

# Biosynthesis, Characterization, and Biological Activities of Iron Nanoparticles using *Sesamum indicum* Seeds Extract

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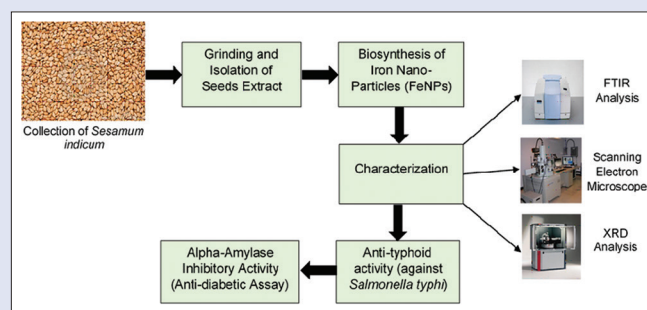
## ABSTRACT

**Background:** Iron nanoparticles (FeNPs) have got many biomedical and health applications because of biocompatible and nontoxic nature to humans. **Objective:** To synthesize the FeNPs using natural sources. **Materials and Methods:** In this study, simple and economical procedure was adopted for FeNPs synthesis. Sesame seeds were processed to obtain seed extract as a biological material for FeNPs production. FeNPs were characterized by Fourier transform infrared spectroscopy, X-ray diffraction, and scanning electron microscopic. **Results:** The average diameter of these FeNPs was 99 nm. These nanoparticles showed significant anti-typhoid activity (30 mm zone of inhibition) as compared to ciprofloxacin (32 mm) as standard. Furthermore, *in vitro* alpha-amylase inhibitory assay also showed moderate antidiabetic activity with more than 50% inhibition. **Conclusion:** This study would be helpful in understanding of nanoparticles synthesis from natural sources and ultimately will be used as potential alternative therapeutic agents.

**Key words:** Biological activities, characterization, iron nanoparticles, *Sesamum indicum*

## SUMMARY

- Iron nanoparticles (FeNPs) were synthesized by *Sesamum indicum* seeds
- FeNPs were characterized by scanning electron microscope with average diameter of 99 nm
- These FeNPs are effective against *Salmonella typhi*, a causative agent of typhoid
- These FeNPs can be used as antidiabetic agent.



**Abbreviations used:** FeNPs: Iron Nano Particles; SEM: Scanning Electron Microscopy; MIC: Minimum Inhibitory Concentration; *S. indicum*: *Sesamum Indicum*.

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## INTRODUCTION

With the advancement in material sciences, nanoparticles have to gain prime importance due to their unique physicochemical and biopharmaceutical properties and are being applied in biomedical, pharmaceutical, and health biotechnological fields. Basically, the approach is to design, characterize, produce, materials, devices, and systems by controlling the shape and size at the nanoscale.<sup>[1]</sup> Recently, this technology is used successfully in drug delivery, especially in targeted drug delivery, molecular imaging, biosensors, delivery of gene, and oligonucleotide, etc., and treatment of infectious diseases and cancer. Various nanoparticulate systems such as nanosuspensions, microemulsions, nanoparticles, nanogels, niosomes, and liposomes are the significant applications of nanotechnology.<sup>[2,3]</sup>

Iron nanoparticles (FeNPs) are more economical, physiological, compatible, safe as compared to gold, silver, and other nanoparticles. Clinical data on biodistribution, clearance, and biocompatibility of FeNPs ensure their safe clinical use.<sup>[4]</sup> These particles have received remarkable attention for its characterization due to its potential applications and hence many analytical methods available for its characterization.<sup>[5]</sup>

A variety of mechanical, physical, and chemical methods could be used for synthesis of FeNPs. However, these methods are stuffed with many disputed points such as generation of hazardous by-products, toxic solvents use, and consumption high energy.<sup>[6]</sup> One of the most

considered methods is the green synthesis of FeNPs, in this method, living organisms are used for synthesis of nanoparticles such as plants, algae, fungi, bacteria, and viruses.<sup>[6]</sup> Plant seems to be the best candidates as compared to other living organisms, for example, microorganisms. Plant extract solutions have the considerable capability to synthesize NPs.<sup>[7,8]</sup>

In this study, pharmacological active FeNPs were readily and efficiently produced using sesame (*Sesamum indicum*) as a reducing agent. Advantage of this procedure is no need of any corrosive, toxic chemical. Products obtained by this method are safe for human use as biomedical applications. This method is more preferable than other methods where high energy, temperature, pressure, and toxic or harmful chemicals required and other significant advantages of this method are easily scaled

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up for bulk synthesis, cost-effective, and environmental friendly.<sup>[9,10]</sup> The biological activities indicate outstanding pharmacological features of these FeNPs particles, therefore, increases its significance and value in biotechnological and pharmaceutical industry.

## MATERIALS AND METHODS

### Collection of plant seeds and biosynthesis of iron nanoparticles

In this study, the *S. indicum* seeds collected from local agriculture store. Seeds samples stored in an amber glass container after crushing and then used for extraction process. FeNPs were synthesized, characterized, and biologically analyzed.

### Preparation of seed extract

*S. indicum* seeds of 100 g were gently grinded in mortar and pestle and then soaked with 300 ml sodium hypochlorite (bleach) solution (30%) for 7 min for sterilization; after that, bleach removed and sample rinsed with sterile distilled water for three times. Seeds sample soaked with 300 ml of 70% ethanol for 2 min and this ethanol was removed. Seed sample further six times rinsed with sterile distilled water. Disinfected seed sample added in sterile distilled water (proportion volume 1:2) and incubated at 25°C for 7 days in amber glass bottle. Now, after incubation, prepared seeds extract filtered by No. 40 Whatman filter papers and then stored in amber glass bottle in refrigerator.<sup>[11]</sup>

### Synthesis of iron nanoparticles

Equal volume ratio of 0.1 M FeCl<sub>3</sub> solution and prepared seed extract (1:1) was added in a beaker by continuous stirring for 60 min and kept mixture for half an hour at room temperature and finally centrifuged for 8 min at 12,000 rpm. Supernatant was removed and precipitates collected.<sup>[5]</sup>

### Fourier transform infrared spectroscopy analysis

Sample was gently ground and diluted in nonabsorbent KBr matrices, and Fourier transform infrared spectroscopy (FTIR) spectrum of FeNPs was recorded over the range of 450–4000 cm<sup>-1</sup> by FTIR spectrophotometer (Alpha-P Bruker, Germany).<sup>[12]</sup>

### Scanning electron microscopic analysis

Scanning electron microscopic (SEM) analysis of FeNPs was carried out using Philips scanning microscope (Philips XL-30 FEG-SEM) type instrument. FeNPs were smeared onto carbon adhesive tapes which supported on metallic disks, and then their images were recorded at different magnifications.<sup>[9]</sup>

### X-ray diffraction analysis

The X-ray diffraction (XRD) analysis was conducted with a Philips X'Pert-Pro (PANalytical X'Pert Diffractometer) instrument. The source comprised Cu K $\alpha$  radiations (wavelength ( $\lambda$ ) = 1.54 Å). FeNPs were placed in glass holder and then scanned from 20 to 80° within the 2 $\theta$  range.<sup>[9,13]</sup>

### Anti-typhoid activity by agar well diffusion method

The synthesized FeNPs were tested against *Salmonella typhi*, a causative agent of typhoid by standard well diffusion assay.

### Preparation of nutrient agar Petri dishes and nutrient broth

We prepared nutrient agar Petri plates by dissolving 14 g agar in 500 ml distilled water and autoclaved.<sup>[14]</sup> Similarly, nutrient

broth (0.45 g) was prepared by taking 0.45 g in 50 ml distilled water and sterilized.

### Preparation of McFarland solution

We added 0.5 ml of BaCl<sub>2</sub> (1.175%) dropwise in 85 ml of H<sub>2</sub>SO<sub>4</sub> (1%) with continuous stirring and volume was adjusted to 100 ml with 1% H<sub>2</sub>SO<sub>4</sub>. This solution was used to analyze the transparency level of diluted inoculum.<sup>[15]</sup>

### Preparation of inoculum

We prepared inoculum culturing *S. typhi* in nutrient broth (25 ml) and incubated at 37°C for 24 h. Sterilized normal saline solution was added into inoculating test tube until transparency level matches McFarland solution.<sup>[16]</sup>

### Preparation of serial dilutions of iron nanoparticles for minimum inhibitory concentration

Stock sample (1000 µg/ml) was serially diluted: 500, 250, 125, and 62.5 µg/ml for evaluation of minimum inhibitory concentration (MIC) of FeNPs.

### Anti-typhoid assay

The inoculum of *S. typhi* was spread onto the surface of sterilized nutrient agar Petri plates using sterile cotton swab; the wells were formed with the help of 6 mm stainless steel borer, and then sample and standard ciprofloxacin were loaded into respective wells. These Petri plates were incubated at 37°C for 24 h and zone of inhibition were measured.<sup>[17]</sup>

### In vitro alpha-amylase inhibitory assay

Starch (1.2 ml) in the phosphate buffer having pH 6.9 also containing 6.7 mM NaCl was added to 600 µl of (10, 20, 40, 60, 80, 100 µg/ml) the synthesized FeNPs. A volume of 60 µl of porcine pancreatic amylase was also added in it, and hence, reaction started and incubated at 37°C. In 600 µl of above mixture, 300 µl 3,5-dinitrosalicylic acid was added and further kept in boiling water bath for 15 min. Furthermore, reaction mixture diluted by adding 2.7 ml water and then finally absorbance was recorded at 540 nm.

Then, blank tubes were also prepared by adding 600 µl distilled water instead of enzyme solution for each concentration. Finally, control was prepared without FeNPs and it representing 100% enzyme activity. This experiment was repeated for three times with the same protocol.<sup>[18]</sup>

### Method for calculation of the alpha-amylase inhibitory activity

The alpha-amylase inhibitory activity was calculated using the given formula:

$$\text{The } \alpha\text{-amylase inhibitory activity} = \frac{(Ac+) - (Ac-) - (As - Ab)}{(Ac+) - (Ac-)} \times 100$$

where

Ac+ = 100% enzyme activity absorbance (only solvent with enzyme)

Ac- = 0% enzyme activity absorbance (only solvent without enzyme)

As = a test sample the absorbance (with enzyme)

Ab = a blank absorbance (a test sample without enzyme).

## RESULTS

### Biosynthesis of iron nanoparticles

Biological active FeNPs were successfully produced using *S. indicum* seeds.

### Fourier transform infrared spectroscopy analysis

FTIR spectroscopy used for identification of the functional groups based on spectral peaks. Figure 1 shows FeNPs spectra and absorbance bands have been observed in the region of 3419.28, 2110.30, 1645.46, 1011.64, 951.82, and 788.40  $\text{cm}^{-1}$  which confirmed O-H group (strong band, broad intensity while stretching), alkyne group (stretching vibration), amide (C = O, strong band, stretching), ether (C-O, stretching), alkene (=C-H, bending), alkyl halide (stretching and strong band), respectively.

### Scanning electron microscopy analysis

SEM micrographs of biosynthesized FeNPs have been shown at magnifications of 1.01 kx, 5.0 kx, 10.0 kx, 25.0 kx, and 50.0 kx [Figure 2a] and the size of FeNPs were found 99 nm [Figure 2b] and round and well monodisperse.

### X-ray diffraction

We observed various peaks at different angles ( $2\theta$ ) 24.6°, 34.8°, 41.9°, 46.1°, 52.5°, 61.8° [Figure 2c]. These peaks exhibited the intensity of FeNPs and the size of these nanoparticles was obtained using the Debye-Scherrer formula:  $D = K\lambda/\beta \text{Cos}\theta$ , where  $D$  is the mean diameter of nanoparticles,  $\beta$  = the full width at half-maximum value of XRD diffraction line,  $\lambda$  = the wavelength of X-ray radiation source 0.15405 nm,  $\theta$  = the half diffraction angle-Bragg angle,  $K$  = the Scherrer constant with the value 0.9.

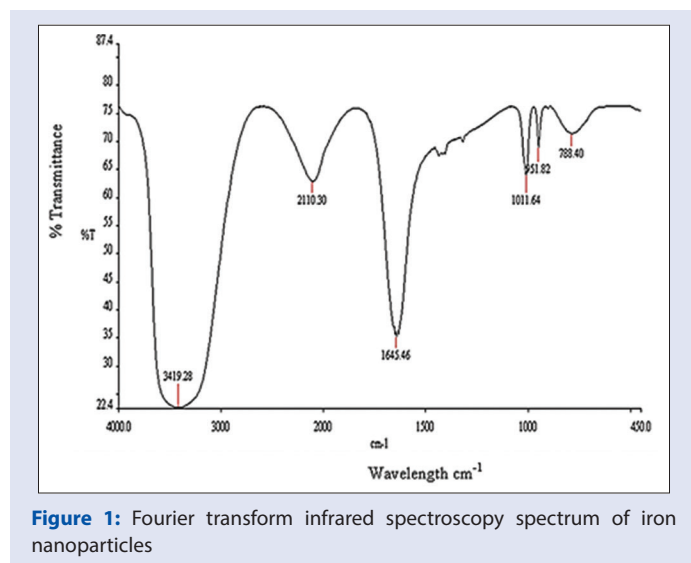


Figure 1: Fourier transform infrared spectroscopy spectrum of iron nanoparticles

### Biological activities of iron nanoparticles

#### Anti-typhoid activity of iron nanoparticles

The synthesized FeNPs were tested against *S. typhi*, a causative agent of typhoid by standard well-diffusion assay. The antibacterial activity of FeNPs showed Figure 3a significant zone of inhibition against *S. typhi*. The ciprofloxacin (standard) showed the clear zone of inhibition, i.e., 32 mm while FeNPs exhibited 30 mm.

The result of MIC of the FeNPs is shown in Figure 3b. 1000  $\mu\text{g/ml}$  (stock) exhibited 26 mm zone of inhibition and 500  $\mu\text{g/ml}$  made 12 mm zone indicating it as the MIC of FeNPs.

#### In vitro alpha-amylase inhibitory assay

Acarbose used as a standard at a concentration of (10–100  $\mu\text{g/ml}$ ) showed  $\alpha$ -amylase inhibitory activity from 57.93  $\pm$  0.31% to 94.62  $\pm$  0.51% with  $\text{IC}_{50}$  value 1.99  $\mu\text{g/ml}$  [Table 1]. FeNPs at a concentration of (10–100  $\mu\text{g/ml}$ ) exhibited  $\alpha$ -amylase inhibition activity in a dose-dependent manner. The FeNPs expressed the highest the inhibitory activity, i.e., 64.39  $\pm$  0.52% with  $\text{IC}_{50}$  value of 21.26  $\mu\text{g/ml}$ .

### DISCUSSION

The methods of preparing FeNPs by chemical means are not satisfactory because these methods are fraught with various problems such as high-energy consumption, toxic chemicals, and unwanted by-products.<sup>[6]</sup> Mercury-based methods used for FeNPs synthesis, but due to toxic mercury vapors, these methods replaced with other methods, i.e., organic solvent-based methods.<sup>[19]</sup>

Table 1: Alpha-amylase inhibitory effects of iron nanoparticles in comparison with the standard drug acarbose

Test samples	Concentrations ( $\mu\text{g/ml}$ )	Percentage of inhibition	$\text{IC}_{50}$ value ( $\mu\text{g/ml}$ )
Iron nanoparticles	10	0	21.26 $\pm$ 0.05*
	20	0	
	40	0	
	60	0	
	80	58.66 $\pm$ 0.64	
	100	64.39 $\pm$ 0.52	
Acarbose (standard)	10	57.93 $\pm$ 0.31	1.99 $\pm$ 0.03
	20	67.63 $\pm$ 0.56	
	40	75.81 $\pm$ 0.72	
	60	83.52 $\pm$ 0.76	
	80	87.31 $\pm$ 0.47	
	100	94.62 $\pm$ 0.51	

\* $P < 0.001$ ;  $n = 3$

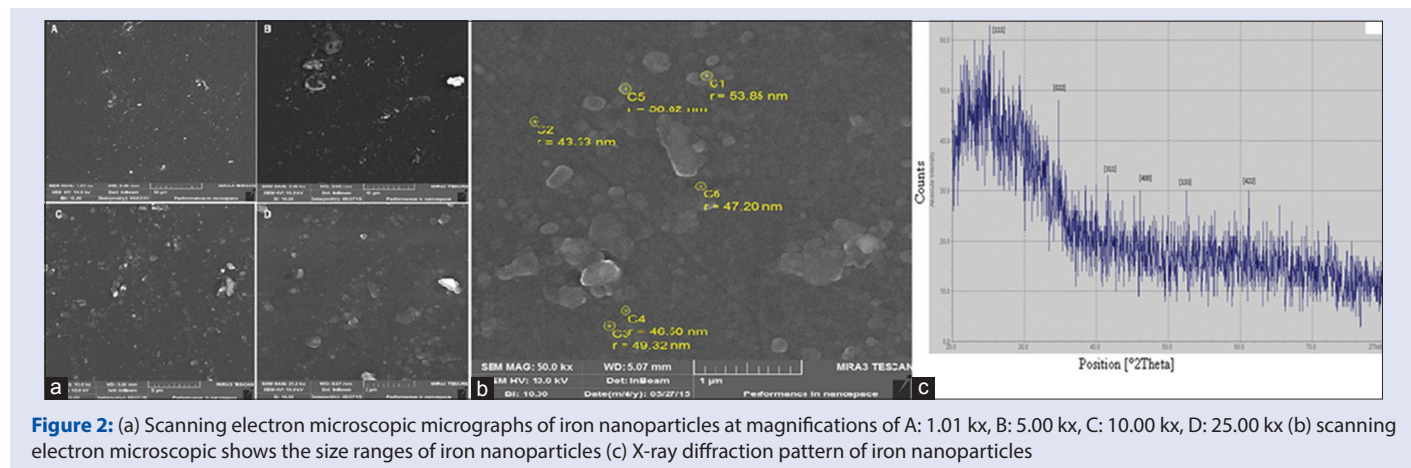
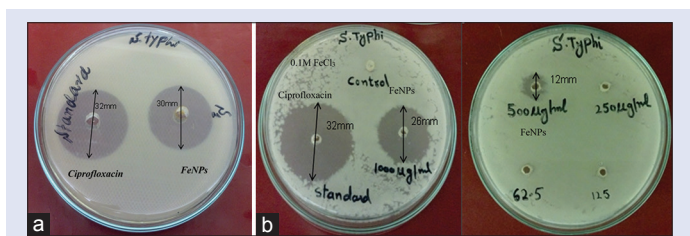


Figure 2: (a) Scanning electron microscopic micrographs of iron nanoparticles at magnifications of A: 1.01 kx, B: 5.0 kx, C: 10.0 kx, D: 25.0 kx (b) scanning electron microscopic shows the size ranges of iron nanoparticles (c) X-ray diffraction pattern of iron nanoparticles



**Figure 3:** (a) The zone of inhibition of iron nanoparticles in comparison with standard drug ciprofloxacin ( $s_1$  = iron nanoparticles, standard = ciprofloxacin, *S. typhi* = *Salmonella typhi*) (b) The minimum inhibitory concentration analysis (consecutively in two Petri plates) of serially diluted iron nanoparticles

Therefore, the safety and health concerns are associated with this reduction approach.<sup>[9,20]</sup> Thermal decomposition of iron pentacarbonyl method for synthesis of FeNPs is the complicated reaction. Surfactant is required for this method that increases the cost of synthetic procedure. Heat and an efficient system are used to remove by-products. Many microemulsion procedures can be employed for the synthesis of FeNPs, but it is not usually preferred due to oxidation problems.

In this study, we used seeds extract as biological material for FeNPs synthesis to avoid all these chemicals hazardous effects reported previously. Seeds extract was used as reducing agent as well as stabilizing agent. We adopted a single step, easy, and rapid procedure for FeNPs synthesis that is the most considered method and simple approach. In this procedure, there is no need of any corrosive, toxic, flammable chemical as in the case of chemical synthetic procedures. Products obtained by this method are safe for human use as biomedical applications because there is no chance of toxic chemical absorption on their surfaces. More importantly, it is inexpensive and environmentally benign, nontoxic products treatment, and size-controlled NPs at mild conditions. The nanoparticles were characterized using XRD technology which is an effective way to confirm the crystal structure of particles. XRD and SEM micrographs show morphological details of FeNPs, revealing spherical shape with average size of 99 nm. The SEM size of FeNPs (99 nm) showed variations in peaks during diffraction analysis at different angles ( $2\theta$ ) 24.6°, 34.8°, 41.9°, 46.1°, 52.5°, 61.8°. The size of these nanoparticles was obtained using the Debye-Scherrer formula. The XRD profile and electron scanning pattern show that FeNPs with body-centered cubic structure are formed. Therefore, this study helps to understand the relationship between the surface topography and functional characteristics. We analyzed and found these nanoparticles biological active against typhoid.

In spite of many anti-typhoid products are available in market such as cephalosporin, fluoroquinolone, azithromycin, and chloramphenicol, these antibiotics have bacterial resistance and costly. We used ciprofloxacin as standard drug, and our results proved that active FeNPs are effective against typhoid. Our results showed significant anti-typhoid activity with 32 mm of zone of inhibition against *S. typhi*. Metabolic disorder such as diabetes mellitus is another major problem in the world. During antidiabetic analysis, these particles showed average to moderate inhibition for  $\alpha$ -amylase enzyme.

## CONCLUSION

FeNPs are successfully biosynthesized by rapid, simple, and cost-effective method using the sesame (*S. indicum*) seeds extract. The average diameter

of FeNPs was 99 nm. Particles were spherical and well monodisperse. These biological active particles are effective for typhoid and diabetes. Such green methods are helpful for the synthesis of safe and nontoxic nanoparticles that can play a potential in biomedical sciences.

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Nil.

## Conflicts of interest

There are no conflicts of interest.

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