

From Neurotoxicity to Neuroprotection: Rethinking GABAAR-

2	Targeting Anesthetics
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ABSTRACT

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Neuroactive Steroids

The brain growth spurt (BGS) represents a pivotal window in neurodevelopment, defined by rapid neurogenesis, heightened synaptogenesis, and the dynamic establishment of neural networks. During this phase, heightened brain plasticity significantly enhances learning and memory abilities, while also increasing the brain's susceptibility to disruptions. Anesthetics, particularly those targeting yaminobuturic acid type A receptors (GABAARs), interfere with GABAergic and glutamatergic systems, disrupt brain-derived neurotrophic factor (BDNF) signaling, and exacerbate neurotoxic effects. These agents activate glial cells, induce inflammation, and contribute to oxidative stress, while also disrupting calcium homeostasis and triggering endoplasmic reticulum stress. Furthermore, anesthetics alter the expression of non-coding RNAs, which affects gene regulation and long-term memory formation. The extent of neurotoxic effects is contingent upon a constellation of factors, including the timing, dosage, and frequency of anesthetic exposure, as well as individual susceptibility. Notably, perioperative administration of anesthetic agents has been implicated in long-term cognitive dysfunction, thereby emphasizing the critical importance of precisely modulated dosing regimens and temporally optimized delivery strategies to mitigate potential neurodevelopmental risks. In contrast, neuroactive steroids demonstrate promising neuroprotective potential by modulating GABAAR activity, enhancing BDNF release, and regulating oxidative stress and inflammation. New strategies for preventing and reversing anesthetic-induced neurotoxicity could include novel anesthetic combinations, anti-apoptotic agents, antioxidants, or nutritional supplements. These findings underscore the complex and multifactorial effects of anesthetic agents on the developing brain and emphasize the urgent need to establish and refine anesthetic strategies that safeguard neural integrity during vulnerable windows of neurodevelopment. Keywords Anesthetics Targeting GABAARs; Neurodevelopment; Neurotoxicity; Neuroprotective;

1. Introduction

A growing body of evidence derived from both animal models and clinical research suggests that the postnatal BGS represents a critical period in neurodevelopment, during which the central nervous system displays exceptional plasticity and heightened susceptibility to external perturbations. During this critical developmental window, neuronal circuits are sculpted through the rapid generation and remodeling of trillions of synapses, accompanied by dynamic neuronal migration, connectivity establishment, and functional maturation(Maksimovic et al., 2022). This crucial period is characterized by an accelerated process of neurogenesis, gliogenesis, and synaptogenesis, which collectively contribute to the brain's high plasticity and vulnerability(Fan et al., 2021; Vanderhaeghen & Polleux, 2023). At this developmental stage, the brain exhibits heightened sensitivity to disturbances arising from both environmental stimuli and pharmacological exposures.

 An expanding corpus of evidence derived from both invertebrate and vertebrate models indicates that administration of general anesthetics during the BGS period can elicit extensive neurodevelopmental disruptions. These include the induction of neuronal apoptosis, inhibition of dendritic arborization, impairment of synaptogenesis, and disruption of synaptic plasticity, collectively attributing to persistent deficits in learning, memory, and long-term neurological function (Schaefer et al., 2019; Xu et al., 2018). It is now well-recognized that such adverse neurodevelopmental outcomes are mechanistically linked to the modulation of synaptic signaling by anesthetic agents. In particular, these drugs enhance inhibitory neurotransmission via GABAARs and/or suppress excitatory activity by antagonizing N-methyl-D-aspartate (NMDA) receptors—105 ations that critically impair synaptic formation and stability during key developmental periods (Forcelli et al., 2551; Zhou et al., 2011). In clinical practice, GABAAR agonists are more commonly administered than NMDA receptor antagonists such as ketazine or nitrous oxide. Frequently utilized GABAAR-targeing anesthetics encompass inhalational agents—such as sevoflurane, isoflurane—as well as intravenous drugs including propofol, etomidate, and a range of benzodiazepines (e.g., midazolam, diazepam, remimazolam) and barbiturates (e.g., thiopental, pentobarbital).

GABA_AR agonists such as propofol disrupt a critical developmental process in which GABA transitions from depolarizing to inhibitory action. By enhancing inhibitory signals, these agents disturb the balance between excitatory and inhibitory signaling, thereby compromising neural network stability and causing long-term damage to developing brain networks. Consequently, these drugs have been included on the FDA's warning list due to their potential neurotoxic risks to children(Food & Administration, 2017).

Neuroactive steroids (NAS), a subclass of endo genous steroid compounds, act as powerful allost modulators of GABA_ARs,s and play a pivotal role in modulating inhibitory neurotransmission within the central nervous system. While NAS agents such as CDNC, alphaxalone, and 3β-hydroxyandrostanes share sedative and anesthetic properties akin to propofol, they differ significantly in their neurotoxicity profiles. Notably, even at equivalent or supratherapeutic concentrations, these compounds do not elicit neuroapoptotic responses. In contrast, conventional anesthetics including isoflurane and propofol have

been shown to induce widespread neuronal apoptosis when administered during critical windows of brain development. These neuropsic effects are intimately linked to enduring impairments in synaptogenesis, reduced neural plasticity, and cognitive decline (Xu et al., 2018; Zhao et al., 2023). Such findings underscore the distinctive neurodevelopmental safety profile of NAS agents relative to traditional anesthetics like propofol(Atluri et al., 2018; Tesic et al., 2020). Furthermore, NAS agents demonstrate more pronounced neuroprotective properties, such as preventing neuronal apoptosis, protecting against neurodegenerative diseases, and exerting anti-neuroinflammatory effects(Diviccaro et al., 2022).

Given the divergent mechanisms of action and neurodevelopmental consequences observed between traditional intravenous anesthetics and emerging NAS, this review systematically examines the neurotoxic risks and neuroprotective potential of GABAergic intravenous anesthetics during early brain development. Particular attention is devoted to the reversibility and long-term consequences of developmental impairments resulting from anesthetic exposure. This discussion provides deeper insights into the molecular and cellular pathways implicated in anesthetic-induced neurotoxicity, while also exploring critical factors that modulate individual susceptibility to these adverse effects. Lastly, we evaluate emerging strategies aimed at preventing or attenuating anesthetic-related neurodevelopmental harm.

2. Neurotoxic Mechanisms of Anesthetics Targeting GABAARs

During key stages of neurode pment, the precisely coordinated activity of GABAergic and glutamatergic signaling cascades is essential for orchestrating neural stem cell proliferation, directing lineage-specific differentiation, and ensuring neuronal survival and maturation. General anesthetics, by profoundly modulating these excitatory and inhibitory neurotransmitter systems, have been demonstrated to disrupt the finely tuned synaptic equilibrium essential for normal brain development during these sensitive periods. This dysregulation is partially mediated by alterations in the NKCC1/KCC2 expression ratio, which governs intracellular chloride gradients and GABAergic polarity (see section 2.1), as well brough direct interference with the functional integrity of GABAARs and NMDA receptors(Cabrera et al., 2020). In addition, anesthetic agents have been reported to modulate the expression patterns of BDNF, a key neurotrophin involved in synaptic regulation, thereby influencing neuron-glia crosstalk and initiation downstream neuroinflammatory responses (Wan et al., 2021). Moreover, anesthetic agents influence the or ression of brain-derived neurotrophic factor (BDNF), thereby modulating the functional dynamics of both neurons and glial cells, and eliciting neuroinflammatory responses. This leads to calcium overload, enhanced oxidative stress, mitochondrial damage, endoplasmic reticulum stress, and alterations in non-coding RNAs (ncRNAs). These interacting mechanisms impair the proper assembly of neural circuits during development and may exert enduring detrimental effects on overall brain function(Fig. 1).

2.1 Disruption of Neurotransmitter Balance by Anesthetics

During early brain development, GABA is released prior to the functional maturation of glutamatergic synapses. In the neonatal cortex, high expression of the sodium-potassium-chloride cotransporter 1 (NKCC1) establishes a depolarizing chloride gradient, thereby rendering GABAergic

signaling excitatory during this critical developmental window. As the brain matures, the expression of potassium-chloride cotransporter 2 (1600) progressively increases, replacing NKCC1 activity and resulting in a developmental transition in GABA function—from excitatory to inhibitory. This dynamic transition between excitatory and inhibitory signals ensures that neurons receive sufficient excitatory inputs during development while progressively establishing inhibitory neural networks(Oh et al., 2016; Watanabe & Fukuda, 2015).GABAAR agonists, such as propofol and etomidate, may disrupt the excitation/inhibition (E/I) balance by increasing NKCC1 expression or inhibiting the maturation of KCC2(Ju et al., 2017). This disruption leads to chloride extrusion and membrane depolarization, ultimately resulting in neuronal hyperexcitability and cognitive dysfunction (DiGruccio et al., 2015). In addition to modulating neurotransmitter systems, general anesthetics have been shown to activate voltage-gated calcium channels—particularly L-type channels—thereby elevating intracellular calcium concentrations. This abnormal calcium influx disrupts neuronal calcium homeostasis, a fundamental regulator of cell viability and intracellular signaling, and initiates a cascade of pathological events that ultimately lead to neuro transfer eneration. These observations underscore calcium dysregulation as a central pathogenic mechanism in anesthetic-induced neurotoxicity (Miao et al., 2022; Schaefer et al., 2019; Soyalp et al., 2022). BZDs like midazolam and diazepam enhance central GABAergic function by binding to GABA_ARs and modulating downer am signaling pathways, including those involving protein kinase G and synaptic marker interactions(Fredriksson et al., 2007; Jevtovic-Todorovic et al., 2003).. For instance, midazolam requires higher doses to induce sedation in neonatal animals because, at elevated concentrations, the anesthetic's effect on KCC2 surpasses its impact on NKCC1, ultimately achieving sedation through neuronal hyperpolarization(Doi et al., 2021). However, excessive dosages increase the risk of neurotoxicity, particularly in infants, as anesthetic doses often exceed those used in adults, rendering them more susceptible to neuronal injury. In contrast, remimazolam, with its lower equivalent dosage, short half-life, and minimal accumulation, results in less memory impairment. This is attributed to the reduced affinity of its metabolites for GABAARs, which is approximately 1/400 of remimazolam's affinity, thereby decreasing NMDAR overexpression and alleviating synaptic damage(Shi et al., 2024).

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During intense synaptic activation, Glutamate (GLU) induces postsynaptic membrane depolarization by activating AMPA receptors, which relieves the magnesium block on NMDA receptors allows calcium ions to enter, triggering long-term potentiation (LTP). Subsequently, Postsynaptic density protein 95 (PSD95) functions as a pivotal structural scaffold at the synapse, anchoring NMDA AMPA receptors at the postsynaptic membrane, facilitating AMPA receptor insertion, and enabling the induction of long-term potentiation (LTP) and synaptic plasticity, thereby strengthening synaptic connectivity. Conversely, GABAAR activation has been shown to impair LTP induction—a fundamental cellular mechanism underpinning learning and memory—by attenuating GLU release at excitatory synapses. This inhibition of excitatory neurotransmission disrupts synaptic strengthening mechanisms essential for cognitive development. In vitro experiments indicate that propofol inhibits GLU release by suppressing presynaptic membrane depolarization, which reduces Na+ influgation while its effect on presynaptic Ca2+ influx and the reverse transport of transporters is not significant (Bademosi et al., 2018; Karunanithi et al., 2020). Evidence from animal models indicates that propofol, at concentrations consistent with clinical use, exerts direct modulatory effects on postsynaptic GABAARs, thereby enhancing inhibitory neurotransmission during critical stages of brain development(Li et al., 2020). At clinically relevant concentrations, both propofol and etomidate have been shown to interact with the vesicle docking protein syntaxin1A, the base by restricting its lateral mobility are impairing glutamate (GLU) exocytosis from glutamatergic neurons in the basal forebrain of mice (Bademosi et al., 2018; Karunanithi et al., 2020). This molecular interaction diminishes neuronal excitability and contributes to functional impairment in cortical networks. Thiopental, another intravenous anesthetic, has been implicated in neonatal neurodegeneration by inhibiting GLU synthesis and promoting apoptotic signaling via upregulation of pro-apoptotic proteins (Naseri et al., 2017). Concurrently, gestational exposure to sevoflurane has been linked to sustained downregulation of key synaptic proteins—most notably PSD95 and synaptophysin—in the hippocampus of both fetal and postnatal offspring, suggesting long-lasting impairments in synaptic architecture and integrity (Zheng et al., 2013). This effect may involve modulation of the ubiquitin—proteasome pathway. Impaired PSD95 expression destabilizes postsynaptic AMPA receptor anchoring, enhancing receptor internalization through clathrin-mediated endocytosis. These alterations compromise hippocampal synaptic potentiation and are associated with memory deficits in rodent behavioral paradigms (Liao et al., 2021; Lu et al., 2017).

2.2 Impact of Anesthetics on BDNF

BDNF, a pivotal modulator of neurodevelopment, governs fundamental processes including neuronal differentiation, axonal and dendritic growth, and synaptic plasticity. Initially synthesized as the precursor protein proBDNF, BDNF is predominantly stored within synaptic vesicles and subsequently converted into its mature form (mBDNF) hrough proteolytic processing—a transformation essential for synaptic remodeling and the refinement of neural circuits during critical windows of brain development. Binding of proBDNF to its receptor p75^{NTR} activates the RhoA signaling cascade, resulting in axonal elongation and growth cone collapse(Head et al., 2009). During neurodevelopment, BDNF regulates the development of GABAergic, glutamatergic, and other synaptic types to ensure the precise establishment and stability of neural networks in the brain(Szymanski & Minichiello, 2022).

General anesthetics such as propofol significantly affect BDNF and its associated signaling pathways by modulating GABAergic and glutamatergic neurotransmission, thereby disrupting neuroplasticity(Cohen-Cory et al., 2010). Experimental studies have demonstrated that propofol attenuates the signaling cascade of BDNF and its receptor TrkB, thereby leading to reduced expression of PSD95 and impairing synaptic plasticity (Wan et al., 2021). Simultaneously, propofol activates the P75^{NTR}/RhoA/ROCK pathway and GABAARs, increasing the ratio of proBDNF to mBDNF, disrupting the homeostasis of neurotrophic factor signaling, and exacerbating neuronal apoptosis, leading to neurological dysfunction(Kahraman et al., 2008). Furthermore, propofol significantly suppresses the expression of BDNF, B-cell lymphoma 2 (BCL-2), and phosphorylated cAMP response element-binding protein (p-CREB) in hippocampal neurons, thereby intensifying its neurotoxic impact(Wei et al., 2016).

Within the context of early neural development, repeated exposure to sevoflurane disrupts the equilibrium between tissue plasminogen activator and its endogenous inhibitor PAI-1, thereby impairing the fibrinolytic cascade and compromising synaptic plasticity. This dysregulation subsequently hinders

the activation of TrkB signaling and reduces the proteolytic processing of proBDNF into mBDNF(Dong et al., 2016). In parallel, sevoflurane engages the proBDNF–p75 neurotrophin receptor (p75^{NTR}) axis to trigger the JNK/c-JUN/AP-1 cascade (Bi et al., 2018), while simultaneously promoting neurodegenerative signaling via β-arrestin 1/2-mediated stimulation of metabotropic GLU receiptrs, leading to ERK1/2-MAPK activation and downstream NRF2/BAC transcriptional modulation (Wang et al., 2016; F. Yang et al., 2020). Disruption of the BDNF–TrkB–PI3K–Akt–mTOR signalling cascade in both cortical and hippocampal circuits is increasingly recognised as a key contributor to the neurotoxic effects observed following exposure to general anaesthetics(Wang & Wang, 2019). In postnatal day 6 (P6) mice, exposure to sevoflurane induces pathological activation of GSK-3β, resulting in aberrant tau hyperphosphorylation, upregulation of IL-6 in the hippocampus, and diminished expression of the synaptic scaffolding protein PSD95. These molecular alterations converge to impair cognitive performance, suggesting a mechanistic link between anesthetic exposure, neuroinflammation, and synaptic dysfunction(Faraco et al., 2019; Tao et al., 2014).

Furthermore, Repeated exposure to zodiazepines, including remimazolam and midazolam, has been demonstrated to disrupt short-term cognitive processes such as learning and memory in juvenile rodent models. This cognitive impairment may be mechanistically attributed to a reduction in BDNF, upregulated caspase-3 expression, diminished phosphorylation of PSD95, and suppression of LTP within

the hippocampus, culminating in the exacerbation of hippocampal neuronal cell death. (Shi et al., 2024).

2.3 Effects of Anesthetics on Glial Cells and Inflammation

Anesthetic agents exert their effects not only on neurons but also extensively influence glial populations in the developing brain, with notable impacts on astrocytes, microglia, and oligodendrocytes. Amplified inflammatory activity within astrocytes and microglia is a defining feature of neuroinflammation, marked by glial cell activation or infiltration of immune cells that subsequently drive the upregulation of pro-inflammatory mediators(Mendiola & Cardona, 2018). Oligodendrocytes further influence neurodevelopment through their role in myelination(Fig. 3).

Microglial activation is broadly acknowledged as a central driver of pro-inflammatory cytokine and chemokine production within the central nervous system. Persistent or excessive activation of these immune mediators is intimately lighed to neuronal injury and is thought to underlie anaesthesia-related neuroinflammatory pathologies (Chen et al., 2018). Repeated exposure to sevoflurane during the neonatal stage has been demonstrated to markedly stigulate microglial activation, leading to heightened expression of pivotal neuroinflammatory mediators such as TNF-α, IL-6, and IL-1β (M. Yang et al., 2020). Moreover, prenatal anaesthetic exposure via mate administration disrupts fetal glial cell signaling, prominently increasing IL-6 production through extracellular signal-regulated kinase (ERK) phosphorylation (Hirotsu et al., 2019). Concurrently, neonatal exposure to sevoflurane significantly upregulates sirtuin 2 (SIRT2) in the hippocampus, promoting microglial polarization towards a pro-inflammatory M1 phenotype (Z. Wu, Y. Zhang, et al., 2020). Beyond inhalational anesthetics, intravenous agents such as propofol and midazolam have also been associated with microglial activation, leading to an upsurge in the release of inflammatory mediators, notably IL-1β and TNF-α (Popić et al., 2015; Ramirez et al., 2016). These responses contribute to neuroinflammatory cascades and interfere

with developmental neurogenesis (Milanovic et al., 2016). Exposure to propofol during early postnatal development significantly elevates the expression of pro-inflammatory markers—including caspase-1 and interleukin-1β—in both cortical and thalamic regions of postnatal day 7 (PND7 86 ts(Milanovic et al., 2016). Importantly, microglial activation induced by sevoflurane has likewise been linked to the pathogenesis of various neurodegenerative conditions (Yeh et al., 2017).

Astrocytes are essential regulators of neural circuit formation and play a fundamental role in sustaining synaptic architecture and functionality throughout neurodevelopment. Anesthetics may disrupt astrocyte density and function, thereby impairing BDNF secretion and indirectly affecting neuronal survival. For example, in high-density co-cultures of astrocytes, BDNF secretion effectively rescues propofol-induced neuronal death, whereas low-density astrocytes, which secrete insufficient BDNF, fail to provide the sample protection (Y. Liu et al., 2017). These observations may offer mechanistic insight into the heightened susceptibility of the developing brain to anesthetic-induced neurotoxicity. Within the hippocampus, astrocytes enriched in phosphoprotein enriched in astrocytes 15 (PEA15) are prominently represented, with PEA15 expression shown to increase progressively as astrocytes undergo maturation... Mounting evidence indicates that propofol interferes with neurodevelopment by suppressing the expression of PEA15, thereby perturbing the ERK/CREB 29 K2 signaling axis. This disruption ultimately results in diminished neuronal proliferation alongside an increase in programmed cell death(Xian et al., 2019).

Oligodendrocytes are central mediators of myelin sheath formation, facilitating efficient axonal conduction and supporting the maturation of neural circuits (Choi et al., 2019; Tarnason et al., 2020). Myelination, a tightly regulated and temporally coordinated process occurring during early postnatal development, plays a pivotal role in the emergence of higher-order cognitive capacities and is intimately associated with the dynamic remodeling of both structural and functional neural circuitry(Deoni et al., 2018). Oligodendrocytes are not fully mature during the neonatal period, and anesthetic-induced inflammation may permanently damage myelination, hindering normal neurodevelopment. Studies have shown that anesthetic drugs have a far greater impact on oligodendrocyte apoptosis than on neurons, with apoptosis occurring at twice the rate in oligodendrocytes, representing 59% of all apoptotic cells(Schenning et al., 2017). Administration of propofol during vulnerable windows of early neurodevelopment markedly amplifies apoptotic activity in both neuronal and oligodendrocyte populations, concurrently triggering a substantial upregulation of pro-inflammatory cytokine expression. These molecular and cellular disruptions exhibit a strong association with enduring behavioural impairments manifesting in later stages of life (Milanovic et al., 2016; Ye et al., 2013). Moreover, gestational exposure to sevoflurane has been shown to impair myelination in offspring by inducing iron deficiency, thereby compromising myelin structure and reducing overall myelin content (Zuo et al., 2020). Similarly, exposure to high concentrations of sevoflurane during the early postnatal phason as been linked to impaired oligodendrocyte maturation and disruption of the normal trajectory of white matter myelination in the developing rat brain. These disturbances in oligodendrocyte lineage development contribute to central nervous system dysmyelination, a pathological process that may underlie long-term cognitive impairments (Z. Wu, H. Xue, et al., 2020).

2.4 Effects of Anesthetics on Calcium Overload, Oxidative Stress, and Endoplasmic Reticulum Stress

Appropriate calcium ion (Ca²⁺) levels are crucial for regulating oxidative phosphorylation in the mitochondrial matrix, thereby maintaining normal ATP production rates. Nevertheless, excessive calcium influx—particularly originating from the cytosol or the endogramic reticulum (ER)—can compromise mitochondrial respiratory chain function, thereby promoting the overproduction of reactive oxygen spries (ROS). The resulting elevation in intracellular ROS levels ultimately culminates in oxidative DNA damage(Piao et al., 2020). Excessive generation and accumulation of ROS compromise the structural integrity of the mitochondrial membrane, disrupt cellular energy homeostasis by impairing ATP synthesis, and increase mitochondrial outer membrane permeability (MOMP), collectively triggering ignaling cascades that culminate in cellular dysfunction(Calvo-Rodriguez & Bacskai, 2021; Marchi et al., 2018).

General anesthetics have been shown to induce excessive activation of inositol 1,4,5-trisphosphate receptors, leading to a pronounced increase in cytosolic Ca²⁺ levels while simultaneously depleting calcium stores within the endoplasmic reticulum. This disruption in intracellular calcium homeostasis may represent a key mechanistic link underlying anesthesia-induced neurotoxicity during brain development(Yang & Wei, 2017). For instance, the combination of propofol and midazolam significantly increases mitochondrial ROS levels, inhibits antioxidant enzyme activity, and impairs synaptic function(Boscolo et al., 2013). Propofol uncouples oxidative phosphorylation, disrupting mitochondrial function and increasing ROS production, thereby exacerbating neurotoxicity, which is difficult to mitigate through pretreatment(Shibuta et al., 2022). When midazolam is combined with nitrous oxide and isoflurane, ROS levels increase by approximately 30%, and an indicate the introduced activity is notably reduced(Boscolo et al., 2013). Anesthetic compounds—particularly benzodiazepines such as midazolam and diazepam—have been associated with detrimental neurographopomental consequences, primarily through the disruption of synaptic architecture, impairment of synaptic transmission, and induction of neuronal apoptosis(Lee et al., 2015).

Exposure to sevoflurane has been demonstrated to activate the PI3K /Akt signaling cascade, thereby perturbing the balance between pro-survival and pro-apoptotic pathways and triggering apoptotic cell death. This dysregulation enhances MOMP, ultimately triggering intrinsic 101 totic pathways and contributing to long-term cognitive impairments observed during adolescence (Hu et al., 2019; Yu et al., 2020). In parallel, mounting evidence implicates ER stress as a key mediator of sevoflurane-inducent neurotoxicity, primarily via the PERK–eIF2α–ATF4–CHOP signaling pathway (B. Liu et al., 2017). In the developing brain, exposure to sevoflurane has also been shown to upregulate the expression of protein tyrosine phosphatase 1B within the ER, thereby initiating ER stress responses and promoting neurodegenerative alterations (Liu et al., 2019).

2.5 Effects of Anesthetics Targeting GABAARs on Non-Coding RNAs (ncRNAs)

Emerging evidence highlights the pivotal influence of ncRNAs in orchestrating diverse neurobiological processes through intricate post-transcriptional regulatory mechanisms. General

anesthetics modulate the expression profiles of microRNAs (miRNAs) and long non-coding RNAs (lncRNAs) in the developing brain, forming complex molecular regulatory networks that may perturb critical neural pathways. Such dysregulation is implicated in the induction of acute neuronal apoptosis persistent memory deficits, potentially resulting in irreversible neurodevelopmental alterations(Jiang et al., 2021; Rodrigues et al., 2020; Shu et al., 2019).

With respect to apoptotic mechanisms, sevoflurane has been shown to upregulate the expression of caspase-3 and Bax, while concurrently downregulation Bcl-2, BDNF, and nerve growth factor (NGF). These changes are accompanied by alterations in long non-coding RNA (lncRNA) expression and a reduction in hippocampa neuronal density, collectively leading to ultrastructural abnormalities and neuronal apoptosis (Hu et al., 2019) (Hu et al., 2019). Moreover, the non-coding RNA LRCF has been identified as a pivotal regulatory element in mediating isoflurane-induced apoptosis of oligodendrocytes.. In neonatal mice, expression levels of LRCF are significantly elevated relative to those in adults. This upregulation promotes apoptotic signaling through activation of the HIF-1α/caspase-3 axis. Congresely, reduced LRCF expression appears to exert neuroprotective effects by recruiting the alternative HIF-1α/miR-138-5pgspase-3 pathway. These expression-dependent regulatory divergences may account for the heightened susceptibility of the developing brain to anesthesia-induced neurotoxicity during the neonatal period. Consequently, gigh LRCF expression may serve as a high-risk biomarker for OLG damage caused by anesthesia(Zeng et al., 2021). Exposure to sevoflurane has been demonstrated to influence synaptic plasticity by upregulating the expression of synaptophysin mRNA, a key presynaptic marker, via a mechanism dependent on N6-methyladenosine RNA modification. Paradoxically, this post-transcriptional modification contributes to a subsequent decline in synaptophysin protein abundance, which is associated with disrupted motor coordination and impaired cognitive performance in juvenile mice(L. Zhang et al., 2022). Isoflurane anesthesia has been reported to downregulate the expression of microRNA-132, thereby ducing dendritic spine density in the hippocampus, a morphological alteration that contributes to impaired learning and memory performance in rodent models.(Zhang et al., 2017).

3. Neuroprotective Potential of Anesthetics

NAS, as sedatives and anesthetics, demonstrate significant neuroprotective potential (Tateiwa & Evers, 2024). By modulating both inhibitory and excitatory neurotransmitter systems, NAS maintain the stability of neural networks. Additionally, NAS promote the release of BDNF, a molecule essential for supporting neuronal growth, viability, and synaptic plasticity. Furthermore, NAS regulate inflammation and oxidative stress, alleviating neuroinflammation and reducing oxidative damage, thereby effectively protecting neuronal cells from injury. These multiple mechanisms collectively highlight the potential of NAS agents as neurodevelopmental protective compounds (Fig. 3).

3.1 Modulation of the Inhibitory Neurotransmitter System

While most NAS serve as potent allosteric modulators of GABA_AR, enhancing or inhibiting GABA-induced currents via subtype-specific binding—such as potentiation by 3α -hydroxylated neurosteroids and suppression by 3β -sulfated derivatives—emerging evidence indicates a divergence in neurotoxic potential among GABA_AR-activating agents(Chen et al., 2019; Laverty et al., 2017). Notably,

compounds like propofol, CDNC24, and alphaxalone all activate GABAARs, yet only propofol has been consistently associated with significant neurotoxicity during brain development, whereas NAS such as alphaxalone exhibit negligible or no overt neurotoxic effects(Tesic et al., 2020). Further investigations revealed that NAS compounds, such as alphaxalone and 3β-OH, do not induce neuronal apoptosis, regardless of their activation of GABA_ARs. The neuroprotective properties of these compounds may be partially attributed to their ability to inhibit T-type calcium channels, particularly Cav3.1, thereby diminishing presynaptic GABA release(Tesic et al., 2020; Timic Stamenic et al., 2021). Intriguingly, the mechanism of action of 3β-OH appears to operate independently of classical modulation of GABAergic or NMDA receptor pathways(Atluri et al., 2018; Timic Stamenic et al., 2021). NAS enhances neuronal regeneration. For instance, allopregnanolone binds to the transmembrane domain of the GABAAR complex, activating the receptor and increasing intracellular Ca2+ concentrations. This calcium influx subsequently activates CaMK IV, which phosphorylates and activates the transcription factor CREB1. Once activated, CREB1 enhances the transcription of genes involved in cell cycle regulation, thereby promoting mitotic activity in neural stem cells and oligodendrocyte precursor cells(G. F. Li et al., 2018). Moreover, NAS demonstrates significant potential for selective regulation of different GABAAR subtypes. These compounds can modulate receptor tonic currents by enhancing GABAAR-mediated current activity. At moderate concentrations, NAS compounds inhibit GABAAR function, whereas at lower and higher concentrations, they enhance GABA regulation. This suggests that NAS compounds exert a complex bidirectional effect on GABAARs within different concentration ranges, potentially providing a new therapeutic strategy for neuroprotection by modulating neural activity (Fig. 4).

3.2 Modulation of the Excitatory Neurotransmitter System

NAS, such as 3β-OH, reduces presynaptic AMPA receptor-mediated excitatory synaptic currents but does not affect NMDAR-mediated excitatory currents, thus modulating the glutamatergic system(Atluri et al., 2018). Gonadal steroids, such as estradiol (E2) and 17 stradiol, potentiate NMDAR-mediated synaptic transmission and facilitate the release of GLU from primary afferent terminals, thereby increasing dendritic spine density in dorsal horn neurons of the spinal cord(Zhang et al., 2012). Estradiol not only enhances the binding of NMDAR agonists and competitive antagonists(Kow et al., 2005) but may also exert a dual action by enhancing cognitive function, promoting synaptic transmission while offering neuroprotective effects(Nilsen et al., 2002). The enantiomer of 17β-estradiol exhibits neuroprotective properties against glutamate-induced toxicity in cultured neuronal cells, implying that its protective effects may be mediated through pathways independent of classical nuclear estrogen receptors(Green et al., 2001). These studies suggest that NAS drugs exhibit neuroprotective potential by modulating NMDARs and the GLU transport system within a specific concentration range(Fig. 4).

3.3 Promotion of BDNF Release

Many studies have indicated that NAS can bind to and regulate various proteins involved in cell protection, including estrogen receptors α/β (ER α/β), the orphan nuclear receptor pregnane-X receptor (PXR), and the GPER1, thereby modulating BDNF levels(Briz & Baudry, 2014; Jakab et al., 2001; Serrao & Goodchild, 2022). For example, the enantiomer of 17 β -estradiol regulates BDNF levels through an ER β -mediated mechanism(Jakab et al., 2001). This modulatory action not only contributes to improved learning and memory performance but also serves a pivotal role in mitigating cognitive impairments induced by anesthesia(Taylor et al., 2017). Additionally, neurosteroid drugs such as alphaxalone activate PXR, further promoting the secretion of BDNF and exerting neuroprotective effects(Frye et al., 2014; Serrao & Goodchild, 2022). Clinical randomized controlled trials (RCTs) have demonstrated that patients treated with alphaxalone during hip replacement surgery show better cognitive performance postoperatively, with significantly higher plasma levels of BDNF than patients treated with propofol and sevoflurane(Serrao & Goodchild, 2022). These findings suggest that NAS, by modulating BDNF and associated pathways, exhibits significant neuroprotective effects, particularly in the recovery of cognitive function following anesthesia and surgery.

3.4 Regulation of Inflammation and Oxidative Stress

NAS, such as pregnenolone, exert anti-inflammatory effects through multiple signaling pathways, including Toll-like receptor signaling, the TRAM-TRIF pathway, BDNF, and CX3CL1 signaling pathways. NAS suppress the initiation of pro-inflammatory signaling cascades by disrupting the binding of Toll-like receptors—specifically TLR2, TLR4, and TLR7—to their adaptor proteins such as My 8, MD2, and TIRAP, thereby reducing downstream synthesis of pro-inflammatory mediators(Balan 2019; Balan, Grusca, et al., 2023; Balan, Patterson, et al., 2023). In addition to its established role in suppressing pro-inflating atory signaling, NAS activate the TLR4-TRAM-TRIF pathway, leading to the upregulation of key anti-inflammatory mediators such as IL-10 and BDNF, thereby contributing to neuroprotection (Aurelian & Balan, 2019). NAS also promortible expression of the anti-inflammatory chemokine CX3CL1, which is essential for maintaining the equilibrium between pro- and antiinflammatory responses within the central nervous system, ultimately alleviating neuroinflammation. Furthermore, NAS have been shown to attenuate microglial pro-inflammatory activation by engaging $GABA_ARs$ and modulating the BDNF-TrkA/TrkB signaling cascade, while concurrently regulating the transcription of pro-inflammatory genes (Alexaki et al., 2018; S. Y. Wu et al., 2020). Beyond antiinflammatory action, NAS support neuronal repair and regeneration through diverse neuroprotective mechanisms, offering potential therapypic benefits in the context of neurodegenerative conditions and traumatic brain injury. Collectively, these findings underscore the central role of inflammation and oxidative stress in anesthetic-induced neurotoxicity. Importantly, NAS modulate antioxidant and antiinflammatory processes not only through GABAergic pathways but also via alternative non-GABAergic mechanisms, highlighting their promise in mitigating the adverse neurodevelopmental effects associated with anesthetic exposure.

4. Susceptibility Factors for Anesthetic-Induced Neurotoxicity

4.1. Dose-, Time-, and Frequency-Dependent Effects of Anesthetics

Anesthetic-induced neurotoxicity demonstrates marked dependence on dose, exposure duration, and administration frequency. Evidence m both cellular and animal models indicates that anesthetic agents, such as propofol, may facilitate the differentiation of neural progenitor cells into neurons when administered at very low concentrations(Qiao et al., 2017). However, at subclinical or medium-to-high doses, these anesthetics inhibit neuronal proliferation, increase pro-inflammatory factor levels, and trigger with read neuronal apoptosis, with the extent of apoptosis gradually increasing with higher doses(K. Chen et al., 2022; Han et al., 2015). Anesthetic neurotoxicity also demonstrate ear time dependence, with neuronal apoptosis worsening as the exposure duration increases(Huang et al., 2012; Zhazet al., 2016). In rodent models, brief exposure to propofol (2 or 4 hours) induced marked activation of caspase-3, a key executor of apoptosis. Notably, a more prolonged exposure of 6 hours elicited not only a pronounced increase in caspase-3 but also elevated calpain activity within the cerebral cortex and thalamus. These molecular alterations were concomitant with the emergence of apoptotic neuror profiles, indicating that extended anesthetic exposure may potentiate neurodegenerative processes in specific brain regions (Milanovic et al., 2010). Prolonged exposure to sevoflurane impairs the maturation of neural circuits by inhibiting synaptogenesis and destabilizing dendritic spine development. This anesthetic disruption is further characterized by structural synaptic alterations, including increased expression of synaptic vesicle-related proteins, reduced density of apical podritic spines, and ultrastructural disorganization within hippocampal synapses, which collectively contribute to cognitive deficits observed in juvenile rats (Yu et al., 2020). Moreover, a study comparing single-dose versus sevendose administrations of propofol with the same total dose showed that rats exposed multiple times exhibited significant impairments in spatial learning and hippocampal function, suggesting that frequent exposures may have a more severe impact on neurological function than a single cumulative dose(Gao et al., 2014). Recurrent administration of sevoflurane during critical periods of cortical development has been implicated in long-term synaptic dysfunction, accompanied by structural synaptic anomalies and persistent cognitive deficits manifesting in adulthood(G. Li et al., 2019; Zhou et al., 2019)

4.2 Synergistic Effects of Combined Anesthesia

Emerging evidence indicates that the combined administration of anesthetic agents may markedly potentiate neuronal apoptosis and disrupt hippocampal synaptic integrity, thereby impairing learning and memory through synergistic neurotoxic effects(Jevtovic-Todorovic et al., 2003). For instance, during critical windows of brain maturation, the combined administration of ketamine with thiopental or propofol significantly amplifies neuronal cell death in neonatal animals and results in enduring functional impairments in adulthood, whereas the administration of these agents individually terms to produce only modest neurodevelopmental effects(Fredriksson et al., 2007). The combined use of anesthetics are incompared to an animal synaptic density (Lunardi et al., 2010), destabilize mitochondria(Sanchez et al., 2011), and increase autophagic exposure to midazolam and nitrous oxide perturbs the homeostasis of mitochondrial dynamics by dysregulating key modulators such as Mfn-2 and Drp-1. This imbalance leads to pronounced mitochondrial morphological alterations, compromised bioenergetic function, and subsequent impairments in neuronal integrity and performance(Boscolo et al., 2013).

4.3 Region-Specific Sensitivity to Anesthetics During Critical Periods of Brain Development

During BGS, exposure to anesthetic drugs can lead to widespread neuronal apoptosis and result in persistent behavioral deficits(Gentry et al., 2013). The temporal window of the BGS varies significantly across mammalian species. In rodents, this critical phase of neurodevelopment unfolds within the first two weeks following birth (Semple et al., 2013), whereas in rhesus macaques, it spans from gestational day 115 to approximately postnatal day 60 (Brambrink et al., 2012); and in humans, the growth spurts occur at varying times across different brain regions, with most regions peaking in the second year after birth and typically reaching their maximum between 3 and 10 years of age(Andropoulos & Greene, 2017). Neuronal apoptosis induced by anesthetic drugs exhibits significant regional specificity at different time points during the BGS. In rodents, postnatal day 0 (P0) is the peak period of early apoptosis, primarily occurring in neurons of the hypothalamus and certain thalamic nuclei; day 3 (P3) marks the peak of intermediate apoptosis, affecting regions such as the Subiculum, Hibrait, caudate nucleus, and thalamic nuclei; day 7 (P7) represents the peak of late apoptosis, predominantly affecting cortical neurons(Maloney et al., 2019). Similarly, In a comparable model using rhesus monkeys, prenatal exposure to propofol predominantly induces apoptosis in neurons and oligodendrocytes within the cerebellum, hypothalamus, and additional posterior and anterior brain regions, whereas exposure during the neonatal period primarily affects the cerebral cortex(Creeley et al., 2013) (Fig. 4).

4.4 Vulnerable Populations for Anesthetic-Induced Neurotoxicity

Neonates—particularly those born prematurely, with low birth weight, reduced gestational age, or complex congenital anomalies—constitute a highly vulnerable population, exhibiting an elevated susceptibility to anesthesia-related neurodevelopmental impairment. Neuroimaging studies have revealed that preterm infants frequently display structural abnormalities in both gray and white matter following neonatal surgery. These alterations, detectable by MRI, have been strongly correlated with subsequent cognitive deficits and delays in neurodevelopmental milestones(Filan et al., 2012; Stolwijk et al., 2017). Weiss et al. noted that neonatal brains are highly sensitive to ischemic injury, placing these groups at a higher risk for neurodevelopmental disorders during the perioperative period. Specifically, during anesthesia, changes in hemodynamics, respiration, and metabolism may have a greater impact on neurodevelopment than the anesthetic drugs themselves(Weiss et al., 2016). Furthermore, factors such as increased surgical frequency, prolonged hospital stays, and prolonged respiratory support further exacerbate the risk of neurodevelopmental disorders. These impairments are not only related to the direct effects of surgery and anesthesia, but may also be closely associated with hemodynamic instability, improper respiratory control, and metabolic imbalance during anesthesia, compounded by surgical stress and primary diseases(Stolwijk et al., 2016).

5 Potential Strategies to Prevent and Reverse Neurotoxicity

5.1 Combination of Newer Anesthetic Agents

Dexmedetomidine (DEX), a highly selective α₂-adrenergic receptor agonist, has emerged as a potent neuroprotective agent against isoflurane-induced neurotoxicity, primarily due to its robust anti-apoptotic

properties. Mechanistically, DEX exerts its neuropertective effects through engagement with several converging molecular cascades. These include the DNMT3a-miR-377-5p-Arc axis and the ERK1/2-CREB-BDNF signaling pathway (Xiao et al., 2018), along with activation of the PI3K-Akt cascade. Concurrently, it inhibits pro-apoptotic processes by downregulating the GSK-3β-CRMP2 and CDK5-CRMP2 pathways (Z. Chen et al., 2022). Pretreatment with DEX has also been shown to counteract the isoflurane-mediated repression of neurotrophic mediators such as BDNF mRNA, phosphorylated ERK1/2, CREB, and BDNF protein, thereby preserving synaptic homeostasis u stress(Tu et al., 2019). Additional studies have confirmed that the suppression of the GSK-3β/CRMP2 and CDK5/CRMP2 pathways by DEX contributes to reduced neuronal apoptosis and cognitive decline in neonatal models (J. Li et al., 2019). Moreover, the neuroprotection afforded by DEX extends beyond apoptosis, encompassing antioxidative and anti-inflammatory actions that mitigate sevoflurane-induced cognitive impairments (Wang et al., 2015). In neonatal rats, DEX may also confer protection via modulation of glutamatergic transmission (Wang et al., 2019). Activation of α₂-adrenergic receptors by DEX and related agonists like clonidine has additionally been found to attenuate tau hyperphosphorylation and its associated cognitive sequelae in neonatal mice exposed to sevoflurane (Sun et al., 2021). Notably, the neuroprotective efficacy of DEX in anesthetic regimens appears dosedependent: at low concentrations, co-administration with sevoflurane enhances neuroprotection; however, higher doses paradoxically increase apoptosis and negretality, potentially due to reduced α2 receptor selectivity and inadvertent α1 receptor activation (Liu et al., 2016; Perez-Zoghbi et al., 2020) (Perez-Zoghbi et al., 2017)(Fig. 4).

Novel BZDs such as remimazolam, when combined with sevoflurane, significantly reduce the incidence of postoperative delirium, suggesting a neuroprotective potential that may be linked to reduced overall anesthetic exposure(Yang et al., 2022). The combined use of anesthetics might also protect the brain by balancing excitatory and inhibitory pathways. For example, midazolam could facilitate the reuptake of excess GLU via EAAT2 and entropy e GABAAR activity, inhibiting the release of glutamate transporters, thereby effectively alleviating excitotoxicity caused by the upregulation of NMDAR NR1 subunits after ketamine withdrawal(Y. Li et al., 2018; Slikker et al., 2007). Moreover, NAS such as 17β-18 adiol may enhance AKT phosphorylation, increase p-GSK-3β levels, and stimulate BDNF release(W. Li et al., 2019; Yang et al., 2022), effective the mitigating the apoptosis induced by combinations of triple cocktails(Lu et al., 2006) or ketamine(Li et al., 2014). Alphaxalone has been reported to mitigate the neurotoxic effects elicited by isoflurane exposure in neonatal rats, highlighting its potential as a neuroprotective agent during early brain development(Zhao et al., 2023).

5.2 Neuroprotective Agents

5.2.1 Anti-Apoptotic Agents

Neuronal apoptosis is a major mechanism of developmental neurotoxicity induced by GABA_AR receptor agonists. Inhibiting apoptotic pathways may serve as a potential intervention. The histamine H_3 receptor antagonist Clobenpropit has been shown to activate the PI3K/Akt signaling cascade, thereby mitigating isoflurane-induced apoptotic injury in hippocampal neurons in vitro (He et al., 2018) (He et al., 2018). Complementarily, the TNF- α inhibitor Etanercept significantly reduced neuronal apoptosis in

the developing rat brain following isoflurane exposure (Chen et al., 2016) then et al., 2016), suggesting a role for inflammatory modulation in anesthetic-induced neurotoxicity. In a separate study, Strait and colleagues demonstrated that the co-administration of lithium with either ketamine or propofol did not significantly alter caspase-3 activation levels compared to control conditions, indicating a limited effect of lithium in modulating anesthetic-induced apoptosis via this pathway(Straiko et al., 2009). Moreover, erythropoietin (EPO) can partially reverse the decrease in BDNF and Bcl-2 levels caused by general anesthetics, thereby mitigating neuronal damage(Pellegrini et al., 2014).

5.2.2 Anti-Inflammatory Agents

Neuroinflammation has emerged as a critical mediator of anesthetic-induced neurotoxicity, making it a compelling target for therapeutic intervention. Recent findings indicate that edaravone confers neuroprotectical gainst isoflurane-induced neuronal injury by attenuating inflammatory and apoptotic ponses via activation of the mBDNF-TrkB-PI3K signaling axis (Yang et al., 2021). These beneficial effects have been validated across both in vitro neuronal culture systems and in vivo preclinical models, highlighting the therapeutic promise of neurotrophic pathway modulation in mitigating anesthesia-related neural damage. In parallel, doxycycline—a semi-synthetic tetracycline antibiotic—has demonstrated potent neuroprotective capabilities beyond its antimicrobial function. Through engagement of the PI3K/Akt pathway, doxycycline has been shown to exert significant anti-inflammatory and anti-apoptotic effects, supporting its candidacy as a therapeutic agent for inflammation-related neurodevelopmental pathologies (Möller et al., 2016).

5.2.3 Anti-Oxidative Stress Neuroprotective Agents

Recent advances in neuropharmacology underscore the neuroprotective potential of natural antioxidants against anesthesia-induced neurotoxicity. Monttonin, for instance, has been shown to activate the PKCa/Nrf2 signaling cascade, upregulate the anti-apoptotic protein Bcl-xL, suppress cytochrome c release, and inhibit mitochondrial-dependent apoptotic pathways. Through these mechanisms, melatonin markedly reduces neuronal apoptosis triggered by combined exposure to azolam, nitrous oxide, and isoflurane, suggesting a protective role in anesthetic neurotoxicity (B. Li et al., 2018; Yon et al., 2006). Similarly, curcumin (Ji et al., 2015) and resveratrol (Tang et al., 2021), both known for their antioxidant properties, have been demonstrated to attenuate sevoflurane-induced oxidative stress by neutralizing ROS and suppressing associated inflammatory responses. Applyl-L-carnitine (L-Ca), a mitochondria-targeted antioxidant, has been identified as neuroprotective in aging and neurodegenerative 75 ditions (Robinson et al., 2019; Yan et al., 2014). Notably, co-administration of L-Ca with propofol has been reported to alleviate propofol-induced cytotoxicity in embryonic neural stem cells, primage by reducing ROS production and downregulating oxidative stress-related signaling pathways (Liu et al., 2014). These findings highlight the promise of L-Ca as a potential adjunct in anesthetic protocols, particularly those involving agents with known oxidative liabilities.

5.2.4 Nutritional Supplements for Neuroprotection

Calcium homeostasis disruption is a key mechanism underlying anesthetic-induced neurotoxicity, involving mitochondrial dysfunction as well as damage to astrocytes and neurons. Emerging evidence indicates that stabilizing intracellular calcium dynamics may represent a promising strategy to counteract the neurodevelopmental toxicity associated with sevoflurane exposure(Zhu et al., 2024). Iron deficiency has been recognized as a pivotal determinant of impaired myelination within the central nervous system, exerting its most detrimental effects during the fetal period and early stages of postnatal development. Disruptions in iron homeostasis during these sensitive periods may compromise oligodendrocyte function and hinder the proper formation of myelin sheaths(Ward et al., 2014). Sevoflurane exposure has been implicated in disrupting myelin development during early brain maturation, potentially through mechanisms involving oligodendrocyte apoptosis, as well as deficiencies in iron and folate metabolism. Supplementing with iron and folate and inhibiting OL apoptosis could be effective preventive measures, although further research is needed.

These findings suggest that strategies aimed at preventing apoptosis and inflammation, reducing oxidative stress, and restoring calcium homeostasis and nutrient balance may offer promising interventions against sevoflurane-induced neurotoxicity (Fig. 3).

6. Concluding Remarks and Future Directions

During the BGS, the finely tuned interplay between the GABAergic and glutamatergic (GABA/GLU) systems is essential for proper neural development, with the developmental switch from NKCC1 to KCC2 transporters serving as a pivotal mechanism in establishing the balance between E/I signaling. Anesthetics, particularly intravenous agents targeting GABAARs, may disrupt this transition, thereby destabilizing neural networks(Cabrera et al., 2020). Additional mechanisms include the imbalance of the BDNF signaling pathway, oxidative stress responses, and inflammatory reactions, all of which agreebate neurotoxicity. Compared with traditional anesthetics, newer agents such as NAS(Tesic et al., 2020; Timic Stamenic et al., 2021), remimazolam(Shi et al., 2024), and DEX(Wei et al., 2016) exhibit milder mechanisms of action, maintaining sufficient anesthetic effects while minimizing interference with the nervous system. However, our understanding remains limited regarding how novel NAS and similar compounds may modulate T-type calcium channels and selectively bind to specific sites on GABAAR, thereby facilitating endogenous regenerative processes in the brain and mitigating the neurotoxic effects of anesthetic agents. Further investigation is essential to elucidate their potential in alleviating neurotoxicity. Concurrently, significant strides a peing made in the development of neuroprotective pharmacotherapies. Agents such as melatonin (B. Li et al., 2018), erythropoietin (EPO) (Pellegrini et al., 2014), and resveratrol (Tang et al., 2021)—all of which exhibit potent antioxidant, antiinflammatory, and neuroprotective properties—have been investigated as promising candidates for mitigating anesthetic-induced neurotoxicity (Fig. 3). Accordingly, the development of next-generation anesthetic agents should prioritise strategies that minimise neurotoxicity, enhance intrinsic neuroprotective mechanisms, and facilitate post-injury repair and functional recovery—thereby ensuring both the safety and therapeutic efficacy of anaesthesia in paediatric and other high-risk populations.

Since the 2016 advisory issued by the FDA regarding the potential neurotoxic effects of anesthetic agents in pediatric populations, there has been a marked increase in clinical investigations evaluating the

neurodevelopmental consequences of anestheric exposure in children. Findings from key studies such as the PANDA study (Sun et al., 2016) and the GAS trial (Davidso 39 tal., 2016; McCann et al., 2019) have shown that a single, bigg exposure—typically under one hour—to general anesthesia in children younger than five years of age is not associated with significant long-term neurodevelopmental deficits. However, the MASK study provided evidence that multiple exposures prior to age three may be linked to slower cognitive processing, impaired fine motor coordination, and increased risk of behavioral and learning difficulties. Supporting this, recent data suggest that general anesthesia administered to infants under one year of age is associated with an elevated risk of developmental delays, with risk escalating as the number of surgical procedures increases (Kobayashi et al., 2020). While the majority of clinical studies suggest that isolated anesthetic exposure is relatively safe, the literature remains inconclusive due to ethical constraints, methodological limitations, and perioperative confounders. Notably, the combined neurotoxic effects of multiple anesthetic agents and their potential protective mechanisms remain underexplored. Future research should adopt an integrative approach, combining mechanistic molecular studies, behavioral analyses in animal models, and rigorously designed clinical trials. Prospective randomized controlled trials (RCTs) with long-term neurodevelopmental follow-up are essential. Moreover, the establishment of standardized dosing frameworks and exposure durations, alongside anesthesia protocols tailored to regional brain vulnerability across developmental stages, will be critical to advancing this field

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Clinical investigations into the long-term cognitive and behavioral consequences of anesthetic exposure in children employ heterogeneous methodologies. However, a significant proportion of these studies rely predominantly on subjective outcome assessments (Maloney et al., 2019), with limited incorporation of objective neuroimaging techniques or biomarker analyses, thereby comprehensive evaluation of anesthetic-induced neurodevelopmental effects (Bethlehem et al., 2022; H. 77 ng et al., 2022). Notably, quantitative ultrasound imaging has revealed increased apoptotic activity in the developing brains of non-human primates following sevoflurane exposure, highlighting the potential neurotoxic impact of this agent during vulnerable periods of brain magation (Rosado-Mendez et al., 2019). Complementary neuroimaging studies suggest that pediatric exposure to inhalational pesthetics such as isoflurane and sevoflurane is associated with cognitive impairments and disruptions in white matter integrity (Banerjee et al., 2019; Bethlehem et al., 2022). Moreover, prospective investigations report that as many as 75% of infants undergoing non-cardiac surgery for congenital anomalies exhibit mild to moderate neuroimaging abnormalities postoperatively (Mongerson et al., 2019; Moran et al., 2019). To delineate the precise neurodevel prental impact of anesthetic agents, future studies should adopt an integrative approach—combining MRI and diffusion tensor imaging (DTI) with high-resolution ultrasound and biomarker profiling (e.g., inflammatory cytokines, BDNF)—in conjunction with longitudinal assessments of cognitive and behavioral function across defined developmental windows. This will help establish preventive or reversal strategies, such as the use of antioxidants, calcium channel blockers, or agents that promote myelination, or through environmental stimulation and functional training to restore the function of impaired neural networks.

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7. conclusion

With the widespread clinical application of intravenous anesthetics targeting $GABA_ARs$, their potential impact on the developing nervous system has increasingly become a focal point of academic attention. Although extensive research has elucidated the neurodevelopmental impact of $GABA_AR$ —targeting agents via modulation of both $GABA_ARs$ and NMDA receptors, the specific influence of anesthetic drugs on synaptogenesis remains a complex and multilayered phenomenon. Deciphering the long-term consequences of such agents, alongside the identification of potential neuroprotective strategies and the refinement of their clinical applicability, continues to pose substantial challenges for future investigations.

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for this work.

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published peer-reviewed literature. No new experimental or clinical data were generated or analyzed

756	References
757	
758	Alexaki, V. I., Fodelianaki, G., Neuwirth, A., Mund, C., Kourgiantaki, A., Ieronimaki, E., Lyroni, K.,
759	Troullinaki, M., Fujii, C., Kanczkowski, W., Ziogas, A., Peitzsch, M., Grossklaus, S., Sönnichsen,
760	B., Gravanis, A., Bornstein, S. R., Charalampopoulos, I., Tsatsanis, C., & Chavakis, T. (2018).
761	DHEA inhibits acute microglia-mediated inflammation through activation of the TrkA-Akt1/2-
762	CREB-Jmjd3 pathway. Mol Psychiatry, 23(6), 1410-1420.
763	https://doi.org/10.1038/mp.2017.167
764	Andropoulos, D. B., & Greene, M. F. (2017). Anesthesia and Developing Brains - Implications of the
765	FDA Warning. N Engl J Med, 376(10), 905-907. https://doi.org/10.1056/NEJMp1700196
766	Atluri, N., Joksimovic, S. M., Oklopcic, A., Milanovic, D., Klawitter, J., Eggan, P., Krishnan, K., Covey, D.
767	F., Todorovic, S. M., & Jevtovic-Todorovic, V. (2018). A neurosteroid analogue with T-type
768	calcium channel blocking properties is an effective hypnotic, but is not harmful to neonatal
769	rat brain. Br J Anaesth, 120(4), 768-778. https://doi.org/10.1016/j.bja.2017.12.039
770	Aurelian, L., & Balan, I. (2019). GABA(A)R α2-activated neuroimmune signal controls binge drinking
771	and impulsivity through regulation of the CCL2/CX3CL1 balance. Psychopharmacology (Berl),
772	236(10), 3023-3043. https://doi.org/10.1007/s00213-019-05220-4
773	Bademosi, A. T., Steeves, J., Karunanithi, S., Zalucki, O. H., Gormal, R. S., Liu, S., Lauwers, E.,
774	Verstreken, P., Anggono, V., Meunier, F. A., & van Swinderen, B. (2018). Trapping of
775	Syntaxin1a in Presynaptic Nanoclusters by a Clinically Relevant General Anesthetic. Cell Rep,
776	22(2), 427-440. https://doi.org/10.1016/j.celrep.2017.12.054
777	Balan, I., Beattie, M. C., O'Buckley, T. K., Aurelian, L., & Morrow, A. L. (2019). Endogenous
778	Neurosteroid $(3\alpha,5\alpha)$ 3-Hydroxypregnan-20-one Inhibits Toll-like-4 Receptor Activation and
779	Pro-inflammatory Signaling in Macrophages and Brain. Sci Rep, 9(1), 1220.
780	https://doi.org/10.1038/s41598-018-37409-6
781	Balan, I., Grusca, A., O'Buckley, T. K., & Morrow, A. L. (2023). Neurosteroid $[3\alpha,5\alpha]$ -3-hydroxy-
782	pregnan-20-one enhances IL-10 production via endosomal TRIF-dependent TLR4 signaling
783	pathway. Front Endocrinol (Lausanne), 14, 1299420.
784	https://doi.org/10.3389/fendo.2023.1299420
785	Balan, I., Patterson, R., Boero, G., Krohn, H., O'Buckley, T. K., Meltzer-Brody, S., & Morrow, A. L.
786	(2023). Brexanolone therapeutics in post-partum depression involves inhibition of systemic
787	inflammatory pathways. EBioMedicine, 89, 104473.
788	https://doi.org/10.1016/j.ebiom.2023.104473
789	Banerjee, P., Rossi, M. G., Anghelescu, D. L., Liu, W., Breazeale, A. M., Reddick, W. E., Glass, J. O.,
790	Phillips, N. S., Jacola, L. M., Sabin, N. D., Inaba, H., Srivastava, D., Robison, L. L., Pui, C. H.,
791	Hudson, M. M., & Krull, K. R. (2019). Association Between Anesthesia Exposure and
792	Neurocognitive and Neuroimaging Outcomes in Long-term Survivors of Childhood Acute
793	Lymphoblastic Leukemia. JAMA Oncol, 5(10), 1456-1463.
794 705	https://doi.org/10.1001/jamaoncol.2019.1094
795 706	Bethlehem, R. A. I., Seidlitz, J., White, S. R., Vogel, J. W., Anderson, K. M., Adamson, C., Adler, S.,
796 707	Alexopoulos, G. S., Anagnostou, E., Areces-Gonzalez, A., Astle, D. E., Auyeung, B., Ayub, M.,
797	Bae, J., Ball, G., Baron-Cohen, S., Beare, R., Bedford, S. A., Benegal, V.,Alexander-Bloch, A. F.

798	(2022). Brain charts for the human lifespan. <i>Nature</i> , 604(7906), 525-533.
799	https://doi.org/10.1038/s41586-022-04554-y
800	Bi, C., Cai, Q., Shan, Y., Yang, F., Sun, S., Wu, X., & Liu, H. (2018). Sevoflurane induces neurotoxicity in
801	the developing rat hippocampus by upregulating connexin 43 via the JNK/c-Jun/AP-1
802	pathway. Biomed Pharmacother, 108, 1469-1476.
803	https://doi.org/10.1016/j.biopha.2018.09.111
804	Boscolo, A., Milanovic, D., Starr, J. A., Sanchez, V., Oklopcic, A., Moy, L., Ori, C. C., Erisir, A., & Jevtovic
805	Todorovic, V. (2013). Early exposure to general anesthesia disturbs mitochondrial fission and
806	fusion in the developing rat brain. Anesthesiology, 118(5), 1086-1097.
807	https://doi.org/10.1097/ALN.0b013e318289bc9b
808	Brambrink, A. M., Back, S. A., Riddle, A., Gong, X., Moravec, M. D., Dissen, G. A., Creeley, C. E.,
809	Dikranian, K. T., & Olney, J. W. (2012). Isoflurane-induced apoptosis of oligodendrocytes in
810	the neonatal primate brain. Ann Neurol, 72(4), 525-535. https://doi.org/10.1002/ana.23652
811	Briz, V., & Baudry, M. (2014). Estrogen Regulates Protein Synthesis and Actin Polymerization in
812	Hippocampal Neurons through Different Molecular Mechanisms. Front Endocrinol
813	(Lausanne), 5, 22. https://doi.org/10.3389/fendo.2014.00022
814	Cabrera, O. H., Tesic, V., Tat, Q. L., Chastain, S., Quillinan, N., & Jevtovic-Todorovic, V. (2020).
815	Sevoflurane-Induced Dysregulation of Cation-Chloride Cotransporters NKCC1 and KCC2 in
816	Neonatal Mouse Brain. Mol Neurobiol, 57(1), 1-10. https://doi.org/10.1007/s12035-019-
817	<u>01751-1</u>
818	Calvo-Rodriguez, M., & Bacskai, B. J. (2021). Mitochondria and Calcium in Alzheimer's Disease: From
819	Cell Signaling to Neuronal Cell Death. Trends Neurosci, 44(2), 136-151.
820	https://doi.org/10.1016/j.tins.2020.10.004
821	Chen, B., Deng, X., Wang, B., & Liu, H. (2016). Etanercept, an inhibitor of TNF-a, prevents propofol-
822	induced neurotoxicity in the developing brain. Int J Dev Neurosci, 55, 91-100.
823	https://doi.org/10.1016/j.ijdevneu.2016.10.002
824	Chen, J., Zhang, D. M., Feng, X., Wang, J., Qin, Y. Y., Zhang, T., Huang, Q., Sheng, R., Chen, Z., Li, M., &
825	Qin, Z. H. (2018). TIGAR inhibits ischemia/reperfusion-induced inflammatory response of
826	astrocytes. Neuropharmacology, 131, 377-388.
827	https://doi.org/10.1016/j.neuropharm.2018.01.012
828	Chen, K., Lu, D., Yang, X., Zhou, R., Lan, L., Wu, Y., Wang, C., Xu, X., Jiang, M. H., Wei, M., & Feng, X.
829	(2022). Enhanced hippocampal neurogenesis mediated by PGC-1α-activated OXPHOS after
830	neonatal low-dose Propofol exposure. Front Aging Neurosci, 14, 925728.
831	https://doi.org/10.3389/fnagi.2022.925728
832	Chen, Z., Ding, Y., Zeng, Y., Zhang, X. P., & Chen, J. Y. (2022). Dexmedetomidine reduces propofol-
833	induced hippocampal neuron injury by modulating the miR-377-5p/Arc pathway. BMC
834	Pharmacol Toxicol, 23(1), 18. https://doi.org/10.1186/s40360-022-00555-9
835	Chen, Z. W., Bracamontes, J. R., Budelier, M. M., Germann, A. L., Shin, D. J., Kathiresan, K., Qian, M. X.
836	Manion, B., Cheng, W. W. L., Reichert, D. E., Akk, G., Covey, D. F., & Evers, A. S. (2019).
837	Multiple functional neurosteroid binding sites on GABAA receptors. PLoS Biol, 17(3),
838	e3000157. https://doi.org/10.1371/journal.pbio.3000157

839	Choi, E. H., Blasiak, A., Lee, J., & Yang, I. H. (2019). Modulation of Neural Activity for Myelination in
840	the Central Nervous System. Front Neurosci, 13, 952.
841	https://doi.org/10.3389/fnins.2019.00952
842	Cohen-Cory, S., Kidane, A. H., Shirkey, N. J., & Marshak, S. (2010). Brain-derived neurotrophic factor
843	and the development of structural neuronal connectivity. Dev Neurobiol, 70(5), 271-288.
844	https://doi.org/10.1002/dneu.20774
845	Creeley, C., Dikranian, K., Dissen, G., Martin, L., Olney, J., & Brambrink, A. (2013). Propofol-induced
846	apoptosis of neurones and oligodendrocytes in fetal and neonatal rhesus macaque brain. Br J
847	Anaesth, 110 Suppl 1(Suppl 1), i29-38. https://doi.org/10.1093/bja/aet173
848	Davidson, A. J., Disma, N., de Graaff, J. C., Withington, D. E., Dorris, L., Bell, G., Stargatt, R., Bellinger,
849	D. C., Schuster, T., Arnup, S. J., Hardy, P., Hunt, R. W., Takagi, M. J., Giribaldi, G., Hartmann, P.
850	L., Salvo, I., Morton, N. S., von Ungern Sternberg, B. S., Locatelli, B. G.,McCann, M. E.
851	(2016). Neurodevelopmental outcome at 2 years of age after general anaesthesia and
852	awake-regional anaesthesia in infancy (GAS): an international multicentre, randomised
853	controlled trial. Lancet, 387(10015), 239-250. https://doi.org/10.1016/s0140-6736(15)00608-
854	<u>x</u>
855	Deoni, S., Dean, D., 3rd, Joelson, S., O'Regan, J., & Schneider, N. (2018). Early nutrition influences
856	developmental myelination and cognition in infants and young children. Neuroimage, 178,
857	649-659. https://doi.org/10.1016/j.neuroimage.2017.12.056
858	DiGruccio, M. R., Joksimovic, S., Joksovic, P. M., Lunardi, N., Salajegheh, R., Jevtovic-Todorovic, V.,
859	Beenhakker, M. P., Goodkin, H. P., & Todorovic, S. M. (2015). Hyperexcitability of rat
860	thalamocortical networks after exposure to general anesthesia during brain development. ${\it J}$
861	Neurosci, 35(4), 1481-1492. https://doi.org/10.1523/jneurosci.4883-13.2015
862	Diviccaro, S., Cioffi, L., Falvo, E., Giatti, S., & Melcangi, R. C. (2022). Allopregnanolone: An overview on
863	its synthesis and effects. J Neuroendocrinol, 34(2), e12996.
864	https://doi.org/10.1111/jne.12996
865	Doi, A., Miyazaki, T., Mihara, T., Ikeda, M., Niikura, R., Andoh, T., & Goto, T. (2021). CLP290 promotes
866	the sedative effects of midazolam in neonatal rats in a KCC2-dependent manner: A
867	laboratory study in rats. PLoS One, 16(3), e0248113.
868	https://doi.org/10.1371/journal.pone.0248113
869	Dong, Y., Hong, W., Tang, Z., Gao, Y., Wu, X., & Liu, H. (2020). Sevoflurane leads to learning and
870	memory dysfunction via breaking the balance of tPA/PAI-1. Neurochem Int, 139, 104789.
871	https://doi.org/10.1016/j.neuint.2020.104789
872	Fan, X. Y., Shi, G., & Zhao, P. (2021). Neonatal Sevoflurane Exposure Impairs Learning and Memory by
873	the Hypermethylation of Hippocampal Synaptic Genes. Mol Neurobiol, 58(3), 895-904.
874	https://doi.org/10.1007/s12035-020-02161-4
875	Faraco, G., Hochrainer, K., Segarra, S. G., Schaeffer, S., Santisteban, M. M., Menon, A., Jiang, H.,
876	Holtzman, D. M., Anrather, J., & ladecola, C. (2019). Dietary salt promotes cognitive
877	impairment through tau phosphorylation. Nature, 574(7780), 686-690.
878	https://doi.org/10.1038/s41586-019-1688-z
879	Filan, P. M., Hunt, R. W., Anderson, P. J., Doyle, L. W., & Inder, T. E. (2012). Neurologic outcomes in
880	very preterm infants undergoing surgery. J Pediatr, 160(3), 409-414.
881	https://doi.org/10.1016/j.jpeds.2011.09.009

882	Food, U., & Administration, D. (2017). FDA Drug Safety Communication: FDA review results in new
883	warnings about using general anesthetics and sedation drugs in young children and pregnant
884	women. Silver Spring, MD, USA: US Food and Drug Administration.
885	Forcelli, P. A., Kim, J., Kondratyev, A., & Gale, K. (2011). Pattern of antiepileptic drug-induced cell
886	death in limbic regions of the neonatal rat brain. Epilepsia, 52(12), e207-211.
887	https://doi.org/10.1111/j.1528-1167.2011.03297.x
888	Fredriksson, A., Pontén, E., Gordh, T., & Eriksson, P. (2007). Neonatal exposure to a combination of N-
889	methyl-D-aspartate and gamma-aminobutyric acid type A receptor anesthetic agents
890	potentiates apoptotic neurodegeneration and persistent behavioral deficits. Anesthesiology,
891	107(3), 427-436. https://doi.org/10.1097/01.anes.0000278892.62305.9c
892	Frye, C. A., Koonce, C. J., & Walf, A. A. (2014). Novel receptor targets for production and action of
893	allopregnanolone in the central nervous system: a focus on pregnane xenobiotic receptor.
894	Front Cell Neurosci, 8, 106. https://doi.org/10.3389/fncel.2014.00106
895	Gao, J., Peng, S., Xiang, S., Huang, J., & Chen, P. (2014). Repeated exposure to propofol impairs spatial
896	learning, inhibits LTP and reduces CaMKIIα in young rats. Neurosci Lett, 560, 62-66.
897	https://doi.org/10.1016/j.neulet.2013.11.061
898	Gentry, K. R., Steele, L. M., Sedensky, M. M., & Morgan, P. G. (2013). Early developmental exposure to
899	volatile anesthetics causes behavioral defects in Caenorhabditis elegans. Anesth Analg,
900	116(1), 185-189. https://doi.org/10.1213/ANE.0b013e31826d37c5
901	Green, P. S., Yang, S. H., Nilsson, K. R., Kumar, A. S., Covey, D. F., & Simpkins, J. W. (2001). The
902	nonfeminizing enantiomer of 17beta-estradiol exerts protective effects in neuronal cultures
903	and a rat model of cerebral ischemia. Endocrinology, 142(1), 400-406.
904	https://doi.org/10.1210/endo.142.1.7888
905	Han, D., Jin, J., Fang, H., & Xu, G. (2015). Long-term action of propofol on cognitive function and
906	hippocampal neuroapoptosis in neonatal rats. Int J Clin Exp Med, 8(7), 10696-10704.
907	He, W., Yuan, Q. H., & Zhou, Q. (2018). Histamine H3 receptor antagonist Clobenpropit protects
908	propofol-induced apoptosis of hippocampal neurons through PI3K/AKT pathway. Eur Rev
909	Med Pharmacol Sci, 22(22), 8013-8020. https://doi.org/10.26355/eurrev_201811_16430
910	Head, B. P., Patel, H. H., Niesman, I. R., Drummond, J. C., Roth, D. M., & Patel, P. M. (2009). Inhibition
911	of p75 neurotrophin receptor attenuates isoflurane-mediated neuronal apoptosis in the
912	neonatal central nervous system. Anesthesiology, 110(4), 813-825.
913	https://doi.org/10.1097/ALN.0b013e31819b602b
914	Hirotsu, A., Iwata, Y., Tatsumi, K., Miyai, Y., Matsuyama, T., & Tanaka, T. (2019). Maternal exposure to
915	volatile anesthetics induces IL-6 in fetal brains and affects neuronal development. Eur J
916	Pharmacol, 863, 172682. https://doi.org/10.1016/j.ejphar.2019.172682
917	Hu, X., Hu, X., & Huang, G. (2019). LncRNA MALAT1 is involved in sevoflurane-induced neurotoxicity in
918	developing rats. J Cell Biochem, 120(10), 18209-18218. https://doi.org/10.1002/jcb.29127
919	Huang, L., Liu, Y., Jin, W., Ji, X., & Dong, Z. (2012). Ketamine potentiates hippocampal
920	neurodegeneration and persistent learning and memory impairment through the PKCγ-ERK
921	signaling pathway in the developing brain. Brain Res, 1476, 164-171.
922	https://doi.org/10.1016/j.brainres.2012.07.059

923	Jakab, R. L., Wong, J. K., & Belcher, S. M. (2001). Estrogen receptor beta immunoreactivity in
924	differentiating cells of the developing rat cerebellum. J Comp Neurol, 430(3), 396-409.
925	https://doi.org/10.1002/1096-9861(20010212)430:3
926	Jevtovic-Todorovic, V., Hartman, R. E., Izumi, Y., Benshoff, N. D., Dikranian, K., Zorumski, C. F., Olney,
927	J. W., & Wozniak, D. F. (2003). Early exposure to common anesthetic agents causes
928	widespread neurodegeneration in the developing rat brain and persistent learning deficits.
929	Neurosci, 23(3), 876-882. https://doi.org/10.1523/jneurosci.23-03-00876.2003
930	Ji, M. H., Qiu, L. L., Yang, J. J., Zhang, H., Sun, X. R., Zhu, S. H., Li, W. Y., & Yang, J. J. (2015). Pre-
931	administration of curcumin prevents neonatal sevoflurane exposure-induced
932	neurobehavioral abnormalities in mice. Neurotoxicology, 46, 155-164.
933	https://doi.org/10.1016/j.neuro.2014.11.003
934	Jiang, C., Arzua, T., Yan, Y., & Bai, X. (2021). Expression Signature of IncRNAs and mRNAs in
935	Sevoflurane-Induced Mouse Brain Injury: Implication of Involvement of Wide Molecular
936	Networks and Pathways. Int J Mol Sci, 22(3). https://doi.org/10.3390/ijms22031389
937	Ju, L. S., Yang, J. J., Gravenstein, N., Seubert, C. N., Morey, T. E., Sumners, C., Vasilopoulos, T., Yang, J.
938	J., & Martynyuk, A. E. (2017). Role of environmental stressors in determining the
939	developmental outcome of neonatal anesthesia. Psychoneuroendocrinology, 81, 96-104.
940	https://doi.org/10.1016/j.psyneuen.2017.04.001
941	Kahraman, S., Zup, S. L., McCarthy, M. M., & Fiskum, G. (2008). GABAergic mechanism of propofol
942	toxicity in immature neurons. J Neurosurg Anesthesiol, 20(4), 233-240.
943	https://doi.org/10.1097/ANA.0b013e31817ec34d
944	Karunanithi, S., Cylinder, D., Ertekin, D., Zalucki, O. H., Marin, L., Lavidis, N. A., Atwood, H. L., & van
945	Swinderen, B. (2020). Proportional Downscaling of Glutamatergic Release Sites by the
946	General Anesthetic Propofol at Drosophila Motor Nerve Terminals. eNeuro, 7(1).
947	https://doi.org/10.1523/eneuro.0422-19.2020
948	Kobayashi, Y., Tokuda, N., Adachi, S., Takeshima, Y., Hirose, M., & Shima, M. (2020). Association
949	between surgical procedures under general anesthesia in infancy and developmental
950	outcomes at 1 year: the Japan Environment and Children's Study. Environ Health Prev Med,
951	25(1), 32. https://doi.org/10.1186/s12199-020-00873-6
952	Kow, L. M., Easton, A., & Pfaff, D. W. (2005). Acute estrogen potentiates excitatory responses of
953	neurons in rat hypothalamic ventromedial nucleus. Brain Res, 1043(1-2), 124-131.
954	https://doi.org/10.1016/j.brainres.2005.02.068
955	Laverty, D., Thomas, P., Field, M., Andersen, O. J., Gold, M. G., Biggin, P. C., Gielen, M., & Smart, T. G.
956	(2017). Crystal structures of a GABA(A)-receptor chimera reveal new endogenous
957	neurosteroid-binding sites. Nat Struct Mol Biol, 24(11), 977-985.
958	https://doi.org/10.1038/nsmb.3477
959	Lee, J. H., Zhang, J., Wei, L., & Yu, S. P. (2015). Neurodevelopmental implications of the general
960	anesthesia in neonate and infants. Exp Neurol, 272, 50-60.
961	https://doi.org/10.1016/j.expneurol.2015.03.028
962	Li, B., Feng, X. J., Hu, X. Y., Chen, Y. P., Sha, J. C., Zhang, H. Y., & Fan, H. G. (2018). Effect of melatonin
963	on attenuating the isoflurane-induced oxidative damage is related to PKC $\!\alpha/Nrf2$ signaling
964	pathway in developing rats. Brain Res Bull, 143, 9-18.
965	https://doi.org/10.1016/j.brainresbull.2018.09.018

966	Li, G., Du, J., Wang, L., & Shi, X. (2019). Developmental neurotoxicity in the context of multiple
967	sevoflurane exposures: Potential role of histone deacetylase 6. Neurotoxicol Teratol, 74,
968	106813. https://doi.org/10.1016/j.ntt.2019.106813
969	Li, G. F., Li, Z. B., Zhuang, S. J., & Li, G. C. (2018). Inhibition of microRNA-34a protects against propofol
970	anesthesia-induced neurotoxicity and cognitive dysfunction via the MAPK/ERK signaling
971	pathway. Neuroscience Letters, 675, 152-159.
972	https://doi.org/https://doi.org/10.1016/j.neulet.2018.03.052
973	Li, J., Guo, M., Liu, Y., Wu, G., Miao, L., Zhang, J., Zuo, Z., & Li, Y. (2019). Both GSK-3β/CRMP2 and
974	CDK5/CRMP2 Pathways Participate in the Protection of Dexmedetomidine Against Propofol-
975	Induced Learning and Memory Impairment in Neonatal Rats. Toxicol Sci, 171(1), 193-210.
976	https://doi.org/10.1093/toxsci/kfz135
977	Li, J., Wang, B., Wu, H., Yu, Y., Xue, G., & Hou, Y. (2014). 17β-estradiol attenuates ketamine-induced
978	neuroapoptosis and persistent cognitive deficits in the developing brain. Brain Res, 1593, 30-
979	39. https://doi.org/10.1016/j.brainres.2014.09.013
980	Li, W., Li, H., Wei, H., Lu, Y., Lei, S., Zheng, J., Lu, H., Chen, X., Liu, Y., & Zhang, P. (2019). 17β-Estradiol
981	Treatment Attenuates Neurogenesis Damage and Improves Behavior Performance After
982	Ketamine Exposure in Neonatal Rats. Front Cell Neurosci, 13, 251.
983	https://doi.org/10.3389/fncel.2019.00251
984	Li, Y., Chen, L., Zhu, D., Chen, Y., Qin, W., & Cui, J. (2020). Propofol downregulates the activity of
985	glutamatergic neurons in the basal forebrain via affecting intrinsic membrane properties and
986	postsynaptic GABAARs. Neuroreport, 31(17), 1242-1248.
987	https://doi.org/10.1097/wnr.000000000001540
988	Li, Y., Li, X., Zhao, J., Li, L., Wang, Y., Zhang, Y., Chen, Y., Liu, W., & Gao, L. (2018). Midazolam
989	Attenuates Autophagy and Apoptosis Caused by Ketamine by Decreasing Reactive Oxygen
990	Species in the Hippocampus of Fetal Rats. Neuroscience, 388, 460-471.
991	https://doi.org/10.1016/j.neuroscience.2018.03.040
992	Liao, Z., Huang, Z., Li, J., Li, H., Miao, L., Liu, Y., Zhang, J., Xu, Y., & Li, Y. (2021). Regulation of CRMP2 b
993	$Cdk5\ and\ GSK-3\beta\ participates\ in\ sevoflurane-induced\ dendritic\ development\ abnormalities$
994	and cognitive dysfunction in developing rats. Toxicol Lett, 341, 68-79.
995	https://doi.org/10.1016/j.toxlet.2021.01.023
996	Liu, B., Ou, G., Chen, Y., & Zhang, J. (2019). Inhibition of protein tyrosine phosphatase 1B protects
997	against sevoflurane-induced neurotoxicity mediated by ER stress in developing brain. Brain
998	Res Bull, 146, 28-39. https://doi.org/10.1016/j.brainresbull.2018.12.006
999	Liu, B., Xia, J., Chen, Y., & Zhang, J. (2017). Sevoflurane-Induced Endoplasmic Reticulum Stress
1000	Contributes to Neuroapoptosis and BACE-1 Expression in the Developing Brain: The Role of
1001	elF2a. Neurotox Res, 31(2), 218-229. https://doi.org/10.1007/s12640-016-9671-z
1002	Liu, F., Rainosek, S. W., Sadovova, N., Fogle, C. M., Patterson, T. A., Hanig, J. P., Paule, M. G., Slikker,
1003	W., Jr., & Wang, C. (2014). Protective effect of acetyl-L-carnitine on propofol-induced toxicity
1004	in embryonic neural stem cells. Neurotoxicology, 42, 49-57.
1005	https://doi.org/10.1016/j.neuro.2014.03.011
1006	Liu, J. R., Yuki, K., Baek, C., Han, X. H., & Soriano, S. G. (2016). Dexmedetomidine-Induced
1007	Neuroapoptosis Is Dependent on Its Cumulative Dose. Anesth Analg, 123(4), 1008-1017.
1008	https://doi.org/10.1213/ane.000000000001527

1009	Liu, Y., Yan, Y., Inagaki, Y., Logan, S., Bosnjak, Z. J., & Bai, X. (2017). Insufficient Astrocyte-Derived
1010	Brain-Derived Neurotrophic Factor Contributes to Propofol-Induced Neuron Death Through
1011	Akt/Glycogen Synthase Kinase 3β/Mitochondrial Fission Pathway. Anesth Analg, 125(1), 241-
1012	254. https://doi.org/10.1213/ane.000000000002137
1013	Lu, H., Liufu, N., Dong, Y., Xu, G., Zhang, Y., Shu, L., Soriano, S. G., Zheng, H., Yu, B., & Xie, Z. (2017).
1014	Sevoflurane Acts on Ubiquitination-Proteasome Pathway to Reduce Postsynaptic Density 95
1015	Protein Levels in Young Mice. Anesthesiology, 127(6), 961-975.
1016	https://doi.org/10.1097/aln.000000000001889
1017	Lu, L. X., Yon, J. H., Carter, L. B., & Jevtovic-Todorovic, V. (2006). General anesthesia activates BDNF-
1018	dependent neuroapoptosis in the developing rat brain. Apoptosis, 11(9), 1603-1615.
1019	https://doi.org/10.1007/s10495-006-8762-3
1020	Lunardi, N., Ori, C., Erisir, A., & Jevtovic-Todorovic, V. (2010). General anesthesia causes long-lasting
1021	disturbances in the ultrastructural properties of developing synapses in young rats. Neurotox
1022	Res, 17(2), 179-188. https://doi.org/10.1007/s12640-009-9088-z
1023	Maksimovic, S., Useinovic, N., Quillinan, N., Covey, D. F., Todorovic, S. M., & Jevtovic-Todorovic, V.
1024	(2022). General Anesthesia and the Young Brain: The Importance of Novel Strategies with
1025	Alternate Mechanisms of Action. Int J Mol Sci, 23(3). https://doi.org/10.3390/ijms23031889
1026	Maloney, S. E., Creeley, C. E., Hartman, R. E., Yuede, C. M., Zorumski, C. F., Jevtovic-Todorovic, V.,
1027	Dikranian, K., Noguchi, K. K., Farber, N. B., & Wozniak, D. F. (2019). Using animal models to
1028	evaluate the functional consequences of anesthesia during early neurodevelopment.
1029	Neurobiol Learn Mem, 165, 106834. https://doi.org/10.1016/j.nlm.2018.03.014
1030	Marchi, S., Patergnani, S., Missiroli, S., Morciano, G., Rimessi, A., Wieckowski, M. R., Giorgi, C., &
1031	Pinton, P. (2018). Mitochondrial and endoplasmic reticulum calcium homeostasis and cell
1032	death. Cell Calcium, 69, 62-72. https://doi.org/10.1016/j.ceca.2017.05.003
1033	McCann, M. E., de Graaff, J. C., Dorris, L., Disma, N., Withington, D., Bell, G., Grobler, A., Stargatt, R.,
1034	Hunt, R. W., Sheppard, S. J., Marmor, J., Giribaldi, G., Bellinger, D. C., Hartmann, P. L., Hardy,
1035	P., Frawley, G., Izzo, F., von Ungern Sternberg, B. S., Lynn, A.,Davidson, A. J. (2019).
1036	Neurodevelopmental outcome at 5 years of age after general anaesthesia or awake-regional
1037	anaesthesia in infancy (GAS): an international, multicentre, randomised, controlled
1038	equivalence trial. Lancet, 393(10172), 664-677. https://doi.org/10.1016/s0140-
1039	<u>6736(18)32485-1</u>
1040	Mendiola, A. S., & Cardona, A. E. (2018). The IL-1 β phenomena in neuroinflammatory diseases. J
1041	Neural Transm (Vienna), 125(5), 781-795. https://doi.org/10.1007/s00702-017-1732-9
1042	Miao, H. H., Liu, W. B., Jiao, X. H., Shao, K. J., Yuan, Y. X., Sha, S., Zhang, Q. Q., Yan, J., Sun, Y. Y., Zhou,
1043	C. H., & Wu, Y. Q. (2022). Neonatal Exposure to Propofol Interferes with the Proliferation and
1044	Differentiation of Hippocampal Neural Stem Cells and the Neurocognitive Function of Rats in
1045	Adulthood via the Akt/p27 Signaling Pathway. Biomed Environ Sci., 35(4), 283-295.
1046	https://doi.org/10.3967/bes2022.040
1047	Milanovic, D., Pesic, V., Loncarevic-Vasiljkovic, N., Pavkovic, Z., Popic, J., Kanazir, S., Jevtovic-
1048	Todorovic, V., & Ruzdijic, S. (2016). The Fas Ligand/Fas Death Receptor Pathways Contribute
1049	to Propofol-Induced Apoptosis and Neuroinflammation in the Brain of Neonatal Rats.
1050	Neurotox Res. 30(3), 434-452, https://doi.org/10.1007/s12640-016-9629-1

1051	Milesente D. Desie I. Desie V. Leavennie Verliterte N. Kennete C. Leaterte Tedennie V. O.
1051	Milanovic, D., Popic, J., Pesic, V., Loncarevic-Vasiljkovic, N., Kanazir, S., Jevtovic-Todorovic, V., &
1052	Ruzdijic, S. (2010). Regional and temporal profiles of calpain and caspase-3 activities in
1053	postnatal rat brain following repeated propofol administration. <i>Dev Neurosci, 32</i> (4), 288-301.
1054	https://doi.org/10.1159/000316970
1055	Möller, T., Bard, F., Bhattacharya, A., Biber, K., Campbell, B., Dale, E., Eder, C., Gan, L., Garden, G. A.,
1056	Hughes, Z. A., Pearse, D. D., Staal, R. G., Sayed, F. A., Wes, P. D., & Boddeke, H. W. (2016).
1057	Critical data-based re-evaluation of minocycline as a putative specific microglia inhibitor.
1058	Glia, 64(10), 1788-1794. https://doi.org/10.1002/glia.23007
1059	Mongerson, C. R. L., Wilcox, S. L., Goins, S. M., Pier, D. B., Zurakowski, D., Jennings, R. W., & Bajic, D.
1060	(2019). Infant Brain Structural MRI Analysis in the Context of Thoracic Non-cardiac Surgery
1061	and Critical Care. Front Pediatr, 7, 315. https://doi.org/10.3389/fped.2019.00315
1062	Moran, M. M., Gunn-Charlton, J. K., Walsh, J. M., Cheong, J. L. Y., Anderson, P. J., Doyle, L. W.,
1063	Greaves, S., & Hunt, R. W. (2019). Associations of Neonatal Noncardiac Surgery with Brain
1064	Structure and Neurodevelopment: A Prospective Case-Control Study. J Pediatr, 212, 93-
1065	101.e102. https://doi.org/10.1016/j.jpeds.2019.05.050
1066	Naseri, M., Parham, A., & Moghimi, A. (2017). The effect of sodium thiopental as a GABA mimetic
1067	drug in neonatal period on expression of GAD65 and GAD67 genes in hippocampus of
1068	newborn and adult male rats. Iran J Basic Med Sci, 20(9), 996-1001.
1069	https://doi.org/10.22038/ijbms.2017.9264
1070	Nilsen, J., Chen, S., & Brinton, R. D. (2002). Dual action of estrogen on glutamate-induced calcium
1071	signaling: mechanisms requiring interaction between estrogen receptors and src/mitogen
1072	activated protein kinase pathway. Brain Res, 930(1-2), 216-234.
1073	https://doi.org/10.1016/s0006-8993(02)02254-0
1074	Oh, W. C., Lutzu, S., Castillo, P. E., & Kwon, H. B. (2016). De novo synaptogenesis induced by GABA in
1075	the developing mouse cortex. Science, 353(6303), 1037-1040.
1076	https://doi.org/10.1126/science.aaf5206
1077	Pellegrini, L., Bennis, Y., Velly, L., Grandvuillemin, I., Pisano, P., Bruder, N., & Guillet, B. (2014).
1078	Erythropoietin protects newborn rat against sevoflurane-induced neurotoxicity. Paediatr
1079	Anaesth, 24(7), 749-759. https://doi.org/10.1111/pan.12372
1080	Perez-Zoghbi, J. F., Zhu, W., Grafe, M. R., & Brambrink, A. M. (2017). Dexmedetomidine-mediated
1081	neuroprotection against sevoflurane-induced neurotoxicity extends to several brain regions
1082	in neonatal rats. Br J Anaesth, 119(3), 506-516. https://doi.org/10.1093/bja/aex222
1083	Perez-Zoghbi, J. F., Zhu, W., Neudecker, V., Grafe, M. R., & Brambrink, A. M. (2020). Neurotoxicity of
1084	sub-anesthetic doses of sevoflurane and dexmedetomidine co-administration in neonatal
1085	rats. Neurotoxicology, 79, 75-83. https://doi.org/10.1016/j.neuro.2020.03.014
1086	Piao, M., Wang, Y., Liu, N., Wang, X., Chen, R., Qin, J., Ge, P., & Feng, C. (2020). Sevoflurane Exposure
1087	Induces Neuronal Cell Parthanatos Initiated by DNA Damage in the Developing Brain via an
1088	Increase of Intracellular Reactive Oxygen Species. Front Cell Neurosci, 14, 583782.
1089	https://doi.org/10.3389/fncel.2020.583782
1090	Popić, J., Pešić, V., Milanović, D., Lončarević-Vasiljković, N., Smiljanić, K., Kanazir, S., & Ruždijić, S.
1091	(2015). Induction of TNF-α signaling cascade in neonatal rat brain during propofol anesthesia.
1092	Int J Dev Neurosci, 44, 22-32. https://doi.org/10.1016/j.ijdevneu.2015.05.003

1093	Qiao, H., Li, Y., Xu, Z., Li, W., Fu, Z., Wang, Y., King, A., & Wei, H. (2017). Propofol Affects
1094	Neurodegeneration and Neurogenesis by Regulation of Autophagy via Effects on Intracellular
1095	Calcium Homeostasis. Anesthesiology, 127(3), 490-501.
1096	https://doi.org/10.1097/aln.00000000001730
1097	Ramirez, K., Niraula, A., & Sheridan, J. F. (2016). GABAergic modulation with classical benzodiazepines
1098	prevent stress-induced neuro-immune dysregulation and behavioral alterations. Brain Behav
1099	Immun, 51, 154-168. https://doi.org/10.1016/j.bbi.2015.08.011
1100	Robinson, B., Gu, Q., Ali, S. F., Dumas, M., & Kanungo, J. (2019). Ketamine-induced attenuation of
1101	reactive oxygen species in zebrafish is prevented by acetyl l-carnitine in vivo. Neurosci Lett,
1102	706, 36-42. https://doi.org/10.1016/j.neulet.2019.05.009
1103	Rodrigues, D. C., Mufteev, M., Weatheritt, R. J., Djuric, U., Ha, K. C. H., Ross, P. J., Wei, W., Piekna, A.,
1104	Sartori, M. A., Byres, L., Mok, R. S. F., Zaslavsky, K., Pasceri, P., Diamandis, P., Morris, Q.,
1105	Blencowe, B. J., & Ellis, J. (2020). Shifts in Ribosome Engagement Impact Key Gene Sets in
1106	Neurodevelopment and Ubiquitination in Rett Syndrome. Cell Rep., 30(12), 4179-4196.e4111.
1107	https://doi.org/10.1016/j.celrep.2020.02.107
1108	Rosado-Mendez, I. M., Noguchi, K. K., Castañeda-Martinez, L., Kirvassilis, G., Wang, S. H., Manzella, F.,
1109	Swiney, B. S., Masuoka, K., Capuano, S., III, Brunner, K. G., Crosno, K., Guerrero, Q. W.,
1110	Whitson, H., Brambrink, A., Simmons, H. S., Mejia, A. F., Zagzebski, J. A., Hall, T. J., &
1111	Ikonomidou, C. (2019). Quantitative ultrasound and apoptotic death in the neonatal primate
1112	brain. Neurobiol Dis, 127, 554-562. https://doi.org/10.1016/j.nbd.2019.03.032
1113	Sanchez, V., Feinstein, S. D., Lunardi, N., Joksovic, P. M., Boscolo, A., Todorovic, S. M., & Jevtovic-
1114	Todorovic, V. (2011). General Anesthesia Causes Long-term Impairment of Mitochondrial
1115	Morphogenesis and Synaptic Transmission in Developing Rat Brain. Anesthesiology, 115(5),
1116	992-1002. https://doi.org/10.1097/ALN.0b013e3182303a63
1117	Schaefer, M. L., Wang, M., Perez, P. J., Coca Peralta, W., Xu, J., & Johns, R. A. (2019). Nitric Oxide
1118	Donor Prevents Neonatal Isoflurane-induced Impairments in Synaptic Plasticity and Memory.
1119	Anesthesiology, 130(2), 247-262. https://doi.org/10.1097/aln.000000000002529
1120	Schenning, K. J., Noguchi, K. K., Martin, L. D., Manzella, F. M., Cabrera, O. H., Dissen, G. A., &
1121	Brambrink, A. M. (2017). Isoflurane exposure leads to apoptosis of neurons and
1122	oligodendrocytes in 20- and 40-day old rhesus macaques. Neurotoxicol Teratol, 60, 63-68.
1123	https://doi.org/10.1016/j.ntt.2016.11.006
1124	Semple, B. D., Blomgren, K., Gimlin, K., Ferriero, D. M., & Noble-Haeusslein, L. J. (2013). Brain
1125	development in rodents and humans: Identifying benchmarks of maturation and
1126	vulnerability to injury across species. Prog Neurobiol, 106-107, 1-16.
1127	https://doi.org/10.1016/j.pneurobio.2013.04.001
1128	Serrao, J. M., & Goodchild, C. S. (2022). Alfaxalone anaesthesia increases brain derived neurotrophic
1129	factor levels and preserves postoperative cognition by activating pregnane-X receptors: an in
1130	vitro study and a double blind randomised controlled trial. BMC Anesthesiol, 22(1), 401.
1131	https://doi.org/10.1186/s12871-022-01940-x
1132	Shi, W., Wu, X., Yuan, C., Kuang, T., Xie, X., Gong, W., Li, F., Shen, L., Zhang, Y., & Liang, X. (2024).
1133	Effect of remimazolam toluene sulfonate on the cognitive function of juveniles and its
1134	mechanism of action. Eur J Med Res, 29(1), 543. https://doi.org/10.1186/s40001-024-02142-
1135	6

1136	Shibuta, S., Morita, T., & Kosaka, J. (2022). Effect of preconditioning on propofol-induced
1137	neurotoxicity during the developmental period. PLoS One, 17(8), e0273219.
1138	https://doi.org/10.1371/journal.pone.0273219
1139	Shu, P., Wu, C., Liu, W., Ruan, X., Liu, C., Hou, L., Zeng, Y., Fu, H., Wang, M., Chen, P., Zhang, X., Yin, B.,
1140	Yuan, J., Qiang, B., & Peng, X. (2019). The spatiotemporal expression pattern of microRNAs in
1141	the developing mouse nervous system. J Biol Chem, 294(10), 3444-3453.
1142	https://doi.org/10.1074/jbc.RA118.004390
1143	Slikker, W., Jr., Zou, X., Hotchkiss, C. E., Divine, R. L., Sadovova, N., Twaddle, N. C., Doerge, D. R.,
1144	Scallet, A. C., Patterson, T. A., Hanig, J. P., Paule, M. G., & Wang, C. (2007). Ketamine-induced
1145	neuronal cell death in the perinatal rhesus monkey. Toxicol Sci, 98(1), 145-158.
1146	https://doi.org/10.1093/toxsci/kfm084
1147	Soyalp, C., Oksuz, E., Gorgisen, G., Gulacar, I. M., Yasar, S., Tuncdemir, Y. E., & Delen, L. A. (2022). Role
1148	of Sedative-Hypnotic Agents in Neurodegeneration: Effects of Midazolam and Thiopental on
1149	Apoptosis and Oxidative Stress Expression in Neonatal and Adult Rat Brains. Turk Neurosurg,
1150	32(3), 378-385. https://doi.org/10.5137/1019-5149.Jtn.32324-20.2
1151	Stolwijk, L. J., Keunen, K., de Vries, L. S., Groenendaal, F., van der Zee, D. C., van Herwaarden, M. Y. A.,
1152	Lemmers, P. M. A., & Benders, M. (2017). Neonatal Surgery for Noncardiac Congenital
1153	Anomalies: Neonates at Risk of Brain Injury. J Pediatr, 182, 335-341.e331.
1154	https://doi.org/10.1016/j.jpeds.2016.11.080
1155	Stolwijk, L. J., Lemmers, P. M., Harmsen, M., Groenendaal, F., de Vries, L. S., van der Zee, D. C.,
1156	Benders, M. J., & van Herwaarden-Lindeboom, M. Y. (2016). Neurodevelopmental Outcomes
1157	After Neonatal Surgery for Major Noncardiac Anomalies. Pediatrics, 137(2), e20151728.
1158	https://doi.org/10.1542/peds.2015-1728
1159	Straiko, M. M., Young, C., Cattano, D., Creeley, C. E., Wang, H., Smith, D. J., Johnson, S. A., Li, E. S., &
1160	Olney, J. W. (2009). Lithium protects against anesthesia-induced developmental
1161	neuroapoptosis. Anesthesiology, 110(4), 862-868.
1162	https://doi.org/10.1097/ALN.0b013e31819b5eab
1163	Sun, L. S., Li, G., Miller, T. L., Salorio, C., Byrne, M. W., Bellinger, D. C., Ing, C., Park, R., Radcliffe, J.,
1164	Hays, S. R., DiMaggio, C. J., Cooper, T. J., Rauh, V., Maxwell, L. G., Youn, A., & McGowan, F. X.
1165	(2016). Association Between a Single General Anesthesia Exposure Before Age 36 Months
1166	and Neurocognitive Outcomes in Later Childhood. Jama, 315(21), 2312-2320.
1167	https://doi.org/10.1001/jama.2016.6967
1168	Sun, M., Dong, Y., Li, M., Zhang, Y., Liang, F., Zhang, J., Soriano, S. G., & Xie, Z. (2021).
1169	Dexmedetomidine and Clonidine Attenuate Sevoflurane-Induced Tau Phosphorylation and
1170	Cognitive Impairment in Young Mice via α-2 Adrenergic Receptor. Anesth Analg, 132(3), 878-
1171	889. https://doi.org/10.1213/ane.00000000005268
1172	Szymanski, J., & Minichiello, L. (2022). NKCC1 Deficiency in Forming Hippocampal Circuits Triggers
1173	Neurodevelopmental Disorder: Role of BDNF-TrkB Signalling. Brain Sci, 12(4).
1174	https://doi.org/10.3390/brainsci12040502
1175	Tang, X. L., Wang, X., Fang, G., Zhao, Y. L., Yan, J., Zhou, Z., Sun, R., Luo, A. L., & Li, S. Y. (2021).
1176	Resveratrol ameliorates sevoflurane-induced cognitive impairment by activating the
1177	SIRT1/NF-кB pathway in neonatal mice. J Nutr Biochem, 90, 108579.
1178	https://doi.org/10.1016/j.jnutbio.2020.108579

1180	Sevoflurane induces tau phosphorylation and glycogen synthase kinase $3eta$ activation in
1181	young mice. Anesthesiology, 121(3), 510-527.
1182	https://doi.org/10.1097/aln.000000000000278
1183	Tateiwa, H., & Evers, A. S. (2024). Neurosteroids and their potential as a safer class of general
1184	anesthetics. J Anesth, 38(2), 261-274. https://doi.org/10.1007/s00540-023-03291-4
1185	Taylor, G. T., Manzella, F. M., Huffman, J., Cabrera, O. H., & Hoffman, J. (2017). Cognition in female
1186	rats after blocking conversion of androgens to estrogens. Horm Behav, 90, 84-89.
1187	https://doi.org/10.1016/j.yhbeh.2017.02.011
1188	Tesic, V., Joksimovic, S. M., Quillinan, N., Krishnan, K., Covey, D. F., Todorovic, S. M., & Jevtovic-
1189	Todorovic, V. (2020). Neuroactive steroids alphaxalone and CDNC24 are effective hypnotics
1190	and potentiators of GABA(A) currents, but are not neurotoxic to the developing rat brain. Br $\it J$
1191	Anaesth, 124(5), 603-613. https://doi.org/10.1016/j.bja.2020.01.013
1192	Thomason, E. J., Escalante, M., Osterhout, D. J., & Fuss, B. (2020). The oligodendrocyte growth cone
1193	and its actin cytoskeleton: A fundamental element for progenitor cell migration and CNS
1194	myelination. Glia, 68(7), 1329-1346. https://doi.org/10.1002/glia.23735
1195	Timic Stamenic, T., Feseha, S., Manzella, F. M., Wallace, D., Wilkey, D., Corrigan, T., Fiedler, H., Doerr,
1196	P., Krishnan, K., Raol, Y. H., Covey, D. F., Jevtovic-Todorovic, V., & Todorovic, S. M. (2021).
1197	The T-type calcium channel isoform Ca(v)3.1 is a target for the hypnotic effect of the
1198	anaesthetic neurosteroid (3 β ,5 β ,17 β)-3-hydroxyandrostane-17-carbonitrile. Br J Anaesth,
1199	126(1), 245-255. https://doi.org/10.1016/j.bja.2020.07.022
1200	Tu, Y., Liang, Y., Xiao, Y., Lv, J., Guan, R., Xiao, F., Xie, Y., & Xiao, Q. (2019). Dexmedetomidine
1201	attenuates the neurotoxicity of propofol toward primary hippocampal neurons in vitro via
1202	Erk1/2/CREB/BDNF signaling pathways. Drug Des Devel Ther, 13, 695-706.
1203	https://doi.org/10.2147/dddt.S188436
1204	Vanderhaeghen, P., & Polleux, F. (2023). Developmental mechanisms underlying the evolution of
1205	human cortical circuits. Nat Rev Neurosci, 24(4), 213-232. https://doi.org/10.1038/s41583-
1206	<u>023-00675-z</u>
1207	Wan, J., Shen, C. M., Wang, Y., Wu, Q. Z., Wang, Y. L., Liu, Q., Sun, Y. M., Cao, J. P., & Wu, Y. Q. (2021).
1208	Repeated exposure to propofol in the neonatal period impairs hippocampal synaptic
1209	plasticity and the recognition function of rats in adulthood. Brain Res Bull, 169, 63-72.
1210	https://doi.org/10.1016/j.brainresbull.2021.01.007
1211	Wang, N., & Wang, M. (2019). Dexmedetomidine suppresses sevoflurane anesthesia-induced
1212	neuroinflammation through activation of the PI3K/Akt/mTOR pathway. BMC Anesthesiol,
1213	19(1), 134. https://doi.org/10.1186/s12871-019-0808-5
1214	Wang, W. Y., Jia, L. J., Luo, Y., Zhang, H. H., Cai, F., Mao, H., Xu, W. C., Fang, J. B., Peng, Z. Y., Ma, Z. W.,
1215	Chen, Y. H., Zhang, J., Wei, Z., Yu, B. W., & Hu, S. F. (2016). Location- and Subunit-Specific
1216	NMDA Receptors Determine the Developmental Sevoflurane Neurotoxicity Through ERK1/2
1217	Signaling. Mol Neurobiol, 53(1), 216-230. https://doi.org/10.1007/s12035-014-9005-1
1218	Wang, X., Shan, Y., Tang, Z., Gao, L., & Liu, H. (2019). Neuroprotective effects of dexmedetomidine
1219	against isoflurane-induced neuronal injury via glutamate regulation in neonatal rats. Drug
1220	Des Devel Ther, 13, 153-160. https://doi.org/10.2147/dddt.S163197

Tao, G., Zhang, J., Zhang, L., Dong, Y., Yu, B., Crosby, G., Culley, D. J., Zhang, Y., & Xie, Z. (2014).

1221	wang, A., Zhao, B., & Li, A. (2013). Dexinedetornidine attendates isondrane-induced cognitive
1222	impairment through antioxidant, anti-inflammatory and anti-apoptosis in aging rat. Int J Clin
1223	Exp Med, 8(10), 17281-17288.
1224	Ward, R. J., Zucca, F. A., Duyn, J. H., Crichton, R. R., & Zecca, L. (2014). The role of iron in brain ageing
1225	and neurodegenerative disorders. Lancet Neurol, 13(10), 1045-1060.
1226	https://doi.org/10.1016/s1474-4422(14)70117-6
1227	Watanabe, M., & Fukuda, A. (2015). Development and regulation of chloride homeostasis in the
1228	central nervous system. Front Cell Neurosci, 9, 371.
1229	https://doi.org/10.3389/fncel.2015.00371
1230	Wei, Y., Hu, J., Liang, Y., Zhong, Y., He, D., Qin, Y., Li, L., Chen, J., Xiao, Q., & Xie, Y. (2016).
1231	Dexmedetomidine pretreatment attenuates propofol-induced neurotoxicity in neuronal
1232	cultures from the rat hippocampus. Mol Med Rep, 14(4), 3413-3420.
1233	https://doi.org/10.3892/mmr.2016.5628
1234	Weiss, M., Hansen, T. G., & Engelhardt, T. (2016). Ensuring safe anaesthesia for neonates, infants and
1235	young children: what really matters. Arch Dis Child, 101(7), 650-652.
1236	https://doi.org/10.1136/archdischild-2015-310104
1237	Wu, S. Y., Pan, B. S., Tsai, S. F., Chiang, Y. T., Huang, B. M., Mo, F. E., & Kuo, Y. M. (2020). BDNF
1238	reverses aging-related microglial activation. J Neuroinflammation, 17(1), 210.
1239	https://doi.org/10.1186/s12974-020-01887-1
1240	Wu, Z., Xue, H., Gao, Q., & Zhao, P. (2020). Effects of early postnatal sevoflurane exposure on
1241	oligodendrocyte maturation and myelination in cerebral white matter of the rat. Biomed
1242	Pharmacother, 131, 110733. https://doi.org/10.1016/j.biopha.2020.110733
1243	Wu, Z., Zhang, Y., Zhang, Y., & Zhao, P. (2020). Sirtuin 2 Inhibition Attenuates Sevoflurane-Induced
1244	Learning and Memory Deficits in Developing Rats via Modulating Microglial Activation. Cell
1245	Mol Neurobiol, 40(3), 437-446. https://doi.org/10.1007/s10571-019-00746-9
1246	Xian, F., Li, Q., & Chen, Z. (2019). Overexpression of phosphoprotein enriched in astrocytes 15
1247	reverses the damage induced by propofol in hippocampal neurons. Mol Med Rep, 20(2),
1248	1583-1592. https://doi.org/10.3892/mmr.2019.10412
1249	Xiao, Y., Zhou, L., Tu, Y., Li, Y., Liang, Y., Zhang, X., Lv, J., Zhong, Y., & Xie, Y. (2018). Dexmedetomidine
1250	attenuates the propofol-induced long-term neurotoxicity in the developing brain of rats by
1251	enhancing the PI3K/Akt signaling pathway. Neuropsychiatr Dis Treat, 14, 2191-2206.
1252	https://doi.org/10.2147/ndt.S169099
1253	Xu, J., Mathena, R. P., Xu, M., Wang, Y., Chang, C., Fang, Y., Zhang, P., & Mintz, C. D. (2018). Early
1254	Developmental Exposure to General Anesthetic Agents in Primary Neuron Culture Disrupts
1255	Synapse Formation via Actions on the mTOR Pathway. Int J Mol Sci, 19(8).
1256	https://doi.org/10.3390/ijms19082183
1257	Yan, J., Huang, Y., Lu, Y., Chen, J., & Jiang, H. (2014). Repeated administration of ketamine can induce
1258	hippocampal neurodegeneration and long-term cognitive impairment via the ROS/HIF-1 α
1259	pathway in developing rats. Cell Physiol Biochem, 33(6), 1715-1732.
1260	https://doi.org/10.1159/000362953
1261	Yang, F., Zhang, Y., Tang, Z., Shan, Y., Wu, X., & Liu, H. (2020). Hemin treatment protects neonatal rate
1262	from sevoflurane-induced neurotoxicity via the phosphoinositide 3-kinase/Akt pathway. Life
1263	Sci 242 117151 https://doi.org/10.1016/j.lfs.2019.117151

1264	Yang, M., Lian, N., Yu, Y., Wang, Y., Xie, K., & Yu, Y. (2020). Coenzyme Q10 alleviates
1265	sevoflurane-induced neuroinflammation by regulating the levels of apolipoprotein E and
1266	phosphorylated tau protein in mouse hippocampal neurons. Mol Med Rep, 22(1), 445-453.
1267	https://doi.org/10.3892/mmr.2020.11131
1268	Yang, M., & Wei, H. (2017). Anesthetic neurotoxicity: Apoptosis and autophagic cell death mediated
1269	by calcium dysregulation. Neurotoxicol Teratol, 60, 59-62.
1270	https://doi.org/10.1016/j.ntt.2016.11.004
1271	Yang, X., Lin, C., Chen, S., Huang, Y., Cheng, Q., & Yao, Y. (2022). Remimazolam for the Prevention of
1272	Emergence Delirium in Children Following Tonsillectomy and Adenoidectomy Under
1273	Sevoflurane Anesthesia: A Randomized Controlled Study. Drug Des Devel Ther, 16, 3413-
1274	3420. https://doi.org/10.2147/dddt.S381611
1275	Yang, Y., Yi, J., Pan, M., Hu, B., & Duan, H. (2021). Edaravone Alleviated Propofol-Induced
1276	Neurotoxicity in Developing Hippocampus by mBDNF/TrkB/PI3K Pathway. Drug Des Devel
1277	Ther, 15, 1409-1422. https://doi.org/10.2147/dddt.S294557
1278	Ye, X., Lian, Q., Eckenhoff, M. F., Eckenhoff, R. G., & Pan, J. Z. (2013). Differential general anesthetic
1279	effects on microglial cytokine expression. PLoS One, 8(1), e52887.
1280	https://doi.org/10.1371/journal.pone.0052887
1281	Yeh, F. L., Hansen, D. V., & Sheng, M. (2017). TREM2, Microglia, and Neurodegenerative Diseases.
1282	Trends Mol Med, 23(6), 512-533. https://doi.org/10.1016/j.molmed.2017.03.008
1283	Yon, J. H., Carter, L. B., Reiter, R. J., & Jevtovic-Todorovic, V. (2006). Melatonin reduces the severity of
1284	anesthesia-induced apoptotic neurodegeneration in the developing rat brain. Neurobiol Dis,
1285	21(3), 522-530. https://doi.org/10.1016/j.nbd.2005.08.011
1286	Yu, Z., Wang, J., Wang, H., Wang, J., Cui, J., & Junzhang, P. (2020). Effects of Sevoflurane Exposure
1287	During Late Pregnancy on Brain Development and Beneficial Effects of Enriched Environment
1288	on Offspring Cognition. Cell Mol Neurobiol, 40(8), 1339-1352.
1289	https://doi.org/10.1007/s10571-020-00821-6
1290	Zeng, Z., Yao, J., Zhong, J., Fan, S., Xue, Y., Chen, X., Luo, Y., & Xue, S. (2021). The Role of the IncRNA-
1291	LRCF in Propofol-Induced Oligodendrocyte Damage in Neonatal Mouse. Neurochem Res,
1292	46(4), 778-791. https://doi.org/10.1007/s11064-020-03205-w
1293	Zhang, H., Li, J., Su, X., Hu, Y., Liu, T., Ni, S., Li, H., Zuo, X. N., Fu, J., Yuan, T. F., & Yang, Z. (2022).
1294	Growth charts of brain morphometry for preschool children. Neuroimage, 255, 119178.
1295	https://doi.org/10.1016/j.neuroimage.2022.119178
1296	Zhang, L., Cheng, Y., Xue, Z., Li, J., Wu, N., Yan, J., Wang, J., Wang, C., Chen, W., Zhou, T., Qiu, Z., &
1297	Jiang, H. (2022). Sevoflurane impairs m6A-mediated mRNA translation and leads to fine
1298	motor and cognitive deficits. Cell Biol Toxicol, 38(2), 347-369.
1299	https://doi.org/10.1007/s10565-021-09601-4
1300	Zhang, S., Liang, Z., Sun, W., & Pei, L. (2017). Repeated propofol anesthesia induced downregulation
1301	of hippocampal miR-132 and learning and memory impairment of rats. Brain Res, 1670, 156-
1302	164. https://doi.org/10.1016/j.brainres.2017.04.011
1303	Zhang, Y., Xiao, X., Zhang, X. M., Zhao, Z. Q., & Zhang, Y. Q. (2012). Estrogen facilitates spinal cord
1304	synaptic transmission via membrane-bound estrogen receptors: implications for pain
1305	hypersensitivity. J Biol Chem, 287(40), 33268-33281.
1306	https://doi.org/10.1074/jbc.M112.368142

1307	Zhao, T., Li, C., Wei, W., Zhang, H., Ma, D., Song, X., & Zhou, L. (2016). Prenatal ketamine exposure
1308	causes abnormal development of prefrontal cortex in rat. Sci Rep, 6, 26865.
1309	https://doi.org/10.1038/srep26865
1310	Zhao, X., Li, Z., Guo, Y., Liu, Q., Qing, M., Sheng, Y., Chen, Y., Xie, L., & Zhou, Z. (2023). Alfaxalone
1311	Alleviates Neurotoxicity and Cognitive Impairment Induced by Isoflurane Anesthesia in
1312	Offspring Rats. Anesth Analg, 136(6), 1206-1216.
1313	https://doi.org/10.1213/ane.0000000000006420
1314	Zheng, H., Dong, Y., Xu, Z., Crosby, G., Culley, D. J., Zhang, Y., & Xie, Z. (2013). Sevoflurane anesthesia
1315	in pregnant mice induces neurotoxicity in fetal and offspring mice. Anesthesiology, 118(3),
1316	516-526. https://doi.org/10.1097/ALN.0b013e3182834d5d
1317	Zhou, B., Chen, L., Liao, P., Huang, L., Chen, Z., Liao, D., Yang, L., Wang, J., Yu, G., Wang, L., Zhang, J.,
1318	Zuo, Y., Liu, J., & Jiang, R. (2019). Astroglial dysfunctions drive aberrant synaptogenesis and
1319	social behavioral deficits in mice with neonatal exposure to lengthy general anesthesia. PLoS
1320	Biol, 17(8), e3000086. https://doi.org/10.1371/journal.pbio.3000086
1321	Zhou, Z. W., Shu, Y., Li, M., Guo, X., Pac-Soo, C., Maze, M., & Ma, D. (2011). The glutaminergic,
1322	GABAergic, dopaminergic but not cholinergic neurons are susceptible to anaesthesia-induced
1323	cell death in the rat developing brain. Neuroscience, 174, 64-70.
1324	https://doi.org/10.1016/j.neuroscience.2010.10.009
1325	Zhu, R., Liu, L., Mao, T., Wang, X., Li, Y., Li, T., Lv, S., Zeng, S., Fu, N., Li, N., Wang, Y., Sun, M., & Zhang,
1326	J. (2024). Mfn2 regulates mitochondria and mitochondria-associated endoplasmic reticulum
1327	membrane function in neurodegeneration induced by repeated sevoflurane exposure. Exp
1328	Neurol, 377, 114807. https://doi.org/10.1016/j.expneurol.2024.114807
1329	Zuo, Y., Li, B., Xie, J., Ma, Z., Thirupathi, A., Yu, P., Gao, G., Zhou, J., Zhou, C., Xu, H., Chang, Y., & Shi, Z.
1330	(2020). Sevoflurane anesthesia during pregnancy in mice induces cognitive impairment in the
1331	offspring by causing iron deficiency and inhibiting myelinogenesis. Neurochem Int, 135,
1332	104693. https://doi.org/10.1016/j.neuint.2020.104693
1222	
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FIGURES LEGENDS

Fig. 1 Mechanisms of Developmental Neurotoxicity Induced by Anesthetics Targeting GABAA

1337 Receptors

Anesthetic drugs target GABA_A receptors (GABA_AR) in immature neurons, leading to developmental neurotoxicity. Black arrows represent the direct effects of anesthetic drugs on immature neurons, while red arrows indicate the subsequent effects triggered by the inhibition of relevant signaling pathways. The underlying mechanisms include: GA upregulating NKCC1 and downregulating KCC2, thereby enhancing GABA-mediated excitatory signaling, which leads to excessive calcium influx and calcium overload. This overload then triggers neuroinflammation and exacerbates oxidative stress. Through NMDAR-mediated excitatory/inhibitory imbalance (E/I imbalance), synaptic plasticity is disrupted, with a reduction in PSD95 and related proteins, as well as inhibition of long-term potentiation (LTP). Furthermore, GA affects the BDNF signaling pathway by enhancing BDNF/ P75^{NTR}, inhibiting BDNF/TrkB signaling, and causing BDNF dysregulation. Simultaneously, it interferes with abnormal ncRNA and PTP-1B signaling pathways, inducing endoplasmic reticulum stress, which ultimately results in neuronal damage.

Fig. 2 The Effects of Anesthetics Targeting GABAA Receptors on Different Neuronal Populations

Anesthetic agents activate microglia, triggering the release of pro-inflammatory cytokines and M1 markers, which in turn exacerbate neuroinflammation. These drugs induce massive apoptosis in oligodendrocytes, leading to neuronal demyelination. Notably, iron and folate have been shown to mitigate this demyelination process. Furthermore, anesthetics interfere with astrocyte function via the ERK pathway, disrupting BDNF metabolism and ultimately leading to neuronal apoptosis. This cascade of events underscores the harmful effects of anesthetics on neurodevelopment, particularly in the context of inflammation-induced apoptosis in both neurons and glial cells.

1358 Fig. 3 Neuroprotective Mechanisms of NAS

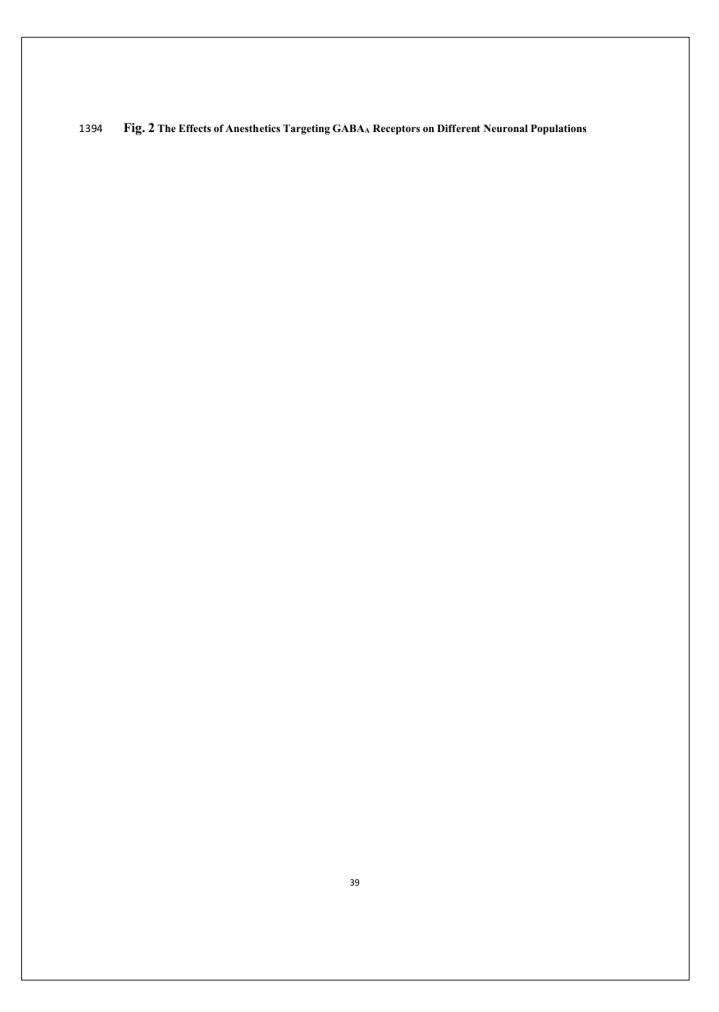
NAS exerts neuroprotective effects through multiple pathways. The mechanisms are as follows: black arrows indicate the direct effects of anesthetics on immature neurons, while red arrows represent subsequent effects induced by the inhibition of related signaling pathways. NAS interacts with TLR2, TLR4, and TLR7 and their adaptor proteins to reduce the production of inflammatory factors, thereby alleviating neuronal inflammation. NAS also regulates the synthesis of mBDNF through receptors such as GPER1, ER α/β , and PXR. mBDNF activates MEK/ERK, PI3K/AKT, and PLC γ /PKC signaling pathways through TrkA/B receptors, inhibiting ROS production, alleviating mitochondrial dysfunction, reducing neuronal apoptosis, enhancing neurotransmitter release, and activating NMDAR to promote LTP formation, thus increasing synaptic plasticity. Additionally, NAS may exert anesthetic effects by modulating GABAARs and calcium ions while enhancing neuronal regeneration to further protect the nervous system. NAS, DEX, and EPO reduce neuroinflammation and exhibit anti-apoptotic and neuroprotective effects through the BDNF/TrkB pathway, while Clobenpropit, Edaravone, and Doxycycline act through the BDNF/PI3K pathway. Melatonin increases BDNF production through Nrf2, while L-Ca, Curcumin, and Resveratrol reduce oxidative stress, and Etanercept reduces the production of inflammatory factors.

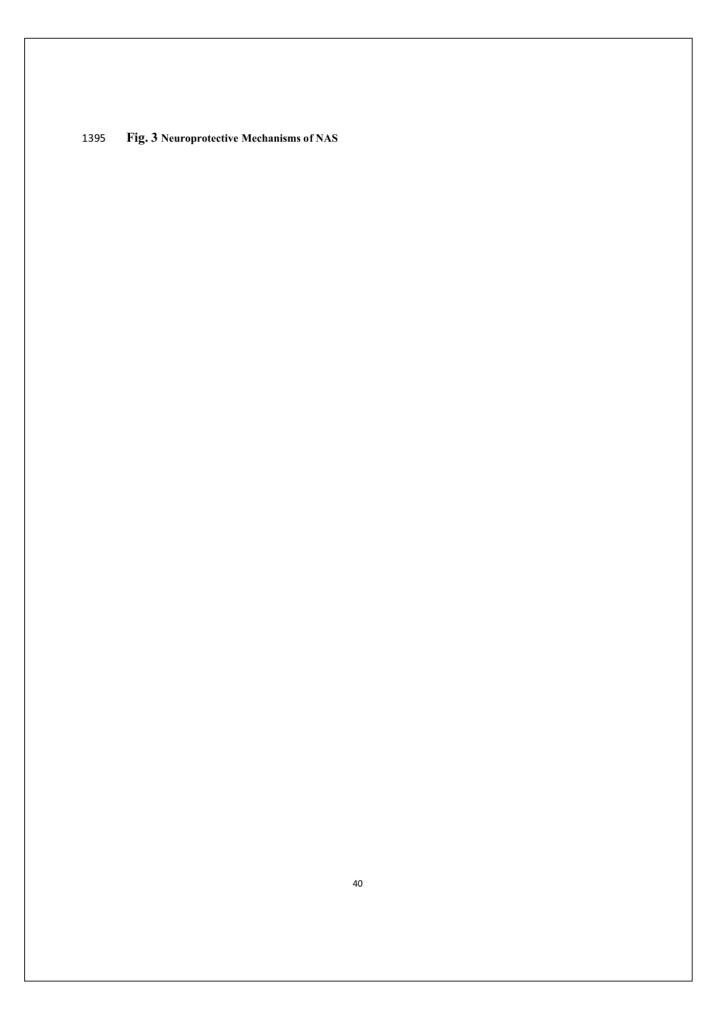
Fig. 4. The Impact of Anesthetic Agents on Different Brain Regions During Development

Neurodevelopmental trajectories based on global pediatric MRI data are presented, with the y-axis representing the percentage of maximum volume attained by each brain region during development, and 1377 the x-axis representing age. The yellow shaded region highlights the period of anesthesia vulnerability, 1378 in comparison to the critical peak developmental periods for various brain functions. The impact of 1379 anesthetic drugs on different brain regions is shown, with highlighted areas indicating regions 1380 particularly vulnerable to anesthesia-induced damage. BGS:brain growth spurt;GMV:total cortical grey 1381 matter volume; WMV: total white matter volume; CSF: total ventricular cerebrospinal fluid volume 1382 (ventricles or cerebrospinal fluid);MCT:mean cortical thickness;TSA:total surface area; TCV:total 1383 cerebrum volume. 1384 Fig. 5 Graphical Abstract: Mechanisms of Anesthetic-Induced Neurotoxicity Targeting GABAA 1385 Receptors and Associated Susceptibility Factors 1386 Neonatal brain development occurs during a critical period, and anesthetic-induced neurotoxicity during 1387 this time is influenced by multiple complex factors, including the mechanisms of anesthetic drugs 1388 targeting GABA receptors and associated susceptibility factors. The interplay of these factors contributes 1389 to the formation of a complex mechanism underlying neonatal neurotoxicity induced by anesthesia.

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1391 1392	Fig. 1 Mechanisms of Developmental Neurotoxicity Induced by Anesthetics Targeting GABA _A Receptors
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1396	Fig. 4 The Impact of Anesthetic Agents on Different Brain Regions During Development
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1399	Fig. 5 Graphical Abstract: Mechanisms of Anesthetic-Induced Neurotoxicity Targeting GABA			
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1402	Fig. 1-5 Created with BioRender.com.			
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- link.springer.com
 Internet

 110 words 1 %
- 2 www.frontiersin.org 72 words 1 %
- Jing Zhang, Yu Li. "Propofol-Induced Developmental Neurotoxicity: From Mechanisms to Therapeutic Strategies", ACS Chemical Neuroscience, 2023
- Mingyang Sun, Xiaoling Wang, Zhongyuan Lu, Yitian Yang, Shuang Lv, Mengrong Miao, Wan-Ming Chen, Szu-Yuan Wu, Jiaqiang Zhang. "Comparative effectiveness of SGLT2 inhibitors and GLP-1 receptor agonists in preventing Alzheimer's disease, vascular dementia, and other dementia types among patients with type 2 diabetes", Diabetes & Metabolism, 2025
- Qiu-Xia Xiao, Min-Jian Geng, Qiu-Lin Wang, Chang-Le Fang, Jing-Han Zhang, Qi Liu, Liu-Liu Xiong. "Unraveling the Effects of Prenatal Anesthesia on Neurodevelopment: A Review of Current Evidence and Future Directions", NeuroToxicology, 2024



on Neuroprotection and its Possible Mechanisms: Evidence from and Studies ", European Journal of Neuroscience, 2021

- Thiago Arzua, Congshan Jiang, Yasheng Yan, Xiaowen Bai. "The importance of non-coding RNAs in environmental stress-related developmental brain disorders: A systematic review of evidence associated with exposure to alcohol, anesthetic drugs, nicotine, and viral infections", Neuroscience & Biobehavioral Reviews, 2021 $_{\text{Crossref}}$
- Wen-Liang Yan, Wenliang Yan, Xitong Liang, Yang
 Li et al. "Genome-wide association analyses for
 revealing QTN, QTN-by-environment and QTN-by-QTN
 interactions in soybean phenology", Springer Science and
 Business Media LLC, 2024
 Crossref Posted Content

www2.mdpi.com 17 words - < 1%

Rosalva Mora-Escobedo, Gloria Dávila-Ortiz, Gustavo F. Gutiérrez López, José J. Berrios.

"Health-Promoting Food Ingredients during Processing", Routledge, 2024

Publications

hdl.handle.net 16 words — < 1 %

onlinelibrary.wiley.com $_{\text{Internet}}$ 16 words -<1%

www.jneurosci.org 16 words — < 1%

- Bonnie Robinson, Qiang Gu, Jyotshna Kanungo. "Antidepressant Actions of Ketamine: Potential Role of L-Type Calcium Channels", Chemical Research in Toxicology, 2021 $_{\text{Crossref}}$
- Chao Ji, Qiang Ni, Wurong Chen, Zeyong Yang,
 Daqing Ma. "General anesthetic neurotoxicity in
 the young: Mechanism and prevention", Neuroscience &
 Biobehavioral Reviews, 2019
 Crossref
- Yi Wei, Junhua Hu, Yubing Liang, Yuling Zhong, Dan He, Yi Qin, Li Li, Jing Chen, Qiang Xiao, Yubo Xie. "Dexmedetomidine pretreatment attenuates propofolinduced neurotoxicity in neuronal cultures from the rat hippocampus", Molecular Medicine Reports, 2016 Crossref
- Danfeng Yang, Junwei Su, Yeru Chen, Gang Chen. $_{14 \text{ words}} < 1\%$ "The NF- κ B pathway: Key players in neurocognitive functions and related disorders", European Journal of Pharmacology, 2024
- Fahad Iqbal, Andrew J. Thompson, Saba Riaz, Marcus Pehar, Tiffany Rice, Naweed I. Syed.

 "Anesthetics: from modes of action to unconsciousness and neurotoxicity", Journal of Neurophysiology, 2019

 Crossref
- Yan Jin, Song Cai, Yuepeng Jiang, Kai Zhong,
 Chengping Wen, Yeping Ruan, Lindsey A. Chew,
 Rajesh Khanna, Zhenghao Xu, Jie Yu. "Tetramethylpyrazine
 Reduces Epileptogenesis Progression in Electrical Kindling
 Models by Modulating Hippocampal Excitatory
 Neurotransmission", ACS Chemical Neuroscience, 2019

Internet

- 12 words -<1%"Handbook of Neurotoxicity", Springer Nature, 28 2014 Crossref 12 words — < 1 % "Mitochondrial Biology and Experimental 29 Therapeutics", Springer Science and Business Media LLC, 2018 Crossref 12 words -<1%Biswas, Bharti. "Effect of Combined Early Life 30 Stressors on Brain and Behaviour in Rat Offspring.", University of New South Wales (Australia) ProQuest 11 words -<1%Annalisa Savardi, Ilias Ziogas, Maria Bolla, Laura 31 Cancedda. "Chloride transporters in physiological brain development and neurodevelopmental disorders: The case of the Down syndrome", Elsevier BV, 2020 Crossref 11 words -<1%Kantarakorn Unchiti, Prangmalee 32 Leurcharusmee, Artid Samerchua, Tanyong Pipanmekaporn et al. "The potential role of dexmedetomidine on neuroprotection and its possible mechanisms: Evidence from in vitro and in vivo studies", European Journal of Neuroscience, 2021 Crossref 11 words -<1%academic.oup.com 11 words -<1%www.medrxiv.org
- $_{10 \text{ words}} = < 1\%$ Andreas W. Loepke. "Developmental 35 neurotoxicity of sedatives and anesthetics: A

concern for neonatal and pediatric critical care medicine?* :", Pediatric Critical Care Medicine, 03/2010

Crossref

- Dan Zhao, Minli Zhang, Lingling Yang, Mingquan Zeng. "GPR68 Improves Nerve Damage and Myelination in an Immature Rat Model Induced by Sevoflurane Anesthesia by Activating cAMP/CREB to Mediate BDNF", ACS Chemical Neuroscience, 2022 Crossref
- Diogo Dominguini, Amanda V. Steckert, Monique Michels, Mariana B. Spies et al. "The effects of anaesthetics and sedatives on brain inflammation", Neuroscience & Biobehavioral Reviews, 2021
- Félicie Lorenc, Luc Dupuis, Raphaelle Cassel. "Impairments of inhibitory neurons in amyotrophic lateral sclerosis and frontotemporal dementia", Neurobiology of Disease, 2024 Crossref
- Laura Castañeda-Martinez, Kevin K. Noguchi,
 Chrysanthy Ikonomidou, James A. Zagzebski,
 Timothy J. Hall, Ivan M. Rosado-Mendez. "Optimization of
 Ultrasound Backscatter Spectroscopy to Assess Neurotoxic
 Effects of Anesthesia in the Newborn Non-human Primate
 Brain", Ultrasound in Medicine & Biology, 2020
 Crossref
- Lili Xu, Qi Xu, Shaobing Dai, Cuicui Jiao, Yingying Tang, Jiaqian Xie, Hui Wu, Xinzhong Chen. "IncRNA Xist regulates sevoflurane-induced social and emotional impairment by modulating miR-98-5p/EDEM1 signaling axis in neonatal mice", Molecular Therapy Nucleic Acids, 2021 Crossref

- Shetty, Ashok K. "Studies on Developmental Exposures to Anesthetic Agents and Stem Cell Derived Models", Neural Stem Cells in Health and Disease, 2015.

 Crossref
- Cheng, Yuan, Qiaoying Li, Yidan Zhang, Quan Wen, and Jianjun Zhao. "Effects of Female Sex Hormones on Expression of Ang-(1-7)-Mas-R-nNOS Pathways in Rat Brain", Canadian Journal of Physiology and Pharmacology, 2015.

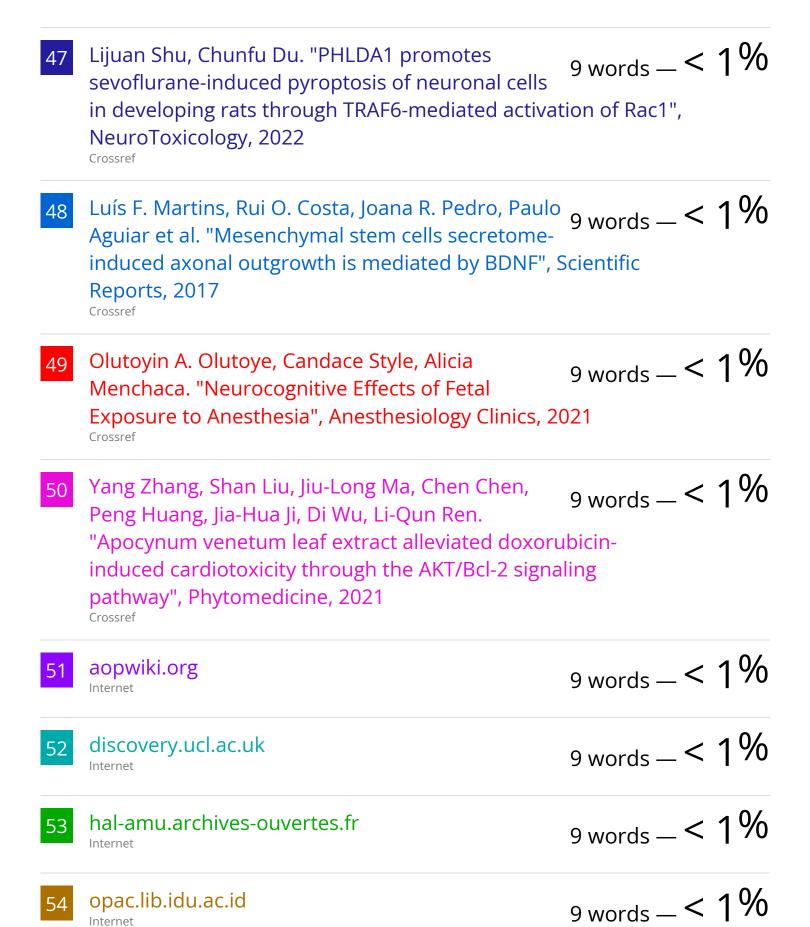
 Crossref
- E.B. Engler-Chiurazzi, C.M. Brown, J.M. Povroznik, J.W. Simpkins. "Estrogens as neuroprotectants: Estrogenic actions in the context of cognitive aging and brain injury", Progress in Neurobiology, 2017

 Crossref
- Genghua Xu, Nuoya Liu, Yaqing Qiu, Jiayu Qi, Danyan Zhu. "Role of Histamine H3 Receptor Antagonist Pitolisant in Early Neural Differentiation of Mouse Embryonic Stem Cells", Stem Cells and Development, 2023 Crossref
- Guo, Peipei, Zhibin Huang, Tao Tao, Xiaohui Chen, Wenqing Zhang, Yiyue Zhang, and Chunshui Lin.

 "Zebrafish as a model for studying the developmental neurotoxicity of propofol in zebrafish", Journal of Applied Toxicology, 2015.

 Crossref
- Jing Lv, Yubing Liang, Youbing Tu, Jing Chen, Yubo Xie. "Hypoxic preconditioning reduces propofolinduced neuroapoptosis via regulation of Bcl-2 and Bax and downregulation of activated caspase-3 in the hippocampus of neonatal rats", Neurological Research, 2018

 Crossref



- "Neonatal Anesthesia", Springer Science and Business Media LLC, 2023
- 8 words = < 1%

Crossref

- Abdelhamid, Ramy, Jia Luo, Lawren VandeVrede, Indraneel Kundu, Bradley Michalsen, Vladislav A.

 Litosh, Isaac T. Schiefer, Teshome Gherezghiher, Ping Yao, Zhihui Qin, and Gregory R. J. Thatcher. "Benzothiophene Selective Estrogen Receptor Modulators Provide Neuroprotection by a Novel GPR30-Dependent Mechanism", ACS Chemical Neuroscience, 2011.
- Anita C. Truttmann, Vanessa Ginet, Julien Puyal.
 "Current Evidence on Cell Death in Preterm Brain
 Injury in Human and Preclinical Models", Frontiers in Cell and
 Developmental Biology, 2020

 Crossref
- Chun-Yun Zhang, Fang-Fang He, Hua Su, Chun Zhang, Xian-Fang Meng. "Association between chronic kidney disease and Alzheimer's disease: an update", Metabolic Brain Disease, 2020 $_{\text{Crossref}}$
- Doboszewska, Urszula, Bernadeta Szewczyk,
 Magdalena Sowa-Kućma, Karolina NoworytaSokołowska, Paulina Misztak, Joanna Gołębiowska, Katarzyna
 Młyniec, Beata Ostachowicz, Mirosław Krośniak, Agnieszka
 Wojtanowska-Krośniak, Krystyna Gołembiowska, Marek
 Lankosz, Wojciech Piekoszewski, and Gabriel Nowak.
 "Alterations of Bio-elements, Oxidative, and Inflammatory
 Status in the Zinc Deficiency Model in Rats", Neurotoxicity
 Research, 2015.

Crossref

- Fei-Fei Ma, Run-Hui Ma, Wang-Wei Zhang, Xiu-Xiu Zhang, Qian Zhong, Kiran Thakur, Jian-Guo Zhang, Jesus Simal-Gandara, Hui Cao. "Neferine induces apoptosis and inhibits metastasis in human endometrial cancer cells by down-regulating miR-1266–5p", Food Bioscience, 2025 Crossref
- Groark, Christina J.; McCarthy, Stephanie K.; Kirk, Afton R. . "Early Childhood Development: From Theory to Practice, Second Edition", UAGC, 2023

 Publications

 8 words < 1%
- Hisham F. Bahmad, Batoul Darwish, Karem Bou Dargham, Rabih Machmouchi et al. "Role of MicroRNAs in Anesthesia-Induced Neurotoxicity in Animal Models and Neuronal Cultures: a Systematic Review", Neurotoxicity Research, 2019
- Hocker, Austin D., Jennifer A. Stokes, Frank L. Powell, and Adrianne G. Huxtable. "The impact of inflammation on respiratory plasticity", Experimental Neurology, 2016.

 Crossref
- Jennifer L. Walters, Merle G. Paule. "Review of preclinical studies on pediatric general anesthesia-induced developmental neurotoxicity", Neurotoxicology and Teratology, 2017
- Kokane, Saurabh S., and Qing Lin. "Impact of Early Life Ketamine Exposure on the Developing Brain and Cognitive Sequelae", Neuropathology of Drug Addictions and Substance Misuse, 2016.

 Crossref

- Martin Mortensen, Yue Xu, Mohamed A. Shehata, Jacob Krall, Margot Ernst, Bente Frølund, Trevor G. Smart. "Pregnenolone sulfate analogues differentially modulate GABA receptor closed/desensitised states ", British Journal of Pharmacology, 2023 Crossref
- Mary Ellen McCann, Sulpicio G. Soriano. "General Anesthetics in Pediatric Anesthesia: Influences on the Developing Brain", Current Drug Targets, 2012

 Crossref
- N. Lunardi. "General Anesthesia Causes Long-Lasting Disturbances in the Ultrastructural Properties of Developing Synapses in Young Rats", Neurotoxicity Research, 07/21/2009
- Ngoc Kim Cuong Tran, Ji Hoon Jeong, Naveen Sharma, Yen Nhi Doan Nguyen et al. "Ginsenoside Re blocks Bay k-8644-induced neurotoxicity via attenuating mitochondrial dysfunction and PKC δ activation in the hippocampus of mice: Involvement of antioxidant potential", Food and Chemical Toxicology, 2023
- Oliver Henschel, Keith Gipson, Angelique Bordey. $_{8 \text{ words}} < 1\%$ "GABAA Receptors, Anesthetics and Anticonvulsants in Brain Development", CNS & Neurological Disorders Drug Targets, 2008
- Omar Hoseá Cabrera, Nemanja Useinovic, Stefan Maksimovic, Michelle Near et al. "Neonatal ketamine exposure impairs infrapyramidal bundle pruning and causes lasting increase in excitatory synaptic transmission in hippocampal CA3 neurons", Neurobiology of Disease, 2022

- Shan Lei, Pan Lu, Yang Lu, Juan Zheng et al.
 "Dexmedetomidine Alleviates Neurogenesis

 Damage Following Neonatal Midazolam Exposure in Rats
 through JNK and P38 MAPK Pathways", ACS Chemical
 Neuroscience, 2020
 Crossref
- Vinayak Mishra, Brian Sims, Margaret Kuper-Sassé, 8 words < 1 % Akhil Maheshwari. "Neurodevelopmental Impairment in Specific Neonatal Disorders", Elsevier BV, 2024 Crossref
- Yafang Liu, Daowei Lin, Chuiliang Liu, Yifan Zhao, Zhiwen Shen, Kun Zhang, Minghui Cao, Yujuan Li.

 "Cyclin-dependent kinase 5/Collapsin response mediator protein 2 pathway may mediate sevoflurane-induced dendritic development abnormalities in rat cortical neurons",

 Neuroscience Letters, 2017

 Crossref
- Youbing Tu, Yubing Liang, Yong Xiao, Jing Lv,
 Ruicong Guan, Fei Xiao, Yubo Xie, Qiang Xiao. "

 Poexmedetomidine attenuates the neurotoxicity of propofol toward primary hippocampal neurons in vitro via Erk1/2/CREB/BDNF signaling pathways
 Poevelopment and Therapy, 2019
 Crossref
- Yu Hou, Shuai Li, Qi Hou, Runjia Wang, Xiyuan Xu, Zhichao Li, Linhui Ma, Qidong Liu, Yuan Shen, Hui Zheng. "Vitamin K2 mitigates cognitive, and motor impairments induced by multiple surgery with anesthesia and analgesia in neonatal stage", Biochemical and Biophysical Research Communications, 2025



- Zachary Fleissner, Antonia Schonwald, Neil Patel, Benjamin F. Gruenbaum, Valerie A. Smart, Shaun E. Gruenbaum. "Neurotoxicity and translational research", Elsevier BV, 2022
- Zhi-Li Yao, Qing-Nan Li, Yong-Ji Li, Hao-Rui Li et al. "CRISPR/Cas12a-based fluorescence-colorimetric dual-mode strategy for sensitive detection of clear cell renal cell carcinoma biomarker miR-210–3p", Sensors and Actuators B: Chemical, 2025 Crossref

d.docksci.com 8 words - < 1%journals.lww.com 8 words - < 1%openrepository.aut.ac.nz 8 words - < 1%

- qspace.library.queensu.ca 8 words < 1 %
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 8 words < 1 %
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- wiredspace.wits.ac.za 8 words < 1%www.biorxiv.org 8 words < 1%www.grafiati.com 8 words < 1%
- A. Leslie Morrow, Minna H. McFarland, Todd K. O'Buckley, Donita L. Robinson. "Emerging evidence 7 words <1% for pregnane steroid therapeutics for alcohol use disorders", Elsevier BV, 2024

 Crossref
- Baptiste Riffault, Igor Medina, Camille Dumon, Carine Thalman et al. "Pro-Brain-Derived Neurotrophic Factor Inhibits GABAergic Neurotransmission by Activating Endocytosis and Repression of GABA Receptors ", The Journal of Neuroscience, 2014 Crossref
- Guiping Xu, Yang Wang, Zhe Chen, Yuxuan Zhang, 7 words < 1% Xuexue Zhang, Guichao Zhang. "Esketamine improves propofol-induced brain injury and cognitive impairment in rats", Translational Neuroscience, 2022
- Hewitt, Danielle. "Neural Changes Associated with $_{7 \text{ words}} < 1\%$ Spinal Cord and External Peripheral Nerve Stimulation for the Treatment of Neuropathic Lower Limb Pain", The University of Liverpool (United Kingdom), 2023 ProQuest
- lan Coulter, Tamara Timic Stamenic, Pierce Eggan, Brier R. Fine et al. "Different roles of T-type calcium 7 words <1%

channel isoforms in hypnosis induced by an endogenous neurosteroid epipregnanolone", Neuropharmacology, 2021

- Kok, Morgan. "A Molecular and Computational Approach to Analyze the Biogenesis of the Potassium-Chloride Cotransporter 2 (KCC2), A Neuron-Specific Protein Implicated in Disease", University of Pittsburgh ProQuest
- Nan Wang, Mingyu Wang. "Dexmedetomidine suppresses sevoflurane anesthesia-induced neuroinflammation through activation of the PI3K/Akt/mTOR pathway", BMC Anesthesiology, 2019

 Crossref
- Refik Servi, Ramazan Fazıl Akkoç, Feyza Aksu,
 Süleyman Servi. "Therapeutic potential of
 enzymes, neurosteroids, and synthetic steroids in
 neurodegenerative disorders: A critical review", The Journal of
 Steroid Biochemistry and Molecular Biology, 2025
 Crossref
- Stephanie L. Merhar, Carla M. Bann, Nicole Mack, Jamie E. Newman et al. "Prenatal Opioid Exposure is Associated with Punctate White Matter Lesions in Term Newborns", The Journal of Pediatrics, 2025

 Crossref
- "Fundamentals of Pediatric Neuroanesthesia", Springer Science and Business Media LLC, 2021 6 words -<1%
- "Total Intravenous Anesthesia and Target Controlled Infusions", Springer Science and Business Media LLC, 2017

 Crossref 6 words -<1%

- A. Leslie Morrow, Giorgia Boero, Irina Balan. "Emerging Evidence for Endogenous Neurosteroid Modulation of Pro-Inflammatory and Anti-Inflammatory Pathways that Impact Neuropsychiatric Disease", Neuroscience & Biobehavioral Reviews, 2024 Crossref
- Hostetler, Rachel Elaine. "The Diversity and Synaptic Connectivity of Somatostatin-containing Inhibitory Interneurons in the Somatosensory (Barrel) Cortex", West Virginia University, 2024

 ProQuest
- Jiaxi Qian, Maorong Jiang, Zihan Ding, Dandan Gu, Huiyuan Bai, Min Cai, Dengbing Yao. "Role of Long Non-coding RNA in Nerve Regeneration", International Journal of Neuroscience, 2023

 Crossref
- Ryan A. Stevens, Brandon D. Butler, Saurabh S. Kokane, Andrew W. Womack, Qing Lin. "Neonatal inhibition of Na + -K + -2Cl -cotransporter prevents ketamine induced spatial learning and memory impairments", Neurotoxicology and Teratology, 2017 Crossref
- Wanping Gu, Tingting Pan, Xuedong Wang, Liheng Kang, Nan Liu, Meihua Piao, Chunsheng Feng.

 "Sevoflurane exposure triggers ferroptosis of neuronal cells initiated by the activation of ATM/p53 in the neonatal brain via JNK/p38 MAPK-mediated oxidative DNA damage", Springer Science and Business Media LLC, 2024

 Crossref Posted Content
- X. Zhang, F. Liu, W. Slikker, C. Wang, M.G. Paule. "Minimally invasive biomarkers of general

anesthetic-induced developmental neurotoxicity", Neurotoxicology and Teratology, 2017 Crossref

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