

■ Medicinal Chemistry | Hot Paper |

A New, Practical One-Pot Synthesis of Unprotected Sulfonimidamides by Transfer of Electrophilic NH to Sulfinamides

Flavia Izzo, [a] Martina Schäfer, [b] Robert Stockman, *[a] and Ulrich Lücking*[b]

Abstract: Unprotected tertiary sulfonimidamides have been prepared in good to excellent yields in a one-pot transformation from tertiary sulfinamides through NH transfer. The reaction is mediated by commercially available (diacetoxyiodo)benzene and ammonium carbamate in methanol under convenient conditions. A wide range of functional groups are tolerated and initial results indicate that the NH transfer

is stereospecific. A small molecule X-ray analysis of NH sulfonimidamide **2a** and its behavior in selected in vitro assays in comparison to the matched sulfonamide are also reported. This new reaction provides a safe, short and efficient approach to sulfonimidamides, which have been the subject of recent, growing interest in the life sciences.

Introduction

The sulfonamide group 1 (Figure 1) has long been a very important pharmacophore in drug discovery, being widely used in anticancer, anti-inflammatory and antiviral agents.^[1] In contrast, sulfonimidamides 2, the mono-aza analogues of sulfonamides, have been rather neglected in the life sciences so far,

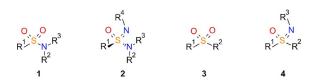


Figure 1. General structures of sulfonamides **1** (1° R², R³=H; 2° R²=H, R³ \neq H; 3° R²+H, R³ \neq H), sulfonimidamides **2** (1° R², R³=H; 2° R²=H, R³ \neq H; 3° R²+H, R³+H), sulfones **3** and sulfoximines **4**.

despite offering very interesting properties such as high stability, favorable physicochemical properties, multiple hydrogenbond acceptor/donor functionalities and structural diversity, as highlighted recently by Arvidsson and co-workers.^[2] Arguably,

the use of sulfonimidamides in the life sciences has been hampered by the lack of their commercial availability and the limited methods available for their synthesis.^[3] The "state of the art" concerning sulfonimidamides in drug discovery resembles that of sulfoximines **4**,^[4] the mono-aza analogues of sulfones **3** (Figure 1), about 15 years ago.

After its late discovery in 1949, the sulfoximine group garnered only a very moderate interest in medicinal chemistry for many decades. In recent years, however, interest in sulfoximine chemistry has increased substantially, as evidenced by the development of new and safe methods for the preparation of sulfoximines and the clinical evaluation of at least four novel sulfoximines, the kinase inhibitors roniciclib (BAY 1000394),^[5] atuveciclib (BAY 1143572),^[6] AZD 6738^[7] and BAY 1251152.^[8] To the best of our knowledge, a sulfonimidamide candidate has yet to be evaluated in clinical trials; however, there are a few recent examples of sulfonimidamides in medicinal chemistry^[2] including an analogue of the clinical anticancer agent tasisulam, with comparable antiproliferative activity against two melanoma cell lines in vitro, [9] and saccharin aza bioisosteres, which are interesting, new scaffolds for drug design^[10] (Figure 2).



[b] Dr. M. Schäfer, Dr. U. Lücking Bayer AG, Pharmaceuticals Division, Drug Discovery Müllerstr. 178, 13353 Berlin (Germany) E-mail: ulrich.luecking@bayer.com

Supporting information and the ORCID number(s) for the author(s) of this article can be found under https://doi.org/10.1002/chem.201703272.

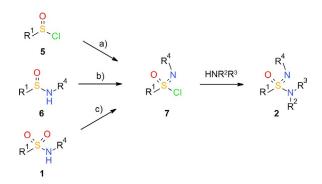
© 2017 The Authors. Published by Wiley-VCH Verlag GmbH & Co. KGaA. This is an open access article under the terms of Creative Commons Attribution NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.



Figure 2. Sulfonimidamide-based structure of the analogue of tasisulam and general structures of novel saccharin aza-bioisosteres.

Sulfonimidamides **2** were first synthesized in 1962 by Levchenko and co-workers.^[11] Since then, only a few synthetic approaches for the preparation of sulfonimidamides have been





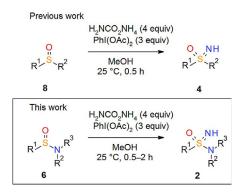
Scheme 1. Common strategy for the synthesis of sulfonimidamides 2 through the reaction of amines with sulfonimidoyl chlorides 7, which can be prepared by a) oxidative imidation of sulfinyl chlorides 5, b) oxidative chlorination of sulfinamides 6 or c) deoxychlorination of secondary sulfonamides 1.

reported, usually involving the nucleophilic substitution of a sulfonimidoyl chloride **7** by an amine (Scheme 1).^[12] The required key chloride intermediates **7** have been prepared through different strategies that generally hinge on the oxidative imidation of sulfinyl chlorides **5**,^[11,13] oxidative chlorination of sulfinamides **6**^[14] or deoxychlorination of secondary sulfonamides **1**.^[3,15] However, most of these protocols require the use of a protecting group, and deprotection to the corresponding unprotected NH sulfonimidamides can be challenging. Furthermore, many protocols have a rather limited substrate scope and/or rely on reagents with an associated safety risk, an example being *tert*-butyl hypochlorite, which is commonly used for oxidative chlorination despite its explosive nature.^[16]

A new synthetic strategy for unprotected tertiary sulfonimidamides **2**, based on a copper-catalyzed reaction of sulfoximines **4a** with secondary amines, was disclosed by Bolm and co-workers in 2016 (Scheme 2).^[17] However, this method requires prior synthesis of the corresponding sulfoximine **4a**, which in our experience can be challenging, depending on the nature of R¹.^[18]

Scheme 2. Redox-neutral S–C to S–N bond-exchange reaction allowing the conversion of sulfoximines **4a** into unprotected tertiary sulfonimidamides **2**; TC = thiophene-2-carboxylate.

During the course of our long-standing interest in sulfoximines in medicinal chemistry,^[19] we have witnessed significant improvements over the last 15 years towards safe and efficient synthetic methods for the preparation of sulfoximines. Our attention was captured by a recent report from Luisi, Bull and co-workers outlining the use of a mixture of ammonium carbamate and (diacetoxyiodo)benzene [Phl(OAc)₂] for the direct conversion of sulfoxides **8** into NH sulfoximines **4** at room temperature (Scheme 3).^[20] Furthermore, this facile and new NH-transfer method was applicable to a wide substrate scope and



Scheme 3. Synthesis of NH sulfoximines **4** by NH transfer to sulfoxides **8** using ammonium carbamate mediated by the hypervalent iodine reagent PhI(OAc)₂, as described by Bull and Luisi,^[20] and proposed synthesis of tertiary NH sulfonimidamides **2** ($R^2 \neq H$, $R^3 \neq H$) from tertiary sulfinamides **6** ($R^2 \neq H$, $R^3 \neq H$) by applying similar reaction conditions.

tolerated a large number of heterocycles and other functionalities.^[21] Attracted by the use of commercial reagents and the robust results of this new one-pot method, we were intrigued as to whether these safe reaction conditions could also be useful for the facile synthesis of sulfonimidamides **2** by NH transfer to sulfinamides **6**.

Results and Discussion

Initially, the reported reaction conditions^[20] for the imination of sulfoxides **8** were applied to a commercially available tertiary sulfinamide. Thus, 1-(phenylsulfinyl)piperidine (**6a**; 100 mg, 0.48 mmol) was treated with ammonium carbamate (4 equiv) in the presence of Phl(OAc)₂ (3 equiv) in methanol at room temperature for 1 hour (Scheme 4). To our delight, the unpro-

Scheme 4. Synthesis of tertiary NH sulfonimidamide 2a by NH transfer to tertiary sulfinamide 6a.

tected tertiary sulfonimidamide **2a** was isolated in 94% yield after column chromatography. A scaled-up reaction of sulfinamide **6a** (4.0 g, 19.1 mmol) also resulted in a clean conversion and very good yield (88%). The structure of sulfonimidamide **2a** was confirmed by X-ray analysis (Figure 3).

Given the clean reaction and high yield, we then elected to explore the substrate scope of this new process. Our initial focus was on variation of the NR^2R^3 group of the sulfinamide $\bf 6$ and thus, tertiary sulfinamides $\bf 6a-k$ were subjected to the standard reaction conditions (Table 1). Gratifyingly, the reactions were generally very clean, resulting in the desired unprotected tertiary sulfonimidamides $\bf 2a-k$ in good to excellent isolated yields. Only the reaction of sulfinamide $\bf 6d$ ($NR^2R^3=NiPr_2$)



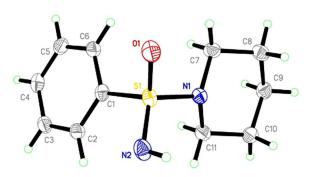


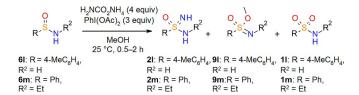
Figure 3. ORTEP plot (50% thermal ellipsoids) of the crystal structure of sulfonimidamide 2a.

Table 1. Exploration of the substrate scope of the NH transfer reaction: Variation of the NR^2R^3 group in tertiary sulfinamides $\bf 6a-k$.						
0 5 N,R ³	H ₂ NCO ₂ NH ₄ (4 equiv) PhI(OAc) ₂ (3 equiv) MeOH 25 °C, 0.5–2 h	S NH R ²	3			
6a–k	=3=3	2a-k				
Sulfonimidamide	NR ² R ³		Yield [%] ^[a]			
2 a	1-piperidinyl		94			
2b	NMe_2		72			
2c	NEt ₂					
2 d	NiPr ₂		17			
2e	1-azetidinyl					
2 f	1-pyrrolidinyl					
2 g	4-morpholinyl					
2h	N N O		65			
2i	·N		67			
2 j	··N N N		78			
2 k ^(b)	N N S N N N N N N N N N N N N N N N N N		50			

resulted in low conversion, presumably due to steric hindrance of the bulky N,N-diisopropylamino group, with the tertiary sulfonimidamide 2d isolated in 17% yield along with recovered starting material. Notably, the reaction of 1,4-bis(phenylsulfinyl)piperazine (6k) resulted in a double NH transfer to give the desired product 2k.

[a] Isolated yield. [b] Starting material 6 k = 1,4-bis(phenylsulfinyl)pipera-

We found, however, that the reaction is limited to the use of tertiary sulfinamides 6. Subjecting the commercial, primary sulfinamide 61 to the standard conditions did not result in formation of the corresponding sulfonimidamide 21; rather, sulfonimidate 91 was isolated in 58% yield along with sulfonamide 11 (15% yield, Scheme 5). Reaction of the secondary ethyl sulfinamide 6 m resulted in a mixture of products from which sulfonimidate 9 m was isolated as the main product. These results are



Scheme 5. Behavior of primary (6 l) and secondary (6 m) sulfinamides under the NH transfer reaction conditions.

in line with previous reports of the preparation of sulfonimidates from primary and secondary sulfinamides 6 and hypervalent iodine reagents in the presence of suitable alcohols. [22] The formation of sulfonamides 1 arising from the standard oxidation of sulfinamides 6 is a side reaction under these condi-

Next, the scope of the R¹ group was explored, with tertiary sulfinamides 6n-y being additionally subjected to the standard reaction conditions (Table 2). Again, the reactions were very clean, giving the desired unprotected tertiary sulfonimidamides 2 n-y in good to excellent isolated yields. Furthermore, a broad variety of substitution patterns and functional groups common in medicinal chemistry were tolerated. Similar to the reaction of the sterically demanding N,N-diisopropylbenzenesulfinamide (6d, Table 1), only the ortho-chloro derivative 6t $(R^1 = 2 - CIC_6H_4)$ resulted in an incomplete but nevertheless clean reaction, providing the tertiary sulfonimidamide 2t in 29% yield under the standard conditions. However, when the reaction was repeated using an additional portion of PhI(OAc)₂ and ammonium carbamate, there was improved conversion which resulted in a 48% isolated yield of 2t, along with un-

Table 2. Exploration of the substrate scope of the NH transfer reaction: Variation of the R¹ group in tertiary sulfinamides 6 n-y.

······································							
R ¹ S N	H ₂ NCO ₂ NH ₄ (4 equiv) PhI(OAc) ₂ (3 equiv) MeOH 25 °C, 0.5–2 h	NH R ¹ S					
6n–y	2n–y						
Sulfonimidamide	R ¹	Yield [%] ^[a]					
2a	Ph	94					
2 n	$4-MeOC_6H_4$	87					
20	4-FC ₆ H ₄	89					
2p	$4-O_2NC_6H_4$	62					
2 q	$3-F_3CC_6H_4$	81					
2r	$4-MeC_6H_4$	95					
2r*	$4-MeC_6H_4$	88 (48 % ee ^[b])					
2 s	3-CIC ₆ H ₄	85					
2t	2-CIC ₆ H ₄	29 (48) ^[c]					
2 u	2-pyridyl	85					
2 v	3-pyridyl	53					
2 w	cyclohexyl	88					
2x	<i>i</i> Pr	70					
2 у	<i>t</i> Bu	83					

[a] Isolated yield. [b] ee determined by chiral-phase HPLC analysis: 2 r* obtained from starting material 6r* with 51% ee. [c] Additional ammonium carbamate and PhI(OAc)2 were used.

zine.



reacted starting material. To our delight, sulfinamides with pyridyl or alkyl substituents also reacted successfully to give the corresponding sulfonimidamides **2u**–**y** in very good yields, illustrating the broad substrate scope of this new, mild reaction. As an initial insight into the stereospecificity of this process, the optically enriched sulfinamide **6r***, which had been prepared with 51% *ee* using the commercial Andersen reagent, was subjected to the standard reaction conditions. The NH transfer proceeded stereospecifically to provide product **2r*** with 48% *ee*.

In a preliminary assessment of the medicinal chemistry properties of sulfonimidamides, the behavior of the fragment-like sulfonimidamide 2a compared to the matched sulfonamide analogue 1a in selected in vitro assays was undertaken (Table 3). Considering that our major concern was a possible inherent instability of the sulfonimidamide group, the hydrolytic stability of the two compounds at five different pH values was investigated, along with the metabolic stability in liver microsomes (human, rat and mouse) and also rat hepatocytes in vitro. Furthermore, the Caco2 permeability and logD values were determined. In line with our expectations, sulfonamide 1a displayed a high hydrolytic stability after 24 hours with stirring in media with pH values ranging from 2.0-7.4 (Table 3). Moderate hydrolytic stability was evident at pH 8.0, with 67% recovered test compound. However, sulfonimidamide 2a also has a very high hydrolytic stability, with 100% recovered test compound after 24 hours with stirring at all tested pH values. In vitro pharmacokinetic studies with sulfonamide 1 a revealed a moderate metabolic stability in human liver microsomes, resulting in a moderate predicted blood clearance (CL_b) of 0.41 Lh⁻¹ kg⁻¹; however, a low metabolic stability and high pre-

Table 3. Comparison of the in vitro properties of sulfonamide **1a** and NH sulfonimidamide **2a**.

Compound	Recovery [%] ^[a] (pH)		CL _b ^[b] rHep [L h ⁻¹ kg ⁻¹]	P_{app} A-B ^[c] [nm s ⁻¹]	Efflux ratio ^[c]	log <i>D</i> pH 7.5 ^[d]
O S N	91 (2.0) 85 (4.5) 100 (6.5) 100 (7.4) 67 (8.0)	3.8 (r)	4.0	393	0.64	2.6
NH S N	100 (2.0) 100 (4.5) 100 (6.5) 100 (7.4) 100 (8.0)	3.0 (r)	3.6	378	0.59	1.9

[a] Hydrolytic stability measured as recovery of test compound after 24 hours with stirring at pH 2.0 (citrate buffer), pH 4.5 (citrate buffer), pH 6.5 (phosphate-buffered saline), pH 7.4 (phosphate-buffered saline), and pH 8.0 (sodium borate buffer). [24] [b] Predicted hepatic metabolic clearance based on a high-throughput metabolic stability assay using 1) pooled human liver microsomes (hLMs), ii) pooled rat liver microsomes (rLMs), iii) pooled mouse liver microsomes (mLMs), and iv) freshly harvested rat hepatocytes (rHep). [25] [c] $P_{\rm app}$ A–B (apical to basolateral) and efflux ratio (ER) data were generated in a bidirectionally performed Caco2 permeability assay in a 24-well format; ER was calculated as $P_{\rm app}$ B–A/ $P_{\rm app}$ A–B [25] [d] Determined by reversed-phase HPLC. [26]

dicted blood clearance of sulfonamide 1 a was evident both in rat and mouse liver microsomes [CL_h=3.8 L h⁻¹ kg⁻¹ (rat), $4.0 \, \text{Lh}^{-1} \, \text{kg}^{-1}$ (mouse)]. A high blood clearance ($4.0 \, \text{Lh}^{-1} \, \text{kg}^{-1}$) was also predicted from in vitro studies with rat hepatocytes. The main sites of metabolism were not determined in these studies. In comparison, the matched sulfonimidamide 2a revealed a trend for higher metabolic stabilities in vitro: studies with human and mouse liver microsomes resulted in low predicted blood clearances of 0.04 and 1.1 Lh⁻¹kg⁻¹, respectively. Sulfonimidamide 2a exhibited a low metabolic stability in rat preparations, with correspondingly high predicted CL_b values [3.0 Lh⁻¹kg⁻¹ (rat liver microsomes), 3.6 Lh⁻¹kg⁻¹ (rat hepatocytes)]; however, these data also revealed a trend in rats for a higher metabolic stability of sulfonimidamide 2a relative to sulfonamide 1 a. In the Caco2 screening assay, sulfonamide 1 a and the matched sulfonimidamide 2a both had high permeability coefficients (P_{app} A-B) of 393 and 378 nm s⁻¹, respectively, and no evidence of efflux. However, the switch from the sulfonamide to the sulfonimidamide results in a remarkable decrease in lipophilicity, with a logD value of 2.6 (1 a) compared to 1.9 (2a).

Conclusion

We have utilized electrophilic NH transfer to achieve the first direct synthesis of tertiary NH sulfonimidamides **2** from tertiary sulfinamides **6**. The developed protocol relies on the readily available nitrogen source ammonium carbamate and the oxidant Phl(OAc)₂, and is compatible with a broad substrate scope, providing the corresponding sulfonimidamides **2** in good to excellent yields. Initial results indicate that this reaction process is stereospecific; additional investigations are currently underway. In vitro studies of the properties relevant to medicinal chemistry for compound **2a** have not revealed any intrinsic flaw of the sulfonimidamide group with respect to its application as a new and versatile pharmacophore in the life sciences.

Experimental Section

General synthetic procedure for sulfonimidamides 2

To the sulfinamide (0.48 mmol, 1.00 equiv) were added (diacetoxyiodo)benzene (1.43 mmol, 3.00 equiv), ammonium carbamate (1.91 mmol, 4.00 equiv) and finally MeOH (0.96 mL, 0.5 m). The mixture was stirred in an open flask at RT for 0.5–2 h. When all the starting material had been consumed (as monitored by TLC and UPLC-MS), saturated aqueous NaHCO3 solution and EtOAc were added, and the mixture was stirred for 5 min. The organic layer was separated and the aqueous layer was extracted with EtOAc (3 x). The organic phases were combined, washed with brine (3 x) and dried over water-repellent filter paper. The volatiles were removed under reduced pressure and the crude product was purified by flash column chromatography.

1-(S-Phenylsulfonimidoyl)piperidine (2a)

Prepared according to the general procedure, from commercial 1-(phenylsulfinyl)piperidine (Sigma–Aldrich); crude purified by flash





column chromatography (KP-Sil, 0-40% EtOAc in hexane) to give 2a as a yellow-orange solid (105 mg, 94%): m.p. 99-100 °C; ¹H NMR (600 MHz, [D₆]DMSO): $\delta = 7.79 - 7.74$ (m, 2H), 7.66–7.61 (m, 1H), 7.60-7.55 (m, 2H), 4.29 (s, 1H), 2.91-2.74 (m, 4H), 1.56-1.44 (m, 4H), 1.30 ppm (q, J=5.82 Hz, 2H); ¹³C NMR (101 MHz, [D₆]DMSO): δ = 136.6, 132.0, 128.7, 127.7, 47.4, 25.2, 23.2 ppm; IR (KBr): $\tilde{\nu}\!=\!3234$, 2989, 2933, 2837, 1445, 1256, 907, 721, 687, 575 cm⁻¹; HRMS (ESI-TOF) m/z [M+H]⁺ calcd for $C_{11}H_{17}N_2OS$: 225.1062, found: 225.1066.

Acknowledgements

The support of U. Ganzer and P. Lienau for the measurement and interpretation of the physicochemical and in vitro pharmacokinetic properties is highly appreciated. We thank M. Bergmann and K. Greenfield for valuable technical support with the manuscript.

Conflict of interest

The authors declare no conflict of interest.

Keywords: drug design ⋅ hypervalent iodine ⋅ medicinal chemistry · sulfinamides · sulfonimidamides

- [1] For recent reviews, see for example: a) Ajeet, A. K. Mishra, A. Kumar, J. Pharmacol. Sci. 2015, 3, 18; b) A. Kołaczek, I. Fusiarz, J. Ławecka, D. Branowska, Chemik 2014, 68, 620; c) S. Shoaib Ahmad Shah, G. Rivera, M. Ashfaq, Mini-Rev. Med. Chem. 2013, 13, 70.
- [2] P. K. Chinthakindi, T. Naicker, N. Thota, T. Govender, H. G. Kruger, P. I. Arvidsson, Angew. Chem. Int. Ed. 2017, 56, 4100.
- [3] Y. Chen, J. Gibson, RSC Adv. 2015, 5, 4171.
- [4] a) C. R. Johnson, Aldrichimica Acta 1985, 18, 3; b) M. Reggelin, C. Zur, Synthesis 2000, 1; c) U. Lücking, Angew. Chem. Int. Ed. 2013, 52, 9399; Angew. Chem. 2013, 125, 9570; d) M. Frings, C. Bolm, A. Blum, C. Gnamm, Eur. J. Med. Chem. 2017, 126, 225; e) J. A. Sirvent, U. Lücking, ChemMedChem 2017, 12, 487.
- [5] a) G. Siemeister, U. Luecking, A. M. Wengner, P. Lienau, W. Steinke, C. Schatz, D. Mumberg, K. Ziegelbauer, Mol. Cancer Ther. 2012, 11, 2265; b) U. Lücking, R. Jautelat, M. Krüger, T. Brumby, P. Lienau, M. Schäfer, H. Briem, J. Schulze, A. Hillisch, A. Reichel, A. M. Wengner, G. Siemeister, ChemMedChem 2013, 8, 1067.
- [6] a) A. Scholz, T. Oellerich, A. Hussain, S. Lindner, U. Luecking, A. O. Walter, P. Ellinghaus, R. Valencia, F. von Nussbaum, D. Mumberg, M. Brands, S. Ince, H. Serve, K. Ziegelbauer, Cancer Res. 2016, 76, 3022; b) A. Scholz, U. Luecking, G. Siemeister, P. Lienau, U. Boemer, P. Ellinghaus, A. O. Walter, R. Valencia, S. Ince, F. von Nussbaum, D. Mumberg, M. Brands, K. Ziegelbauer, Cancer Res. 2015, 75, DDT02-02; c) U. Luecking, A. Scholz, P. Lienau, G. Siemeister, D. Kosemund, R. Bohlmann, K. Eis, M. Gnoth, I. Terebesi, K. Meyer, K. Prelle, R. Valencia, S. Ince, F. von Nussbaum, D. Mumberg, K. Ziegelbauer, B. Klebl, A. Choidas, P. Nussbaumer, M. Baumann, C. Schultz-Fademrecht, G. Ruehter, J. Eickhoff, M. Brands, Cancer Res. 2015, 75, 2828.
- [7] K. M. Foote, A. Lau, J. W. M. Nissink, Future Med. Chem. 2015, 7, 873.
- [8] U.T. Luecking, A. Scholz, D. Kosemund, R. Bohlmann, H. Briem, P. Lienau, G. Siemeister, I. Terebesi, K. Meyer, K. Prelle, R. Valencia, S. Ince, F. von Nussbaum, D. Mumberg, K. Ziegelbauer, M. Brands, AACR Annual Meeting, Washington, DC, April, 2017, MS.CH01.01 #984.
- [9] A.-D. Steinkamp, L. Schmitt, X. Chen, K. Fietkau, R. Heise, J. M. Baron, C. Bolm, Skin Pharmacol. Physiol. 2016, 29, 281.

- [10] Y. Chen, C.-J. Aurell, A. Pettersen, R. J. Lewis, M. A. Hayes, M. Lepistö, A. C. Jonson, H. Leek, L. Thunberg, ACS Med. Chem. Lett. 2017, 8, 672.
- [11] E. S. Levchenko, N. Y. Derkach, A. V. Kirsanov, Zh. Obshch. Khim. 1962, 32,
- [12] For a comprehensive review, see: E. S. Levchenko, L. N. Markovskii, Y. G. Shermolovich, Russ. J. Org. Chem. 2000, 36, 143.
- [13] a) E. S. Levchenko, N. Y. Derkach, A. V. Kirsanov, Zh. Obshch. Khim. 1960, 30, 1971; b) E. S. Levchenko, N. Y. Derkach, A. V. Kirsanov, Zh. Obshch. Khim. 1961, 31, 1971; c) S. Tsushima, Y. Yamada, T. Onami, K. Oshima, M. O. Chaney, N. D. Jones, J. K. Swartzendruber, Bull. Chem. Soc. Jpn. 1989, 62, 1167; d) P. H. Di Chenna, F. Robert-Peillard, P. Dauban, R. H. Dodd, Org. Lett. 2004, 6, 4503; e) C. Fruit, F. Robert-Peillard, G. Bernardinelli, P. Müller, R. H. Dodd, P. Dauban, Tetrahedron: Asymmetry 2005, 16, 3484; f) T. A. Khan, J. D. Scott, J. N. Cumming, Merck Sharp & Dohme Corp., WO 2014/150340, 2014.
- [14] a) E. U. Jonsson, C. C. Bacon, C. R. Johnson, J. Am. Chem. Soc. 1971, 93, 5306; b) E. U. Jonsson, C. C. Bacon, C. R. Johnson, J. Am. Chem. Soc. 1971, 93, 5308; c) C. R. Johnson, E. U. Jonsson, C. C. Bacon, J. Org. Chem. 1979, 44, 2055; d) C. R. Johnson, A. Wambsgans, J. Org. Chem. 1979, 44, 2278; e) S. G. Pyne, J. Org. Chem. 1986, 51, 81; f) K. Okuma, T. Kolke, H. Ohta, J. Org. Chem. 1988, 53, 4190; g) M. Harmata, Tetrahedron Lett. 1989, 30, 437; h) R. Kluge, H. Hocke, M. Schulz, F. Schilke, Phosphorus Sulfur Silicon Relat. Elem. 1999, 149, 179; i) M. Reggelin, B. Junker, Chem. Eur. J. 2001, 7, 1232; j) O. García Mancheño, C. Bolm, Beilstein J. Org. Chem. 2007, 3, 25; k) M. Funes Maldonado, F. Sehgelmeble, F. Bjarnemark, M. Svensson, J. Ahman, P.I. Arvidsson, Tetrahedron 2012, 68, 7456; I) S. R. K. Battula, G. V. Subbareddy, I. E. Chakravarthy, Tetrahedron Lett. 2014, 55, 517; m) E. Langkopf, A. Blum, Boehringer Ingelheim International GMBH, WO 2016/023789, 2016.
- [15] a) E. S. Levchenko, A. V. Kirsanov, Zh. Obshch. Khim. 1960, 30, 1553; b) E. S. Levchenko, I. E. Sheinkman, A. V. Kirsanov, Zh. Obshch. Khim. 1960, 30, 1941; c) W. Ried, J. Valentin, Justus Liebigs Ann. Chem. 1966, 699, 183; d) A. K. Roy, J. Am. Chem. Soc. 1993, 115, 2598.
- [16] See refs. [3] and [14j], and references therein.
- [17] J. Wen, H. Cheng, S. Dong, C. Bolm, Chem. Eur. J. 2016, 22, 5547.
- [18] J. A. Sirvent, D. Bierer, R. Webster, U. Lücking, Synthesis 2017, 49, 1024.
- [19] See ref. [4e], and references therein.
- [20] M. Zenzola, R. Doran, L. Degennaro, R. Luisi, J. Bull, Angew. Chem. Int. Ed. 2016, 55, 7203; Angew. Chem. 2016, 128, 7319.
- [21] For recent developments concerning the use of PhI(OAc)2 in the synthesis of sulfoximines, see also: a) A. Tota, M. Zenzola, S. J. Chawner, S. St. John-Campbell, C. Carlucci, G. Romanazzi, L. Degennaro, J. A. Bull, R. Luisi, Chem. Commun. 2017, 53, 348; b) J.-F. Lohier, T. Glachet, H. Marzag, A.-C. Gaumont, V. Reboul, Chem. Commun. 2017, 53, 2064; c) Y. Xie, B. Zhou, S. Zhou, S. Zhou, W. Wei, J. Liu, Y. Zhan, D. Cheng, M. Chen, Y. Li, B. Wang, X.-S. Xue, Z. Li, ChemistrySelect 2017, 2, 1620.
- [22] a) D. Leca, L. Fensterbank, E. Lacôte, M. Malacria, Org. Lett. 2002, 4, 4093; b) D. Leca, K. Song, M. Amatore, L. Fensterbank, E. Lacôte, M. Malacria, Chem. Eur. J. 2004, 10, 906; c) A. Felim, A. Toussaint, C. R. Phillips, D. Leca, A. Vagstad, L. Fensterbank, E. Lacôte, M. Malacria, Org. Lett. **2006**, *8*, 337.
- [23] a) K. K. Andersen, Tetrahedron Lett. 1962, 3, 93; b) J. R. Fulton, L. M. Kamara, S. C. Morton, G. J. Rowlands, Tetrahedron 2009, 65, 9134.
- [24] For the assay description, see: E. H. Kerns, L. Di, Drug-like Properties: Concepts, Structure Design and Methods, Elsevier, Burlington, 2008, pp. 353-356.
- [25] For the assay description, see: U. Lücking, P. Wasnaire, A. Scholz, P. Lienau, G. Siemeister, C. Stegmann, U. Boemer, K. Zheng, P. Gao, G. Chen, J. Xi (Bayer Pharma Aktiengesellschaft), WO 2015155197, 2015.
- [26] For the assay description, see: D. J. Minick, J. H. Frenz, M. A. Patrick, D. A. Brent, J. Med. Chem. 1988, 31, 1923.

Manuscript received: July 14, 2017

Accepted manuscript online: August 22, 2017 Version of record online: October 9, 2017