

Review

Open Access

Biochemical prevention and treatment of viral infections – A new paradigm in medicine for infectious diseases

Hervé Le Calvez*¹, Mang Yu² and Fang Fang²

Address: ¹Abgent, Inc. 6310 Nancy Ridge Drive, Suite 106, San Diego, CA 92121 USA and ²NexBio, Inc. 6330 Nancy Ridge Drive, Suite 105, San Diego, CA 92121 USA

Email: Hervé Le Calvez* - lecalvez@abgent.com; Mang Yu - myu@nexbio.com; Fang Fang - ffang@nexbio.com

* Corresponding author

Published: 23 November 2004

Received: 10 November 2004

Virology Journal 2004, 1:12 doi:10.1186/1743-422X-1-12

Accepted: 23 November 2004

This article is available from: <http://www.virologyj.com/content/1/1/12>

© 2004 Le Calvez et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

For two centuries, vaccination has been the dominating approach to develop prophylaxis against viral infections through immunological prevention. However, vaccines are not always possible to make, are ineffective for many viral infections, and also carry certain risk for a small, yet significant portion of the population. In the recent years, FDA's approval and subsequent market acceptance of Synagis, a monoclonal antibody indicated for prevention and treatment of respiratory syncytial virus (RSV) has heralded a new era for viral infection prevention and treatment. This emerging paradigm, herein designated "Biochemical Prevention and Treatment", currently involves two aspects: (1) preventing viral entry via passive transfer of specific protein-based anti-viral molecules or host cell receptor blockers; (2) inhibiting viral amplification by targeting the viral mRNA with anti-sense DNA, ribozyme, or RNA interference (RNAi). This article summarizes the current status of this field.

Introduction

A landmark in the battle against viral infectious diseases was made in 1798 when Jenner first inoculated humans against smallpox with the less virulent cowpox. For about two centuries since then, humans relied almost exclusively on vaccines for protection against viruses. Only in the recent years, new strategies for controlling viral infectious diseases have emerged, which have so far led to a couple of viral prophylaxis/therapeutics on the market. These strategies are fundamentally different from vaccines in that they attempt to directly interrupt viral infectious life cycle at molecular level by using proteins or oligonucleotides. To differentiate them from the conventional vaccines that prevent viral infection by boosting immune

system, we refer the new antiviral approaches as "Biochemical Prevention and Treatment" (see figure 1). Biochemical Prevention and Treatment, as an alternative to vaccines and chemical compound based antiviral drugs, may prove to be particularly valuable in the areas where vaccines and/or chemical drugs can not be generated or have not been successful in human, including diseases caused by some common pathogenic viruses, such as HIV, hepatitis C virus (HCV), RSV and human rhinovirus (HRV). In this review, we will discuss various molecular intervention approaches.

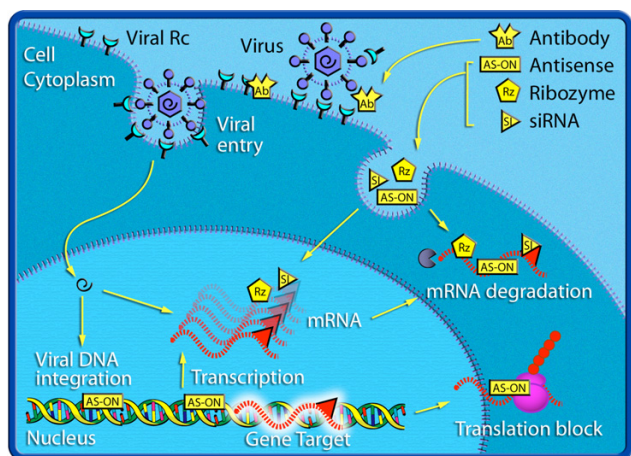


Figure 1
Targets of different Biochemical Prevention and Treatment strategies. Antibodies (Ab) or soluble receptors (Rc) can inhibit the viral entry. Antisense oligonucleotides (AS-ONs), ribozymes (Rz) or siRNA (SI) pair with their complementary target genomic DNA, RNA or mRNA. AS-ONs can block recombination, transcription of the mRNA or induce its degradation by RNaseH. Rz possess catalytic activity and cleave their targets. SiRNAs (SI) induce degradation of the target mRNA via RNA-induced silencing complex (RISC).

I. Biochemical Prevention and Treatment via Protein targeting

Among the biochemical therapeutics currently in clinical trials, the majority consists of monoclonal antibodies (MAbs). Soluble receptor drug candidates have gradually lost favor over the past several years due to issues relating to low potency and cost. Peptide-based drug candidates are limited by insufficient efficacy and unfavorable pharmacokinetics. MAbs have increasingly gained favor in large part because of the development of chimeric, humanized, and human antibodies have reduced the immunogenicity of antibody therapies. The MAbs that are currently in clinical trials for viral infection prophylaxis and treatment are listed in Table 1.

Biochemical Prevention and Treatment of Respiratory Syncytial Virus Infection

The respiratory syncytial virus (RSV) is a major cause of lower respiratory tract infection in infants and young children producing bronchiolitis and pneumonia worldwide. RSV infection leads to more than 90,000 hospitalizations and a 2% mortality rate among infants nationwide [2-5]. Approximately two-thirds of infants are infected with RSV during the first year of life and approximately 95% of children test seropositive for RSV by the age of two [6]. Unfortunately, even natural RSV infection produces limited immunity at best. In fact, an inactivated RSV vaccine paradoxically resulted in more severe disease instead of protection [7].

The most successful approach to date has been Biochemical Prevention and Treatment with anti-viral antibodies. In 1996, RespiGam™ (respiratory syncytial virus immune

Table 1: Monoclonal Antibodies in Clinical Trials

Product	Company	Disease	Status
MEDI-501	MedImmune	Genital Warts HPV	II
Nabi-HB	Nabi Biopharmaceuticals	Hepatitis B	Market
Ostavir	Protein Design Labs	Hepatitis B	II
XTL-002	XTL Biopharmaceuticals Ltd.	Hepatitis C	I
Civacir	Nabi Biopharmaceuticals	Hepatitis C	I/II
IF7 Antibody	Immune Network Ltd.	Hepatitis C, HIV/AIDS	Preclinical
PRO 140	Progenics Pharmaceuticals	HIV/AIDS	Preclinical
hNM01	AbNovo Inc., Immune Network Ltd.	HIV/AIDS	I
PRO 367	Roche Holding Progenics Pharmaceuticals	HIV/AIDS	I/II
TNX-355	Tanox, Inc., Biogen, Inc. (Massachusetts)	HIV/AIDS	I
OraQuick HIV-1	OraSure Technologies, Inc.	HIV/AIDS	Market
Cytolin	CytoDyn Amerimmune Pharmaceuticals, Inc.	HIV/AIDS	I/II
Tipranavir	TIPRANAVIR	HIV/AIDS	III
HXB	AAI International, AnaaiPharma Company	Herpes Simplex Virus type 2	Preclinical
MEDI-491	MedImmune	Human B19 parvovirus	I
Synagis™ (Palivizumab)	MedImmune	Respiratory Syncytial Virus	Approved in 1998
Numax	MedImmune	Respiratory Syncytial Virus	Preclinical
INS37217 Intranasal	Inspire Pharmaceuticals	Rhinovirus (common cold)	II

globulin or RSV-IG) became available for use in children less than two years of age with high-risk factors [8-10]. The use of RespiGam™ was largely supplanted with the approval of Synagis™ (Palivizumab) in 1998. Palivizumab is an IgG1 MAb administered IM monthly that selectively binds to the RSV surface glycoprotein F [1,51]. The drug specifically inhibits RSV replication by preventing the virus from fusing with the respiratory endothelial cell membrane. Palivizumab has been shown to reduce the rate of hospitalization of at-risk infants by about 55% in clinical studies and now serves as the primary medical means of RSV prevention [11-13].

Prevention of Human Rhinovirus infections

Human rhinovirus (HRV) causes over 80% of the common cold in the fall [14]. Developing vaccines against HRV is unfeasible because HRVs have at least 115 antigenically distinct serotypes [15,16]. One of the proven methods to prevent and inhibit viral infections is to block host cell receptors that are used by viruses to gain cell entry. Receptor blockage is commonly achieved via application of MABs that bind to specific epitopes on the receptor molecules. A plethora of *in vitro* studies have reported effective viral inhibition by receptor-blocking MABs. However, these works have not yielded yet any approved drug on the market.

In HRV infection, about 90% of HRV serotypes utilize a single cell surface receptor exclusively, which is the intercellular adhesion molecule-1 (ICAM-1), for viral attachment and subsequent viral entry [17,18]. As such, ICAM-1 has become a very promising target for biochemical prevention. A receptor blocking approach has shown that the soluble ICAM-1 and an anti-ICAM-1 monoclonal antibody, Mab 1A6, could prevent infections by a broad spectrum of rhinovirus serotypes in human cells *in vitro* [19-21]. Administration of soluble ICAM-1 and MABs in human clinical trials had indeed achieved reduction in symptoms, but did not prevent the incidence of the disease [22-24]. For the MABs, the limited efficacy is most likely due to its low functional affinity (or avidity) for ICAM-1 when compared to that of the multivalent HRV particles [25].

High avidity is achieved by multivalency. To improve avidity of HRV receptor blocking antibody, a novel tetravalent recombinant antibody, CFY196, has been generated against ICAM-1 [26]. CFY196 is composed of Fab fragment of a humanized version of Mab 1A6 fused with a linker derived from human immunoglobulin D (IgD) hinge and a tetramerization domain derived from the coiled-coil sequence of human transcription factor ATF α . CFY196 is expressed in bacteria and purified as a homogeneous tetrameric molecular complex. CFY196 exhibited almost two-orders-of-magnitude improvement in

functional affinity compared with its bivalent counterpart based on the kinetic parameters measured by BIAcore analysis. Such kinetic improvement also directly leads to functional superiorities of CFY196. In *in vitro* assays, CFY196 consistently and significantly outpaced the best commercial anti-ICAM-1 MABs in preventing HRV infection as measured by reduction of cytopathic effects and HRV viral titers [26]. The preclinical findings of CFY196 bode well its efficacy in human since Mab 1A6, from which CFY196 is derived, has already exhibited positive effects in a human trial. Moreover, to prevent possible immunogenicity, CFY196 is humanized [27]. Further preclinical and clinical development of CFY196 is warranted to fully evaluate its potential as a prophylaxis and therapeutics for the HRV induced common colds.

2. Biochemical Prevention and Treatment via targeting on viral mRNA

Targeting viral mRNA is one of the most active areas of research and development. Several strategies have emerged over the years and are being tested pre-clinically and clinically. They include: antisense-oligonucleotides (AS-ONs), ribozymes, and recently, RNA interference (RNAi). All these strategies share the features of conceptual simplicity, straightforward drug design and quick route to identify drug leads. However, the challenges have been to improve potency, pharmacokinetics and, most importantly, intracellular delivery of the drug candidates. As the oldest strategy, AS-ON technology has produced to date one drug in the market place, Vitravene®. A number of clinical trials of drug candidates from these technologies are currently ongoing.

Antisense-oligonucleotides

Antisense-oligonucleotides (AS-ONs) are short synthetic oligonucleotides that form complementary pair with specific viral mRNA targets. AS-ONs inhibit viral protein production by both blocking viral mRNA translation and triggering its degradation. Since the discovery of viral inhibition effect of AS-ONs by Zamecnik and Stephenson in 1978 [28], antisense technology has been developed as a powerful tool for target validation and therapeutic purposes.

Vitravene is the first AS-ON based drug approved by FDA. Vitravene, or fomivirsin sodium, is a 21-base phosphorothioate oligodeoxynucleotide complementary to the messenger RNA of the major immediate-early region proteins of human cytomegalovirus, and is a potent and selective antiviral agent for cytomegalovirus retinitis, a herpes-like eye disease that afflicts the immune-suppressed [29,30]. A number of clinical trials as well as one approved therapy based on AS-ON technologies are summarized in Table 2.

Table 2: Clinical trials and an approved therapy based on AS-ON technologies [31-33].

Product	Company	Target	Disease	Chemistry	Status
Vitravene (Fomivirsen)	ISIS Pharmaceuticals	CMV IE2	CMV retinitis	PS DNA	Approved in 1998
Affinitac (ISIS 3521)	ISIS	PKC- α	Cancer	PS DNA	Phase III
Genasense	Genta	Bcl2	Cancer	PS DNA	Phase III
Alicaforsen (ISIS 2302)	ISIS	ICAM-1	Psoriasis, Crohn's disease, Ulcerative colitis	PS DNA	Phase II/III
ISIS 14803	ISIS	Antiviral	Hepatitis C	PS DNA	Phase II
ISIS 2503	ISIS	H-ras	Cancer	PS DNA	Phase II
MG98	Methylgene	DNA methyl transferase	Solid tumors	PS DNA	Phase II
EPI-2010	EpiGenesis Pharmaceuticals	Adenosine A1 receptor	Asthma	PS DNA	Phase II
GTI 2040	Lorus Therapeutics	Ribonucleotide reductase (R2)	Cancer	PS DNA	Phase II
ISIS 104838	ISIS	TNF α	Rheumatoid Arthritis, Psoriasis	2nd generation	Phase II
Avi4126	AVI BioPharma	c-myc	Restenosis, cancer, Polycystic kidney disease	3rd generation	Phase I/II
Gem231	Hybridon	PKA R1 α	Solid tumors	2nd generation	Phase I/II
Gem92	Hybridon	HIV gag	AIDS	2nd generation	Phase I
GTI 2051	Lorus Therapeutics	Ribonucleotide reductase (R1)	Cancer	PS DNA	Phase I
Avi4557	AVI BioPharma	CYP3A4	Metabolic redirection of approved drugs	3rd generation	Phase I

Phosphorothioate (PS) oligodeoxynucleotides are the 'first generation' DNA analogs. The 'second generation' ONs contain nucleotides with alkyl modifications at the 2' position of the ribose. They are less toxic than PS-DNAs and have a slightly enhanced affinity. DNA and RNA analogs with modified phosphate linkages, or different sugar residues substituting the furanose ring have been referred as 'third generation' [34]. For instance, peptide nucleic acids and their analogs display superior sequence specificity and are resistant to nuclease degradation. These third generation AS-ON have limited non-specific interactions with other genes and, therefore, have shown great potentials in clinical trials.

Ribozymes

Ribozymes (Rz) are catalytically active ONs that both bind and cleave target RNAs. They were discovered after the AS-ON technology. Initial findings on ribozymes raised the hope that they may offer a more potent alternative to AS-ONs. Many cell based and animal tests have performed on anti-viral effects of ribozymes, including HIV, hepatitis B, hepatitis C, influenza, etc. Results from these tests have shown that ribozymes are promising viral inhibitors [35-38]. However, further progress in the field has been hampered by difficulties to achieve satisfactory potency and efficient intracellular delivery of ribozymes in vivo. HEPTAZYME is a modified ribozyme that cleaves the internal ribosome entry site of the Hepatitis C virus.

The Rz was demonstrated to inhibit viral replication up to 90% in cell culture [39]. HEPTAZYME was tested in a Phase II clinical trial, but was later withdrawn from further clinical trials due to insufficient efficacy. So far, there is no anti-viral ribozymes that are being actively tested in advanced clinical trials.

RNA Interference (RNAi)

RNA interference, or RNAi, is the inhibition of expression of specific genes by double-stranded RNAs (dsRNAs). It is becoming the method of choice to knockdown gene expression rapidly and robustly in mammalian cells. Comparing to the traditional antisense method, RNAi technology has the advantage of significantly enhanced potency; therefore, only lower concentrations may be needed to achieve same level of gene knockdown. RNAi gained rapid acceptance by researchers after Tuschl and coworkers discovered that *in vitro* synthesized small interfering RNAs (siRNAs) of 21 to 23 nucleotides in length can effectively silence targeted genes in mammalian cells without triggering interferon production [40,41]. In mammalian cells, the level of gene inhibition mediated by siRNA routinely reaches an impressive 90% [42].

Several initial studies, which test the potential application of synthetic siRNAs as antiviral agents, have shown very promising results. To date, RNAi has been used effectively

to inhibit the replication of several different pathogenic viruses in culture, including: RSV (respiratory syncytial virus) [43], influenza virus [44], poliovirus [45] and HIV-1 [46-48]. In the case of HIV-1, several specific mRNAs have been successfully targeted for siRNA-mediated silencing, including those that encode Gag, Pol, Vif and the small regulatory proteins Tat and Rev. These studies show that RNAi can effectively trigger the degradation of not only viral mRNAs, but also genomic RNAs at both the pre- and post-integration stages of the viral lifecycle. In addition to targeting viruses directly, alternative strategies have employed siRNAs that silence the expression of essential host factors including Tsg101, required for vacuolar sorting and efficient budding of HIV-1 progeny [49], and the chemokine receptor CCR5, required as a co-receptor for HIV-1 cell entry [50].

Conclusions

Currently, our understanding of the biological mechanisms underlying RNAi lags behind the movement to apply this technology to human diseases such as viral infections. Some major technical hurdles need to be overcome before siRNA-based anti-viral prophylaxis and treatments move into the clinics. Especially, intracellular delivery of siRNA needs to be greatly improved. The next few years of research will indicate whether RNAi technology will realize its potential as the next wave of Biochemical Prevention and Treatment.

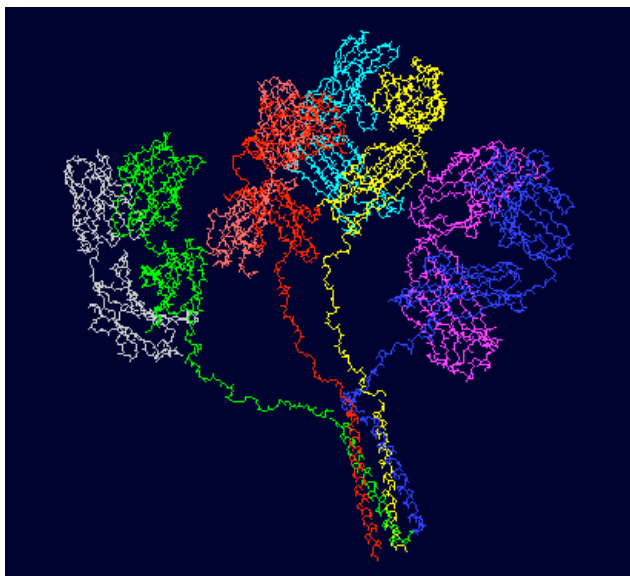


Figure 2
3D model of the tetrameric Fab anti-ICAM-1 molecule CFY196 [26]. Each identical subunit is represented by a different color.

Competing Interests

Dr. Hervé Le Calvez declares that he has no competing interest. Dr. Mang Yu and Dr. Fang Fang are the co-founders and current share holders of Perlan Therapeutics who has developed CFY196.

Acknowledgements

The authors wish to thank Kosi Gramatikoff for graphic assistance and helpful discussions. They are grateful to Libby Weber for the critical assistance on the completion of this manuscript.

References

- Anderson LJ, Bingham P, Hierholzer J: **Neutralization of respiratory syncytial virus by individual and mixtures of F and G protein monoclonal antibodies.** *J Virol* 1988, **62**:4232-4238.
- Chanock RM, Kim HW, Vargosko AJ, Deleva A, Johnson KM, Cumming C, Parrott RH: **Respiratory syncytial virus: I. Virus recovery and other observations during 1960 outbreak of bronchiolitis, pneumonia, and minor respiratory diseases in children.** *JAMA* 1961, **176**:647-653.
- Parrott RH, Vargosko AJ, Kim HW, Cumming C, Turner H, Huebner RJ, Chanock RM: **Respiratory syncytial virus. II. Serologic studies over a 34-month period of children with bronchiolitis, pneumonia, and minor respiratory diseases.** *JAMA* 1961, **176**:653-657.
- Prober CG, Wang EE: **Reducing the morbidity of lower respiratory tract infections caused by respiratory syncytial virus: still no answer.** *Pediatrics* 1997, **99**:454-61.
- Simoes EAF, Rieger CHL: **RSV infection in developed and developing countries.** *Infect Med* 1999, **16**:11-17.
- Parrott RH, Kim HW, Arrobbio JO, Hodes DS, Murphy BR, Brandt CD, Camargo E, Chanock RM: **Epidemiology of respiratory syncytial virus infection in Washington D.C. II. Infection and disease with respect to age, immunological status, race and sex.** *J Epidemiol* 1973, **98**:289-300.
- Levin MJ: **Treatment and prevention options for respiratory syncytial virus infections.** *J Pediatrics* 1994, **125**:S22-S27.
- Groothuis JR, Levin NJ, Rodriguez W, Hall CB, Long CE, Kim HW, Lauer BA, Hemming VG: **Use of intravenous gamma globulin to passively immunize high-risk children against RSV: safety and pharmacokinetics.** *Antimicrob Agents Chemother* 1991, **35**(7):1469-1473.
- Meissner HC, Fulton DR, Groothuis JR, Geggel RL, Marx GR, Hemming VG, Hougren T, Snyderman DR: **Controlled trial to evaluate protection of high-risk infants against RSV disease by using standard intravenous immune globulin.** *Antimicrob Agents Chemother* 1993, **37**:1655-1658.
- MedImmune Inc: **RespiGam™: Respiratory Syncytial Virus Immune Globulin Intravenous (Human), [RSV-IGIV].** Gaithersburg, MD 1996.
- The Impact-RSV Study Group: **Palivizumab, a humanized respiratory syncytial virus monoclonal antibody, reduces hospitalization from respiratory syncytial virus infection in high-risk infants.** *Pediatrics* 1998, **102**:531-537.
- Cohen AH, Sorrentino M, Powers T: **Effectiveness of palivizumab for preventing serious RSV disease.** *J Resp Dis* 2000, **2**:S30-S32.
- MedImmune Inc: **Synagis™: Palivizumab for intramuscular administration.** Gaithersburg, MD 1996.
- Arruda E, Pitkaranta A, Witek TJ Jr, Doyle CA, Hayden FG: **Frequency and natural history of rhinovirus infections in adults during autumn.** *J Clin Microb* 1997, **35**:2864-2868.
- Stanway G: **Rhinoviruses.** In: Webster RG ed. *In Encyclopedia of Virology* New York: Academic Press; 1994:1253-1259.
- Skern T, Duechler M, Sommergruber W, Blaas D, Kuechler E: **The molecular biology of human rhinoviruses.** *Biochem Soc Symp* 1987, **53**:63-73.
- Staunton DE, Merluzzi VJ, Rothlein R, Barton R, Marlin SD, Springer TA: **A cell adhesion molecule, ICAM-1, is the major surface receptor for rhinoviruses.** *Cel* 1989, **56**:849-853.
- Uncapher CR, Dewitt CM, Colonno RJ: **The major and minor group receptor families contain all but one human rhinovirus serotype.** *Virology* 1991, **180**:814-817.

19. Marlin SD, Ltaunton DE, Springer TA: **A soluble form of intercellular adhesion molecule-1 inhibits rhinovirus infection.** *Nature* 1990, **344**:70-72.
20. Huguenel ED, Cohn D, Dockum DP, Greve JM, Fournel MA, Hammond L, Irwin R, Mahoney J, McClelland A, Muchmore E, Ohlin AC, Scuderi P: **Prevention of rhinovirus infection in chimpanzees by soluble intercellular adhesion molecule-1.** *Am J Resp Critical Care Med* 1997, **155**:1206-1210.
21. Colonna RJ, Callahan PL, Long WJ: **Isolation of a monoclonal antibody that blocks attachment of the major group of human rhinoviruses.** *J Virol* 1986, **57**:7-12.
22. Turner RB, Wecker MT, Pohl G, Witek TJ, McNally E, St George R, Winther B, Hayden FG: **Efficacy of tremacamra, a soluble intercellular adhesion molecule 1, for experimental rhinovirus infection: a randomized clinical trial.** *JAMA* 1999, **281**:1797-1804.
23. Colonna RJ: **Virus receptors: the Achilles' heel of human rhinoviruses.** *Adv Exp Med Biol* 1992, **312**:61-70.
24. Hayden FG, Gwaltney JM, Colonna RJ: **Modification of experimental rhinovirus colds by receptor blockade.** *Antiviral Res* 1988, **9**:233-247.
25. Casanovas JM, Springer TA: **Kinetics and thermodynamics of virus binding to receptor. Studies with rhinovirus, intercellular adhesion molecule-1 (ICAM-1), and surface plasmon resonance.** *J Biol Chem* 1995, **270**:13216-13224.
26. Charles CH, Luo GX, Kohlstaedt LA, Gorfain E, Morantte I, Williams JH, Fang F: **Prevention of Human Rhinovirus Infection by Multivalent Fab Molecules Directed against ICAM-1.** *Antimicrobial Agents and Chemotherapy* 2003, **47**:1503-1508.
27. Luo GX, Kohlstaedt LA, Charles CH, Gorfain E, Morantte I, Williams JH, Fang F: **Humanization of an anti-ICAM-1 antibody with over 50-fold affinity and functional improvement.** *J Immunol Methods* 2003, **275**:31-40.
28. Zamecnik PC, Stephenson ML: **Inhibition of Rous sarcoma virus replication and cell transformation by a specific oligodeoxynucleotide.** *Proc Natl Acad Sci USA* 1978, **75**:280-284.
29. Orr RM: **Technology evaluation: fomivirsen. Isis Pharmaceuticals Inc/CIBA vision.** *Curr Opin Mol Ther* 2001, **3**:288-294.
30. Roehr B: **Fomivirsen approved for CMV retinitis.** *J Int Assoc Physicians AIDS Care* 1998, **4**:14-16.
31. Dove A: **Antisense and sensibility.** *Nat Biotechnol* 2002, **20**:121-124.
32. Braasch DA, Corey DR: **Novel antisense and peptide nucleic acid strategies for controlling gene expression.** *Biochemistry* 2002, **41**:4503-4509.
33. Opalinska JB, Gewirtz AM: **Nucleic acids therapeutics: Basic principles and recent applications.** *Nat Rev Drug Discov* 2002, **1**:503-514.
34. Kurreck J: **Antisense technologies: Improvement through novel chemical modifications.** *Eur J Biochem* 2003, **270**:1628-1644.
35. Yu M, Ojwang J, Yamada O, Hampel A, Rappaport J, Looney D, Wong-Staal F: **A Hairpin Ribozyme Inhibits Expression of Diverse Strains of HIV-1.** *Proc Natl Acad Sci USA* 1993, **90**:6341.
36. Welch P, Tritz R, Yei S, Barber JR, Yu M: **Intracellular Application of Hairpin Ribozyme Genes Against Hepatitis B Virus.** *Gene Therapy* 1997, **4**:736.
37. Welch PJ, Tritz R, Yei S, Leavitt M, Yu M, Barber J: **Apotential therapeutic application of hairpin ribozymes: In vitro and in vivo studies of gene therapy for hepatitis C virus infection.** *Gene Ther* 1996, **3**:994.
38. Tang XB, Hobom G, Luo D: **Ribozyme mediated destruction of influenza A virus in vitro and in vivo.** *J Med Virol* 1994, **42**:385.
39. Macejak D, Jensen KL, Jamison S, Domenico K, Roberts EC, Chaudhary N, von Carlowitz I, Bellon L, Tong MJ, Conrad A, Pavco PA, Blatt LM: **Inhibition of Hepatitis C Virus (HCV)-RNA-dependent translation and replication of a chimeric HCV Poliovirus using synthetic stabilized ribozymes.** *Hepatology* 2000, **31**:769-776.
40. McManus MT, Sharp PA: **Gene silencing in mammals by small interfering RNAs.** *Nature Rev* 2002, **3**:737-747.
41. Thompson JD: **Applications of antisense and siRNAs during preclinical drug development.** *Drug Discovery Today* 2002, **7**:912-917.
42. Shi Y: **Mammalian RNAi for the masses.** *Trends in Genetics* 2003, **19**:9-12.
43. Bitko V, Barik S: **Phenotypic silencing of cytoplasmic genes using sequence-specific double-stranded short interfering RNA and its application in the reverse genetics of wild type negative-strand RNA viruses.** *BMC Microbiology* 2001, **1**:34-46.
44. Ge O, McManus MT, Nguyen T, Shen CH, Sharp PA, Eisen HN: **RNA interference of influenza virus production by directly targeting mRNA for degradation and indirectly inhibiting all viral RNA transcription.** *Proc Natl Acad Sci USA* 2003, **100**:2718-2723.
45. Gitlin L, Karelsky S, Andino R: **Short interfering RNA confers intracellular antiviral immunity in human cells.** *Nature* 2002, **418**:430-434.
46. Coburn GA, Cullen BR: **Potent and specific inhibition of human immunodeficiency virus type 1 replication by RNA interference.** *Journal of Virology* 2002, **76**:9225-9231.
47. Jacque JM, Triques K, Stevenson M: **Modulation of HIV-1 replication by RNA interference.** *Nature* 2002, **418**:435-438.
48. Novina CD, Murray MF, Dykxhoorn DM, Beresford PJ, Riess J, Lee SK, Collman RG, Lieberman J, Shankar P, Sharp PA: **siRNA-directed inhibition of HIV-1 infection.** *Nature Medicine* 2002, **8**:681-686.
49. Garrus JE, von Schwedler UK, Pornillos OW, Morham SG, Zavitz KH, Wang HE, Wettstein DA, Stray KM, Cote M, Rich RL, Myszka DG, Sundquist WL: **Tsg101 and the vacuolar protein sorting pathway are essential for HIV-1 budding.** *Cell* 2001, **107**:55-65.
50. Martinez MA, Gutierrez A, Armand-Ugon M, Blanco J, Parera M, Gomez J, Clotet B, Este JA: **Suppression of chemokine receptor expression by RNA interference allows for inhibition of HIV-1 replication.** *AIDS* 2002, **16**:2385-2390.
51. Johnson S, Oliver C, Prince GA, Hemming VG, Pfarr DS, Wang SC, Dormitzer M, O'Grady J, Koenig S, Tamura JK, Woods R, Bansal G, Couchenour D, Tsao E, Hall WC, Young JF: **Development of a humanized monoclonal antibody (MEDI-493) with potent in vitro and in vivo activity against respiratory syncytial virus.** *J Infect Dis* 1997, **176**:1215-1224.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

