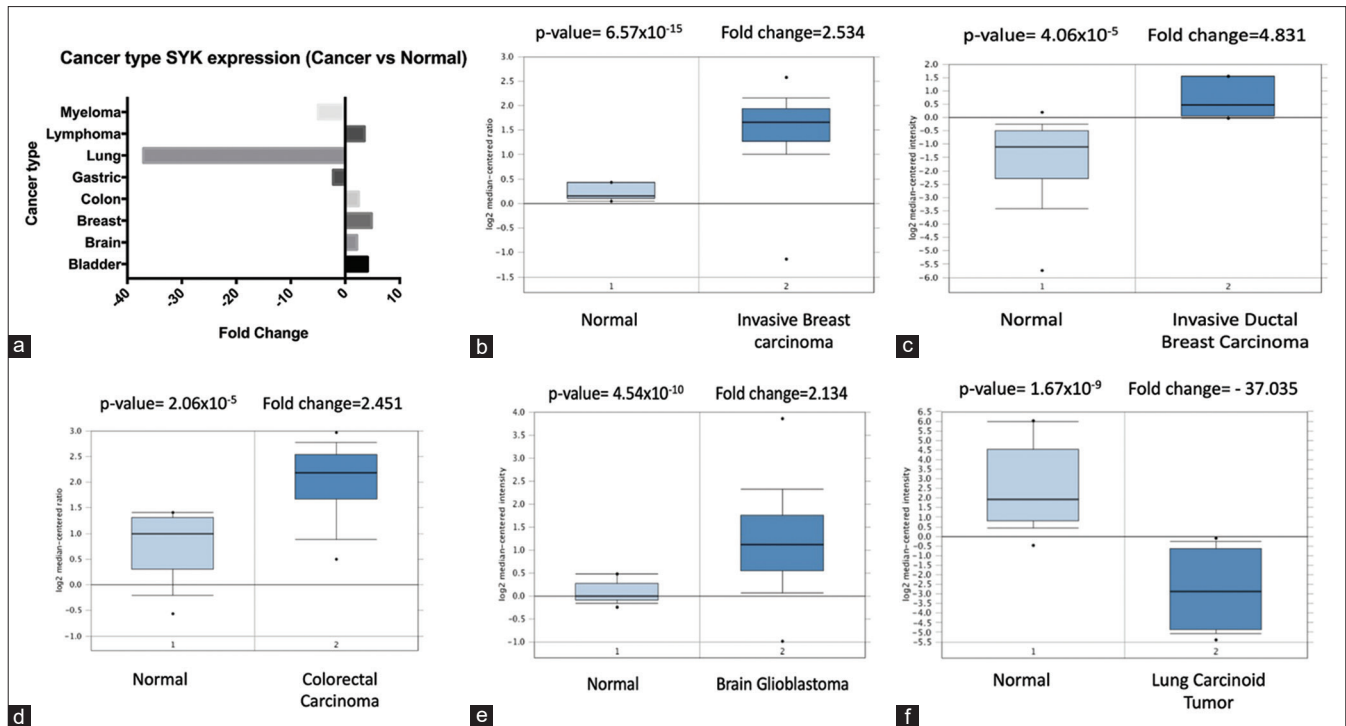


Figure 2: Spleen tyrosine kinase analysis in different cancer types using the OncoPrint database. (a) Fold change of spleen tyrosine kinase in various identified cancers. The box plot illustrates the expression of spleen tyrosine kinase gene in normal tissue (left) versus tumor tissue (right). Spleen tyrosine kinase expression in (b and c) breast carcinoma, (d and e) colorectal carcinoma and brain glioblastoma and (f) lung carcinoid tumor

Table 1: Spleen tyrosine kinase expression in different cancer types

Cancer	Cancer subtype	P	Fold change	Rank (%)	Sample	Study
Bladder	Infiltrating bladder urothelial carcinoma	1.47 × 10 ¹²	4.072	2	157	Sanchez-Carbayo <i>et al.</i> ^[15]
Brain	Brain glioblastoma	4.54 × 10 ¹⁰	2.134	4	557	Levine and The Cancer Genome Atlas Research Network ^[16]
Breast	Invasive ductal breast carcinoma	4.06 × 10 ⁵	4.831	1	22	Karnoub <i>et al.</i> ^[17]
	Invasive breast carcinoma	6.57 × 10 ¹⁵	2.534	6	59	Finak <i>et al.</i> ^[18]
	Medullary breast carcinoma	6.09 × 10 ⁸	2.093	9	176	Curtis <i>et al.</i> ^[19]
Colon	Colorectal carcinoma	2.06 × 10 ⁵	2.451	5	30	Graudens <i>et al.</i> ^[20]
Gastric	Gastric intestinal type adenocarcinoma	3.17 × 10 ⁹	-2.203	4	57	D'Errico <i>et al.</i> ^[21]
Leukemia	Chronic lymphocytic leukemia	1.59 × 10 ⁹	4.529	1	50	Rosenwald <i>et al.</i> ^[22]
	T-cell childhood acute lymphoblastic leukemia	2.02 × 10 ²⁰	-31.556	1	50	Coustan-Smith <i>et al.</i> ^[23]
	B-cell childhood acute lymphoblastic leukemia	1.65 × 10 ⁷	-11.069	2	242	Maia <i>et al.</i> ^[24]
	B-cell acute lymphoblastic leukemia	7.37 × 10 ⁹	-3.856	1	22	Haferlach <i>et al.</i> ^[25]
Lung	T-cell acute lymphoblastic leukemia	1.57 × 10 ²⁸	-2.263	6	248	Piccaluga <i>et al.</i> ^[26]
	Lung carcinoid tumor	1.67 × 10 ⁹	-37.035	3	37	Alizadeh <i>et al.</i> ^[27]
Lymphoma	Follicular lymphoma	1.02 × 10 ⁷	3.515	1	40	Bhattacharjee <i>et al.</i> ^[28]
	T-cell lymphoma	1.05 × 10 ⁹	3.096	7	48	Brune <i>et al.</i> ^[29]
	Hodgkin's lymphoma	1.44 × 10 ¹³	-5.647	1	37	
	Burkitt lymphoma	2.79 × 10 ⁹	-5.598	1	30	
	Diffuse large B-cell lymphoma	2.74 × 10 ⁷	-2.856	2	36	
Myeloma	Multiple myeloma	4.20 × 10 ¹³	-4.975	1	118	Zhan <i>et al.</i> ^[30]
Ovarian	Ovarian serious adenocarcinoma	7.11 × 10 ⁶	2.695	6	53	Yoshihara <i>et al.</i> ^[31]
Sarcoma	Synovial sarcoma	6.94 × 10 ⁷	-12.82	1	19	Detwiller <i>et al.</i> ^[32]

tissue is shown in Figure 6. Compared with normal tissues, SYK promoter hypermethylation was significantly higher ($P < 0.005$) in breast, renal, liver, lung, pancreatic, prostatic, skin and stomach cancers, but there was no differences in bladder, colon, head-and-neck and rectal cancers.

DISCUSSION

SYK is involved in many hematopoietic cell responses, especially immune signaling, that control cellular proliferation and differentiation. SYK has also been reported to be expressed in nonhematopoietic cells,

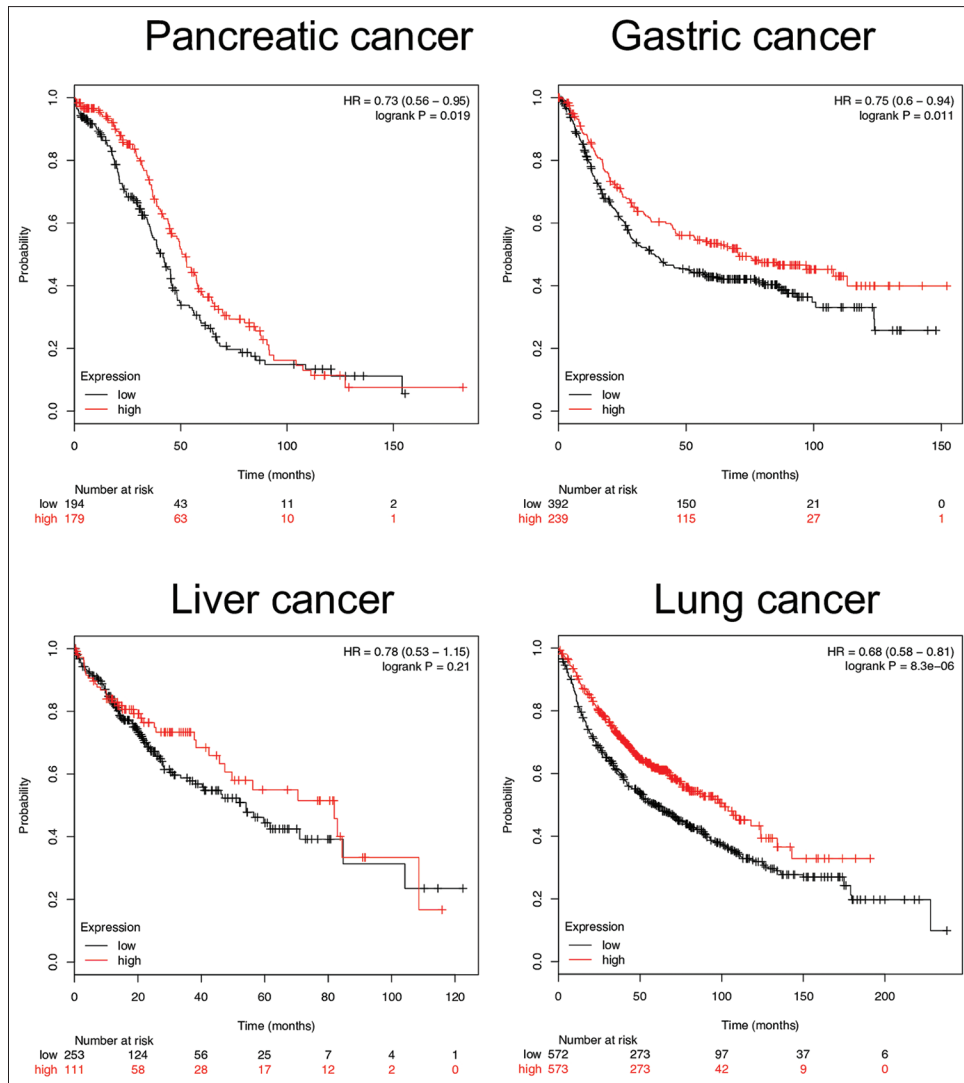


Figure 3: Correlations between spleen tyrosine kinase expression and survival rates of patients with different cancer types using Kaplan–Meier plotter database. Survival curves of patients with pancreatic, liver, gastric and lung cancers, segregated according to high (red) or low (black) expression of spleen tyrosine kinase

including epithelial cells, fibroblast cells and endothelial cells.^[33] It is functionally important in many cell types and plays a central role in a wide variety of cells. In cancers, numerous studies have shown the dual role of *SYK*: as a tumor promoter^[34–36] and as a malignant cell growth suppressor.^[37] The current study found that the mRNA expression of the *SYK* gene varied among different solid cancer types; *SYK* was overexpressed in breast, brain and pancreatic tumors as compared with normal cells. These findings suggest that *SYK* does play a role in the tumorigenesis of certain types of cancer. However, in liquid tumors, *SYK* seems to drive cell transformation.^[7]

SYK undergo autophosphorylation at the Tyr-518 site. After ligand binding to receptors, tyrosine residues are phosphorylated by Lyn, which is a Src-family nonreceptor.

The phosphorylation of tyrosine residues of *SYK* produces binding sites for CBL, VAV1 and PLC- γ and controls B-cell receptor cascades.^[38] These result in increasing the IP3 concentration, which in turn increases calcium ion internalization. In addition, *SYK* can activate several molecules such as a Bruton's tyrosine kinase (BTK) during the B-cell receptor-signaling pathway.^[39] BTK has been proposed to play a tumor-suppressive action in the epithelial cancer cells through p53-dependent mechanisms.^[40,41] Therefore, *SYK* could play a role as an antitumor gene in the epithelial cells though BTK regulation.

This study found that compared with normal cells, *SYK* is overexpressed in chronic lymphoblastic leukemia but underexpressed in acute childhood lymphoblastic leukemia. In line with this observation, the high expression of *SYK* in chronic lymphoblastic leukemia has been reported in a

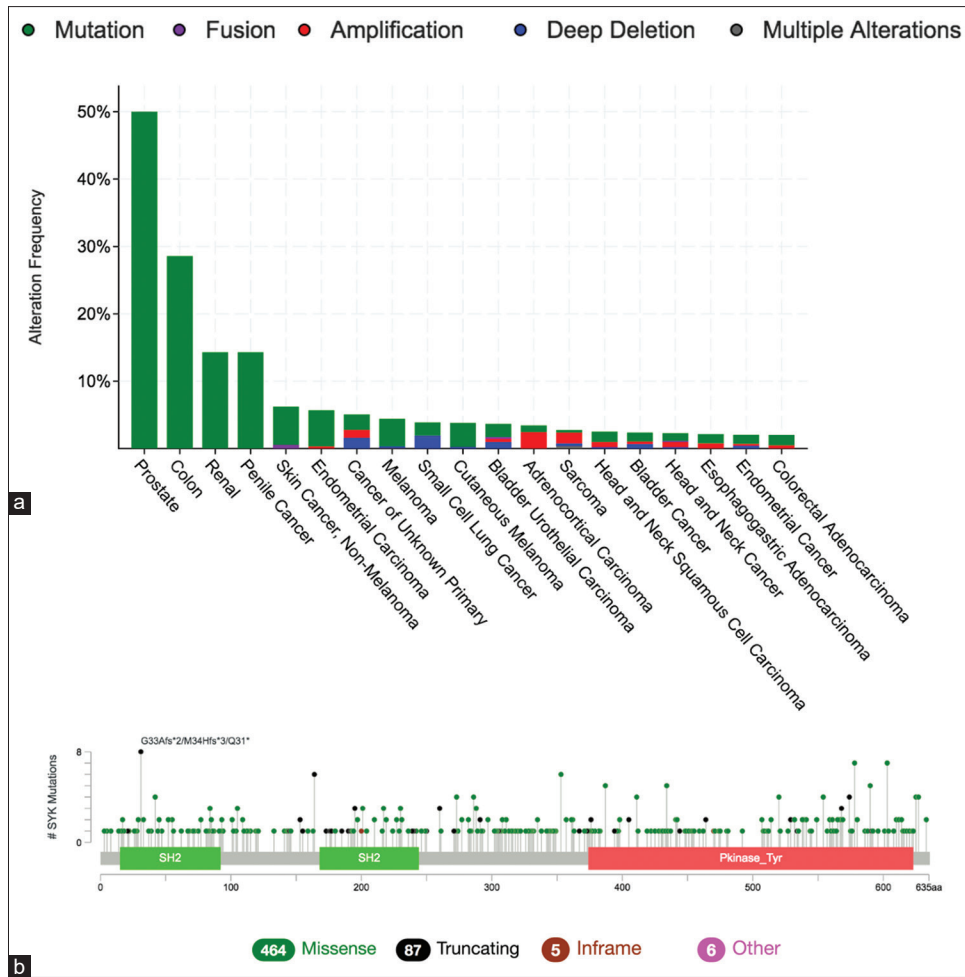


Figure 4: Mutation diagram of spleen tyrosine kinase in different cancer types across protein domains showing that spleen tyrosine kinase mutation mainly occurred in prostate cancer (a) and existed in different protein sites (b)

number of studies.^[34,35] However, there is no consensus in the literature regarding the *SYK* expression levels in acute leukemia: Some studies have shown elevated levels of *SYK* in this cancer,^[42,43] while others have shown downregulation of *SYK* expression in childhood acute leukemia.^[7] The reason for such underexpression of *SYK* is yet to be explained.

This study found that *SYK* expression is highly downregulated in lung carcinoid tumors compared with those in normal tissues, which is in contrast with the findings of several studies, where *SYK* expression was found to be higher in cancer cells. For instance, Fotheringham *et al.*^[44] showed that *SYK* is elevated in lung tumor. Similarly, *SYK* expression was significantly upregulated in the primary samples of neuroendocrine tumors compared with in the normal alveolar epithelium samples.^[36,45] However, given that lung carcinoids account for only 1%–2% of lung cancer cases, it would be difficult to conclude that the status of *SYK* expression differ across tumors in different organs.

The current study found a positive correlation between high expression of *SYK* and survival of patients in different solid tumors. This is consistent with the findings of other clinical studies. For instance, Nakashima *et al.*^[46] found a strong association between *SYK* expression and absence of lymph node metastasis in gastric cancer. Moreover, Toyama *et al.*^[47] showed that low mRNA expression of *SYK* in patients diagnosed with breast cancer was significantly associated with distant metastasis and poor prognosis.^[47] Taken together, these findings clearly indicate a relationship between low *SYK* expression and poor prognosis of many types of cancer.

The analysis in this study revealed that, in general, there are low number of mutations in the *SYK* gene in many cancers such as breast, colon, pancreatic and ovarian cancers. In addition, these alterations in the *SYK* gene did not appear to play a pivotal role in the overall survival. Although a growing body of evidence has shown that the loss of *SYK* clearly contributes to malignant phenotypes, no gene alterations such as mutations, deletions or translocation in

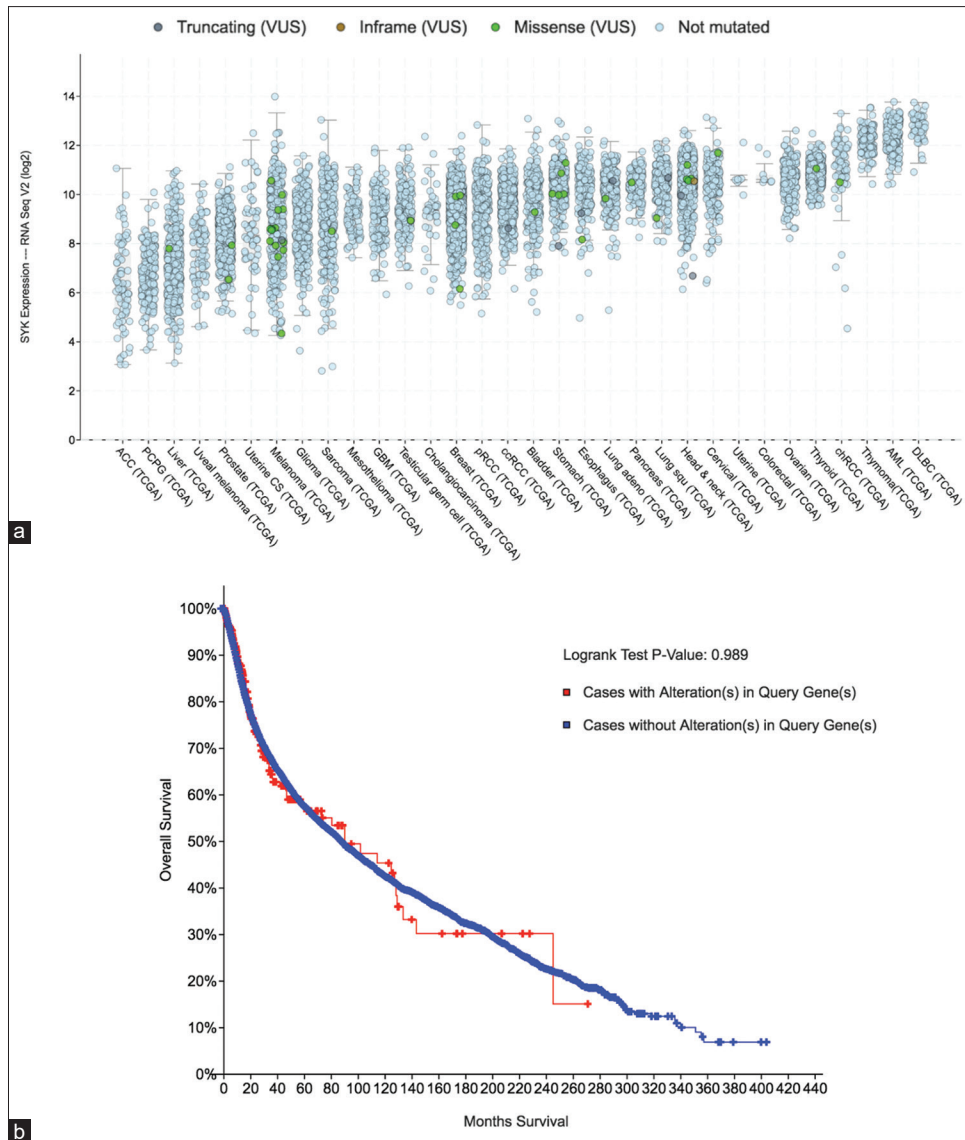


Figure 5: Wild-type and mutated-type spleen tyrosine kinase expression in different cancer types. (a) Mean spleen tyrosine kinase expression in different cancers. (b) The prognostic value of mutated- and/or wild-type spleen tyrosine kinase in different cancer type

the *SYK* gene have been found to be involved in naturally occurring tumor. For instance, Coopman and Mueller^[48] reported that in invasive breast cancer, the loss of SYK protein is associated with reduction in its mRNA levels, clearly explaining that the loss of *SYK* expression occurs at the transcription level. Moreover, mutation frequencies vary in different tumor stages.^[49,50]

Methylation is considered as a major mechanism for silencing tumor-suppressor genes.^[51] DNA hypermethylation, increasing of methyltransferase activity and global hypomethylation have been observed in human cancers. For instance, one of the most important tumor-suppressing genes in colorectal cancer is *CDKN2A*.^[52] *CDKN2A* has been found to be silenced by hypermethylation, and a recent meta-analysis suggested that *CDKN2A* promoter

hypermethylation is associated with unfavorable prognosis in colorectal cancer patients.^[53]

DNA methylation has also been used to predict and discover cancer origin from biological samples.^[54] Brock *et al.*^[55] discovered that four hypermethylated genes are associated with early tumor recurrence in lung cancer patients. These DNA methylation candidates can be targeted by certain demethylating agents to treat lung cancer. Indeed, Juergens *et al.*^[56] tested the efficacy of a combination of epigenetic inhibitors for lung cancer patients who had hypermethylated genes, and significant progression-free survival and overall survival were reported in the clinical trial. The study highlighted the possible role of methylation of SYK promoter in solid cancer, but no data were provided

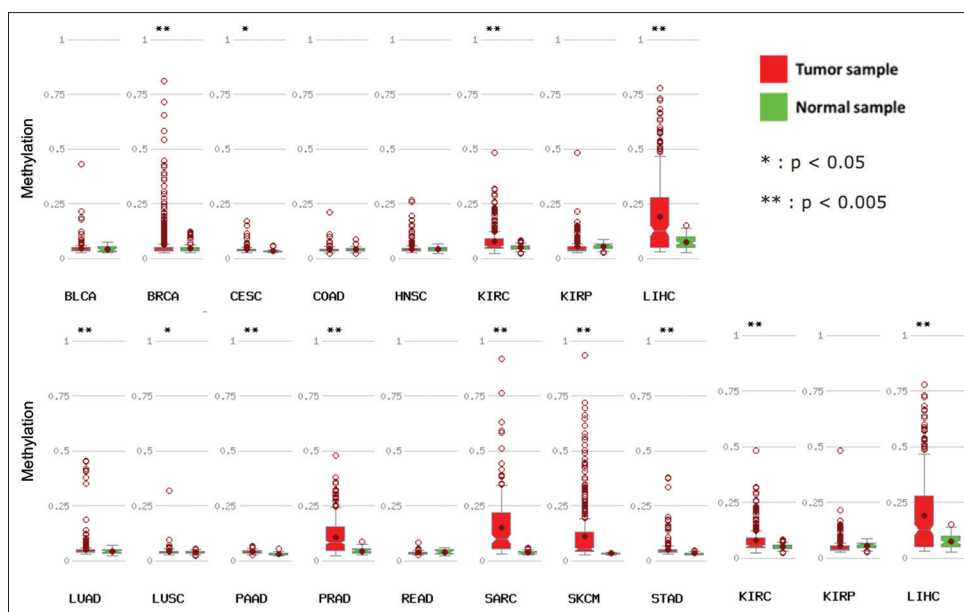


Figure 6: Average beta value of methylation of spleen tyrosine kinase promoter in different cancer types. The level of CpG methylation across the spleen tyrosine kinase proximal promoter (from – 1500 bp to TSS +1) upstream of the first exon analyzed using an *in silico* bioinformatics software MethHC49. Methylation across the spleen tyrosine kinase proximal promoter was determined in different tumors type (red) and the paired normal tissue (green)

regarding its status in blood cells. Previous work in monomorphic epitheliotropic intestinal T-cell lymphoma showed that *SYK* overexpression was associated with promoter hypomethylation.^[57] Therefore, *in vitro* and *in vivo* epigenetic studies should be conducted to validate the bioinformatics data of this study before *SYK* demethylation is used as a therapy in clinical trials.

Limitations

Although the systematic analysis revealed the dual role of *SYK* in different cancer types, it has some limitations that should be highlighted. First, analyses were conducted using different databases; therefore, there are likely to be differences in ethnicity, tumor stage and other cancer genetic factors in these genomic databases. In addition, there are multiple CpG sites in the promoter region of *SYK*, but the MethHC software does not provide the exact CpG methylation site of *SYK*.

CONCLUSION

In this analytical study, *SYK* was found to be overexpressed in bladder, colon and breast cancers and downregulated in leukemia and lymphoma. High expression of *SYK* was correlated with better prognosis of pancreatic, gastric, liver and lung patients. It was also found that in different cancers types, there was hypermethylation in the *SYK* promoter compared with the normal tissue. Therefore, using demethylating agents for *SYK* promoter could be exploited for increasing the sensitivity of

conventional cancer therapy. However, further analysis should be conducted to analyze the mutation patterns in each cancer and to validate the results before initiating the use of *SYK* in clinical settings.

Peer review

This article was peer reviewed by three independent and anonymous reviewers.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Keller B, Stumpf I, Strohmeier V, Usadel S, Verhoeven E, Eibel H, *et al.* High SYK Expression Drives Constitutive Activation of CD21 low B Cells. *J Immunol* 2017;198:4285-92.
2. Sadras T, Cutler J, Aguade-Gorgorio J, Chen Z, Coşgun KN, Pandey A, *et al.* Cooperation between SYK and ZAP70 kinases As a driver of oncogenic BCR-signaling in B-Cell malignancies. *Blood* 2018;132 Suppl 1:3922.
3. Böhmer R, Neuhaus B, Bühren S, Zhang D, Stehling M, Böck B, *et al.* Regulation of developmental lymphangiogenesis by Syk(+) leukocytes. *Dev Cell* 2010;18:437-49.
4. Meltzer EO, Berkowitz RB, Grossbard EB. An intranasal Syk-kinase inhibitor (R112) improves the symptoms of seasonal allergic rhinitis in a park environment. *J Allergy Clin Immunol* 2005;115:791-6.
5. Braselmann S, Taylor V, Zhao H, Wang S, Sylvain C, Baluom M, *et al.* R406, an orally available spleen tyrosine kinase inhibitor blocks fc receptor signaling and reduces immune complex-mediated inflammation. *J Pharmacol Exp Ther* 2006;319:998-1008.

6. Sharman J, Hawkins M, Kolibaba K, Boxer M, Klein L, Wu M, *et al.* An open-label phase 2 trial of entospletinib (GS-9973), a selective spleen tyrosine kinase inhibitor, in chronic lymphocytic leukemia. *Blood* 2015;125:2336-43.
7. Goodman PA, Wood CM, Vassilev A, Mao C, Uckun FM. Spleen tyrosine kinase (Syk) deficiency in childhood pro-B cell acute lymphoblastic leukemia. *Oncogene* 2001;20:3969-78.
8. Althubiti M. Spleen tyrosine kinase inhibition modulates p53 Activity. *J Cell Death* 2017;10:1179066017731564.
9. Moroni M, Soldatenkov V, Zhang L, Zhang Y, Stoica G, Gehan E, *et al.* Progressive loss of syk and abnormal proliferation in breast cancer cells. *Cancer Res* 2004;64:7346-54.
10. Rhodes DR, Yu J, Shanker K, Deshpande N, Varambally R, Ghosh D, *et al.* ONCOMINE: A cancer microarray database and integrated data-mining platform. *Neoplasia* 2004;6:1-6.
11. Lániczky A, Nagy Á, Bottai G, Munkácsy G, Szabó A, Santarpia L, *et al.* miRpower: A web-tool to validate survival-associated miRNAs utilizing expression data from 2178 breast cancer patients. *Breast Cancer Res Treat* 2016;160:439-46.
12. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, *et al.* Integrative analysis of complex cancer genomics and clinical profiles using the BioPortal. *Sci Signal* 2013;6:p11.
13. Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO, Aksoy BA, *et al.* The cBio cancer genomics portal: An open platform for exploring multidimensional cancer genomics data. *Cancer Discov* 2012;2:401-4.
14. Huang WY, Hsu SD, Huang HY, Sun YM, Chou CH, Weng SL, *et al.* MethHC: A database of DNA methylation and gene expression in human cancer. *Nucleic Acids Res* 2015;43:D856-61.
15. Sanchez-Carbayo M, Socci ND, Lozano J, Saint F, Cordon-Cardo C. Defining molecular profiles of poor outcome in patients with invasive bladder cancer using oligonucleotide microarrays. *J Clin Oncol* 2006;24:778-89.
16. Cancer Genome Atlas Research Network, Kandoth C, Schultz N, Cherniack AD, Akbani R, Liu Y, *et al.* Integrated genomic characterization of endometrial carcinoma. *Nature* 2013;497:67-73.
17. Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, *et al.* Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 2007;449:557-63.
18. Finak G, Bertos N, Pepin F, Sadekova S, Souleimanova M, Zhao H, *et al.* Stromal gene expression predicts clinical outcome in breast cancer. *Nat Med* 2008;14:518-27.
19. Curtis C, Shah SP, Chin SF, Turashvili G, Rueda OM, Dunning MJ, *et al.* The genomic and transcriptomic architecture of 2,000 breast tumours reveals novel subgroups. *Nature* 2012;486:346-52.
20. Graudens E, Boulanger V, Mollard C, Mariage-Samson R, Barlet X, Grémy G, *et al.* Deciphering cellular states of innate tumor drug responses. *Genome Biol* 2006;7:R19.
21. D'Errico M, de Rinaldis E, Blasi MF, Viti V, Falchetti M, Calcagnile A, *et al.* Genome-wide expression profile of sporadic gastric cancers with microsatellite instability. *Eur J Cancer* 2009;45:461-9.
22. Rosenwald A, Alizadeh AA, Widhopf G, Simon R, Davis RE, Yu X, *et al.* Relation of gene expression phenotype to immunoglobulin mutation genotype in B cell chronic lymphocytic leukemia. *J Exp Med* 2001;194:1639-47.
23. Coustan-Smith E, Song G, Clark C, Key L, Liu P, Mehrpooya M, *et al.* New markers for minimal residual disease detection in acute lymphoblastic leukemia. *Blood* 2011;117:6267-76.
24. Maia S, Haining WN, Ansén S, Xia Z, Armstrong SA, Seth NP, *et al.* Gene expression profiling identifies BAX-delta as a novel tumor antigen in acute lymphoblastic leukemia. *Cancer Res* 2005;65:10050-8.
25. Haferlach T, Kohlmann A, Wiczorek L, Basso G, Kronnie GT, Béné MC, *et al.* Clinical utility of microarray-based gene expression profiling in the diagnosis and subclassification of leukemia: Report from the international microarray innovations in Leukemia study group. *J Clin Oncol* 2010;28:2529-37.
26. Piccaluga PP, Agostinelli C, Califano A, Rossi M, Basso K, Zupo S, *et al.* Gene expression analysis of peripheral T cell lymphoma, unspecified, reveals distinct profiles and new potential therapeutic targets. *J Clin Invest* 2007;117:823-34.
27. Alizadeh AA, Eisen MB, Davis RE, Ma C, Lossos IS, Rosenwald A, *et al.* Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. *Nature* 2000;403:503-11.
28. Bhattacharjee A, Richards WG, Staunton J, Li C, Monti S, Vasa P, *et al.* Classification of human lung carcinomas by mRNA expression profiling reveals distinct adenocarcinoma subclasses. *Proc Natl Acad Sci U S A* 2001;98:13790-5.
29. Brune V, Tiacchi E, Pfeil I, Döring C, Eckerle S, van Noesel CJ, *et al.* Origin and pathogenesis of nodular lymphocyte-predominant Hodgkin lymphoma as revealed by global gene expression analysis. *J Exp Med* 2008;205:2251-68.
30. Zhan F, Hardin J, Kordsmeier B, Bumm K, Zheng M, Tian E, *et al.* Global gene expression profiling of multiple myeloma, monoclonal gammopathy of undetermined significance, and normal bone marrow plasma cells. *Blood* 2002;99:1745-57.
31. Yoshihara K, Tajima A, Komata D, Yamamoto T, Kodama S, Fujiwara H, *et al.* Gene expression profiling of advanced-stage serous ovarian cancers distinguishes novel subclasses and implicates ZEB2 in tumor progression and prognosis. *Cancer Sci* 2009;100:1421-8.
32. Detwiler KY, Fernando NT, Segal NH, Ryeom SW, D'Amore PA, Yoon SS. Analysis of hypoxia-related gene expression in sarcomas and effect of hypoxia on RNA interference of vascular endothelial cell growth factor A. *Cancer Res* 2005;65:5881-9.
33. Coopman PJ, Do MT, Barth M, Bowden ET, Hayes AJ, Basyuk E, *et al.* The syk tyrosine kinase suppresses malignant growth of human breast cancer cells. *Nature* 2000;406:742-7.
34. Chen L, Huynh L, Apgar J, Tang L, Rassenti L, Weiss A, *et al.* ZAP-70 enhances IgM signaling independent of its kinase activity in chronic lymphocytic leukemia. *Blood* 2008;111:2685-92.
35. Baudot AD, Jeandel PY, Mouska X, Maurer U, Tartare-Deckert S, Raynaud SD, *et al.* The tyrosine kinase Syk regulates the survival of chronic lymphocytic leukemia B cells through PKCdelta and proteasome-dependent regulation of Mcl-1 expression. *Oncogene* 2009;28:3261-73.
36. Udyavar AR, Hoeksema MD, Clark JE, Zou Y, Tang Z, Li Z, *et al.* Co-expression network analysis identifies Spleen Tyrosine Kinase (SYK) as a candidate oncogenic driver in a subset of small-cell lung cancer. *BMC Syst Biol* 2013;7 Suppl 5:S1.
37. Sung YM, Xu X, Sun J, Mueller D, Sentissi K, Johnson P, *et al.* Tumor suppressor function of Syk in human MCF10A *in vitro* and normal mouse mammary epithelium *in vivo*. *PLoS One* 2009;4:e7445.
38. Liu D, Mamorska-Dyga A. Syk inhibitors in clinical development for hematological malignancies. *J Hematol Oncol* 2017;10:145.
39. Wu J, Liu C, Tsui ST, Liu D. Second-generation inhibitors of Bruton tyrosine kinase. *J Hematol Oncol* 2016;9:80.
40. Althubiti M, Rada M, Samuel J, Escorsa JM, Najeeb H, Lee KG, *et al.* BTK modulates p53 activity to enhance apoptotic and senescent responses. *Cancer Res* 2016;76:5405-14.
41. Rada M, Althubiti M, Ekpenyong-Akiba AE, Lee KG, Lam KP, Fedorova O, *et al.* BTK blocks the inhibitory effects of MDM2 on p53 activity. *Oncotarget* 2017;8:106639-47.
42. Perova T, Grandal I, Nutter LM, Papp E, Matei IR, Beyene J, *et al.* Therapeutic potential of spleen tyrosine kinase inhibition for treating high-risk precursor B cell acute lymphoblastic leukemia. *Sci Transl Med* 2014;6:236a62.
43. Uckun FM, Dibirdik I, Qazi S, Yiv S. Therapeutic nanoparticle constructs of a JAK3 tyrosine kinase inhibitor against human B-lineage ALL cells. *Arzneimittelforschung* 2010;60:210-7.
44. Fotheringham JA, Coalson NE, Raab-Traub N. Epstein-Barr virus latent membrane protein-2A induces ITAM/Syk- and Akt-dependent epithelial migration through α v-integrin membrane translocation. *J Virol* 2012;86:10308-20.

45. Barretina J, Caponigro G, Stransky N, Venkatesan K, Margolin AA, Kim S, *et al.* The cancer cell line encyclopedia enables predictive modelling of anticancer drug sensitivity. *Nature* 2012;483:603-7.
46. Nakashima H, Natsugoe S, Ishigami S, Okumura H, Matsumoto M, Hokita S, *et al.* Clinical significance of nuclear expression of spleen tyrosine kinase (Syk) in gastric cancer. *Cancer Lett* 2006;236:89-94.
47. Toyama T, Iwase H, Yamashita H, Hara Y, Omoto Y, Sugiura H, *et al.* Reduced expression of the Syk gene is correlated with poor prognosis in human breast cancer. *Cancer Lett* 2003;189:97-102.
48. Coopman PJ, Mueller SC. The Syk tyrosine kinase: A new negative regulator in tumor growth and progression. *Cancer Lett* 2006;241:159-73.
49. Berg M, Nordgaard O, Kørner H, Oltedal S, Smaaland R, Søreide JA, *et al.* Molecular subtypes in stage II-III colon cancer defined by genomic instability: Early recurrence-risk associated with a high copy-number variation and loss of RUNX3 and CDKN2A. *PLoS One* 2015;10:e0122391.
50. Althubiti MA. Mutation frequencies in endometrial cancer patients of different ethnicities and tumor grades: An analytical study. *Saudi J Med Med Sci* 2019;7:16-21.
51. Kazanets A, Shorstova T, Hilmi K, Marques M, Witcher M. Epigenetic silencing of tumor suppressor genes: Paradigms, puzzles, and potential. *Biochim Biophys Acta* 2016;1865:275-88.
52. Liang JT, Chang KJ, Chen JC, Lee CC, Cheng YM, Hsu HC, *et al.* Hypermethylation of the p16 gene in sporadic T3N0M0 stage colorectal cancers: Association with DNA replication error and shorter survival. *Oncology* 1999;57:149-56.
53. Xing X, Cai W, Shi H, Wang Y, Li M, Jiao J, *et al.* The prognostic value of CDKN2A hypermethylation in colorectal cancer: A meta-analysis. *Br J Cancer* 2013;108:2542-8.
54. Kang S, Li Q, Chen Q, Zhou Y, Park S, Lee G, *et al.* Cancer locator: Non-invasive cancer diagnosis and tissue-of-origin prediction using methylation profiles of cell-free DNA. *Genome Biol* 2017;18:53.
55. Brock MV, Hooker CM, Ota-Machida E, Han Y, Guo M, Ames S, *et al.* DNA methylation markers and early recurrence in stage I lung cancer. *N Engl J Med* 2008;358:1118-28.
56. Juergens RA, Wrangle J, Vendetti FP, Murphy SC, Zhao M, Coleman B, *et al.* Combination epigenetic therapy has efficacy in patients with refractory advanced non-small cell lung cancer. *Cancer Discov* 2011;1:598-607.
57. Mutzbauer G, Maurus K, Buszello C, Pischmarov J, Roth S, Rosenwald A, *et al.* SYK expression in monomorphic epitheliotropic intestinal T-cell lymphoma. *Mod Pathol* 2018;31:505-16.