

Downregulated miR-23b-3p expression acts as a predictor of hepatocellular carcinoma progression: A study based on public data and RT-qPCR verification

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Abstract. Mounting evidence has shown that miR-23b-3p, which is associated with cell proliferation, invasion, and apoptosis, acts as a biomarker for diagnosis and outcomes in numerous cancers. However, the clinicopathological implication of miR-23b-3p in hepatocellular carcinoma (HCC) remains unclear. Our study evaluated the role of miR-23b-3p in HCC and investigated its potential application as a marker for preliminary diagnosis and therapy in HCC. High-throughput data from the NCBI Gene Expression Omnibus (GEO) and The Cancer Genome Atlas (TCGA) were collected and analyzed. One hundred and one tissue sections of HCC were paired with adjacent non-cancerous HCC as further supplements. miR-23b-3p expression was detected using quantitative real-time PCR. Additionally, the relationship between miR-23b-3p expression and HCC progression and Time-to-recurrence (months) was explored. Ten algorithms were applied to predict the prospective target genes of miR-23b-3p. Next, we conducted bioinformatics analysis for further study. miR-23b-3p expression was pronouncedly decreased in HCC tissues in contrast with their paired adjacent non-cancerous HCC ($P < 0.001$) with RT-qPCR. In total, 405 targets, acquired with consistent prediction from at least five databases, were used for the bioinformatics analysis. According to the Gene Ontology (GO) analysis, all targets were classified into biological processes, cellular components and molecular functions.

In the pathway analysis, targets of miR-23b-3p were primarily enriched in the signaling pathways of renal cell carcinoma, hepatitis B and pancreatic cancer (corrected P -value < 0.05). In the protein-protein interaction (PPI) network for miR-23b-3p, a total of 8 targets, including SRC, AKT1, EGFR, CTNNB1, BCL2, SMAD3, PTEN and KDM6A, were located in the key nodes with high degree (> 35). In conclusion, this study provides impressive illumination of the potential role of miR-23b-3p in HCC tumorigenesis and progression. Furthermore, miR-23b-3p may act as a predictor of HCC and could be a new treatment target.

Introduction

Hepatocellular carcinoma (HCC) is a malignant tumor that has one of the highest morbidity and mortality rates, and its tumorigenesis is closely related to HBV and HCV infection (1). There were estimated 782,500 hepatic carcinoma cases and 745,500 related deaths globally in 2012, according to Torre *et al* (2). Moreover, estimated 70-90% of primary liver cancers are HCC. Despite advances in chemotherapy, radiotherapy and surgery, the low 5-year survival and quality of life observed in HCC patients remain intractable issues. Many etiological factors, such as alcohol consumption and aflatoxin B1 exposure, are widely accepted to facilitate HCC occurrence (3,4); however, the precise molecular mechanisms remain unknown. Thus, there is an urgent need for a reliable biomarker that can be used to predict HCC progression and prognosis.

MicroRNAs (miRNAs), which are limited to a length of 19-22 nucleotides, are small non-coding RNAs that post-transcriptionally influence gene expression (5). A vast array of oncogenes and anti-oncogenes are potentially regulated by miRNAs, which act as gene regulators by conjugating the 3'-untranslated region (3'-UTR) of their mRNAs. In regulating genes, miRNAs play a crucial biological role in tumor development, especially during initiation, proliferation, differentiation, invasion and metastasis (6). Thus, miRNAs may serve as promising diagnostic and prognostic indexes.

Validating a potential target of miRNA is arduous and time-consuming due to the tremendous amounts of target

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sites in miRNA. However, predicting the targets of miRNA, which is a vital step in performing miRNA-target interactions, contributes to narrowing down prospective target sites and promoting experimental verification. Following the principle of sequence complementarity, many algorithms were manipulated to figure out the predicted miRNA targets.

miR-23b-3p was identified as a tumor suppressor that showed a tendency toward downregulated expression in different classes of human malignant tumors such as prostate cancer, renal cell carcinoma, acute myeloid leukemia and osteosarcoma (7-10). Nevertheless, increasing evidence has indicated that the upregulation of miR-23b-3p promoted cell proliferation and invasion in glioma, gastric cancer and breast cancer (11-13). miR-23b-3p was also found to be involved in liver stem cell differentiation, and its reduced expression may contribute to liver regeneration after partial hepatectomy (14,15). However, to our knowledge, only one study (16) has explored the relationship between miR-23b-3p expression and HCC but did not comment on the clinicopathological significance and prognosis. Therefore, further inquiry is urgently needed.

Our objectives were as follows: to study the pathophysiological expression quantity of miR-23b-3p in HCC tissue through comparison with matching adjacent tissues as well as to elucidate the correlation between miR-23b-3p expression and HCC clinicopathological parameters. Additionally, we aimed to identify the probable target genes of miR-23b-3p and determine the potential role of miR-23b-3p in HCC development and progression.

Materials and methods

miR-23b-3p expression in HCC based on GEO datasets

Data acquisition and exclusion criteria. Twenty-one microarray datasets were obtained from the GEO database (<http://www.ncbi.nlm.nih.gov/geo/>), and 11 were eliminated after screening. The search strategy was formulated as follows: (malignan* OR cancer OR tumor OR tumour OR neoplas* OR carcinoma) AND (hepatocellular OR liver OR hepatic OR HCC). The last dataset search was on April 2016. The exclusion criteria were as follows: i) datasets without information on miR-23b-3p; ii) datasets without complete data for analysis; iii) samples based on cell lines; iv) not all subjects of the included studies were human; or v) miR-23b-3p was determined in the HCC patients without a comparison. To identify the clinical relevance of miR-23b-3p in HCC, we collected data on miR-23b-3p expression from the 6 datasets and analyzed their association with the clinicopathological characteristics of HCC. The clinical features, which could be obtained from more than two microarray datasets, were used for the meta-analysis.

Statistical analysis. The meta-analysis was carried out with Stata 12.0 (StataCorp LP, College Station, TX, USA). A standard mean difference (SMD) and a 95% confidence interval (CI) were utilized to measure continuous outcomes, including age, sex, HBV infection, invasion and metastasis. Fixed or random effects models were applied to pool the effect sizes. Cochrane's Q test (Chi-square test; Chi2) (17) and inconsistency (I^2) (18) were conducted to assess heterogeneity. A $P < 0.05$ or $I^2 > 50\%$ indicates significant heterogeneity (19), and a random effects model (20) was applied. Otherwise,

the fixed effects model would be adopted. Begg's funnel plot for asymmetry and Egger's funnel plot for quantitation were generated to evaluate publication bias. In addition, sensitivity analysis was performed to evaluate the reliability of results by elimination of a study each time.

miR-23b-3p expression in HCC based on TCGA dataset

Retrieval of public data. Altogether, 377 anonymized HCC tissues and 50 normal tissues were retrieved from the TCGA database (<http://cancergenome.nih.gov/publications/publicationguidelines>). The non-HCC samples or samples with data deficiency were excluded; 361 HCC patients were finally included in this study. Additionally, a total of 50 normal liver tissues were retrieved for comparison. The clinicopathological features of HCC patients in TCGA are available in Table II.

Statistical analysis. The statistics were analyzed with SPSS 22.0 software (IBM Corp., Armonk, NY, USA). The final data after calculation are shown as the mean \pm standard deviation (SD). Student's t-test was conducted for a comparative analysis of two independent groups. A one-way analysis of variance (ANOVA) test was utilized to study the data, which were divided into three or four groups, such as T status, N status or M status. Spearman's method was performed to evaluate the association between miR-23b-3p expression and other clinicopathological parameters. The relationship between miR-23b-3p and recurrence was obtained with the Kaplan-Meier survival approach along with a log-rank test. To differentiate between controls and HCC tissues, a diagnostic value was identified using a receiver operator characteristic (ROC) curve. A P -value < 0.05 denoted a statistically significant difference. At least 5 individual tests were applied without ambiguity.

miR-23b-3p expression in HCC hospital tissues

Patients and data collected from medical records. Regarding the cases recruited, 101 FFPE HCC tissues and their counterpart adjacent non-cancerous HCC were obtained from the First Affiliated Hospital of the Guangxi Medical University March, 2010 and December, 2011. Before curative hepatectomy, none had received radiotherapy or chemotherapy. Formalin-fixed and paraffin embedded (FFPE) tissues were assessed in both the HCC tissues and adjacent non-cancerous HCC; we retrospectively acquired the following clinicopathological parameters: age, sex, cirrhosis, tumor size, tumor nodes, differentiation, clinical TNM stages, metastasis, existence or inexistence of portal vein tumor embolus, capsular infiltration, vaso-invasion, as well as other biomarkers, such as serum AFP level, microvessel density (MVD). Two independent pathologists diagnosed all the cases. The project had already gained ethics approval from the Ethics Committee of First Affiliated Hospital of Guangxi Medical University and the patients signed informed consents and agreed their samples to be used in scientific research. The clinicopathological characteristics of the population are shown in Table III.

RT-qPCR. The total RNAs were isolated from clinical tissue samples with a miRNeasy FFPE kit (Qiagen, Limburg, The Netherlands) according to the manufacturer's instructions. To assess the miRNA purity and concentration, a quantitative real-time PCR method was utilized in 101 HCC tissues and their counterpart adjacent non-cancerous HCC.

Statistical analysis. The statistical software and methods that were used to analyze the data from clinical patient files downloaded from TCGA were also used to identify the relationship between miR-23b-3p expression and HCC based on qPCR data.

Meta-analysis for GEO datasets, TCGA datasets and PCR verification in-house

Data acquisition. We next added the data from TCGA datasets and PCR verification in-house into meta-analysis to expand the sample size and enhance the credibility. The expression of hsa-miR-23b-3p in HCC tissues and adjacent non-cancerous tissues were extracted in meta-analysis.

Statistical analysis. As for statistical analysis of meta-analysis of GEO datasets, the same statistical methods and software were used to perform the meta-analysis of GEO datasets, TCGA datasets and qPCR data.

Biological information analysis

Prediction targets of miR-23b-3p collection. Ten algorithms, including those from TargetScan (<http://www.targetscan.org>), microRNA.org (<http://www.microrna.org>), RNA22 (<https://cm.jefferson.edu/rna22/Precomputed>), PicTar-vert (http://pictar.mdc-berlin.de/cgi-bin/PicTar_vertebrate.cgi), miRDB (<http://mirdb.org/miRDB>), PolymiRTS Database (<http://compbio.uthsc.edu/miRSNP/>), PITA (http://genie.weizmann.ac.il/pubs/mir07/mir07_dyn_data.html#), TargetMiner (http://www.isical.ac.in/~bioinfo_miu/targetminer20.htm), TarBase (<http://diana.imis.athenainnovation.gr/DianaTools/index.php?r=tarbase>), and miRTarBase (<http://mirtarbase.mbc.nctu.edu.tw>), were applied. Only target genes appearing more than or equal to five times amongst all 10 algorithms were finally applied for investigation of the next step.

Validation targets of miR-23b-3p collection. We searched the PubMed database using the following search strategy: (MIRN23 OR microRNA23 OR microRNA-23 OR miR-23 OR hsa-mir-23 OR miR-23b OR microRNA-23b OR miRNA-23b OR 'miR23b' OR 'miRNA23b' OR 'microRNA23b' OR miR-23b-3p OR miRNA-23b-3p OR microRNA-23b-3p) and target*. The validation targets of hsa-miR-23b-3p were recorded from the identified articles by two authors (P-R Wu and X-L Xiang), and the third author resolved controversies. These targets were integrated with prediction targets after combination.

Bioinformatics analysis of miR-23b-3p. Gene Ontology (GO) analysis. GO analysis was conducted using the Database for Annotation, Visualization and Integrated Discovery (DAVID) (<http://David.abcc.ncifcrf.gov/>). The targets of miR-23b-3p were divided into three primary classes: biological processes, cellular components and molecular functions.

Pathway analysis. For pathway analysis, we downloaded pathway data from the Kyoto Encyclopedia of Genes and Genomes (KEGG) (<http://www.genome.ad.jp/KEGG>) to investigate gene interactions. miR-23b-3p targets were mapped to the KEGG pathway database using KOBAS 2.0 (<http://www.genome.jp/kegg/pathway.html>), and P-values [and a corresponding false discovery rate (FDR)] were applied to estimate each enriched pathway.

Network analysis. The Search Tool for the Retrieval of Interacting Genes (STRING), which provides information for experimental and predicted interactions, is an online database. STRING was applied to search and to determine an interaction network of targets of miR-23b-3p via confidence score calculation.

Results

miR-23b-3p expression in HCC based on GEO datasets

Characteristics of the included datasets. The characteristics of the GEO datasets included are presented in Table I. A total of 10 datasets, including GSE6857 (USA, 2013) (21), GSE10694 (China, 2008) (22), GSE21362 (Japan, 2010) (23), GSE12717 (China, 2008) (24), GSE54751 (USA, 2014) (25), GSE57555 (Japan, 2015) (26), GSE41874 (Japan, 2013) (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE41874>), GSE67138 (USA, 2015) (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67138>), GSE69580 (China 2015) (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE69580>), and GSE67139 (USA, 2015) (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE67139>), were identified to meet the standards. All of the datasets were derived from tissues.

Value of miR-23b-3p as a biomarker for HCC. In 7 datasets presenting miR-23b-3p expression with comparable groups, 414 patients with HCC and 343 healthy subjects were recruited. The expression of miR-23b-3p was lower in HCC patients than healthy controls (Fig. 1A); however, no statistical significance was found (SMD=-0.081; 95% CI, -0.216 to 0.054; P=0.239). A fixed effects model was selected to pool the effect variables that did not show heterogeneity (P=0.807, I²=0%). The results of Begg's and Egger's test shown in Fig. 1B and C was 0.133 and 0.251, respectively. As a consequence, no publication bias was discovered. According to sensitivity analysis shown in Fig. 1D, the result was altered when we eliminated the study of GSE6857, suggesting that this study may influence the pooled SMD and 95% CI. The forest plot after the removing of GSE6857 is shown in Fig. 2A; however, no statistical significance was found (SMD=-0.145; 95% CI, -0.352 to 0.061; P=0.168). According to Begg's and Egger's test (Fig. 2B and C), no publication bias was observed (P>0.05). The sensitivity analysis did not show any alteration when we removed a single study (Fig. 2D).

Six datasets were enrolled to assess the association between miR-23b-3p expression and the clinical aspects of HCC. However, five parameters, including HBV invasion (83 patients), age (90 patients), sex (8 patients), HBV infection (66 patients) and metastasis (27 patients) were estimated in this study. The pooled SMD were -1.302 (95% CI, -3.055 to 0.451, P=0.146) (Fig. 3A), 0.101 (95% CI, -0.203 to 0.405, P=0.515) (Fig. 3B), -0.266 (95% CI, -1.293 to 0.762, P=0.613) (Fig. 3C), -0.186 (95% CI, -1.093 to 1.465, P=0.775) (Fig. 3D), and -0.288 (95% CI, -0.787 to 0.210, P=0.257) (Fig. 3E), respectively. None of the five parameters were associated with the level of miR-23b-3p expression.

miR-23b-3p expression in HCC based on the TCGA dataset

The expression level of miR-23b-3p was downregulated in HCC samples. To explore the relationship between miR-23b-3p and HCC, we further obtained data for 361 HCC

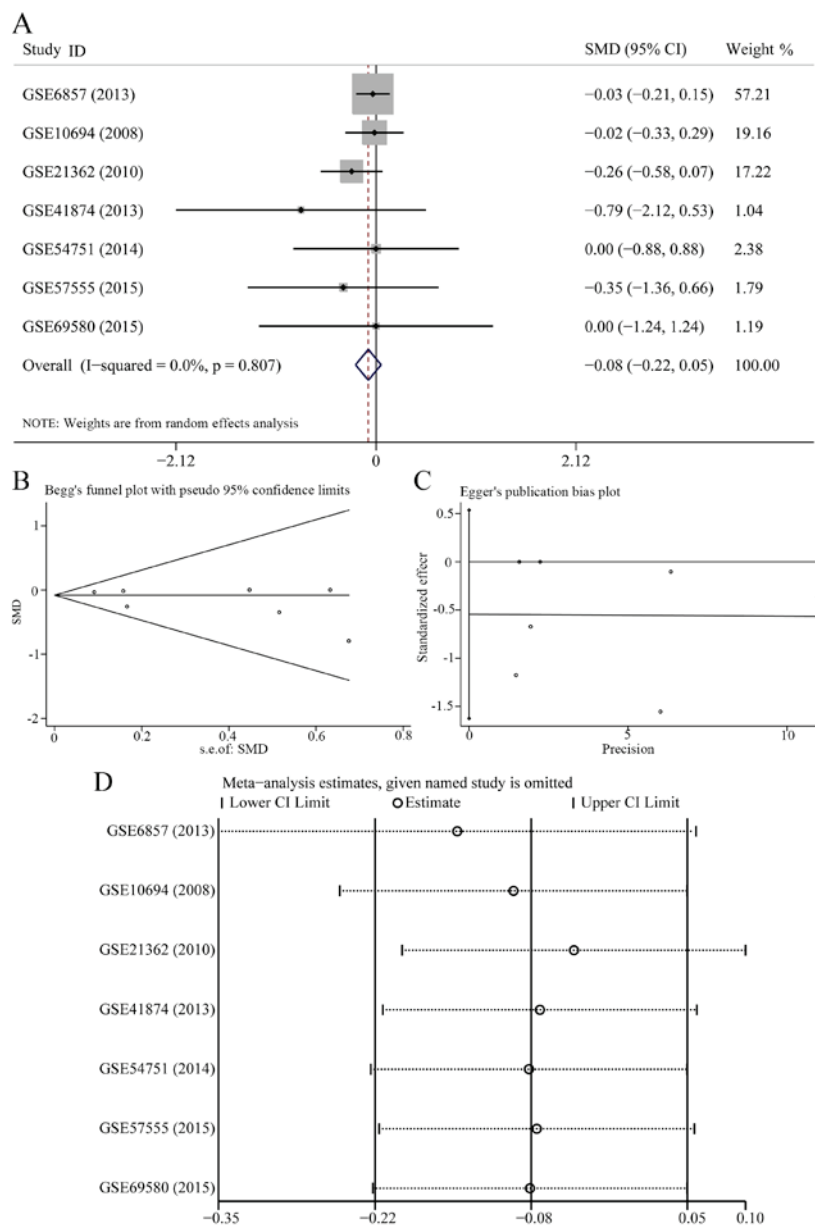


Figure 1. Meta-analysis of miR-23b-3p expression in hepatocellular carcinoma (HCC) patients and healthy controls retrieved from Gene Expression Omnibus (GEO) datasets. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg's funnel plot. Hazard ratios are presented on a logarithmic scale. (C) Egger's publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis.

tissues and 50 normal liver tissues from TCGA. The clinicopathological features are presented in Table II. According to the TCGA data, the expression level of miR-23b-3p was substantially downregulated in HCC (12.6722 ± 0.978150) being compared to normal liver tissues (13.4039 ± 0.51072 , $P < 0.001$) (Fig. 4A).

Relationships between miR-23b-3p and clinicopathological features. Due to the downregulated miR-23b-3p expression observed in HCC, the association between clinicopathological features and miR-23b-3p was studied (Table II and Fig. 5). The data indicated that a higher level of miR-23b-3p was closely related with hepatitis B (12.8831 ± 0.96070), in contrast to that without hepatitis B infection (12.5999 ± 0.97009 , $P = 0.013$). miR-23b-3p was also downregulated in smoking patients (12.2025 ± 1.11847) compared to those without smoking history (12.7127 ± 0.96181 , $P = 0.035$). The results

also gave a demonstration that the downregulated expression of miR-23b-3p was closely associated with vascular invasion (12.5905 ± 0.95709) compared with non-vascular invasion (12.8530 ± 0.94514 , $P = 0.032$). According to the universal standard pathologic neoplasm staging methods of AJCC, there was an obvious negative relationship between tumor node metastasis and miR-23b-3p. The expression level of miR-23b-3p was screened in regional lymphatic metastasis patients (11.7684 ± 0.75393). The expression level was 12.7780 ± 0.96867 in the patients who never had lymphatic metastasis; those with samples that remained in an undefined stage had a level of 12.4733 ± 0.96540 ($P = 0.006$). No other clinicopathological parameters had a significant relationship with miR-23b expression, such as HCV infection or alcohol consumption, nor did neoadjuvant therapy, radiation therapy or pharmaceutical treatment. In accordance with Table II, Spearman's method was

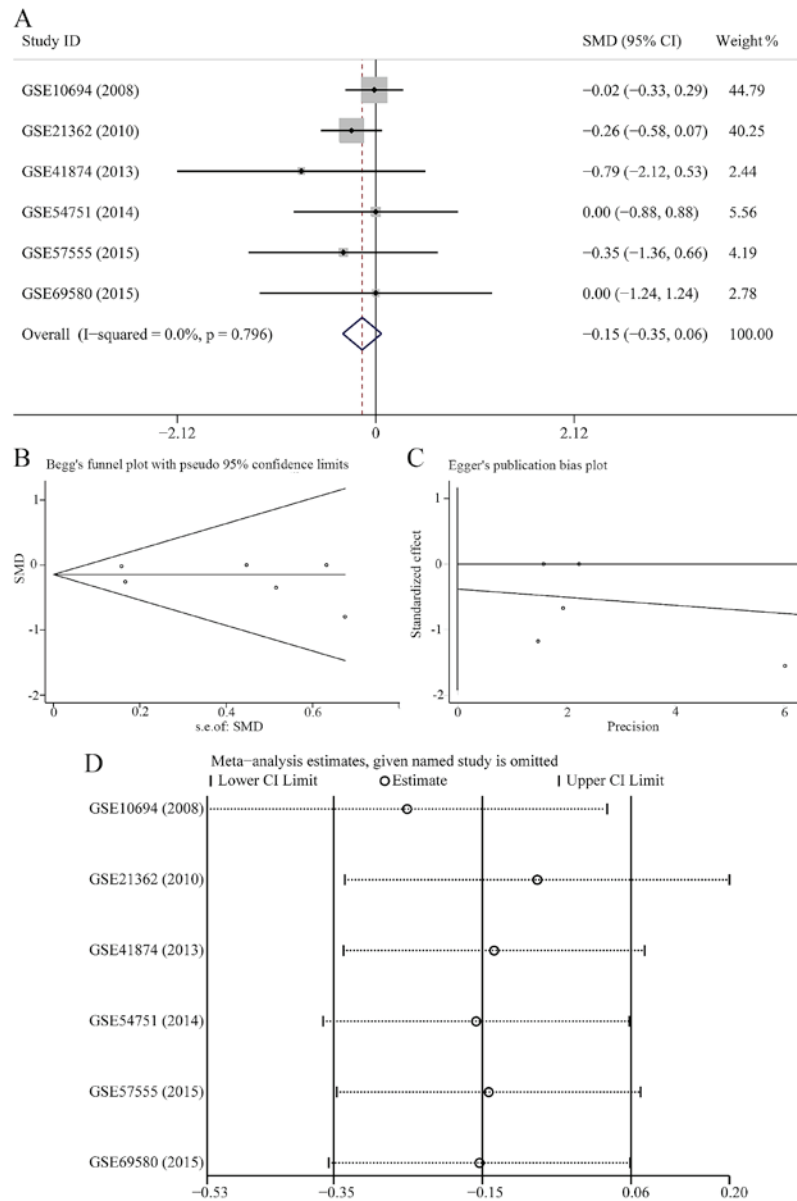


Figure 2. Meta-analysis of removing a single study from Gene Expression Omnibus (GEO) datasets. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg's funnel plot. Hazard ratios are presented on a logarithmic scale. (C) Egger's publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis. Pooled SMDs and 95% CIs by eliminating each study.

applied when correlating miR-23b-3p with clinicopathological parameters, including hepatitis B ($r=0.134$, $P=0.013$), smoking ($r=-0.014$, $P=0.035$), vascular invasion ($r=-0.122$, $P=0.032$), and N status ($r=-0.158$, $P=0.003$). There were no other positive correlations with the remaining clinical features.

The diagnostic accuracy of miR-23b-3p in HCC tissues. To further determine miR-23b-3p as a biomarker in diagnosing HCC, we performed ROC for verification. As shown in Fig. 4B, the AUC of miR-23b-3p in HCC and its counterpart normal tissues was 0.737 (95% CI, 0.680 to 0.794; $P<0.001$). The cut-off value reached 0.429; the sensitivity and specificity were 82.0 and 60.9%, respectively. The result suggested that miR-23b-3p may be treated as a reliable biomarker in diagnosing HCC. As shown in Fig. 6, the AUC to judge hepatitis B was 0.597 (95% CI, 0.532 to 0.662; $P=0.004$). The cut-off

value for miR-23b-3p was 0.174; the sensitivity and specificity were 65.1 and 52.3% respectively. The AUC to judge smoking was 0.626 (95% CI, 0.485 to 0.767; $P=0.080$). The cut-off value for miR-23b-3p was 0.259; the sensitivity and specificity were 78.8 and 47.1%, respectively. The AUC to judge vascular invasion was 0.566 (95% CI, 0.499 to 0.633, $P=0.057$). The cut-off value for miR-23b-3p was 0.125; the sensitivity and specificity were 45.2 and 67.3%, respectively. The AUC to judge N status was 0.601 (95% CI, 0.538 to 0.664, $P=0.002$). The cut-off value for miR-23b-3p was 0.125; the sensitivity and specificity were 63.2 and 55.8%, respectively.

miR-23b-3p expression in surviving HCC. According to the data from TCGA, 359 of the 361 patients were followed up. The Kaplan-Meier analysis in Fig. 7 showed that 176 patients had a lower miR-23b-3p expression (lower than the mean expression

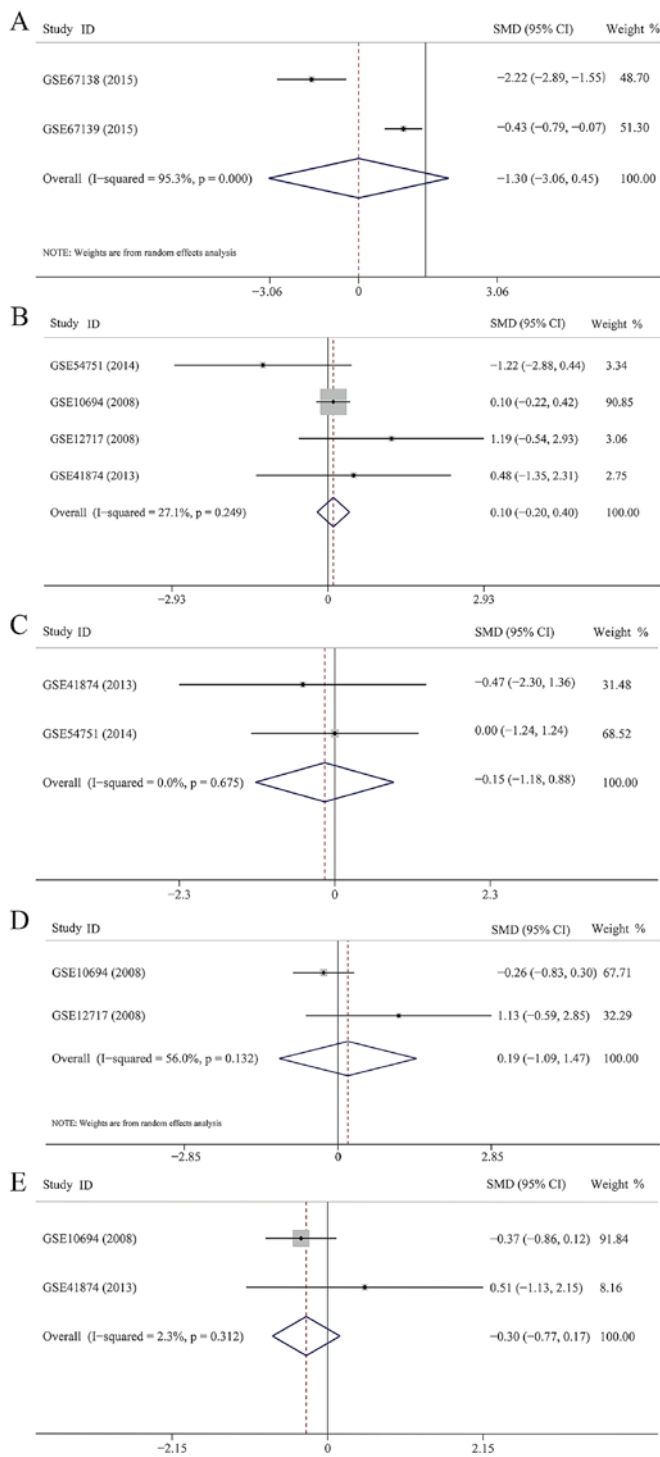


Figure 3. Meta-analysis of the relationships between miR-23b-3p and clinicopathological parameters in hepatocellular carcinoma (HCC) patients. (A) Invasion; (B) age; (C) sex; (D) HBV infection; (E) metastasis. Pooled standard mean differences (SMDs) and 95% confidence intervals (CIs) by eliminating each study.

level of 12.6797), whereas the remaining 183 patients had a higher level (higher than the mean expression level of 12.6797). In contrast, the lower group had an average survival time of 1,617.668±151.722 days, whereas the higher group had a survival time of 1,944.626±160.587 days. However, the survival times of the high and low miR-23b-3p expression groups were not significantly different (Chi-square =3.351, P=0.061).

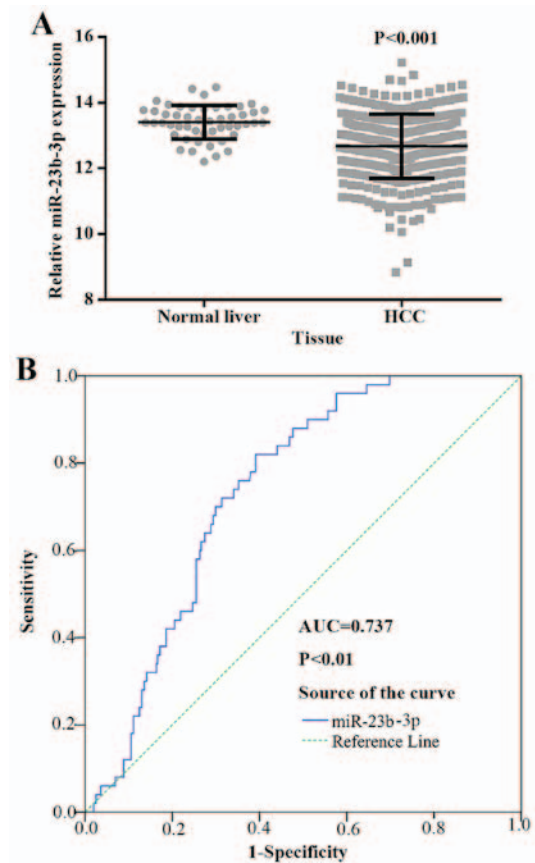


Figure 4. miR-23b-3p expression in hepatocellular carcinoma (HCC) tissues from The Cancer Genome Atlas (TCGA). (A) The difference in relevant miR-23b-3p expression between HCC and adjacent non-cancerous HCC; (B) ROC curve to distinguish HCC from normal tissues. Error bars represented standard deviation (SD).

miR-23b-3p expression in HCC tissues from PCR verification in-house

The expression of miR-23b-3p was downregulated in HCC. To study the expression level of miR-23b-3p, a quantitative real-time PCR method was performed in 101 HCC tissues and counterpart adjacent non-cancerous HCC. The clinicopathological features of the patients, obtained from medical records, are available in Table III. The expression of miR-23b-3p was obviously downregulated in HCC cases (2.7376 ± 1.99328) in contrast to their counterpart adjacent non-cancerous HCC (2.7376 ± 1.57739 , P<0.001) (Fig. 8A). The TCGA and PCR data provided similar results.

The relationships between miR-23b-3p and clinicopathological features. As noted above, the expression of miR-23b-3p was remarkably lower in the HCC tissues than their counterpart adjacent non-cancerous HCC, which indicated that miR-23b-3p may perform the function of a feasible tumor suppressive miRNA in HCC. We investigated the relationships between miR-23b-3p expression and clinicopathological features. The results shown in Table III and Fig. 9 validated that the down-regulated level of miR-23b-3p was positively related to patients without metastasis (3.3959 ± 1.57506), in contrast with those with metastasis (2.1173 ± 1.31762 , P<0.001). Subsequently, the miR-23b-3p level was significantly positively associated with multiple tumor nodes (3.0158 ± 1.45979) compared with single nodes (2.3773 ± 1.66606 , P=0.043). The presence of a portal

Table I. Characteristics of hsa-miR-23b-3p gene expression profiling datasets included in meta-analysis.

Citation	Country	Data source	HCC patients	Healthy controls	Platform
Budhu <i>et al</i> 2013 (21)	USA	GEO: GSE6857	240	241	GPL4700
Li <i>et al</i> 2008 (22)	China	GEO: GSE10694	75	87	GPL6542
Sato <i>et al</i> 2011 (23)	Japan	GEO: GSE21362	73	73 ^a	GPL10312
Su <i>et al</i> 2009 (24)	USA	GEO: GSE12717	5	3	GPL7274
Shen <i>et al</i> 2015 (25)	USA	GEO: GSE54751	10	10 ^a	GPL18262
Murakami <i>et al</i> 2015 (26)	Japan	GEO: GSE57555	5	5 ^a +11	GPL16699 GPL18044
Morita 2013 ^b	Japan	GEO: GSE41874	6	4	GPL7722
Hung 2015 ^b	China	GEO: GSE69580	5	5 ^a	GPL10850
Barry 2015 ^b	USA	GEO: GSE67138	23	34	GPL8786
Barry 2015 ^b	USA	GEO: GSE67139	60	60	GPL8786

^aCases from adjacent tumor tissues; ^bno relevant references. HCC, hepatocellular carcinoma.

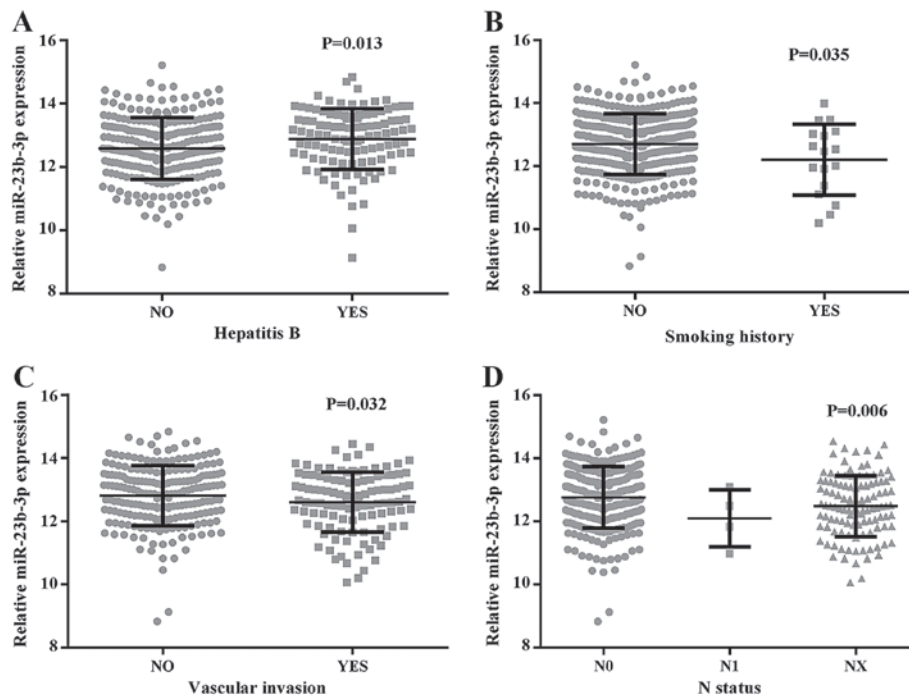


Figure 5. The relationship between miR-23b-3p expression and clinicopathological parameters of hepatocellular carcinoma (HCC) patients from The Cancer Genome Atlas (TCGA). (A) hepatitis B; (B) smoking history; (C) vascular invasion; (D) N status. Error bars represent standard deviation (SD).

vein tumor embolus showed a lower expression of miR-23b-3p (1.9500 ± 1.19055) than did its absence (3.1029 ± 1.60797 , $P < 0.001$). Furthermore, we used Spearman's method to record the correlations between the expression of miR-23b-3p and clinicopathological parameters, including tumor nodes ($r = -0.255$, $P = 0.010$), metastasis ($r = -0.447$, $P < 0.001$), and portal vein embolus ($r = -0.340$, $P = 0.001$). Nonetheless, we failed to gain any positive correlations between miR-23b-3p expression and other clinical features.

The diagnostic value of the miR-23b-3p level in HCC tissues. To verify the diagnostic value of the miR-23b-3p level in HCC tissues, a ROC curve was utilized. As shown in Fig. 8B, the area under the curve (AUC) of miR-23b-3p in HCC and adjacent non-cancerous HCC was 0.765 (95% CI,

0.700 to 0.830, $P < 0.001$). At 0.396 (the cut-off value of miR-23b-3p), the sensitivity and specificity was 88.2 and 55.4%, respectively. In Fig. 10, the AUC to judge tumor nodes was 0.648 (95% CI, 0.538 to 0.758, $P = 0.011$). The cut-off value for miR-23b-3p was 0.272; the sensitivity and specificity was 52.6 and 68.2%, respectively. The AUC to judge metastasis was 0.758 (95% CI, 0.664 to 0.851, $P < 0.001$). The cut-off value for miR-23b-3p was 0.468; the sensitivity and specificity was 77.6 and 69.2%, respectively. The AUC to judge portal vein tumor embolus was 0.711 (95% CI, 0.601 to 0.820, $P = 0.001$). The cut-off value for miR-23b-3p was 0.29; the sensitivity and specificity was 88.4 and 40.6%, respectively.

miR-23b-3p expression in HCC recurrence. Of the 101 patients recruited for the present study, 76 completed the

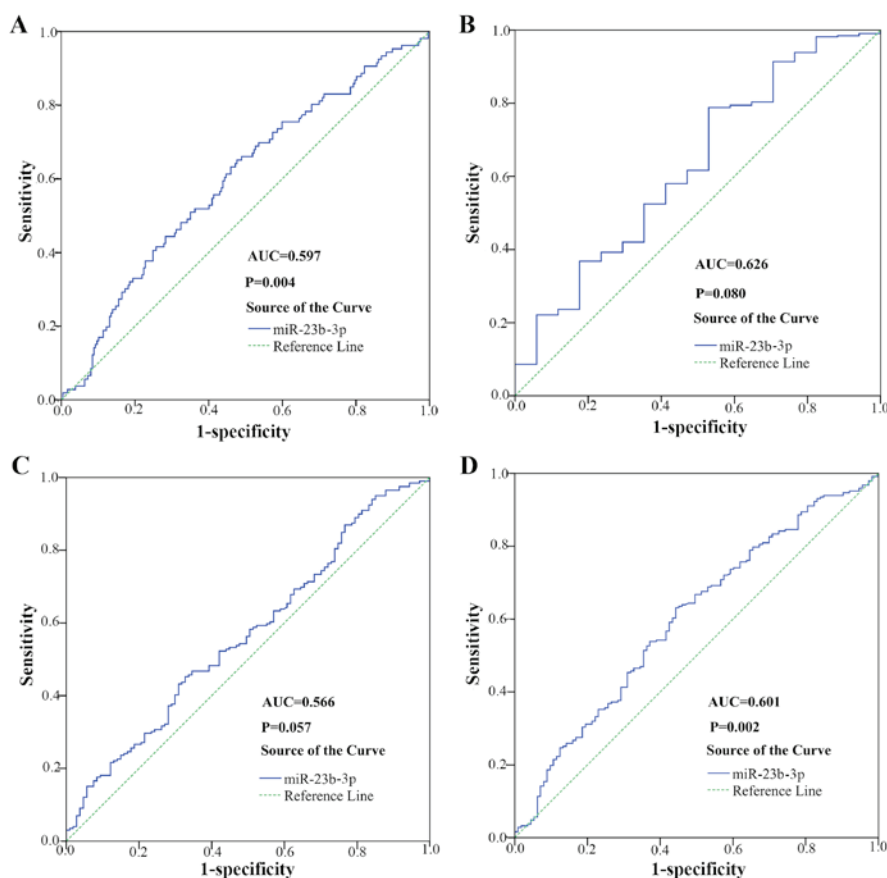


Figure 6. Diagnostic significance between miR-23b-3p and different parameters on hepatocellular carcinoma (HCC) tissues from The Cancer Genome Atlas (TCGA). (A) Hepatitis B; (B) smoking history; (C) vascular invasion; (D) N status. Error bars represented standard deviation (SD).

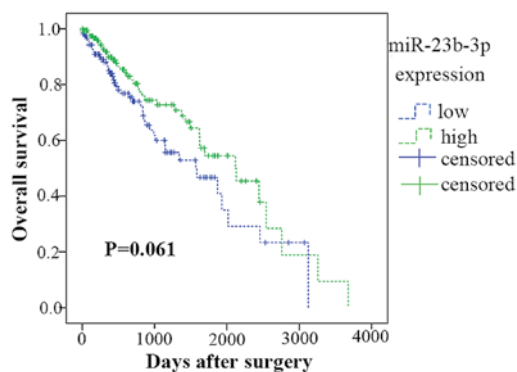


Figure 7. Kaplan-Meier curve for overall survival (months) of hepatocellular carcinoma (HCC) patients included in The Cancer Genome Atlas (TCGA) and grouped by the level of miR-23b-3p expression. There was no significant association between miR-203 expression and survival times in patients with HCC ($P=0.061$). Error bars represented standard deviation (SD).

follow-up. A Kaplan-Meier analysis is presented in Fig. 11. Among the 76 patients, 45 had a lower expression of miR-23b-3p (lower than the median level of 2.600), whereas 31 had higher miR-23b-3p expression. The lower expression group had 54.257 ± 2.943 months of survival time without recurrence; the higher expression group had 54.509 ± 3.962 months. Thus, there were no significant differences in survival time between high and low miR-23b-3p expression (Chi-Square =0.706, $P=0.401$).

Meta-analysis for GEO datasets, TCGA datasets and PCR verification in-house

Characteristics of included studies. Ten GEO datasets, TCGA dataset and the data derived from medical records, as mentioned above with 886 HCC patients and 587 normal persons in total, were included in the new meta-analysis. The characteristics of the studies were previously described (which were shown in sections ‘Characteristics of the included datasets’, ‘The expression level of miR-23b-3p was downregulated in HCC samples’ and ‘The expression of miR-23b-3p was downregulated in HCC’) in detail.

Value of miR-23b-3p as a biomarker for HCC. The pooled SMD and its 95% CI (Fig. 12A) indicated that people with lower expression of miR-23b-3p had a significant risk for HCC (SMD=-0.368; 95% CI, -0.689 to -0.048; $P=0.024$). A random effects model was selected to pool the effect variables with heterogeneity ($P=0.000$, $I^2=81.3\%$). The results of Begg’s and Egger’s test were 0.754 and 0.687, respectively (Fig. 12B and C); however, no publication bias was discovered. The sensitivity analysis indicated that our own PCR data exercised a certain influence on that result (Fig. 12D). Therefore, we removed PCR data and constructed the forest plot again (Fig. 13A). The SMD was -0.258 (95% CI, -0.529 to 0.014) and no statistical significance was found ($P=0.063$). The results of Begg’s and Egger’s test were 0.386 and 0.602, respectively (Fig. 13B and C); therefore, no publication bias was observed. The sensitivity analysis showed that the results were altered when we removed the study of TCGA dataset (Fig. 13D).

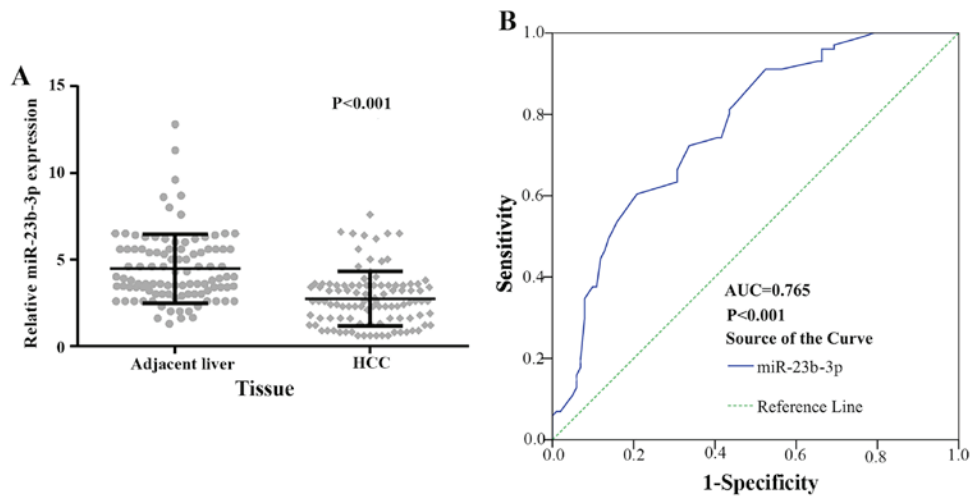


Figure 8. Expression of miR-23b-3p in hepatocellular carcinoma (HCC) and adjacent non-cancerous HCC. Quantitative real-time PCR was performed to detect the expression of miR-23b-3p. (A) The difference of relevant miR-23b-3p expression between HCC and adjacent non-cancerous HCC; (B) ROC curve of miR-23b-3p expression to distinguish HCC from comparative cancerous liver. Error bars represent standard deviation (SD).

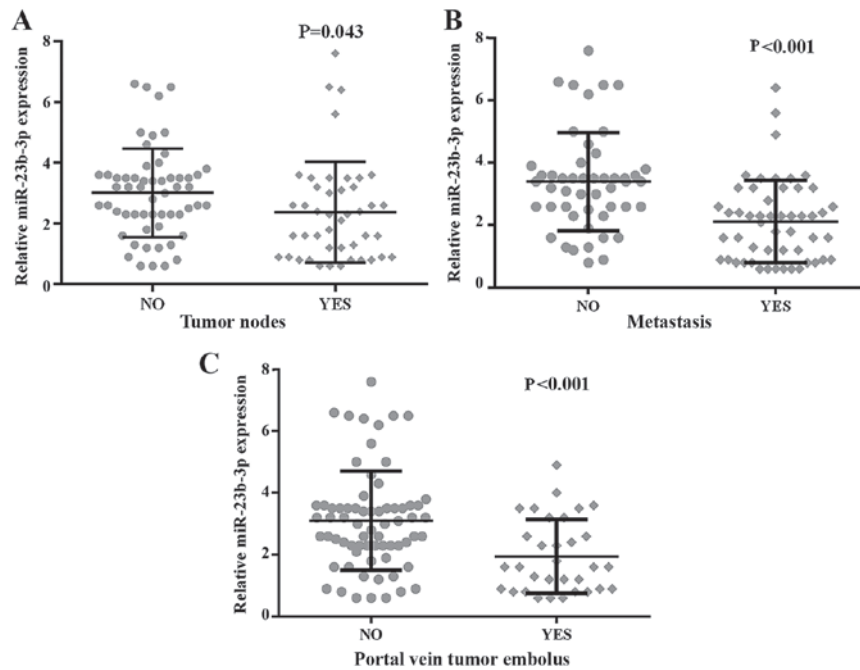


Figure 9. The relationship between miR-23b-3p expression and clinicopathological parameters of hepatocellular carcinoma (HCC). (A) tumor nodes, (B) metastasis; (C) portal vein tumor embolus. Error bars represent standard deviation (SD).

Bioinformatics analysis

Target genes of miR-23b-3p collection and integration. Using the 10 algorithms, 14,810 target genes were predicted to be modulated potentially by miR-23b-3p. The number of miR-23b-3p prediction targets of the 10 algorithms varied, ranging from 74 to 6,192 with an average of 2,375. Only targets that appeared more than or equal to five times among all 10 algorithms were finally applied for further analysis. In total, 357 targets from the algorithms were included in the next step.

After screening 203 articles from PubMed, we ultimately gathered 69 validation targets and found that 21 targets (ATG12, CA2, CFL2, CHUK, GLS, HMGB2, IL6R, KIAA1467, MET, NOTCH2, PLA2, PNR2, PRDM1, PTEN, PTK2B, RRAS2, SEMA6D, SPRY2, TAB3, VHL and WBP2) were duplicated

with prediction targets. By combining targets from algorithms and articles, we finally used 405 targets to conduct the bioinformatics analysis.

Bioinformatics analysis of miR-23b-3p targets. Using GO analysis, 405 targets regulated by miR-23b-3p were classified as biological processes, cellular components, and molecular functions (Table IV and Fig. 14). In the pathway analysis, targets of miR-23b-3p were primarily enriched in the signaling pathways of renal cell carcinoma, hepatitis B (shown in http://www.genome.jp/kegg-bin/show_pathway?hsa05161/hsa:6777%09red/hsa:4318%09red/hsa:8503%09red/hsa:353376%09red/hsa:596%09red/hsa:317%09red/hsa:1147%09red/hsa:5728%09red/hsa:6714%09red/hsa:4214%09red/hsa:355%09red/hsa:4088%09red/hsa:4089%09red/hsa:2185%09red/hsa:1869%09red/

Table II. Characteristics of the HCC patients included in TCGA.

Clinicopathological features	N	miR-23b-3p relevant expression ($2^{-\Delta C_q}$)			Correlation	
		Mean \pm SD	t-value	P-value	r-value	P-value
Tissue						
Normal	50	13.4039 \pm 0.51072	-8.286	<0.001	0.246	<0.001
HCC	371	12.6722 \pm 0.97815				
Age (years)						
<60	166	12.7336 \pm 0.97231	0.85	0.396	-0.045	0.396
\geq 60	194	12.6464 \pm 0.9000				
Sex						
Male	246	12.6581 \pm 0.93294	-0.613	0.54	0.032	0.54
Female	115	12.7259 \pm 1.0132				
Neoadjuvant treatment						
No	359	12.6798 \pm 0.97924	0.036	0.972	-0.002	0.972
Yes	2	12.6550 \pm 1.07244				
Radiation therapy						
No	231	12.6570 \pm 0.95382	0.998	0.319	-0.065	0.319
Yes	4	12.1762 \pm 1.03021				
Pharmaceutical treatment						
No	218	12.6475 \pm 0.96107	0.041	0.967	-0.003	0.967
Yes	12	12.6358 \pm 0.90350				
Alcohol consumption						
-	226	12.7394 \pm 0.99963	1.374	0.17	-0.074	0.17
+	117	12.5870 \pm 0.92032				
Hepatitis B						
-	237	12.5999 \pm 0.97009	-2.506	0.013	0.134	0.013
+	106	12.8831 \pm 0.96070				
Hepatitis C						
-	289	12.6963 \pm 0.97544	0.388	0.698	-0.021	0.698
+	54	12.6401 \pm 0.97811				
Non-alcoholic fatty liver						
-	324	12.6763 \pm 0.98327	-0.874	0.383	0.047	0.383
+	19	12.8774 \pm 0.81188				
Smoking						
-	326	12.7127 \pm 0.96181	2.115	0.035	-0.114	0.035
+	17	12.2025 \pm 1.11847				
Cirrhosis						
-	337	12.6956 \pm 0.95014	0.564	0.597	-0.063	0.248
+	6	12.2312 \pm 2.01359				
Grade						
GI-II	23	12.6505 \pm 0.76988	1.493	0.147	-0.214	0.256
GIII-IV	7	12.0712 \pm 1.26499				
Vascular invasion						
No	199	12.8350 \pm 0.94514	2.149	0.032	-0.122	0.032
Yes	107	12.5905 \pm 0.95709				
Recurrence after treatment						
No	166	12.5805 \pm 0.98050	-0.589	0.556	0.037	0.556
Yes	94	12.6537 \pm 0.92925				
Survival status						
Dead	86	12.6050 \pm 1.01283	-0.811	0.418	0.043	0.418
Alive	275	12.7031 \pm 0.96776				

Table II. Continued.

Clinicopathological features	N	miR-23b-3p relevant expression ($2^{-\Delta Cq}$)			Correlation	
		Mean \pm SD	t-value	P-value	r-value	P-value
T status						
T1	177	12.7653 \pm 0.93707	F=1.605 ^a	0.202 ^a	-0.083	0.117
T2-T4	181	12.5926 \pm 1.01626				
TX	1	12.0451 \pm 0.00000				
N status						
N0	247	12.7780 \pm 0.96867	F=5.116 ^a	0.006 ^a	-0.158	0.003
N1	3	11.7684 \pm 0.75393				
NX	110	12.4733 \pm 0.96540				
M status						
M0	262	12.7389 \pm 0.96998	F=1.778 ^a	0.170 ^a	-0.101	0.055
M1	4	12.6180 \pm 0.12946				
MX	95	12.5190 \pm 1.00762				

^aOne-way analysis of variance (ANOVA) test was utilized. t, Student's t-test; SD, standard deviation. HCC, hepatocellular carcinoma; T, tumor; N, nodes; M, metastasis. TCGA, The Cancer Genome Atlas.

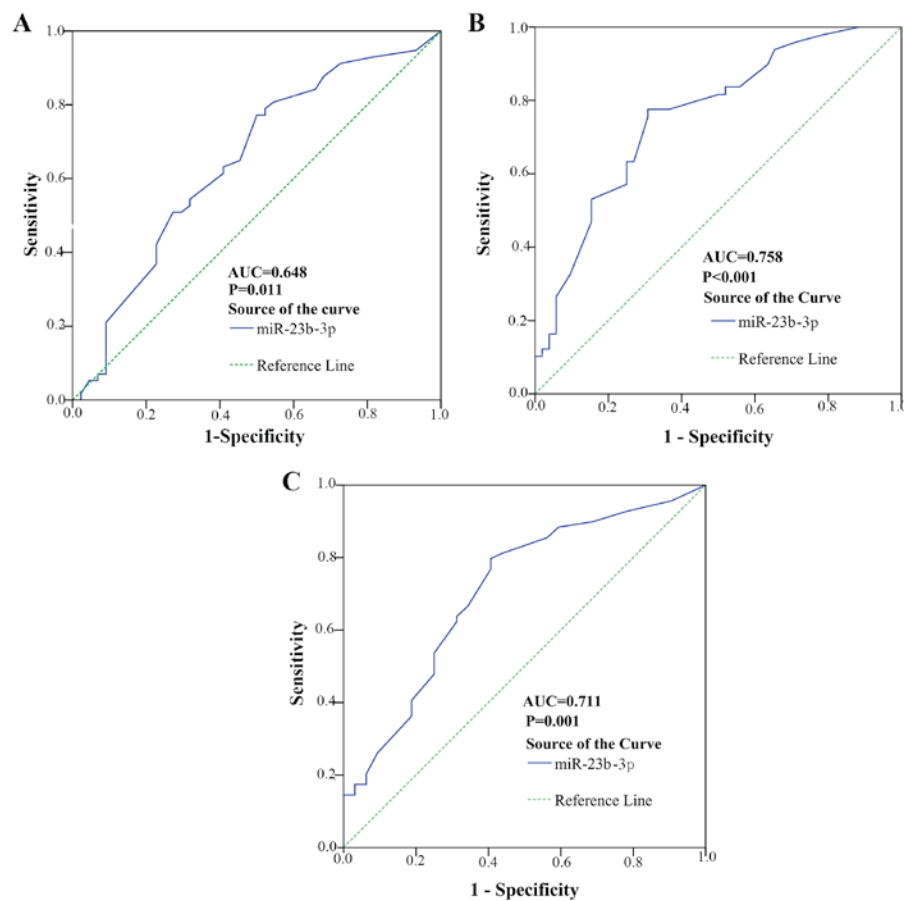


Figure 10. Diagnostic significance between miR-23b-3p and various parameters for hepatocellular carcinoma (HCC). (A) tumor nodes; (B) metastasis; (C) portal vein tumor embolus. Error bars represent standard deviation (SD).

hsa:207%09red) and pancreatic cancer (corrected $P < 0.05$). In the protein-protein interaction (PPI) network for miR-23b-3p (Fig. 15), 338 target genes were identified to be involved in 1,074 nodes. A

total of 8 targets, including SRC, AKT1, EGFR, CTNBN1, BCL2, SMAD3, PTEN and KDM6A, were located in the key nodes with high degree (>35).

Table III. The clinicopathological characteristics related to miR-23b-3p expression in patients.

Clinicopathological features	N	miR-23b-3p relevant expression ($2^{-\Delta C_q}$)			Correlation	
		Mean \pm SD	t-value	P-value	r-value	P-value
Tissue						
Adjacent non-cancerous	101	4.4694 \pm 1.99328	-6.847	<0.001	0.46	<0.001
HCC	101	2.7376 \pm 1.57739				
Age (years)						
<50	50	2.7160 \pm 1.53973	0.136	0.892	-0.019	0.85
\geq 50	51	2.7588 \pm 1.62852				
Sex						
Male	80	2.7162 \pm 1.55445	-0.265	0.792	0.057	0.568
Female	21	2.8190 \pm 1.69930				
Cirrhosis						
-	54	2.8426 \pm 1.53274	-0.715	0.476	0.112	0.256
+	47	2.6170 \pm 1.63539				
Tumor size						
<5 cm	21	2.5905 \pm 1.59371	-0.478	0.633	0.067	0.505
\geq 5 cm	80	2.7762 \pm 1.58092				
Tumor nodes						
Single	57	3.0158 \pm 1.45979	2.049	0.043	-0.255	0.01
Multiple	44	2.3773 \pm 1.66606				
Differentiation						
High	7	2.9857 \pm 1.65673	F=0.394 ^a	0.675 ^a	-0.044	0.66
Moderate	64	2.8062 \pm 1.71722				
Low	30	2.5333 \pm 1.23995				
Clinical TNM stage						
I-II	25	3.0520 \pm 1.9796	1.151	0.253	-0.192	0.055
III-IV	76	2.6342 \pm 1.67766				
Metastasis						
-	49	3.3959 \pm 1.57506	4.435	<0.001	-0.447	<0.001
+	52	2.1173 \pm 1.31762				
Portal vein tumor embolus						
-	69	3.1029 \pm 1.60797	3.618	<0.001	-0.34	0.001
+	32	1.9500 \pm 1.19055				
Tumor capsular infiltration						
With complete capsule	49	2.7449 \pm 1.60663	0.045	0.964	0.001	0.989
Infiltration or no capsule	52	2.7308 \pm 1.56500				
Vaso-invasion						
-	63	2.9635 \pm 1.63917	1.876	0.064	-0.169	0.092
+	38	2.3632 \pm 1.41123				

^aOne-way analysis of variance (ANOVA) test was utilized; t, Student's t-test; SD, standard deviation; HCC, hepatocellular carcinoma; TNM, tumor node metastasis; ANOVA, analysis of variance.

Discussion

HCC is one of the most prevalent forms of liver cancer, and nearly one million cases of HCC occur annually. Thus, it is urgent to explore the mechanism of HCC tumorigenesis and progression and to determine treatment and prevention strategies. Exploiting a new self-assembled cell microarray, Zhang *et al* carried out

high-throughput screening for miRNAs that are involved in cell transplantation. They discovered miR-23b-3p, which functioned as a tumor suppressor in human colon cancer (27). Since then, substantial research has studied the functions of miR-23b-3p in many physiological and pathological processes, especially tumor progression. As noted above, several studies showed that miR-23b-3p played a dual role in various cancers; these studies

Table IV. Four hundred five genes were categorized in GO as biological processes, cellular components and molecular functions.

Term	Count	P-value	FDR
Biological process			
Regulation of transcription	111	3.87E-10	6.73E-07
Regulation of transcription from RNA polymerase II promoter	47	1.98E-09	3.44E-06
Regulation of transcription, DNA-dependent	83	2.93E-09	5.10E-06
Regulation of RNA metabolic process	83	8.33E-09	1.45E-05
Transcription	90	3.70E-08	6.44E-05
Positive regulation of transcription from RNA polymerase II promoter	29	1.03E-07	1.80E-04
Negative regulation of gene expression	34	2.12E-07	3.68E-04
Positive regulation of gene expression	36	6.48E-07	0.001128
Positive regulation of transcription	35	9.33E-07	0.001624
Regulation of cell motion	19	1.07E-06	0.001868
Protein kinase cascade	27	1.19E-06	0.002065
Positive regulation of transcription, DNA-dependent	31	1.79E-06	0.003114
Positive regulation of RNA metabolic process	31	2.12E-06	0.003697
Positive regulation of nitrogen compound metabolic process	37	2.57E-06	0.004477
Regulation of cell proliferation	42	2.93E-06	0.005101
Regulation of cell migration	17	3.52E-06	0.006119
Positive regulation of macromolecule biosynthetic process	37	3.65E-06	0.006356
Positive regulation of cellular biosynthetic process	38	4.10E-06	0.007132
Positive regulation of macromolecule metabolic process	44	4.31E-06	0.007507
Regulation of locomotion	18	4.36E-06	0.007589
Response to hypoxia	15	4.57E-06	0.007949
Positive regulation of biosynthetic process	38	5.68E-06	0.009878
Negative regulation of transcription	29	7.16E-06	0.012463
Response to oxygen levels	15	8.29E-06	0.014432
Positive regulation of nucleobase, nucleoside, nucleotide and nucleic acid metabolic process	35	8.58E-06	0.014931
Negative regulation of macromolecule biosynthetic process	32	1.01E-05	0.01756
Response to hormone stimulus	25	1.05E-05	0.018246
Positive regulation of cell differentiation	19	1.21E-05	0.021016
Response to endogenous stimulus	26	1.84E-05	0.032008
Embryonic organ development	16	1.91E-05	0.033319
Cell morphogenesis	24	1.96E-05	0.034027
Positive regulation of cell motion	12	2.46E-05	0.042821
Negative regulation of biosynthetic process	32	2.51E-05	0.04366
Cellular component			
Nuclear lumen	54	1.14E-05	0.015418
Nuclear envelope	16	1.76E-05	0.023821
Molecular function			
Transcription regulator activity	70	6.80E-07	9.94E-04
DNA binding	94	2.12E-06	0.0031
Transcription factor activity	48	1.39E-05	0.020262

FDR, false discovery rate.

also investigated the mechanisms of miR-23b-3p. This study focused on determining the role of miR-23b-3p in HCC and its effect on patient prognosis.

In the meta-analysis of data downloaded from GEO datasets, the pooled SMD indicated that miR-23b-3p expression

showed no remarkable difference between HCC patients and normal subjects. No heterogeneity was noted. Additionally, based on GEO datasets, we investigated the association between miR-23b-3p and the clinicopathological aspects of HCC and observed that miR-23b-3p was significantly correlated with

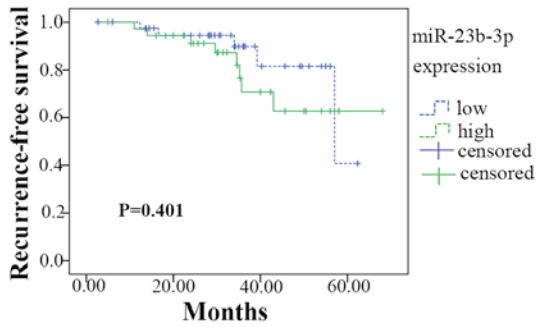


Figure 11. Kaplan-Meier curve for recurrence-free survival in hepatocellular carcinoma (HCC) patients grouped by the level of miR-23b-3p expression. There was no significant association between miR-203 expression and recurrence free survival in patients with HCC (P=0.401). Error bars represented standard deviation (SD).

invasion. We failed to prove the relevance of miR-23b-3p and other clinical parameters. Therefore, diagnosis and targeted therapy based on the miR-23b-3p expression level required further research.

We identified miR-23b-3p as a HCC biomarker by analyzing TCGA data. miR-23b-3p expression was obviously down-regulated in HCC tissues compared to normal liver tissues. Additionally, miR-23b-3p expression was lower (P<0.05) in patients with hepatitis B, smoking history, vascular invasion and lymphatic metastasis. However, only hepatitis B history and lymphatic metastasis of HCC revealed diagnostic values. HCC patients with high miR-23b-3p expression presented longer survival; however, this finding was non-significant.

According to the results above, we propose that miR-23b-3p may play a crucial part in HCC tumorigenesis and

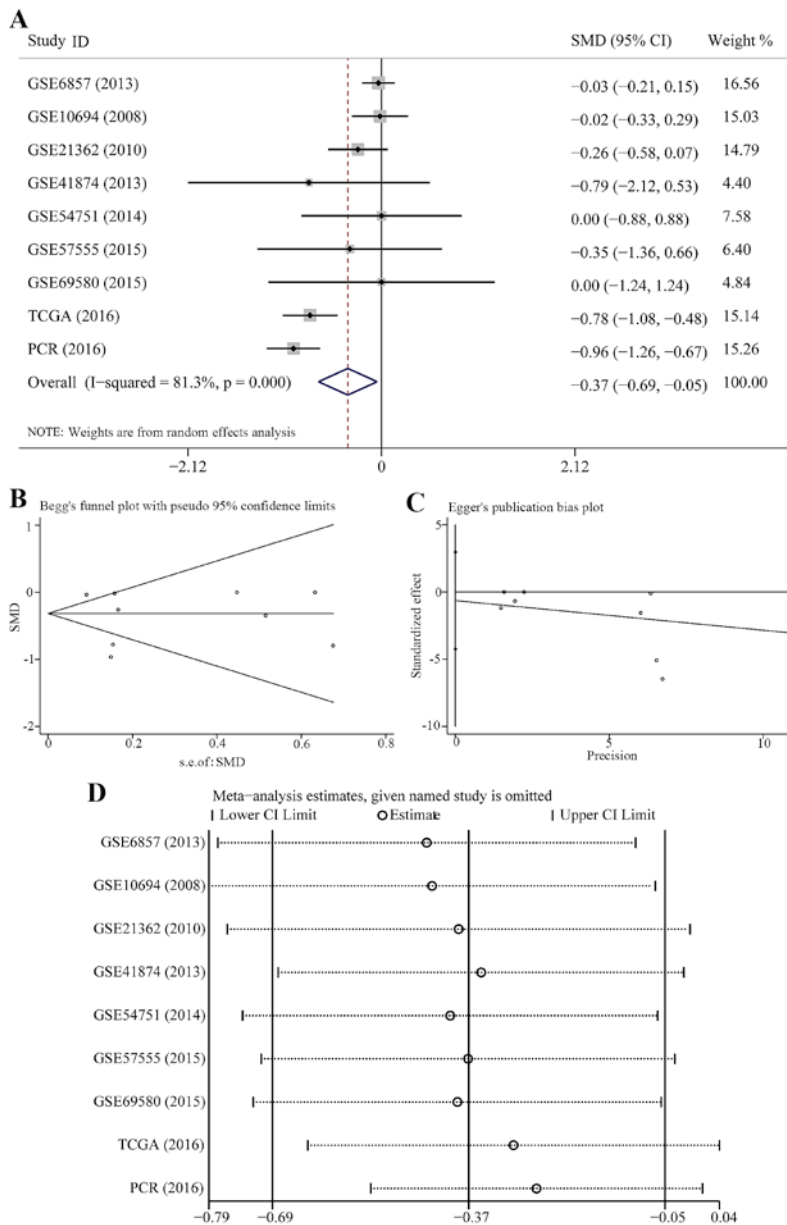


Figure 12. Meta-analysis of miR-23b-3p expression in hepatocellular carcinoma (HCC) patients and healthy controls retrieved from Gene Expression Omnibus (GEO) datasets, The Cancer Genome Atlas (TCGA) datasets and PCR data. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg's funnel plot. Hazard ratios are presented on a logarithmic scale. (C) Egger's publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis. Pooled SMDs and 95% CIs by eliminating each study.

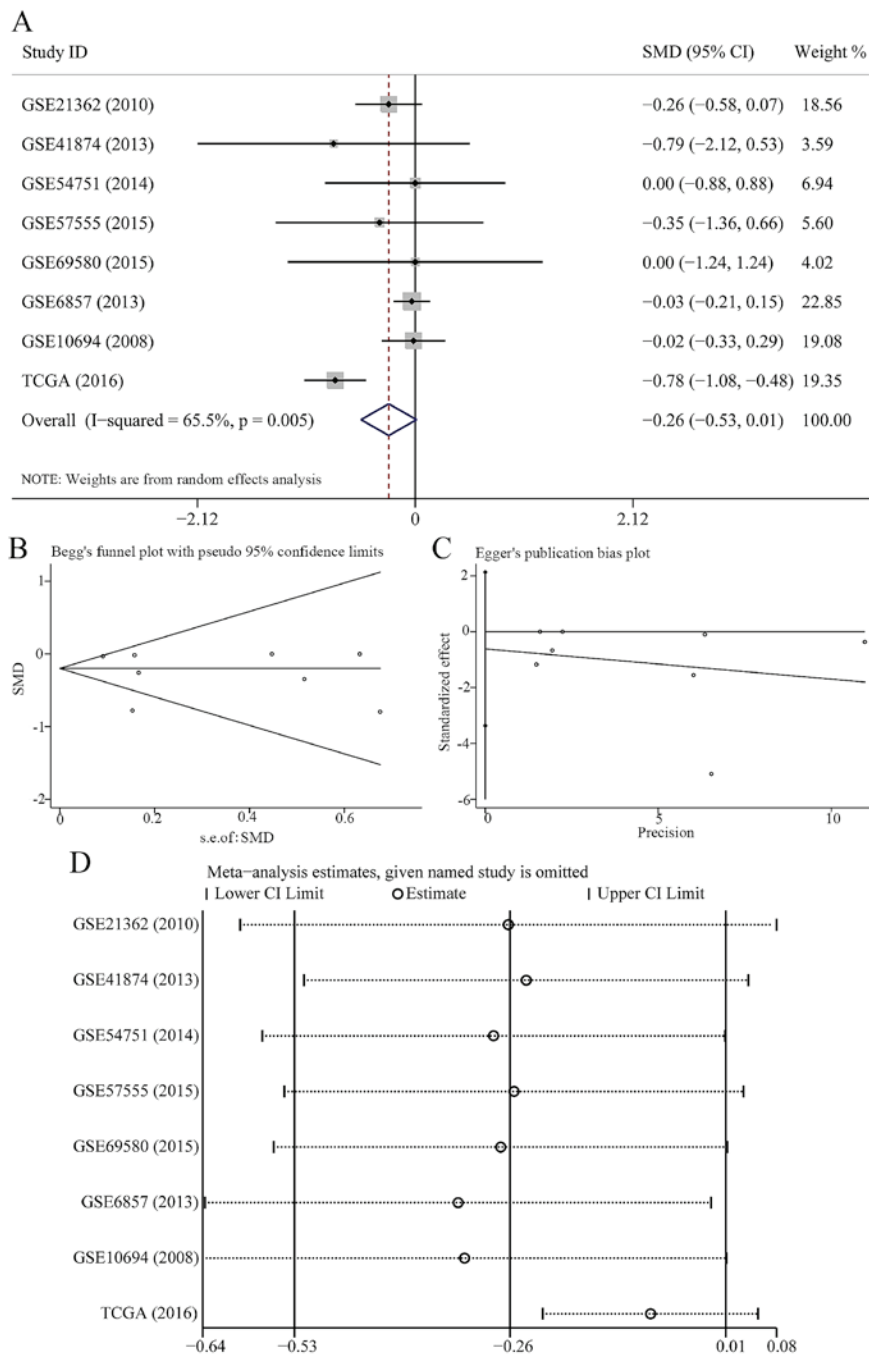


Figure 13. Meta-analysis of removing PCR data from all the studies. (A) Forest plot. The center dot and the horizontal line represent the standard mean difference (SMD) and 95% confidence interval (CI), respectively. The pooled SMD and its corresponding 95% CI are presented below the list of studies; (B) Begg's funnel plot. Hazard ratios are presented on a logarithmic scale. (C) Egger's publication bias plot. Hazard ratios are presented on a logarithmic scale; (D) sensitivity analysis. Pooled SMDs and 95% CIs by eliminating each study.

development. Therefore, we recruited 101 fresh HCC tissues and counterpart adjacent non-cancerous HCC to inspect the expression of miR-23b-3p in HCC tissues and its association with the clinicopathological aspects and prognosis of HCC patients.

However, in contrast with the counterpart adjacent non-cancerous HCC, we observed that miR-23b-3p was aberrantly downregulated in HCC tissue ($P < 0.001$). This consequence was supported by a statistical analysis of TCGA data but in contrast to the microarray meta-analysis. Additionally, HCC patients with multiple tumor nodes showed a lower level of miR-23b-3p expression than those with single

tumor nodes ($P < 0.05$). In addition to other parameters that indicated HCC progression, the deterioration of metastasis and the portal vein tumor embolus were strongly associated with a decrease in miR-23b-3p ($P < 0.001$). In line with the aforementioned findings, miR-23b-3p was closely linked to HCC aggressiveness and metastasis. Additionally, Salvi *et al* transfected miR-23b-3p into HCC cells (SKHep1C3), which resulted in a reduction in motility and proliferative potential; they also found that miR-23b-3p was deregulated in HCC (28). To some extent, our study also revealed that increased miR-23b-3p was associated with an increased time-to-recurrence (months) in HCC patients. However, there was no obvious alteration

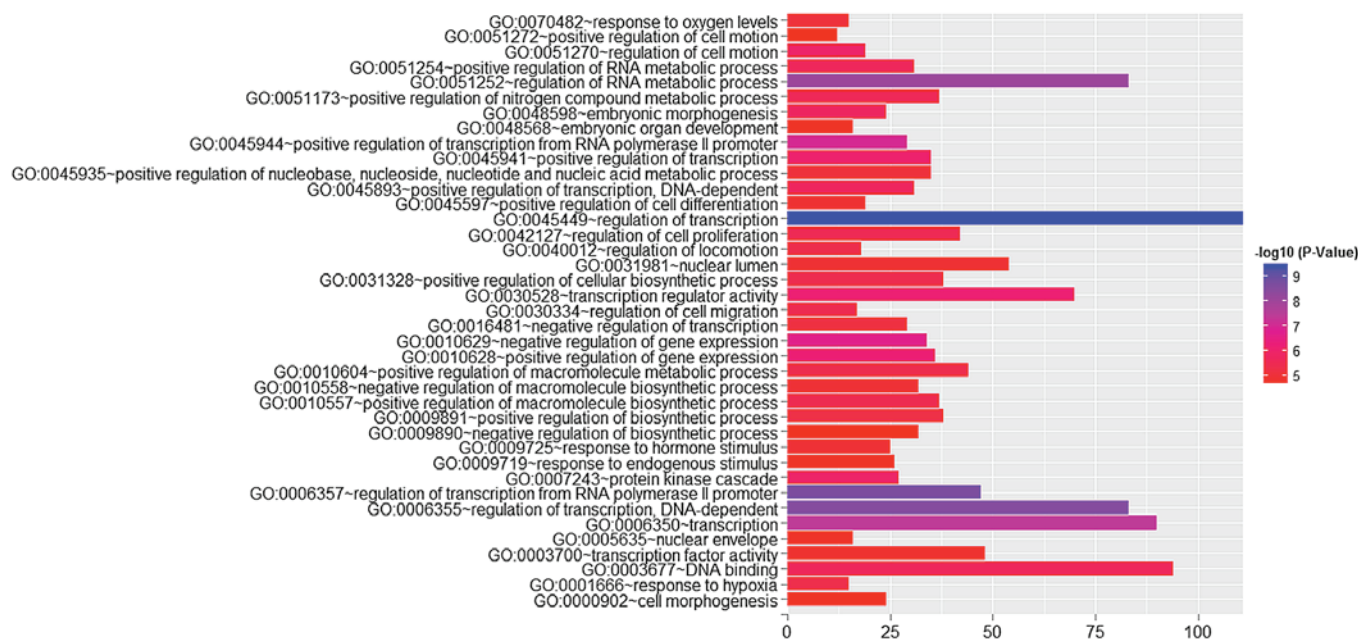


Figure 14. GO analysis for target genes of miR-23b-3p. The length of horizontal column represents the number of genes and their color stand for P-value.

between the two groups concerning the levels of miR-23b-3p. The lack of a study population may explain this result. Thus, a larger cohort of patients is required to study this in the near future. Additionally, only tumor nodes, metastasis portal vein tumor embolus and EGFR expression of HCC were found to correlate with miR-23b-3p expression and presented diagnostic values.

Altogether, the data derived from GEO datasets, TCGA datasets and qPCR were included to perform a comprehensive meta-analysis. The pooled SMD indicated that pronounced downregulation of miR-23b-3p expression was noted in HCC patients as compared to healthy subjects, which indicated that low level of miR-23b-3p may play an important role in HCC tumorigenesis and diagnosis. Nevertheless, high heterogeneity indicated that this conclusion was less reliable. According to sensitivity analysis, the data from TCGA and qPCR contributed to the high heterogeneity. The difference of source of specimen and the detection methods of miR-23b-3p expression may be the major factors.

Ten prediction algorithms (utilizing different matching criteria and computational algorithms) were employed in miR-23b-3p target gene prediction. In contrast to the algorithms that provided validated targets and literature extraction, the prediction algorithms could not predict with accuracy. Some factors, such as varying approaches and rules for miRNA targeting, can affect the target prediction results. To reduce the defects associated with target prediction, we combined the results of these algorithms and stipulated that only the genes that appeared no fewer than five times would be involved in the next analysis.

In addition to targets based on experimental validation, prediction targets combined with algorithms were enrolled for GO analysis, pathways analysis and network interaction. A bioinformatics analysis predicted that miR-23b-3p was involved in extensive target regulation. Because these targets took part in multiple phases of tumorigenesis and tumor

progression, we speculated that miR-23b-3p was dysregulated in various tumors and led to changes in biological characteristics by regulating a series of target genes in different phases. In a GO analysis, the targets of miR-23b-3p were involved in biological processes (like the regulation of transcription and transcription), cellular components (such as the nuclear lumen and nuclear envelope) and molecular functions (such as transcription regulator activity and DNA binding). These targets play a vital role in major cell processes, and a number of them participate in the carcinogenesis of various cancers, including HCC. According to KEGG analysis, miR-23b-3p participated in the hepatitis B pathway, which was recognized as a key factor of hepatic cirrhosis. Moreover, hepatic cirrhosis could result in hepatic carcinoma. We also found that miR-23b-3p was correlated with hepatitis B in the HCC patients who were included in TCGA. Thus, we speculated that miR-23b-3p may play an important role in HCC tumorigenesis.

A network analysis was ultimately conducted to predict the interaction networks of potential targets. Highly connected genes were regarded to play critical roles in stabilization, interaction, and gene network regulation. A connectivity analysis showed the highest connectivity between SRC and AKT1. SRC is a proto-oncogene encoding a tyrosine-protein kinase that belongs to Src family kinases, which are considered vital to cell proliferation, differentiation, apoptosis and invasion (29,30). Src family kinases have been shown to play a vital role in several malignancies, including breast cancer (30), colon cancer (31), ovarian cancer (32) and HCC (33), as well as melanoma, glioma, and various types of sarcoma (34). Akt1 or protein kinase B (PKB) α , which is regularly activated in human cancers, can phosphorylate downstream molecules and cope with a wide range of cell processes (35). Moreover, Akt1 was directly or indirectly controlled by Src kinase activity (35,36). We conjectured that Src kinase and Akt1 may function in the same signal transduction pathways that result in carcinogenesis and development. This suggests that

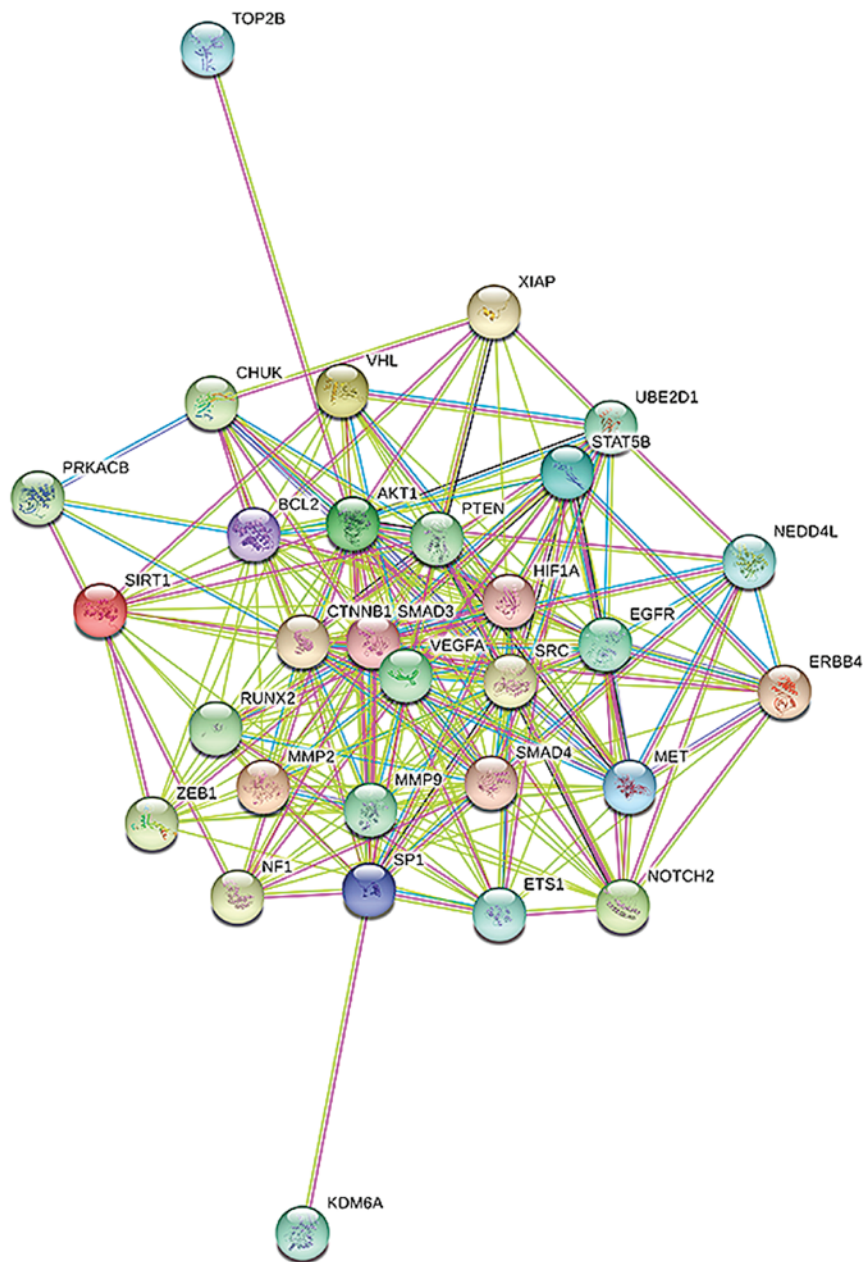


Figure 15. Network analysis with the prospective target genes of miR-23b-3p. Network nodes represent proteins. Colored nodes, query proteins and first shell of interactors; white nodes, second shell of interactors. Small nodes, protein of unknown 3D structure; large nodes, some 3D structure is known or predicted.

researchers should consider factors related to the hub genes for improving diagnosis and treatment.

Inquiries in various cancers indicated that miR-23b-3p, functioning as an oncogenic miRNA, can accelerate tumor processes such as proliferation and invasion. Jin *et al* reported that miR-23b/27b expression was elevated in breast cancer and that it indicated a poor prognosis (12). Nischarin partially participated in reducing the level of miR-23b/27b expression by inhibiting NF- κ B phosphorylation and disturbing cancer aggressiveness *in vivo* (2). Additionally, the AKT/NF- κ B pathway, which responds to TNF α and EGF, is the pivotal signaling cascade for Her2/neu-dependent miR-23b/27b *in vitro* (2). Li *et al* demonstrated that Fas mRNA, which enhances the cell multiplication and apoptosis rates, was directly silenced by miR-23b-3p upregulation in thymic lymphoma cells (37).

Many studies also noted that the role of miR-23b-3p as a tumor suppressor was downregulated in several cancers. Majid *et al* discovered that miR-23b-3p was downregulated in prostate cancer, and its decreasing function of directly inhibiting proto-oncogene Src kinase may lead to the acceleration of neoplasm growth (38). The same author in another study stated that miR-23b-3p played a key role in migration of bladder cancer cells via posttranscriptionally directly regulating at Zeb1, which could modulate the epithelial-to-mesenchymal transition (EMT) (39). All of the mechanisms by which miR-23b-3p acts as a tumor suppressor may contribute to HCC aggressiveness; further studies are required.

Furthermore, extensive study suggested that miR-23b-3p was associated with other clinicopathological features in HCC. Bisio *et al* confirmed the cis-mediated regulation of miR-23 by p53 in the MCF7 cell line (40). However, we

detected the reverse (upregulated miR-23b-3p) in p53-positive tissue ($P > 0.05$). There were two possible reasons for this. First, miR-23b-3p served as oncogenic miRNA in the MCF7 cell line but acted as a tumor suppressor in HCC. Notably, the specific mechanism remained unclear. Secondly, our data was less trustworthy due to its lack of significance. Chen *et al* reported that the downregulated expression of miR-23b-3p inhibited HIF-1 α /VEGF and β -catenin/Tcf-4 signaling by enhancing the level of VHL (41). The quantity of VEGF in our study was also identical with miR-23b-3p expression; however, this was without significance ($P > 0.05$). Patients with occult hepatitis virus B infection (OBI), who are generally negative for HBsAg, were discovered to have a differential expression of miR-23b-3p (42). Notably, as Salvi *et al* reported, miR-23b-3p overexpression contributed to a decrease in urokinase-type plasminogen (uPA) and c-met. This was determined by identifying the 3'-UTR of uPA and c-met in two and four sites, respectively, as well as by observing a reduction in cell proliferation and metastasis in HCC (28). The present study partially supports the above study. To our knowledge, the present study is also the first one to explore the clinicopathological significance of miR-23b-3p and contributed to a more comprehensive understanding of the correlation between miR-23b-3p and HCC progression.

Expression of miR-23b-3p was studied in HCC and their relationship explored. A preliminary analysis of the biological characteristics and function of miR-23b-3p was performed via bioinformatical methods. Despite existing limitations, miR-23b-3p was thought to be a tumor suppressor affecting the carcinogenesis and aggressiveness of HCC and may represent a predictive biomarker and therapeutic target for HCC. These results may contribute to a theoretical basis for HCC diagnosis and therapy.

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Availability of data and material

All data generated or analyzed during this study are included in this published article.

Authors' contributions

RQH and PRW designed the study, did the experiments, analyzed and interpreted the data, and wrote the manuscript. They contributed equally to this work. XLX, XY and HWL performed the statistical analysis, designed and filled-out the figures and tables. XHQ, LHY, ZGP recruited the specimens, carried out the miRNA isolation and real-time RT-qPCR. LHY, ZGP and GC participated in designing the study, supervised

all experiments and corrected the manuscript. All the authors read and approved the final manuscript.

Ethics approval and consent to participate

All experiments referred to patient tissues were authorized by the Ethics Committee of the First Affiliated Hospital of Guangxi Medical University (no. 2016-KY-NSFC-094). This study was a sub-study of the project mentioned in ethics approval. Method and participants are utilized in the same way.

Consent for publication

The consents for publication were obtained from all the patients.

Competing interests

The authors declare that they have no competing interests.

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