# Neurorestoration Induced by Mesenchymal Stem Cells: Potential Therapeutic Mechanisms for Clinical Trials

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Stem cells are emerging as therapeutic candidates in a variety of diseases because of their multipotent capacities. Among these, mesenchymal stem cells (MSCs) derived from bone marrow, umbilical cord blood or adipose tissue, comprise a population of cells that exhibit extensive proliferative potential and retain the ability to differentiate into multiple tissue-specific lineage cells including osteoblasts, chondrocytes, and adipocytes. MSCs have also been shown to enhance neurological recovery, although the therapeutic effects seem to be derived from an indirect paracrine effect rather than direct cell replacement. MSCs secrete neurotrophic factors, promote endogenous neurogenesis and angiogenesis, encourage synaptic connection and remyelination of damaged axons, decrease apoptosis, and regulate inflammation primarily through paracrine actions. Accordingly, MSCs may prevail as a promising cell source for cell-based therapy in neurological diseases.

Key Words: Mesenchymal stem cells, paracrine effect, cell-based therapy

# INTRODUCTION

Stem cells are emerging as therapeutic candidates in a variety of disease because of their multipotent capacities. Embryonic stem cells are pluripotent and can differentiate into all specialized cell types derived from the three embryonic germ layers. Nevertheless, both ethical and technical considerations limit the clinical availability of these cells. Other potential cell sources, especially for central nervous system (CNS) repair, include fetal neural stem cells (NSCs) and neural precursor cells (NPCs). NSCs can be expanded over multiple passages and do not require the recapitulation of early developmental signals to induce neuroectodermal commitment, as they are already neuralized and committed to a CNS cell fate.<sup>1</sup> However, transplantation of fetal NSCs into the adult brain encompasses numerous ethical and scientific hurdles.<sup>1-3</sup> Because of their plastic ability to survive as undifferentiated cells in ectopic perivascular niches,<sup>4</sup> NPCs have been tested in animal models of neurological diseases. These cells also release paracrine factors that foster survival and proliferation of endogenous neural progenitor cells. However, the application of terminal differentiation of NPCs into neural-lineage cells to

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This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/ licenses/by-nc/3.0) which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited. replace damaged cells remains controversial.5

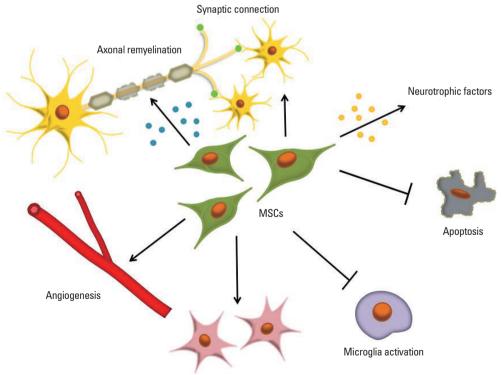
In contrast, adult stem cells, responsible for maintaining the homeostasis of a specific tissue with fewer ethical problems. One of the most extensively studied populations of multipotent adult stem cells is mesenchymal stem cells (MSCs), which are derived from bone marrow (BM). They can also be isolated from other tissues such as umbilical cord blood.<sup>6</sup> synovium.<sup>7</sup> periosteum.<sup>8</sup> peripheral blood.<sup>9</sup> adipose tissue,10 skeletal muscle,11 and placental tissue.12 MSCs are an excellent candidate for cell therapy because they are easily accessible; can be easily isolated and expanded rapidly in vitro;13 are multipotent;14,15 involve minimal loss of potency;<sup>16,17</sup> form supportive stroma for hematopoiesis and support stem cell engraftment;<sup>18</sup> may not require immune suppression;<sup>19,20</sup> seem to be largely immunologically inert, paving the way for allogeneic transplantation;<sup>21</sup> secrete numerous trophic factors that modulate inflammation and apoptosis.22 The large body of work that has accumulated since the discovery of human MSCs has convincingly shown that MSCs from diverse sources retain the ability to differentiate into the mesodermal lineage cells including osteoblasts, chondrocytes and adipocytes.23,24 They also exhibit the ability to differentiate into neurons-like cells,<sup>25</sup> myocytes and skeletal muscle,<sup>23</sup> although there is a lack of definitive evidence as to the functionality of these differentiated cells.<sup>26,27</sup> Nevertheless, MSCs have considerably contributed to tissue repair in myocardial infarction (MI),<sup>28</sup> stroke,<sup>29,30</sup> meniscus injury,<sup>31</sup> and limb ischemia.<sup>32</sup>

In this review, we will discuss the therapeutic mechanisms of MSCs for neurorestoration and neural regeneration. Thereafter, we will review the published reports of clinical trials for a variety of neurological diseases including stroke, traumatic brain injury, spinal cord injury, Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), and multiple sclerosis (MS).

# THERAPEUTIC MECHANISMS OF MSCs FOR NEURORESTORATION

#### Secretion of neurotrophic factors

MSCs secrete a variety of cytokines and growth factors that promote endogenous neuronal growth, neurogenesis and angiogenesis, encourage synaptic connection and remyelination of damaged axons, decrease apoptosis, and regulate inflammation primarily through paracrine actions (Fig. 1).



Neurogenesis and astroglial activation

Fig. 1. Potential therapeutic mechanisms of neurorestoration using mesenchymal stem cells. MSCs secrete a variety of neurotrophic factors that promote endogenous neuronal growth, induce angiogenesis, neurogenesis and astroglial activation, encourage synaptic connection and axonal remyelination, decrease apoptosis, and regulate microglial activation primarily through paracrine actions. MSCs, mesenchymal stem cells. Human MSCs are known to secrete neurotrophic factors including brain-derived neurotrophic factor (BDNF), ciliary neurotrophic factor, glial cell line-derived neurotrophic factor (GDNF), and nerve growth factor (NGF). After direct transplantation in an animal model of stroke, human MSCs were shown to integrate into host brain, survive, differentiate into neurons and astrocytes, and induce neurobehavioral improvement.<sup>33</sup> BM-derived MSCs can secrete various trophic factors, the secretion of which is enhanced under postischemic conditions.<sup>34,35</sup> In our previous study using a rat model of spinal cord injury, neurally induced cells derived from umbilical cord blood also exhibited better functional recovery *in vivo* and secreted more NGFs *in vitro*.<sup>36</sup>

# Induction of neurogenesis and astroglial activation

MSCs induce the proliferation of endogenous neural stem/ progenitor cells in the subventricular zone (SVZ) and are critical to the survival of newborn cells.<sup>37</sup> They have been shown to be directly involved in neural differentiation after engraftment into damaged tissue and migrate to the CNS to a limited extent.<sup>38</sup> Of particular note, genetically modified MSCs expressing Neurogenin1, a proneuronal gene that directs neural differentiation, increased the therapeutic effects of MSCs in ischemic brain.<sup>39</sup> In addition, MSCs promote the plasticity of damaged neurons and activate astroglial cells to secrete neurotrophins such as BDNF, GDNF and NGF.<sup>34</sup> In an animal model of stroke, intravenous transplantation of BM stromal cells improved functional outcomes by promoting endogenous repair.<sup>29</sup>

## Axonal sprouting and synaptic connection

Previous study suggested that extracellular matrix components derived from MSCs can enhance nervous system repair.<sup>40</sup> For example, fibronectin prominently performs essential roles in neuronal survival, axonal sprouting and synaptogenesis following cerebral ischemia.<sup>41</sup> Moreover, extracellular matrix molecules and cell adhesion molecules such as integrin, cadherin, and selectin can promote axonal growth and regeneration.<sup>42</sup>

#### Anti-apoptotic effect

Reportedly intravenous transplantation of MSCs reduced apoptotic cells stained with terminal deoxynucleotidyl transferase-mediated dUTP nick-end labeling in an animal model of middle cerebral artery occlusion.<sup>29</sup> This anti-apoptotic effect, together with the previously described capacity to release neurotrophic molecules, may well explain the remarkable functional recovery obtained with the administration of MSCs in experimental models of stroke as well as spinal cord injury.<sup>43,44</sup>

#### Immunomodulatory effect

MSCs were shown to exert immunomodulatory properties *in vitro*.<sup>45</sup> These features were exploited by researchers in the treatment models of MS and experimental autoimmune encephalomyelitis.<sup>46,47</sup> In PD, MSCs act as neuroprotectors via an anti-inflammatory response to regulate the activity of microglia and to protect dopaminergic neurons.<sup>48,49</sup> In ischemic brain, MSCs were also useful as an immunomodulator.<sup>50</sup> MSCs reduced the numbers of Iba-1<sup>+</sup> and ED1<sup>+</sup> inflammatory cells.<sup>39</sup> In our previous study, intravenously transplanted MSCs did not only decrease the level of pro-inflammatory cytokine IL-1 $\beta$  and the proportion of activated microglia, but also increased the level of anti-inflammatory cytokine IL-10, potentially suggesting that early immunomodulation by MSCs was an underlying mechanism of functional recovery in spinal cord injured rats.<sup>51</sup>

# Induction of angiogenesis

MSCs secrete a number of growth factors and cytokines, which normally support the proliferation of hematopoietic stem/progenitor cells.<sup>22</sup> In experimental models of cardiovascular diseases such as MI and limb ischemia, the secretion of multiple angiogenic cytokines such as hepatocyte growth factor, basic fibroblast growth factor, insulin-like growth factor 1 and vascular endothelial growth factor was induced from MSCs.<sup>52-56</sup> Existing evidence suggests that these cells can ameliorate ischemic tissue injury, produce appropriate cytokine milieu to promote angiogenesis, and possibly differentiate into endothelial cells.<sup>54,55,57</sup> The evidence seems to point toward the theory that a complex set of trophic factors secreted by MSCs significantly contributes to injury repair *in vivo* by stimulating angiogenesis.

#### Host cell effects stimulated by MSC transplantation

MSCs are believed to secrete neurotrophic factors, immunomodulatory cytokines, pro-angiogenic factors, extracellular matrix molecules and so on. Thus, the therapeutic effects of transplanted MSCs seem to be derived from paracrine effects rather than direct cell replacement. It is conceivable that MSCs or host brain cells stimulated by grafts may produce such proteins to induce functional recovery and reorganization.<sup>58</sup> MSC-induced secretion of beneficial cytokines by host cells in subjects with neurological diseases such as stroke<sup>29</sup> as well as spinal cord injury<sup>33,44,59</sup> would be a more plausible mechanism given that activation of recipient host cells after cell transplantation has been previously described in other disease models such as MI and ischemic vascular disease.<sup>60,61</sup>

# CLINICAL TRIALS IN NEUROLOGICAL DISEASES

Several clinical reports on MSC-based treatments have been published in the past decade, and evoked great excitement for therapeutic candidates for several diseases.<sup>62</sup> As early as the 1990s, cultured MSCs have already been supplemented to reduce acute and chronic graft-versus-host disease among patients receiving allogenic hematopoietic stem cell transplantation, as well as to ameliorate clinical symptoms in osteogenesis imperfecta and glycogen storage disease.<sup>62-64</sup> Currently, the effective therapeutic benefits of MSCs have been supported by increasing numbers of clinical trials on various disorders such as MI, cancer, diabetes mellitus, and Crohn's disease.<sup>65-68</sup>

We will discuss the role of MSCs in neurological diseases, spanning the clinical trials on stroke, spinal cord injury, PD, ALS, and MS. A summary of the cell source, route of delivery, number of patients, study design, dose of transplant, outcome measures and main results for each therapeutic application of MSCs is provided in Table 1.

# MSC in stroke and traumatic brain injury

Neuronal and astroglial damages can occur in cerebrovascular disease resulting from the blockage of blood flow in selected brain areas, leading to motor, sensory and cognitive dysfunctions.<sup>3</sup> It has been shown that stroke-induced endogenous neurogenesis and migration of neural stem or progenitor cells into regions of ischemic damage occurs in humans, but the extent to which neurogenesis is able to replace lost neurons or contributes to functional improvement in stroke patients is largely limited.<sup>3,69,70</sup> The limited therapeutic efficacy of endogenous repair processes has encouraged clinicians to incorporate MSCs or BM-derived cells in restorative strategies.

Clinical trials for MSC transplantation to treat stroke and traumatic brain injury are currently ongoing. In patients with middle cerebral artery infarction, the use of autologous MSCs derived from BM has indicated no safety concerns for death, stroke recurrence, or serious adverse events up to 1 year, and trends towards increased functional recovery.<sup>71</sup> This group also reported as a long-term follow-up study for 5 years no serious adverse effects following MSC treatment.<sup>72</sup> Direct administration of MSCs to an injured region following traumatic brain injury has also been performed without adverse events. Briefly, seven patients each received up to 10<sup>9</sup> expanded MSCs as part of a cranial repair operation. The patients were followed up for six months and demonstrated significant improvements in neurological function.<sup>73,74</sup>

# MSC in spinal cord injury

Depending on the severity and location of injury, patients present with a varying range of functional impairments, arising from both damage to the local circuitry of the spinal cord and disruption of the ascending and descending fiber tracts.<sup>75,76</sup> All groups who have tested the safety of the transplantation of BM-derived mononuclear cells and stromal cells, or adipose tissue-derived MSCs in patients with spinal cord injury indicate that administration of these cells does not cause any serious adverse effects.<sup>77,80</sup> Geffner, et al.<sup>77</sup> investigated the improvement in quality of life and bladder function without pain or tumor up to 2 years. Syková, et al.<sup>78</sup> reported that five patients who received cells intra-arterially showed improvement up to 1 year.

## MSC in parkinson's disease

PD is a progressive neurodegenerative disease whose dopaminergic neurons selectively degenerate in the substantia nigra. Although a variety of drugs such as L-dopa are available, they only remain effective for a certain period in most patients. The limitation of pharmacologic agents increases the need for cell-based therapy as a restorative strategy. In a study recently reported by Li, et al.,81 two subjects with PD who underwent transplantation of fetal mesencephalic dopaminergic neurons, which had survived for over 10 years, but later developed  $\alpha$ -synuclein-positive Lewy bodies in the engrafted donor neurons, suggesting that the disease can propagate from host to graft cells. On the other hand, when autologous BM-derived MSCs were transplanted into the SVZ by stereotaxic surgery, the results suggested the treatment to be safe, and no serious adverse events occurred after transplantation in PD.82 Additionally, when patients with multiple system atrophy (MSA) were treated with MSCs, greater improvement was noted on the unified MSA rating scale than in untreated control patients, and no delayed adverse effects related to MSC infusion occurred during the

	Treatment 5; control 25	Phase 1-2 randomized,				
I ntic brain I I I I I I I I I I I I I I I I I I I		controlled safety, efficacy	$5 \times 10^7$ cells in two doses	Safety: no death, stroke recurrence, or serious adverse events after 1 year Efficacy: trend towards increased functional recovery	Middle cerebral artery infarcts	Bang, et al. $^{71}$
	Treatment 16; control 36	Phase 1-2 randomized, controlled safety, efficacy	5×10 <sup>7</sup> cells in two doses	Safety: no death, stroke recurrence, or serious adverse events after 5 years Efficacy: trend towards increased functional recovery	Middle cerebral artery infarcts	Lee JS, et al. $72$
	Treatment 7	Phase 1, open, safety	$1 \text{ st } 10^7  10^9 \text{ cells}$ 2nd $10^8  10^{10} \text{ cells}$	Safety: no death, cell-related serious AE, no toxicity related to MSC within 6 months		Zhang, et al. <sup>73</sup>
	Treatment 30	Phase 1-2a, open, safety, efficacy	1×10 <sup>6</sup> cells/kg	Safety: no serious adverse events Efficacy: no change in ASIA scale; improvement in Barthel's index in thoracic injury occurred within 6 months	Patients with complete SCI at cervical or thoracic level	Pal, et al. <sup>79</sup>
S II .	Treatment 8	Phase 1, open, safety	$4 \times 10^8$ cells	No serious adverse events	Adipose tissue	Ra, et al. <sup>80</sup>
	Treatment 8 (4 acute, 4 chronic)	Phase 1, open, safety	4×10 <sup>8</sup> mononuclear cells	No tumors or pain, improvement in quality of life and bladder function		Geffner, et al. $^{77}$
	Treatment 20 (7 acute, 13 chronic)	Phase 1, open, safety	104±55.3×10 <sup>8</sup> mononuclear cells	No complications up to 1 year, 5 patients treated intra-arterially showed improvement	Only 1 chronic SCI reported improvement	Syková, et al. <sup>78</sup>
Parkinson's disease Intra-ventricle	Treatment 7	Phase 1, open, safety	1×10 <sup>6</sup> cells/kg	Safe and no serious adverse events for 12-36 months.	Feasibility study	Venkataramana, et al. <sup>82</sup>
Intra-spinal cord	Treatment 10	Phase 1, open, safety	11.4-120×10 <sup>6</sup> cells	Pain $(n=7)$ , localized sensory impairment $(n=5)$ , localized tingling sensation $(n=1)$	Feasibility study, two-five injections	Mazzini, et al. <sup>87</sup>
Amyotrophic lateral Intra-spinal cord sclerosis	Treatment 9	Phase 1, open, safety	$57 \times 10^6$ cells	Transient pain $(n=4)$ , transient sensory disturbances $(n=6)$		Mazzini, et al. <sup>86</sup>
Intrathecal, intrathecal plus intravenous	Treatment 19	Phase 1-2a, open, safety, efficacy	23.4-54.7×10 <sup>6</sup> cells	Safety: no major adverse events Efficacy: neurological disorders unchanged during 6 months	Fever (n=11), headache (n=5), dyspnea (n=1)	Karussis, et al. <sup>85</sup>
Intrathecal	Treatment 10	Phase 1-2a, open, safety, efficacy	8.73×10 <sup>6</sup> cells	Safety: iatrogenic meningitis (n=2), headache (n=9) Efficacy: EDSS unchanged (n=4), worsened (n=5), improved (n=1), MRI showed no change (n=7), increased lesion (n=2), decreased lesion (n=1)	Progressive MS with baseline EDSS ⊲6.0 mean follow- up of 19 months	Mohyeddin, et al. <sup>88</sup>
Multiple sclerosis (MS) Intrathecal	Treatment 10	Phase 1-2a, open, safety, efficacy	32-100×10 <sup>6</sup> cells	Safety: encephalopathy, seizure Efficacy: clinical improvement (n=6), worsening of MRI (n= 2)	EDSS 4.0-7.5	Yamout, et al. <sup>89</sup>
Intrathecal, intrathecal plus intravenous	Treatment 15	Phase 1-2a, open, safety, efficacy	24.5-63.2×10 <sup>6</sup> cells	Safety: fever (n=10), headache (n=10), aseptic meningitis (n=1), no serious adverse events Efficaey: reduction of EDSS	Active MS that did not respond to treatments	Karussis, et al. <sup>85</sup>

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# MSCs Induce Neurorestoration

12-month study period.83

# MSC in amyotrophic lateral sclerosis

ALS involves a pathology that causes a selective loss of motor neurons leading to a progressive decline in muscle function and poor prognosis. When Nagano, et al. completed a small double-blind clinical trial to assess the effect of intrathecal administration of IGF-1 on disease progression in nine patients with ALS, the high-dose treatment slowed the decline of motor functions, but not bulbar function or vital capacity.<sup>84</sup> On the other hand, both intravenous and intrathecal administration of autologous MSCs were well tolerated, with some preliminary evidence of efficacy in patients with ALS.<sup>85-87</sup> However, large controlled clinical studies are needed to assess possibility for this therapeutic strategy.

# MSC in multiple sclerosis

Most phase 1 studies for the safety of MSCs have been conducted in MS.<sup>85,88,89</sup> As a result, Mohyeddin, et al.<sup>88</sup> reported iatrogenic meningitis and headache; Yamout, et al.<sup>89</sup> reported transient encephalopathy and seizure; and Karussis, et al.<sup>85</sup> reported fever, headache and aseptic meningitis. Although serious adverse events related with cell transplantation are likely to be extremely uncommon in MS, the therapeutic efficacy in regards to clinical improvement remains controversial.<sup>85,88,89</sup>

# CONCLUSIONS

Experimental evidence in preclinical models of neurological diseases suggests that MSCs are a promising candidate for achieving neural repair and protection. However, current data do not support the possibility that most of the reported effects occur as a result of direct cell replacement. Instead, indirect paracrine mechanisms, including a potent anti-inflammatory capacity, the release of anti-apoptotic and neurotrophic factors, and the ability to induce proliferation of local neural stem/progenitor cells, are likely to promote neurorestoration. Despite tremendous advancements, major unresolved issues concerning therapeutic application still exist. Especially, transplanted MSCs suffer from poor survival and engraftment into host tissue. Further studies are necessary to evaluate in depth the efficacy and safety of MSC-based therapy and whether such treatment would involve a high benefit-to-risk ratio in neurological diseases.

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# REFERENCES

- 1. Gogel S, Gubernator M, Minger SL. Progress and prospects: stem cells and neurological diseases. Gene Ther 2011;18:1-6.
- Lovell-Badge R. The regulation of human embryo and stem-cell research in the United Kingdom. Nat Rev Mol Cell Biol 2008;9: 998-1003.
- Mathews DJ, Sugarman J, Bok H, Blass DM, Coyle JT, Duggan P, et al. Cell-based interventions for neurologic conditions: ethical challenges for early human trials. Neurology 2008;71:288-93.
- Uccelli A, Laroni A, Freedman MS. Mesenchymal stem cells for the treatment of multiple sclerosis and other neurological diseases. Lancet Neurol 2011;10:649-56.
- 5. Martino G, Pluchino S. The therapeutic potential of neural stem cells. Nat Rev Neurosci 2006;7:395-406.
- Erices AA, Allers CI, Conget PA, Rojas CV, Minguell JJ. Human cord blood-derived mesenchymal stem cells home and survive in the marrow of immunodeficient mice after systemic infusion. Cell Transplant 2003;12:555-61.
- De Bari C, Dell'Accio F, Tylzanowski P, Luyten FP. Multipotent mesenchymal stem cells from adult human synovial membrane. Arthritis Rheum 2001;44:1928-42.
- Fukumoto T, Sperling JW, Sanyal A, Fitzsimmons JS, Reinholz GG, Conover CA, et al. Combined effects of insulin-like growth factor-1 and transforming growth factor-beta1 on periosteal mesenchymal cells during chondrogenesis in vitro. Osteoarthritis Cartilage 2003;11:55-64.
- Villaron EM, Almeida J, López-Holgado N, Alcoceba M, Sánchez-Abarca LI, Sanchez-Guijo FM, et al. Mesenchymal stem cells are present in peripheral blood and can engraft after allogeneic hematopoietic stem cell transplantation. Haematologica 2004;89:1421-7.
- Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, Katz AJ, et al. Multilineage cells from human adipose tissue: implications for cell-based therapies. Tissue Eng 2001;7:211-28.
- 11. Cao B, Zheng B, Jankowski RJ, Kimura S, Ikezawa M, Deasy B, et al. Muscle stem cells differentiate into haematopoietic lineages but retain myogenic potential. Nat Cell Biol 2003;5:640-6.
- Parolini O, Alviano F, Bagnara GP, Bilic G, Bühring HJ, Evangelista M, et al. Concise review: isolation and characterization of cells from human term placenta: outcome of the first international Workshop on Placenta Derived Stem Cells. Stem Cells 2008;26: 300-11.
- 13. Sekiya I, Larson BL, Smith JR, Pochampally R, Cui JG, Prockop DJ. Expansion of human adult stem cells from bone marrow stroma: conditions that maximize the yields of early progenitors and

evaluate their quality. Stem Cells 2002;20:530-41.

- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage potential of adult human mesenchymal stem cells. Science 1999;284:143-7.
- Phinney DG, Prockop DJ. Concise review: mesenchymal stem/ multipotent stromal cells: the state of transdifferentiation and modes of tissue repair--current views. Stem Cells 2007;25:2896-902.
- Lee MW, Yang MS, Park JS, Kim HC, Kim YJ, Choi J. Isolation of mesenchymal stem cells from cryopreserved human umbilical cord blood. Int J Hematol 2005;81:126-30.
- Kotobuki N, Hirose M, Takakura Y, Ohgushi H. Cultured autologous human cells for hard tissue regeneration: preparation and characterization of mesenchymal stem cells from bone marrow. Artif Organs 2004;28:33-9.
- Dazzi F, Ramasamy R, Glennie S, Jones SP, Roberts I. The role of mesenchymal stem cells in haemopoiesis. Blood Rev 2006;20: 161-71.
- Rasmusson I, Ringdén O, Sundberg B, Le Blanc K. Mesenchymal stem cells inhibit the formation of cytotoxic T lymphocytes, but not activated cytotoxic T lymphocytes or natural killer cells. Transplantation 2003;76:1208-13.
- DelaRosa O, Lombardo E. Modulation of adult mesenchymal stem cells activity by toll-like receptors: implications on therapeutic potential. Mediators Inflamm 2010;2010:865601.
- Le Blanc K, Tammik C, Rosendahl K, Zetterberg E, Ringdén O. HLA expression and immunologic properties of differentiated and undifferentiated mesenchymal stem cells. Exp Hematol 2003;31: 890-6.
- Caplan AI, Dennis JE. Mesenchymal stem cells as trophic mediators. J Cell Biochem 2006;98:1076-84.
- Bernardo ME, Locatelli F, Fibbe WE. Mesenchymal stromal cells. Ann N Y Acad Sci 2009;1176:101-17.
- Vemuri MC, Chase LG, Rao MS. Mesenchymal stem cell assays and applications. Methods Mol Biol 2011;698:3-8.
- Bae KS, Park JB, Kim HS, Kim DS, Park DJ, Kang SJ. Neuronlike differentiation of bone marrow-derived mesenchymal stem cells. Yonsei Med J 2011;52:401-12.
- Horwitz EM, Le Blanc K, Dominici M, Mueller I, Slaper-Cortenbach I, Marini FC, et al. Clarification of the nomenclature for MSC: The International Society for Cellular Therapy position statement. Cytotherapy 2005;7:393-5.
- Bianco P, Kuznetsov SA, Riminucci M, Gehron Robey P. Postnatal skeletal stem cells. Methods Enzymol 2006;419:117-48.
- Laflamme MA, Murry CE. Regenerating the heart. Nat Biotechnol 2005;23:845-56.
- Chen J, Li Y, Katakowski M, Chen X, Wang L, Lu D, et al. Intravenous bone marrow stromal cell therapy reduces apoptosis and promotes endogenous cell proliferation after stroke in female rat. J Neurosci Res 2003;73:778-86.
- Li Y, Chen J, Zhang CL, Wang L, Lu D, Katakowski M, et al. Gliosis and brain remodeling after treatment of stroke in rats with marrow stromal cells. Glia 2005;49:407-17.
- Murphy JM, Fink DJ, Hunziker EB, Barry FP. Stem cell therapy in a caprine model of osteoarthritis. Arthritis Rheum 2003;48: 3464-74.
- Rosová I, Dao M, Capoccia B, Link D, Nolta JA. Hypoxic preconditioning results in increased motility and improved therapeutic potential of human mesenchymal stem cells. Stem Cells 2008; 26:2173-82.

- Nagai A, Kim WK, Lee HJ, Jeong HS, Kim KS, Hong SH, et al. Multilineage potential of stable human mesenchymal stem cell line derived from fetal marrow. PLoS One 2007;2:e1272.
- 34. Li Y, Chen J, Chen XG, Wang L, Gautam SC, Xu YX, et al. Human marrow stromal cell therapy for stroke in rat: neurotrophins and functional recovery. Neurology 2002;59:514-23.
- Qu R, Li Y, Gao Q, Shen L, Zhang J, Liu Z, et al. Neurotrophic and growth factor gene expression profiling of mouse bone marrow stromal cells induced by ischemic brain extracts. Neuropathology 2007;27:355-63.
- Cho SR, Yang MS, Yim SH, Park JH, Lee JE, Eom YW, et al. Neurally induced umbilical cord blood cells modestly repair injured spinal cords. Neuroreport 2008;19:1259-63.
- 37. Yoo SW, Kim SS, Lee SY, Lee HS, Kim HS, Lee YD, et al. Mesenchymal stem cells promote proliferation of endogenous neural stem cells and survival of newborn cells in a rat stroke model. Exp Mol Med 2008;40:387-97.
- Kopen GC, Prockop DJ, Phinney DG. Marrow stromal cells migrate throughout forebrain and cerebellum, and they differentiate into astrocytes after injection into neonatal mouse brains. Proc Natl Acad Sci U S A 1999;96:10711-6.
- 39. Kim SS, Yoo SW, Park TS, Ahn SC, Jeong HS, Kim JW, et al. Neural induction with neurogenin1 increases the therapeutic effects of mesenchymal stem cells in the ischemic brain. Stem Cells 2008;26:2217-28.
- Maltman DJ, Hardy SA, Przyborski SA. Role of mesenchymal stem cells in neurogenesis and nervous system repair. Neurochem Int 2011;59:347-56.
- 41. Sakai T, Johnson KJ, Murozono M, Sakai K, Magnuson MA, Wieloch T, et al. Plasma fibronectin supports neuronal survival and reduces brain injury following transient focal cerebral ischemia but is not essential for skin-wound healing and hemostasis. Nat Med 2001;7:324-30.
- Giger RJ, Hollis ER 2nd, Tuszynski MH. Guidance molecules in axon regeneration. Cold Spring Harb Perspect Biol 2010;2: a001867.
- Akiyama Y, Radtke C, Honmou O, Kocsis JD. Remyelination of the spinal cord following intravenous delivery of bone marrow cells. Glia 2002;39:229-36.
- 44. Satake K, Lou J, Lenke LG. Migration of mesenchymal stem cells through cerebrospinal fluid into injured spinal cord tissue. Spine (Phila Pa 1976) 2004;29:1971-9.
- Uccelli A, Moretta L, Pistoia V. Mesenchymal stem cells in health and disease. Nat Rev Immunol 2008;8:726-36.
- 46. Zappia E, Casazza S, Pedemonte E, Benvenuto F, Bonanni I, Gerdoni E, et al. Mesenchymal stem cells ameliorate experimental autoimmune encephalomyelitis inducing T-cell anergy. Blood 2005;106:1755-61.
- 47. Gerdoni E, Gallo B, Casazza S, Musio S, Bonanni I, Pedemonte E, et al. Mesenchymal stem cells effectively modulate pathogenic immune response in experimental autoimmune encephalomyelitis. Ann Neurol 2007;61:219-27.
- Kim YJ, Park HJ, Lee G, Bang OY, Ahn YH, Joe E, et al. Neuroprotective effects of human mesenchymal stem cells on dopaminergic neurons through anti-inflammatory action. Glia 2009;57: 13-23.
- 49. Chao YX, He BP, Tay SS. Mesenchymal stem cell transplantation attenuates blood brain barrier damage and neuroinflammation and protects dopaminergic neurons against MPTP toxicity in the substantia nigra in a model of Parkinson's disease. J Neuroimmunol

2009;216:39-50.

- 50. Ohtaki H, Ylostalo JH, Foraker JE, Robinson AP, Reger RL, Shioda S, et al. Stem/progenitor cells from bone marrow decrease neuronal death in global ischemia by modulation of inflammatory/immune responses. Proc Natl Acad Sci U S A 2008;105:14638-43.
- Seo JH, Jang IK, Kim HB, Yang MS, Lee JE, Kim HE, et al. Immunomodulation from intravenous transplantation of mesenchymal stem cells promotes functional recovery in spinal cord injured rats. Cell Med 2011;2:55-67.
- Gnecchi M, He H, Noiseux N, Liang OD, Zhang L, Morello F, et al. Evidence supporting paracrine hypothesis for Akt-modified mesenchymal stem cell-mediated cardiac protection and functional improvement. FASEB J 2006;20:661-9.
- 53. Cai L, Johnstone BH, Cook TG, Liang Z, Traktuev D, Cornetta K, et al. Suppression of hepatocyte growth factor production impairs the ability of adipose-derived stem cells to promote ischemic tissue revascularization. Stem Cells 2007;25:3234-43.
- 54. Nakagami H, Maeda K, Morishita R, Iguchi S, Nishikawa T, Takami Y, et al. Novel autologous cell therapy in ischemic limb disease through growth factor secretion by cultured adipose tissuederived stromal cells. Arterioscler Thromb Vasc Biol 2005;25: 2542-7.
- Rehman J, Traktuev D, Li J, Merfeld-Clauss S, Temm-Grove CJ, Bovenkerk JE, et al. Secretion of angiogenic and antiapoptotic factors by human adipose stromal cells. Circulation 2004;109: 1292-8.
- Sadat S, Gehmert S, Song YH, Yen Y, Bai X, Gaiser S, et al. The cardioprotective effect of mesenchymal stem cells is mediated by IGF-I and VEGF. Biochem Biophys Res Commun 2007;363:674-9.
- Miranville A, Heeschen C, Sengenès C, Curat CA, Busse R, Bouloumié A. Improvement of postnatal neovascularization by human adipose tissue-derived stem cells. Circulation 2004;110:349-55.
- Crigler L, Robey RC, Asawachaicharn A, Gaupp D, Phinney DG. Human mesenchymal stem cell subpopulations express a variety of neuro-regulatory molecules and promote neuronal cell survival and neuritogenesis. Exp Neurol 2006;198:54-64.
- Cho SR, Kim YR, Kang HS, Yim SH, Park CI, Min YH, et al. Functional recovery after the transplantation of neurally differentiated mesenchymal stem cells derived from bone barrow in a rat model of spinal cord injury. Cell Transplant 2009;18:1359-68.
- Cho HJ, Lee N, Lee JY, Choi YJ, Ii M, Wecker A, et al. Role of host tissues for sustained humoral effects after endothelial progenitor cell transplantation into the ischemic heart. J Exp Med 2007; 204:3257-69.
- Tateno K, Minamino T, Toko H, Akazawa H, Shimizu N, Takeda S, et al. Critical roles of muscle-secreted angiogenic factors in therapeutic neovascularization. Circ Res 2006;98:1194-202.
- Si YL, Zhao YL, Hao HJ, Fu XB, Han WD. MSCs: biological characteristics, clinical applications and their outstanding concerns. Ageing Res Rev 2011;10:93-103.
- Koc ON, Lazarus HM. Mesenchymal stem cells: heading into the clinic. Bone Marrow Transplant 2001;27:235-9.
- 64. Horwitz EM, Gordon PL, Koo WK, Marx JC, Neel MD, McNall RY, et al. Isolated allogeneic bone marrow-derived mesenchymal cells engraft and stimulate growth in children with osteogenesis imperfecta: implications for cell therapy of bone. Proc Natl Acad Sci U S A 2002;99:8932-7.
- Dimmeler S, Zeiher AM, Schneider MD. Unchain my heart: the scientific foundations of cardiac repair. J Clin Invest 2005;115: 572-83.

- 66. Loebinger MR, Eddaoudi A, Davies D, Janes SM. Mesenchymal stem cell delivery of TRAIL can eliminate metastatic cancer. Cancer Res 2009;69:4134-42.
- 67. Aguayo-Mazzucato C, Bonner-Weir S. Stem cell therapy for type 1 diabetes mellitus. Nat Rev Endocrinol 2010;6:139-48.
- Pistoia V, Raffaghello L. Potential of mesenchymal stem cells for the therapy of autoimmune diseases. Expert Rev Clin Immunol 2010;6:211-8.
- Minger SL, Ekonomou A, Carta EM, Chinoy A, Perry RH, Ballard CG. Endogenous neurogenesis in the human brain following cerebral infarction. Regen Med 2007;2:69-74.
- Ekonomou A, Ballard CG, Pathmanaban ON, Perry RH, Perry EK, Kalaria RN, et al. Increased neural progenitors in vascular dementia. Neurobiol Aging 2011;32:2152-61.
- Bang OY, Lee JS, Lee PH, Lee G. Autologous mesenchymal stem cell transplantation in stroke patients. Ann Neurol 2005;57:874-82.
- Lee JS, Hong JM, Moon GJ, Lee PH, Ahn YH, Bang OY; START-ING collaborators. A long-term follow-up study of intravenous autologous mesenchymal stem cell transplantation in patients with ischemic stroke. Stem Cells 2010;28:1099-106.
- Zhang ZX, Guan LX, Zhang K, Zhang Q, Dai LJ. A combined procedure to deliver autologous mesenchymal stromal cells to patients with traumatic brain injury. Cytotherapy 2008;10:134-9.
- Joyce N, Annett G, Wirthlin L, Olson S, Bauer G, Nolta JA. Mesenchymal stem cells for the treatment of neurodegenerative disease. Regen Med 2010;5:933-46.
- Schwab ME. Repairing the injured spinal cord. Science 2002;295: 1029-31.
- Sahni V, Kessler JA. Stem cell therapies for spinal cord injury. Nat Rev Neurol 2010;6:363-72.
- 77. Geffner LF, Santacruz P, Izurieta M, Flor L, Maldonado B, Auad AH, et al. Administration of autologous bone marrow stem cells into spinal cord injury patients via multiple routes is safe and improves their quality of life: comprehensive case studies. Cell Transplant 2008;17:1277-93.
- Syková E, Homola A, Mazanec R, Lachmann H, Konrádová SL, Kobylka P, et al. Autologous bone marrow transplantation in patients with subacute and chronic spinal cord injury. Cell Transplant 2006;15:675-87.
- Pal R, Venkataramana NK, Bansal A, Balaraju S, Jan M, Chandra R, et al. Ex vivo-expanded autologous bone marrow-derived mesenchymal stromal cells in human spinal cord injury/paraplegia: a pilot clinical study. Cytotherapy 2009;11:897-911.
- Ra JC, Shin IS, Kim SH, Kang SK, Kang BC, Lee HY, et al. Safety of intravenous infusion of human adipose tissue-derived mesenchymal stem cells in animals and humans. Stem Cells Dev 2011;20:1297-308.
- Li JY, Englund E, Holton JL, Soulet D, Hagell P, Lees AJ, et al. Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. Nat Med 2008;14: 501-3.
- Venkataramana NK, Kumar SK, Balaraju S, Radhakrishnan RC, Bansal A, Dixit A, et al. Open-labeled study of unilateral autologous bone-marrow-derived mesenchymal stem cell transplantation in Parkinson's disease. Transl Res 2010;155:62-70.
- Lee PH, Kim JW, Bang OY, Ahn YH, Joo IS, Huh K. Autologous mesenchymal stem cell therapy delays the progression of neurological deficits in patients with multiple system atrophy. Clin Pharmacol Ther 2008;83:723-30.
- 84. Nagano I, Shiote M, Murakami T, Kamada H, Hamakawa Y, Mat-

subara E, et al. Beneficial effects of intrathecal IGF-1 administration in patients with amyotrophic lateral sclerosis. Neurol Res 2005;27:768-72.

- Karussis D, Karageorgiou C, Vaknin-Dembinsky A, Gowda-Kurkalli B, Gomori JM, Kassis I, et al. Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis. Arch Neurol 2010;67:1187-94.
- Mazzini L, Mareschi K, Ferrero I, Vassallo E, Oliveri G, Nasuelli N, et al. Stem cell treatment in Amyotrophic Lateral Sclerosis. J Neurol Sci 2008;265:78-83.
- 87. Mazzini L, Ferrero I, Luparello V, Rustichelli D, Gunetti M,

Mareschi K, et al. Mesenchymal stem cell transplantation in amyotrophic lateral sclerosis: A Phase I clinical trial. Exp Neurol 2010;223:229-37.

- Mohyeddin Bonab M, Yazdanbakhsh S, Lotfi J, Alimoghaddom K, Talebian F, Hooshmand F, et al. Does mesenchymal stem cell therapy help multiple sclerosis patients? Report of a pilot study. Iran J Immunol 2007;4:50-7.
- Yamout B, Hourani R, Salti H, Barada W, El-Hajj T, Al-Kutoubi A, et al. Bone marrow mesenchymal stem cell transplantation in patients with multiple sclerosis: a pilot study. J Neuroimmunol 2010;227:185-9.