# Environmental toxins and the impact of other endocrine disrupting chemicals in women's reproductive health

Mauri José Piazza<sup>1</sup>, Almir Antônio Urbanetz<sup>1</sup>

<sup>1</sup>Tocogynecology Department, Universidade Federal do Paraná - UFPR - Curitiba (PR), Brazil

#### ABSTRACT

This review aimed to look into agents and mechanisms characterized as endocrine disrupting chemicals (EDCs). These agents are known to cause several harmful effects to the reproductive system of women and wildlife. There is a wide range of chemicals, developed for commercial use mainly in agriculture, which may cause endocrine disruption. Numerous studies show evidence of environmental contamination. However, no one is being held liable for the damages. The most important potentially harmful agents are identified and described, along with the different effects they have on the female genital area. Brazil is a large consumer of pesticides and others chemicals that may interfere with a normal women's life. We analyzed and described the mode of action and the impacts of different EDCs (bisphenols, phthalates, atrazine, polychlorinated and polybrominated biphenyls, DDT-dichlorodiphenyltrichloroethane; DDE-dichlorodiphenyldichloroethylene; DDD-dichlorodiphenyldichloroethane; and DES-diethylstilbestrol) on the genital area, ovarian steroidogenesis, polycystic ovary syndrome, endometriosis, the structure of the uterus and the vagina, and on the formation of leiomyomas.

**Keywords:** human reproduction, environmental toxicants, environmental pollution, endocrine disrupting chemicals, reproductive health

#### INTRODUCTION

A growing number of scientific evidence has been collected over the past few years suggesting that human reproductive capacity has been affected by a wide range of recurrent substances present in a wide array of everyday products. Several indicators are showing increased incidence of cardiovascular disorders, obesity, hormone-dependent cancers, and chronic diseases, not to mention early puberty development, pregnancy length disorders, and other reproductive health abnormalities.

Among the acting agents are substances such as bisphenol A (BPA) and its byproducts bisphenol B, tetrabromobisphenol A, and bisphenols F and S. All of these and more have been defined as endocrine disrupting chemicals (EDCs). Endocrine disruptors are chemicals that may interfere and cause adverse effects on the endocrine system at any life-stage on account of their resemblance with endogenous steroid hormones. Birnbaum (2013) showed that the global production of these chemicals increased 23.5 fold between 1947 and 2007. In 2012 alone, the US produced 9.5 trillion pounds - 2.09 trillion kilograms - of these chemicals embedded in products such as pesticides, plastics, chemical drugs, and even personal hygiene products.

Deserving more attention are DDT (dichlorodiphenyltrichloroethane), DDE (dichlorodiphenyldichloroethylene), DDD (dichlorodiphenyldichloroethane) and their byproducts such as atrazine and 2,4-dichlorophenoxyacetic acid found in toys, and others containing lead and cadmium, materials used in the production of plastic bottles containing BPA, phthalates, and several other substances employed in the textile and apparel industries (Gore *et al.*, 2015). Numerous studies examined the effects of EDCs and their adverse effects against different areas of the female reproductive system.

#### **Historical Biochemical Features**

BPA or 2,2-bis(4-hydroxyphenyl) propane was first synthesized by Dianin, in 1891, and its estrogenic properties were found by Dodds & Lawson (1936). DES (diethylstilbestrol) was also found to have a powerful estrogenic effect (Dodds *et al.*, 1938). Later, in 1950, it was observed that BPA could be polymerized for the manufacturing of plastics given its lightweight, moldability, and impact resistance. Diethylstilbestrol was defined as the first "endocrine disrupting chemical", since abnormalities such as later development of vaginal adenosis, clear cell adenocarcinoma of the vagina, and/or uterine anomalies, were found in the exposed female offspring of pregnant women treated to prevent miscarriage.

#### **Endocrine Disruptors**

Examples of endocrine disruptors:

#### 1. Bisphenols

Bisphenol A (BPA) was the first to be synthesized, but evidences gathered in 1936 showed a low estrogen effect with affinity for the nuclear estrogen receptor. Its effects depend on dosage, targeted tissue, and tissue development on the site where it acts. The occurrence of estrogenic or anti-estrogenic effects depends on the tissue targeted and on their impact on receptors (Rochester et al., 2015). Global production of BPA has steadily grown in recent years on account of its multiple applications in the plastic and manufacturing industries, in food packaging and toys, causing a constant and permanent poisoning of food, water, and the environment. In 1950, it was found that bisphosphonates could be polymerized and, since then, they have been used to make polycarbonate plastics. These plastics have convenient features such as lightweight, moldability, and impact and heat resistance, and are not susceptible to changes over time. About 20% of these plastics are used as a component of epoxy resin, serving as internal coating for plastic containers and bottles. Therefore, it is a liquid and food contaminant present in abnormal levels in human serum analysis according to the literature. BPA is rapidly metabolized to inactive forms with a mean life cycle of approximately 4-5 hours in adults, while in fetuses and children the metabolic rate is relatively low (Gerona et al., 2013; Sartain & Hunt, 2016). BPA can easily accumulate in adipose tissue for having lipophilic properties. Measurements of human serum have determined varied and controversial toxicity rates. Currently, the United States Environmental Protection Agency has established a safe level of 50µg/kg/day and the European Food Safety Authority has established a tolerable daily intake below 4µg/kg/day.

#### 2. Phthalates

Phthalates and their esters consist of a large group of chemical compounds frequently used in the plastic, coating, cosmetic, and toy industries, including the manufacturing of medical equipment such as syringes and blood bags. Phthalates are byproducts of phthalic acid and are used in the plastics industry for their excellent moldability. There are no regulations restricting the use of phthalates in the United States or in Brazil, but the European Community has banned phthalates. In the roster of phthalates, three esters are considered endocrine disruptors with estrogenic effects: DHEP (diethyl-hexyl phthalate), BBP (benzyl-butyl phthalate), and DBP (dibutyl phthalate). Phthalates can be found not only in serum and human urine, but also in milk samples. Tolerable daily intake ranges between 3-30ug/ kg/day (Hines et al., 2009; Fromme et al., 2011; Hannon & Flaws, 2015).

#### 3. Atrazine

Atrazine (2-chloro-4-ethylamino-6-isopropylamino-1,3,5-s-triazine), as chlorotriazine, is largely used in agriculture as a herbicide. It has been used to reduce the growth of leaves and weeds in wheat, soy, and sugar cane crops due to the inhibition of photosynthesis (Gianessi, 1998). Its metabolites remain active for long periods of time and, as pesticides, they cause water contamination, including water sources for human consumption (Solomon *et al.*, 2013).

## 4. Esters of Polychlorinated and Polybrominated Bisphenols

Polychlorinated bisphenols (PCBs) are chemical substances with a phenolic ring and different degrees of chlorination. They were first manufactured in 1920, and were used in the rubber, resin, adhesives, and paint industries (Soto et al., 1995). These chemicals were extensively used around the world and contaminated schools and construction sites. They build up both in the environment and in adipose tissue, and are considered endocrine disruptors affecting the thyroid hormone with estrogenic and anti-androgenic activity. PCBs were banned in 1979 for their persistent pollutant effects. The polybrominated esters of bisphenols were first used as flame retardants and in mattresses and blankets (ATSDR, 2004; 2017). Of all 209 synthesized products categorized as polybrominated aromatic compounds, five esters top the list of toxicity: tetra BDE-47, penta BDE99 -100, -153 and deca BDE-209 or PBDE= Polybrominated diphenyl ethers. (Zota et al., 2011; Costa & Giordano, 2007).

## 5. DDT (Dichlorodiphenyltrichloroethane) - DDE (Dichlorodiphenyldichloroethylene) - DDD (Dichlorodiphenyldichloroethane)

These are chemical compounds once widely used as insecticides with a long life and strong lipophilic properties. Evidenced as contaminants to the environment, exposure to these chemicals can lead to several endocrine diseases, although they have been used to control insects that carry malaria (National Toxicology Program, 2011; McGlynn *et al.*, 2008; Hardell *et al.*, 2004; Safe & Zacharewski, 1997). DDT was banned in 1972 due to its high toxicity levels.

In addition to DDT, other pesticides deserve to be mentioned such as hexachlorocyclohexane, chlordane, and hexachlorobenzene. These products have been closely studied not only for persistently building up in nature but also for being endocrine disrupting chemicals. However, there are new pesticides being launched in the market with shorter mean lives and similar effects, such as 2,4-dichlorophenol, 2,5-dichlorophenol, and 1-naphthol, present in 50% of pregnant women in the Salinas Valley, California, USA.

#### 6. Heavy metals or organometallic compounds

Elements such as cadmium, lead, and mercury have been widely used in various scenarios leading to a great number of reproductive anomalies. Cadmium is used in batteries, metallic pigments, and plastics, but exposure to this chemical may cause harmful effects to the placental DNA and fetal umbilical cord, in addition to accumulating in the liver and kidneys. Lead was once extensively used in the paint, oil, and toy industries. Its adverse effects include genomic methylation and a number of different abnormalities in brain development. Mercury was once used in several industrial processes and emissions have been linked to burning charcoal. Human exposure occurs mainly through the intake of contaminated fish from sites such as Minamata Bay, Japan, the Faroe Islands in the Northern Atlantic, and Nunavik in Canada.

#### 7. Diethylstilbestrol (DES)

This powerful synthetic non-steroidal estrogen was used in the USA from 1940 to 1975 to prevent miscarriage and/or its complications. Initially, low doses of 5mg/day were administered, but they were progressively increased to 125mg/day or more, and eventually got to a mean dose of 3650-4000mg. Dieckmann et al. (1953) proved this treatment was ineffective. Herbst et al. (1971) assessed young women and noticed a correlation between the use of DES and the appearance of clear cell vaginal adenocarcinoma. In 1976, the same author (Herbst, 1976) described other abnormalities in the genital tract of young women whose mothers had been treated with DES. Harris & Waring (2012) and Troisi et al. (2013) described increased numbers of reproductive system disorders in the male and female children of mothers treated with DES. The disorders included cryptorchidism, uterine abnormalities such as T-shaped uterus, and some types of hormone-dependent cancers.

#### Pathological findings

This review lists a number of reproductive abnormalities associated to endocrine disruptors and their different effects:

## **1.** Effects of exposure to bisphenols and other toxins on ovarian steroidogenesis

Endocrine glands secrete different hormones that regulate the development, physiologic processes, and homeostasis of all organisms. These hormones interact with various receptors on target cells, according to their affinity, and have dissociation constants ranging between 10-12 and 10-9, associated with their low circulating concentrations. BPA is an endocrine disruptor that binds to estrogen receptors alpha and beta with a binding affinity 1000 to 10000 times lower than that of endogenous estradiol (Kuiper *et al.*, 1997; Mlynarcíková *et al.*, 2005). BPA further binds to the gramma and G-protein membrane receptors and to the pregnane X receptor, thus activating ion channels and inducing pro-inflammatory responses of cytokines and chemokines (Chapin *et al.*, 2008; Huang & Leung, 2009)

Cholesterol is the substrate needed for enzyme CYP-450scc to complete its cleavage and catalyze the conversion from cholesterol to pregnenolone. Pregnenolone is then converted into an androgenic precursor, DHEA (dehydroepiandrosterone), including the intermediary product, 17-Hydroxipregnenolone, involving two enzymes in this conversion: 17 alpha-hydroxylase and 17-20 desmolase. Subsequently, DHEA is converted into androstenedione, which, while in the theca cell compartments, is converted to testosterone, so that both can migrate through the basal lamina of the antral follicle to the granulosa cells. Since there is aromatase CYP450 in the granulosa cells, both androgens are converted into estrone and estradiol (E2) (Two cell theory by Hillier *et al.*, 1994).

An experimental study by Peretz & Flaws (2013) evaluating female rat ovarian follicles in the antral stage revealed that depending on the dose of BPA administered and the time of action, there was a reduction in the synthesis of estradiol, estrone, testosterone, androstenedione, and DHEA sulfate after 120 hours of exposure to 100µg/ml of BPA. This high level of BPA compromised follicular growth, but no effect was observed following a dose reduction of 1µg/ ml (Takayanagi *et al.*, 2006). Other experimental studies (Mlynarcíková *et al.*, 2005; Huang & Leung, 2009; Watanabe *et al.*, 2012) showed BPA inhibits the mechanism of aromatase CYP450 in the granulosa cells, thus reducing the production of E2.

Only a few studies in humans associated BPA with ovarian follicle synthesis. Mok-Lin *et al.* (2010), Ehrlich *et al.* (2012), and Manikkam *et al.* (2012a) evaluated the relationship between BPA and hormone levels in the granulosa cells of patients submitted to in vitro fertilization, and found a low E2 peak prior to ovum pickup. Lee *et al.* (2014) described found that young people on early puberty exposed to BPA had significant increases in testosterone, estradiol, and pregnenolone levels. On the other hand, Mínguez-Alarcón *et al.* (2015) found no statistical significance between BPA levels found in urine, serum E2 levels, and endometrial thickness measured by ultrasound examination after adjusting the findings for age, body mass index, race, smoking habit, and diagnosis of infertility.

Only a few studies evaluating the harmful effects of exposure to phthalates and the negative effects on steroidogenesis where performed, with insufficient data collection and inadequate statistical methods. On a study called "The Western Australian Pregnancy Cohort Study", Hart et al. (2014) described the negative effects of phthalate metabolites on maternal serum SHBG levels, while the association between those same metabolites with the maternal androgen levels was inconsistent. Other animal and in vitro studies described the impact of phthalates on normal steroidogenesis. Xu et al. (2010) showed that in rats, aromatase inhibition in the granulosa cells led to decreased E2 levels. Svechnikova et al. (2007) and Liu et al. (2014) reported decreased progesterone levels in rats exposed to DEHP (diethylhexyl phthalate) and decreased E2 levels in female rats at 20 days of age, but also decreased sexual hormone levels in adult rats even though the results are still controversial.

Claims that exposure to pesticides may change steroidogenesis in women also have their limitations. A previous study by Luderer et al. (2013) including 457 Hawaiian participants exposed to heptachlor epoxide, observed a shorter luteal phase and decreased serum levels of progesterone and estradiol metabolites. Atrazine effects in steroidogenesis seem to differ according to age, dose, and experimental model. In vitro studies by Fa et al. (2013) showed that Atrazine might alter enzyme expression in steroidogenesis and E2 levels with immature granulosa cells of female rats. In vivo studies with adult animals by Taketa et al. (2011), Quignot et al. (2012), Tinfo et al. (2011), and Buck Louis et al. (2014) showed that repeated doses of Atrazine increased enzyme expression in steroidogenesis and sexual hormone levels. An insufficient number of studies have been performed with female patients. Further research is needed to verify whether these pesticides are harmful to steroidogenesis and fecundity. To this day, only Buck Louis *et al.* (2014) with the LIFE Study showed some evidence on the adverse effects of different DEC to female fecundity.

Environmental Toxicants: Su *et al.* (2012) conducted studies in humans showing that high concentrations of dioxins and polychlorinated biphenyl byproducts decreased plasma estradiol levels. Experimental trials with different animals demonstrated that exposure to dioxins adversely affected ovarian steroidogenesis. Further effects included decreases in estradiol production and synthesis in the antral follicles of female rats, and loss of enzyme synthesis (Karman *et al.*, 2012a; 2012b). However, there are clear limitations in these trials and further research in humans is required.

#### 2. Polycystic ovary syndrome and bisphenols

Polycystic ovary syndrome (PCOS) is an endocrine disorder that includes multiple clinical conditions such as anovulatory cycles, hyperandrogenism, obesity, and regular insulin resistance associated with hypercholesterolemia, dyslipidemia and other metabolic alterations. Today, more than 800 chemical products categorized as endocrine disruptors may strongly affect hormone receptors, act as agonists or antagonists, and lead to anovulation. An evaluation of different phenotypes of patients with PCOS revealed a wide ethnical, geographical, and familial diversity even among twin siblings. Recent studies showed many women exposed to chemical compounds have a genetic susceptibility to developing PCOS and several related metabolic disorders. Time of exposure to these endocrine disruptors is crucial to determine their effect, especially during fetal development, given their potential harmful effects to pregnancy hormones and fetal cellular programming (Palioura & Diamanti-Kandarakis, 2015; Diamanti-Kandarakis et al., 2007).

Prenatal androgenization leads to epigenetic changes and future development of PCOS phenotypes (Xu *et al.*, 2011). Plastic substances caused DNA methylation in animals, and exposure to biphenyl and phthalates in the F0-generation Zero led to trans-generational changes up to the third generation (F3) (Nilsson *et al.*, 2012; Manikkam *et al.*, 2012b; 2014). An observational study with female rats given high doses of EDCs such as BPA in the neonatal stage resulted in adult PCOS phenotypes with increased plasma testosterone and estradiol levels, decreased progesterone levels, and development of ovarian cysts (Fernández *et al.*, 2010). Fernández *et al.* (2009) also described alterations in the pulsatile secretion of GNRH and LH secretion from the pituitary gland.

Although there is a large number of findings in animal studies, translating their conclusions to humans is rather difficult, once the cystic aspect and the different pheno-types found in animals differ from the findings in humans of PCOS antral follicles. Exposure to testosterone in the beginning of gestation in rhesus monkeys provokes metabolic disorders similar to many women with PCOS. Late exposure to dihydrotestosterone and EDCs causes hyper-androgenism and irregular menstrual cycles, similarly to women with PCOS (Abbott *et al.*, 2013).

Coexistence of insulin resistance is found in 50-80% of women with PCOS, leading to decreased insulin sensitivity and hyperglycemia and hyperinsulinemia, followed by hyperandrogenism and chronic anovulation. The mechanisms linked to insulin resistance are still unclear. However, Polyzos *et al.* (2012) suggested the involvement of EDCs in the etiology of insulin resistance. BPA is considered a causal factor of child obesity linked to decreased adiponectin levels, onset of inflammation, and greater risk of developing diabetes type 2 and cardiovascular disorders (Menale *et al.*, 2017). Other mechanisms have been connected to BPA-related hyperglycemia and hyperinsulinemia with direct effect on pancreatic cells, although no alterations on the pancreas islets have been documented (Alonso-Magdalena *et al.*, 2005; 2006).

### 3. Endocrine disrupting chemicals and endometriosis

An enigmatic condition, endometriosis is an estrogen-dependent disease with numerous endocrine disruptors affecting the ectopic endometrial tissue and a wide array of etiological factors. EDCs such as TCDD (2,3,7,8-tetrachlorodibenzo-p-dioxin) and PCBs (dioxin-like polychlorinated biphenyl) may induce the development of peritoneal endometriosis in female Rhesus monkeys, and the magnitude of the effect depends on the level of contamination (Rier *et al.*, 2001). Bredhult *et al.* (2007) evidenced by the proliferation of endometrial cells triggered by the angiogenic effect of TCCD's estrogenization action.

Attention should be paid to The studies with humans performed by Pauwels *et al.* (2001), Eskenazi *et al.* (2002), Fierens *et al.* (2003), Heilier *et al.* (2005), and Simsa *et al.* (2010) deserve attention for the correlations drawn between the effects of dioxin-like products and the genesis of pelvic endometriosis. However, other authors were unable to verify their findings or describe a correlation between PCB and endometriosis (Porpora *et al.*, 2009; Buck Louis *et al.*, 2012).

Phthalates may also have a proliferative effect on endometrial tissue. A prospective case-control study by Kim *et al.* (2011) showed that women with advanced pelvic endometriosis had increased plasma levels of DEHP (di-(2-ethylhexyl) phthalate) and MEHP (mono-(2-ethylhexyl) phthalate) compared to endometriosis-free controls. An additional case-control study found that women with endometriosis had significantly higher concentrations of mono-n-butyl-phthalates in urine than controls (Huang *et al.*, 2010). Buck Louis *et al.* (2013) reported levels twice as high of six phthalate metabolites in women with pelvic endometriosis.

Nevertheless, other epidemiological studies failed to validate these findings. Upson *et al.* (2013), in a study including women from the Northeast of the USA, showed an inverse association between the risk of developing endometriosis and levels of MEHP. Itoh *et al.* (2009) confirmed these findings in a study enrolling infertile women, although the authors included only 57 cases of endometriosis and 80 endometriosis-free controls. The mechanism triggering the development of endometriosis by phthalates remains unclear. Only Kim *et al.* (2010) in an in vitro study showed that DEHP (di-(2-ethylhexyl) phthalate) stimulated the stroma of endometrial cells and increased the viability of Ishikawa cells.

#### 4. The effects of endocrine disruptors in the ovaries, uterine structure, uterine myomas, and vagina.

In recent years, a number of animal and in vitro studies looked into abnormalities in the development of the ovaries linked to endocrine disruptors. The impact of BPA in the development of human ovaries remains unclear. Previous studies by Rivera *et al.* (2011) and Veiga-Lopez *et al.* (2013) demonstrated that low doses of BPA in sheep might lead to increased ovarian follicles with multiple oocytes and altered ovarian steroidogenesis. Hunt *et al.* (2012) described the impact of BPA in the fetal development of female monkey ovaries affecting early meiosis, causing synaptic alterations, and interfering with the recombination between homologous chromosomes.

Insufficient data is available on the impact of phthalates, pesticides, and other environmental toxicants in human prenatal ovarian development. Studies on human postnatal ovarian development are also limited. Sheep and rats exposed to BPA during pregnancy had ovarian anomalies. Low doses of BPA decreased the number of follicles and increased follicular atresia in female rats. On the other hand, high doses of BPA might lead to increased follicular cystification, corpus luteum depletion, and decreased antral follicle counts (Rodríguez *et al.*, 2010; Delclos *et al.*, 2014). According to Chen *et al.* (2012), human ovaries exposed after birth to phthalates such as benzyl butyl phthalate have greater chances of developing granulosa cell apoptosis.

Numerous pesticides such as endosulfan, malathion chlorpyriphos, and cypermethrin cause postnatal ovary anomalies, inducing decreased follicle counts and increased follicular atresia as described by Koç *et al.* (2009) and Nandi *et al.* (2011). While looking into another noteworthy environmental toxicant, Petro *et al.* (2012) linked increased levels of chlorinated bisphenols in humans and in ovarian follicular liquid to decreased fertilization rates and poor conditions for oocyte development.

The uterus is a muscle organ consisting of two main elements: the body, which includes the endometrium, and the caudal end, or cervix, both exposed to significant hormonal influence from their early stages of development. After puberty, the uterus has periodical cycles of hormonal variation and greatly expands during pregnancy, while after menopause the uterus involutes and decreases in size.

The prospective effects of BPA and pesticides in uterine structure and function remain unknown, but abnormalities have been reported in animal studies. Exposure to BPA in the gestational and neonatal periods causes the development of endometrial glands and stroma, as seen in the adipose tissue next to the genital tract of type Balb-c-adult female rats (Signorile et al., 2010; 2012). A single Australian study found that human exposure to mono (carboxy-isooctyl) phthalate changed the uterine volume (Hart et al., 2014). As previously demonstrated (Dieckmann et al., 1953; Herbst et al., 1971; Herbst, 1976; Harris & Waring, 2012; Troisi et al., 2013), diethylstilbestrol caused a great number of uterine abnormalities in the exposed daughters of pregnant women treated to prevent miscarriage during gestation. Along the same lines, other recent studies showed that exposure to DES induced endometrial hyperplasia/dysplasia and increased the chances of endometrial adenocarcinoma and uterine anomalies in female rats and hamsters (Alwis et al., 2011; Yoshida et al., 2011).

Other environmental toxicants have been tested in animals and in humans. Su et al. (2012) found that dioxin and polychlorinated aromatic byproducts of biphenyls caused anomalies in the uterine structure, function, and fundus of 33 young girls. Uterine myomas or fibromyomas are mostly benign tumors, affecting approximately 70-80% of the female population throughout their lives. The growth of nodular tumors and multiple myomas is hormone-dependent and connected to the estradiol and progesterone receptors in the myometrium. The estradiol produced in the granulosa cells of the ovarian follicles regulates the endometrium and myometrium cells by activating their alpha and beta cellular receptors. The binding affinity between estradiol and its receptor can trigger a number of events, mostly in the cell nucleus, by recruiting important proteins to cellular reproduction.

Following ovulation, the corpus luteum produces progesterone, an essential hormone to female reproduction, binding to the A and B progesterone receptors. These receptors promote and regulate the expression of several genes, leading to different cellular responses. No significant correlation has been found between BPA activity and its capacity to promote the development of fibromas. Two Chinese case-control studies by Shen *et al.* (2013) and Zhou *et al.* (2013) showed an association between higher levels of BPA, nonylphenol and octylphenol in women with uterine myomas.

Other authors found positive correlations between disease and phthalates. In 2010, the NHANES study showed that mono-benzyl phthalate increased the risk of myoma in 1227 women. However, other phthalates such as MEHP (mono-(2-ethylhexyl) phthalate), MEHHP (mono-(2-ethyl-5-hydrohexyl) phthalate), and EOP (mono- (2-ethyl-oxohexyl) phthalate) were inversely correlated (Weuve *et al.*, 2010) with the onset of disease.

Meanwhile, few studies reported that prenatal exposure to DES (diethylstilbestrol) increased the number of fibromas. The NURSES study included 11831 and followed them for over 20 years. DES exposure during gestation increased the risk of fibroma by 13%, while DES exposure during the first trimester of pregnancy increased the risk of fibroma by 21% in comparison to non-exposed women (Baird & Newbold, 2005; Mahalingaiah et al., 2014). Another NIEHS Uterine Fibroid Study completed in Washington-DC including 1364 exposed women aged 35-49 reported an odds ratio of 2.4 for Caucasian women (Baird & Newbold, 2005). Subsequently, the NIEHS Sister Study evaluated a group of 3534 African-American women and showed increased risk of developing fibromas after exposure to DES in women with maternal and gestational diabetes and women pregnant with monozygotic twins, with respective odds ratios of 2.02, 1.54, and 1.94 according to D'Aloisio et al. (2012). This study included 19972 Caucasian women and documented five significant risk factors: prenatal exposure to DES; gestational diabetes; getting pregnant while having a history of diabetes; use of soy protein-based formula; and advanced maternal age. All these factors represented an increase of more than 20% in the risk of having fibromas.

As described in 1953, chemical disruptors such as DES (diethylstilbestrol) induce the development of neoplasms such as vaginal adenosis and clear cell adenocarcinoma on the vaginal walls (Dieckmann *et al.*, 1953; Herbst *et al.*, 1971; Herbst, 1976; Harris & Waring, 2012; Troisi *et al.*, 2013). DES (Diethylstilbestrol) inhibits the vaginal stroma causing a persistent down-regulation of the transcription factors (Laronda *et al.*, 2013; Katoh *et al.*, 2013; Nakamura *et al.*, 2012a; 2012b).

## 5. Effects of exposure to endocrine disrupting chemicals on the pituitary gland, menstrual cycles, and fertility

Only a handful of human studies have looked into the correlation between bisphenols, phthalates, and pesticides and harmful effects on the anterior pituitary gland compartment. On the other hand, Xi *et al.* (2011) and Brannick *et al.* (2012) showed that prenatal and immediate postnatal exposure to BPA stimulated the hypothalamic-pituitary axis and increased the number and replication of pituitary gonadotropins in female rats. A recent study by Souter *et al.* (2013) found no correlation between exposure to BPA and FSH levels in women on the third day of IVF cycles. Miao *et al.* (2015) described a positive correlation between exposure to BPA and preserve to BPA and urinary levels of prolactin, although a negative correlation was established with FSH levels.

Animal studies have occasionally described harmful effects of other toxicants such as DES and dioxins (Yoshida *et al.*, 2011; Ishikawa *et al.*, 2014) as having rather controversial effects on the pituitary gland and human gonadotropins. Insufficient human studies were made in the last five years with regard to the effects of toxicants on fertility and menstrual cycles. According to Buck Louis *et al.* (2011), organochlorine pollutants led to an increase of three additional days on the interval between the periods

of women willing to conceive. Few studies demonstrated abnormalities in the menstrual cycles due to exposure to phthalates. Exposure to BPA induced abnormalities in the estrous cycle of rats according to Fernández *et al.* (2009), and Nah *et al.* (2011). Vélez *et al.* (2015), in the MIREC Study the Maternal-Infant Research on Environmental Chemicals, included 2001 women in the first trimester of pregnancy exposed to bisphenols and phthalates and the time of conception. Higher concentrations of Triclosan (>72ng/ml), a bactericide with phenolic compounds, decreased fertility, but no correlation was found with bisphenols and phthalates.

In Brazil, for many years several authors have shown their concern and have looked into the effects of environmental EDCs on the population and as a factor in occupational health (Branco, 1984; Nogueira *et al.*, 1987; Della Rosa & Gomes, 1988; Assunção & Pesquero, 1999; Sanseverino *et al.*, 2001). Studies by Peres *et al.* (2001), Lara *et al.* (2011), and Cremonese *et al.* (2012) also evaluated EDC potential harmful effects in the field of reproductive health.

#### CONCLUSIONS

Growing outspread exposure to a number of different chemicals has caused multiple abnormalities in the reproductive system of women and different animal species. Tens of millions of these chemicals are available in the global market, and even when administered in minimal doses, they have been described as endocrine disrupting chemicals or potential conditioners. Exposure to EDCs is extremely toxic and harmful to the reproductive system. Despite the occasional absence of a match between experimental and epidemiological data, it is clear that EDCs can produce adverse effects on the genital tract. The reproductive effects may be dose-dependent and associated to extended exposure and the level of activity of the compound at hand. Unfortunately, a significant number of these compounds are already a part of the environmental chain, making it difficult to assess their potential harmful effects.

Di Renzo *et al.* (2015) in the FIGO (International Federation of Gynecology and Obstetrics) drafted the following guidelines that must be observed by gynecologists, obstetricians, nurses, and other healthcare workers in this field:

- Extensive guidance and education to ensure preventive measures are in place with respect to exposure to toxicants;
- Appropriate measures to ensure healthy food is provided in the entire food chain; make sure pesticides are banned from farms so as not to contaminate vegetables, fresh fruits, or whole wheat grains; reduce the intake of animal fat or fish contaminated by heavy metals;
- Educational and participatory guidance involving the entire society to ensure they are aware of the risks related to the intake of these toxicants.

Further prospective studies are needed to better elucidate the possible abnormalities affecting women by millions of EDCs.

#### **CONFLICT OF INTEREST**

The authors have no conflict of interest to declare.

#### **Corresponding author:**

Mauri José Piazza Tocogynecology Department Universidade Federal do Paraná - UFPR Curitiba (PR), Brazil. E-mail: mauripiazza@hotmail.com

#### REFERENCES

Abbott DH, Nicol LE, Levine JE, Xu N, Goodarzi MO, Dumesic DA. Nonhuman primate models of polycystic ovary syndrome. Mol Cell Endocrinol. 2013;373:21-8. PMID: 23370180 DOI: 10.1016/j.mce.2013.01.013

Alonso-Magdalena P, Laribi O, Ropero AB, Fuentes E, Ripoll C, Soria B, Nadal A. Low doses of bisphenol A and diethylbestrol impair Ca2+signals in pancreatic alpha-cells through a noclassical membrane estrogen receptor within intact islets of Langerhans. Environ Health Perspect. 2005;113:969-77. PMID: 16079065 DOI: 10.1289/ ehp.8002

Alonso-Magdalena P, Morimoto S, Ripoll C, Fuentes E, Nadal A. The estrogenic effect of bisphenol A disrupts pancreatic beta-cell function in vivo and induces insulin resistance. Environ Health Perspect. 2006;114:106-12. PMID: 16393666 DOI: 10.1289/ehp.8451

Alwis ID, Maroni DM, Hendry IR, Roy SK, May JV, Leavitt WW, Hendry WJ. Neonatal diethylstilbestrol exposure disrupts female reproductive tract structure/function via both direct and indirect mechanisms in the hamster. Reprod Toxicol. 2011;32:472-83. PMID: 21963885 DOI: 10.1016/j. reprotox.2011.09.006

Assunção JV, Pesquero CR. Dioxins and furans: origins and risks. Rev Saúde Pública. 1999;33:523-30. DOI: 10.1590/ S0034-89101999000500014

ATSDR - Agency for Toxic Substances and Disease Registry. Toxicological profile for Polybrominated Biphenyls. Atlanta GA: US Department of Health and Human Services. Public Health Service; 2004. Available at: https://www.atsdr.cdc. gov/ToxProfiles/tp68.pdf. Accessed: 11/07/2018.

ATSDR - Agency for Toxic Substances and Disease Registry. Toxicological profile for Polybrominated Diphenyl ethers (PBDEs). Atlanta GA: US Department of Health and Human Services. Public Health Service; 2017. Available at: https://www.atsdr.cdc.gov/toxprofiles/tp207.pdf. Accessed: 11/07/2018.

Baird DD, Newbold R. Prenatal diethylstilbestrol (DES) exposure is associated with uterine leiomyoma development. Reprod Toxicol. 2005;20:81-4. PMID: 15808789 DOI: 10.1016/j.reprotox.2005.01.002

Birnbaum LS. When environmental chemicals act like uncontrolled medicine. Trends Endocrinol Metab. 2013;24:321-3. PMID: 23660158 DOI: 10.1016/j.tem.2012.12.005

Branco SM, ed. O fenômeno de Cubatão. São Paulo: Convênio Cetesb Ascetesb; 1984.

Brannick KE, Craig ZR, Himes AD, Peretz JR, Wang W, Flaws JA, Raetzman LT. Prenatal exposure to low doses of bisphenol A increases pituitary proliferation and gonadotroph number in female mice offspring at birth. Biol Reprod. 2012;87:82. PMID: 22875908 DOI: 10.1095/biolreprod.112.100636

Bredhult C, Bäcklin BM, Olovsson M. Effects of some endocrine disruptors on the proliferation and viability of human endometrial endothelial cells in vitro. Reprod Toxicol. 2007;23:550-9. PMID: 17493787 DOI: 10.1016/j.reprotox.2007.03.006 Buck Louis GM, Rios LI, McLain A, Cooney MA, Kostyniak PJ, Sundaram R. Persistent organochlorine pollutants and menstrual cycle characteristics. Chemosphere. 2011;85:1742-8. PMID: 22018858 DOI: 10.1016/j.chemosphere.2011.09.027

Buck Louis GM, Chen Z, Peterson CM, Hediger ML, Croughan MS, Sundaram R, Stanford JB, Varner MW, Fujimoto VY, Giudice LC, Trumble A, Parsons PJ, Kannan K. Persistent lipophilic environmental chemicals and endometriosis: the ENDO Study. Environ Health Perspect. 2012;120:811-6. PMID: 22417635 DOI: 10.1289/ehp.1104432

Buck Louis GM, Peterson CM, Chen Z, Croughan M, Sundaram R, Stanford J, Varner MW, Kennedy A, Giudice L, Fujimoto VY, Sun L, Wang L, Guo Y, Kannan K. Bisphenol A and phthalates and endometriosis: the Endometriosis: Natural History, Diagnosis and Outcomes Study. Fertil Steril. 2013;100:162-9 e1-2. PMID: 23579005 DOI: 10.1016/j. fertnstert.2013.03.026

Buck Louis GM, Sundaram R, Sweeney AM, Schisterman EF, Maisog J, Kannan K. Urinary bisphenol A, phthalates, and couple fecundity: the Longitudinal Investigation of Fertility and the Environment (LIFE) Study. Fertil Steril. 2014;101:1359-66. PMID: 24534276 DOI: 10.1016/j. fertnstert.2014.01.022

Chapin RE, Adams J, Boekelheide K, Gray LE Jr, Hayward SW, Lees PS, McIntyre BS, Portier KM, Schnorr TM, Selevan SG, Vandenbergh JG, Woskie SR. NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. Birth Defects Res B Dev Reprod Toxicol. 2008;83:157-395. PMID: 18613034 DOI: 10.1002/bdrb.20147

Chen HS, Chiang PH, Wang YC, Kao MC, Shieh TH, Tsai CF, Tsai EM. Benzyl butyl phthalate induces necrosis by AhR mediation of CYP1B1 expression in human granulosa cells. Reprod Toxicol. 2012;33:67-75. PMID: 22138065 DOI: 10.1016/j.reprotox.2011.11.004

Costa LG, Giordano G. Developmental neurotoxicity of polibrominated diphenyl ether (PBDE) flame retardants. Neurotoxicology. 2007;28:1047-67. PMID: 17904639 DOI: 10.1016/j.neuro.2007.08.007

Cremonese C, Freire C, Meyer A, Koifman S. Pesticide exposure and adverse pregnancy events, Southern Brazil, 1996-2000. Cad Saúde Pública. 2012;28:1263-72. PMID: 22729257 DOI: 10.1590/S0102-311X2012000700005

D'Aloisio AA, Baird DD, DeRoo LA, Sandler DP. Early-life exposures and early-onset uterine leiomyomata in black women in the Sister Study. Environ Health Perspect. 2012;120:406-12. PMID: 22049383 DOI: 10.1289/ ehp.1103620

Delclos KB, Camacho L, Lewis SM, Vanlandingham MM, Latendresse JR, Olson GR, Davis KJ, Patton RE, Gamboa da Costa G, Woodling KA, Bryant MS, Chidambaram M, Trbojevich R, Juliar BE, Felton RP, Thorn BT. Toxicity evaluation of bisphenol A administered by gavage to Sprague Dawley rats from gestation day 6 through postnatal day 90. Toxicol Sci. 2014;139:174-97. PMID: 24496637 DOI: 10.1093/toxsci/kfu022

Della Rosa HV, Gomes JR. Cadmium: toxicokinetics. Rev Bras Saude Ocup. 1988;16:39-42.

Di Renzo GC, Conry JA, Blake J, DeFrancesco MS, DeNicola N, Martin JN Jr, McCue KA, Richmond D, Shah A, Sutton P, Woodruff TJ, van der Poel SZ, Giudice LC. International Federation of Gynecology and Obstetrics opinion on reproductive health impacts of exposure to toxic environmental chemicals. Int J Gynecol Obstet. 2015;131:219-25. PMID: 26433469 DOI: 10.1016/j.ijgo.2015.09.002

Diamanti-Kandarakis E, Piperi C, Patsouris E, Korkolopoulou P, Panidis D, Pawelczyk L, Papavassiliou AG, Duleba AJ. Immunohistochemical localization of advanced glycation end-products (AGEs) and their receptor (RAGE) in polycystic and normal ovaries. Histochem Cell Biol. 2007;127:581-9. PMID: 17205306 DOI: 10.1007/s00418-006-0265-3

Dieckmann WJ, Davis ME, Rynkiewicz LM, Pottinger RE. Does the administration of diethylbestrol during pregnancy have therapeutic value? Am J Obstet Gynecol. 1953;66:1062-81. PMID: 13104505 DOI: 10.1016/S0002-9378(99)70410-2

Dodds EC, Lawson W. Synthetic oestrogenic agentes without the phenantrene nucleus. Nature. 1936;137:996.

Dodds EC, Goldberg L, Lawson W, Robinson R. Estrogenic activity of certain synthetic compounds. Nature. 1938;141:247-8.

Ehrlich S, Williams PL, Missmer SA, Flaws JA, Ye X, Calafat AM, Petrozza JC, Wright D, Hauser R. Urinary bisphenol A concentrations and early reproductive health outcomes among women undergoing IVF. Hum Reprod. 2012;27:3583-92. PMID: 23014629 DOI: 10.1093/humrep/des328

Eskenazi B, Mocarelli P, Warner M, Samuels S, Vercellini P, Olive D, Needham LL, Patterson DG Jr, Brambilla P, Gavoni N, Casalini S, Panazza S, Turner W, Gerthoux PM. Serum dioxin concentrations and endometriosis: a cohort study in Seveso, Italy. Environ Health Perspect. 2002;110:629-34. PMID: 12117638 DOI: 10.1289/ehp.02110629

Fa S, Pogrmic-Majkic K, Samardzija D, Glisic B, Kaisarevic S, Kovacevic R, Andric N. Involvement of ERK1/2 signaling pathway in atrazine action on FSH-stimulated LHR and CY-P19A1 expression in rat granulosa cells. Toxicol Appl Pharmacol. 2013;270:1-8. PMID: 23583632 DOI: 10.1016/j. taap.2013.03.031

Fernández M, Bianchi M, Lux-Lantos V, Libertun C. Neonatal exposure to bisphenol a alters reproductive parameters and gonadotropin releasing hormone signaling in female rats. Environ Health Perspect. 2009;117:757-62. PMID: 19479018 DOI: 10.1289/ehp.0800267

Fernández M, Bourguignon N, Lux-Lantos V, Libertun C. Neonatal exposure to bisphenol a and reproductive and endocrine alterations resembling the polycystic ovarian syndrome in adult rats. Environ Health Perspect. 2010;118:1217-22. PMID: 20413367 DOI: 10.1289/ehp.0901257

Fierens S, Mairesse H, Heilier JF, De Burbure C, Focant JF, Eppe G, De Pauw E, Bernard A. Dioxin/polychlorinated biphenyl body burden, diabetes and endometriosis: findings in a population-based study in Belgium. Biomarkers. 2003;8:529-34. PMID: 15195683 DOI: 10.1080/1354750032000158420 Fromme H, Gruber L, Seckin E, Raab U, Zimmermann S, Kiranoglu M, Schlummer M, Schwegler U, Smolic S, Völkel W; HBMnet. Phthalates and their metabolites in breast milk--results from the Bavarian Monitoring of Breast Milk (BAMBI). Environ Int. 2011;37:715-22. PMID: 21406311 DOI: 10.1016/j.envint.2011.02.008

Gerona RR, Woodruff TJ, Dickenson CA, Pan J, Schwartz JM, Sen S, Friesen MW, Fujimoto VY, Hunt PA. Bisphenol-A (BPA), BPA glucuronide, and BPA sulfate in midgestation umbilical cord serum in a northern and central California population. Environ Sci Technol. 2013;47:12477-85. PMID: 23941471 DOI: 10.1021/es402764d

Gianessi LP. Benefits of triazine herbecides. In: Ballantine LG, McFarland JE, Hackett DE, eds. Triazine Herbicides: Risk Assesment. Washington DC: American Chemical Society; 1998. p. 1-8.

Gore AC, Chappell VA, Fenton SE, Flaws JA, Nadal A, Prins GS, Toppari J, Zoeller RT. EDC-2: The Endocrine Society's Second Scientific Statement on Endocrine-Disrupting Chemicals. Endocr Rev. 2015,36:E1-150. PMID: 26544531 DOI: 10.1210/er.2015-1010

Hannon PR, Flaws JA. The effects of phtalates on the ovary. Front Endocrinol (Lausanne). 2015;6:8. PMID: 25699018 DOI: 10.3389/fendo.2015.00008

Hardell L, van Bavel B, Lindström G, Björnfoth H, Orgum P, Carlberg M, Sörensen CS, Graflund M. Adipose tissue concentrations of p,p'-DDE and the risk for endometrial cancer. Gynecol Oncol. 2004;95:706-11. PMID: 15581986 DOI: 10.1016/j.ygyno.2004.08.022

Harris RM, Waring RH. Diethylstilboestrol--a long-term legacy. Maturitas. 2012;72:108-12. PMID: 22464649 DOI: 10.1016/j.maturitas.2012.03.002

Hart R, Doherty DA, Frederiksen H, Keelan JA, Hickey M, Sloboda D, Pennell CE, Newnham JP, Skakkebaek NE, Main KM. The influence of antenatal exposure to phthalates on subsequent female reproductive development in adolescence: a pilot study. Reproduction. 2014;147:379-90. PMID: 24025997 DOI: 10.1530/REP-13-0331

Heilier JF, Nackers F, Verougstraete V, Tonglet R, Lison D, Donnez J. Increased dioxin-like compounds in the serum of women with peritoneal endometriosis and deep endometriotic (adenomyotic) nodules. Fertil Steril. 2005;84:305-12. PMID: 16084869 DOI: 10.1016/j.fertnstert.2005.04.001

Herbst AL, Ulfelder H, Poskanzer DC. Adenocarcinoma of the vagina. Association of maternal stilbestrol therapy with tumor appearance in young women. N Engl J Med. 1971;284:878-81. PMID: 5549830 DOI: 10.1056/ NEJM197104222841604

Herbst AL. Summary of the changes in the human female genital tract as a consequence of maternal diethylstilbestrol therapy. J Toxicol Environ Health Suppl. 1976;1:13-20. PMID: 994230

Hillier SG, Whitelaw PF, Smyth CD. Follicular oestrogen synthesis: the 'two-cell, two-gonadotrophin' model revisited. Moll Cell Endocrinol. 1994;100:51-4. PMID: 8056158 DOI: 10.1016/0303-7207(94)90278-X

Hines EP, Calafat AM, Silva MJ, Mendola P, Fenton SE. Concentrations of phthalate metabolites in milk, urine, saliva, and Serum of lactating North Carolina women. Environ Health Perspect. 2009;117:86-92. PMID: 19165392 DOI: 10.1289/ehp.11610

Huang H, Leung LK. Bisphenol A downregulates CYP19 transcription in JEG-3 cells. Toxicol Lett. 2009;189:248-52. PMID: 19539015 DOI: 10.1016/j.toxlet.2009.06.853

Huang PC, Tsai EM, Li WF, Liao PC, Chung MC, Wang YH, Wang SL. Associaton between phtalate exposure and glutathione S-transferase M1 polymorphism in adenomyosis, leiomioma and endometriosis. Hum Reprod. 2010;25:986-94. PMID: 20147336 DOI: 10.1093/humrep/deq015

Hunt PA, Lawson C, Gieske M, Murdoch B, Smith H, Marre A, Hassold T, VandeVoort CA. Bisphenol A alters early oogenesis and follicle formation in the fetal ovary of the rhesus monkeys. Proc Natl Acad Sci USA. 2012;109:17525-30. PMID: 23012422 DOI: 10.1073/pnas.1207854109

Ishikawa M, Murai E, Hashiguchi Y, Iguchi T, Sato T. Effects of diethylstilbestrol on luteinizing hormone-producing cells in the mouse anterior pituitary. Exp Biol Med (Maywood). 2014;239:311-9. PMID: 24521563 DOI: 10.1177/1535370213519722

Itoh H, Iwasaki M, Hanaoka T, Sasaki H, Tanaka T, Tsugane S. Urinary phtalates monoesters and endometriosis in infertile Japonese women. Sci Total Environ. 2009;408:37-42. PMID: 19811803 DOI: 10.1016/j.scitotenv.2009.09.012

Karman BN, Basavarajappa MS, Hannon P, Flaws JA. Dioxin exposure reduces the steroidogenic capacity of mouse antral follicles mainly at the level HSD17BI without altering atrésia. Toxicol Appl Pharmacol. 2012a;264:1-12. PMID: 22889882 DOI: 10.1016/j.taap.2012.07.031

Karman BN, Basavarajappa MS, Craig ZR, Flaws JA. 2,3,7,8-Tetrachlorodibenzeno-p-dioxin activates the aryl hydrocarbon receptor and alters sex steroid hormose secretion without affecting growth of mouse antral follicles in vitro. Toxicol Appl Pharmacol. 2012b;261:88-96. PMID: 22483799 DOI: 10.1016/j.taap.2012.03.015

Katoh T, Hayashi S, Iguchi T, Sato T. Epithelial-stromal interactions in the mouse vagina exposed neonatally to diethylstilbestrol. In vivo. 2013;27:333-7. PMID: 23606688

Kim YH, Kim SH, Lee HW, Chae HD, Kim CH, Kang BM. Increased viability of endometrial cells by in vitro treatment with di-(2-ethylhexyl) phtalate. Fertil Steril. 2010;94:2413-6. PMID: 20493477 DOI: 10.1016/j.fertnstert.2010.04.027

Kim SH, Chun S, Jang JY, Chae HD, Kim CH, Kang BM. Increased plasma levels of phthalate esters in women with advanced-stage endometriosis: a prospective case-control study. Fertil Steril. 2011;95:357-9. PMID: 20797718 DOI: 10.1016/j.fertnstert.2010.07.1059

Koç ND, Kayhan FE, Sesal C, Muslu MN. Dose-dependent effects of endosulfan and malathion on adult Wistar albino rat ovaries. Pak J Biol Sci. 2009;12:498-503. PMID: 19579998 DOI: 10.3923/pjbs.2009.498.503

Kuiper GG, Carlsson B, Grandien K, Enmark E, Häggblad J, Nilsson S, Gustafsson JA. Comparison of the ligand binding specifity and transcript tissue distribution of estrogen receptors and alpha and beta. Endocrinology. 1997;138:863-70. PMID: 9048584 DOI: 10.1210/endo.138.3.4979

Lara LA, Duarte AA, Reis RM. Impact of endocrine disruptors on the reproductive function and sexual response of men and women. Rev Bras Ginecol Obstet. 2011;33:377-80. PMID: 22282024 DOI: 10.1590/S0100-72032011001200001

Laronda MM, Unno K, Ishi K, Serna VA, Butler LM, Mills AA, Orvis GD, Behringer RR, Deng C, Sinha S, Kurita T. Diethylstilbestrol induces vaginal adenosis by disrupting SMAD/ RUNX1-mediated cell fate decision in the Müllerian duct epithelium. Dev Biol. 2013;381:5-16. PMID: 23830984 DOI: 10.1016/j.ydbio.2013.06.024

Lee SH, Kang SM, Choi MH, Lee J, Park MJ, Kim SH, Lee WY, Hong J, Chung BC. Changes in steroid metabolism among girls with precocious puberty may not be associated with urinary levels of bisphenol A. Reprod Toxicol. 2014;44:1-6. PMID: 23557689 DOI: 10.1016/j.reprotox.2013.03.008

Liu T, Li N, Zhu J, Yu G, Guo K, Zhou L, Zheng D, Qu X, Huang J, Chen X, Wang S, Ye L. Effects of di-(2-ethylhexyl) phthalate on the hypothalamus-pituitary-ovarian axis in adult female rats. Reprod Toxicol. 2014;46:141-7. PMID: 24675100 DOI: 10.1016/j.reprotox.2014.03.006

Luderer U, Kesner JS, Fuller JM, Krieg EF Jr, Meadows JW, Tramma SL, Yang H, Baker D. Effects of gestational and lactational exposure to heptachlor epoxide on age at puberty and reproductive function in men and women. Environ Res. 2013;121:84-94. PMID: 23194642 DOI: 10.1016/j. envres.2012.11.001

Mahalingaiah S, Hart JE, Wise LA, Terry KL, Boynton-Jarrett R, Missmer SA. Prenatal diethylstilbestrol exposure and risk of uterine leiomyomata in the Nurses' Health Study II. Am J Epidemiol. 2014;179:186-91. PMID: 24142917 DOI: 10.1093/aje/kwt250

Manikkam M, Tracey R, Guerrero-Bosagna C, Skinner MK. Dioxin (TCDD) induces epigenetic transgenerational inheritance of adult onset disease and sperm epimutations. PLoS One. 2012a;7:e46249. PMID: 23049995 DOI: 10.1371/ journal.pone.0046249

Manikkam M, Guerrero-Bosagna C, Tracey R, Haque MM, Skinner MK. Transgenerational actions of environmental compounds on reproductive disease and identification of epigenetic biomarkers of ancestral exposure. PLoS One. 2012b;7:e31901. PMID: 22389676 DOI: 10.1371/journal. pone.0031901

Manikkam M, Haque MM, Guerrero-Bosagna C, Nilsson EE, Skinner MK. Pesticide metoxychlor promotes the epigenetic transgenerational inheritance of adult-onset through the female germline. PLoS One. 2014;9:e102091. PMID: 25057798 DOI: 10.1371/journal.pone.0102091

McGlynn KA, Quraishi SM, Graubard BI, Weber JP, Rubertone MV, Erickson RL. Persistent organochlorine pesticides and risk of testicular gem cells tumors. J Natl Cancer Inst. 2008;100:663-71. PMID: 18445826 DOI: 10.1093/jnci/ djn101 Menale C, Grandone A, Nicolucci C, Cirillo G, Crispi S, Di Sessa A, Marzuillo P, Rossi S, Mita DG, Perrone L, Diano N, Miraglia Del Giudice E. Bisphenol A is associated with insulin resistance and modulates adiponectin and resistin gene expression in obese children. Pediatr Obes. 2017;12:380-7. PMID: 27187765 DOI: 10.1111/ijpo.12154

Miao M, Yuan W, Yang F, Liang H, Zhou Z, Li R, Gao E, Li DK. Associations between Bisphenol A Exposure and Reproductive Hormones among Female Workers. Int J Environ Res Public Health. 2015;12:13240-50. PMID: 26506366 DOI: 10.3390/ijerph121013240

Mínguez-Alarcón L, Gaskins AJ, Chiu YH, Williams PL, Ehrlich S, Chavarro JE, Petrozza JC, Ford JB, Calafat AM, Hauser R; EARTH Study Team. Urinary bisphenol A concentrations and association with in vitro fertilization outcomes among women from a fertility clinic. Hum Reprod. 2015;30:2120-8. PMID: 26209788 DOI: 10.1093/humrep/dev183

Mlynarcíková A, Kolena J, Ficková M, Scsuková S. Alterations in steroid hormone production by porcine ovarian granulosa cells caused by bisphenol A and bisphenol A dimethacrylate. Moll Cell Endocrinol. 2005;244:57-62. PMID: 16225985 DOI: 10.1016/j.mce.2005.02.009

Mok-Lin E, Ehrlich S, Williams PL, Petrozza J, Wright DL, Calafat AM, Ye X, Hauser R. Urinary bisphenol A concentrations and ovarian response among women undergoing IVF. Int J Androl. 2010;33:385-93. PMID: 20002217 DOI: 10.1111/j.1365-2605.2009.01014.x

Nah WH, Park MJ, Gye MC. Effects of early prepubertal exposure to bisphenol A on the onset of puberty, ovarian weight and estrous cycle in female mice. Clin Exp Reprod Med. 2011;38:75-81. PMID: 22384422 DOI: 10.5653/ cerm.2011.38.2.75

Nakamura T, Miyagawa S, Katsu Y, Watanabe H, Mizutani T, Sato T, Morohashi K, Takeuchi T, Iguchi T, Ohta Y. Wnt family genes and their modulation in the ovary-independent and persistent vaginal epithelial cell proliferation and keratinization induced by neonatal diethylstilbestrol exposure in mice. Toxicology. 2012a;296:13-9. PMID: 22445810 DOI: 10.1016/j.tox.2012.02.010

Nakamura T, Miyagawa S, Katsu Y, Mizutani T, Sato T, Takeuchi T, Iguchi T, Ohta Y. p21 and Notch signalings in the persistently altered vagina induced by neonatal diethylstilbestrol exposure in mice. J Vet Med Sci. 2012b;74:1589-95. PMID: 22850433 DOI: 10.1292/jvms.12-0182

Nandi S, Gupta OS, Roy SC, Selvaraju S, Ravindra JP. Chlorpyrifos and endosulfan affect buffalo oocyte maturation, fertilization and embryo development in vitro directly and through cumulus cells. Environ Toxicol. 2011;26:57-67. PMID: 19725121 DOI: 10.1002/tox.20529

National Toxicology Program. NTP 12th Report on Carcinogens. Rep Carcinog. 2011;12:iii-499. PMID: 21822324

Nilsson E, Larsen G, Manikkam M, Guerrero-Bosagna C, Savenkova MI, Skinner MK. Environmentally induced epigenetic transgenerational inheritance of ovarian disease. PLoS One. 2012;7:e36129. PMID: 22570695 DOI: 10.1371/journal.pone.0036129 Nogueira DP, Gomes JR, Brandão JB, Souza MLA, Colacioppo S. Policloretos de bifenila: Um grave ocupacional e ambiental. Rev Bras Saude Ocup. 1987;57:32-50.

Palioura E, Diamanti-Kandarakis E. Polycystic ovary syndrome (PCOS) and endocrine-disrupting chemicals (EDCs). Rev Endocr Metab Disord. 2015;16:365-71. PMID: 26825073 DOI: 10.1007/s11154-016-9326-7

Pauwels A, Schepens PJ, D'Hooghe T, Delbeke L, Dhont M, Brouwer A, Weyler J. The risk of endometriosis and exposure to dioxins and polychlorinated bisphenyls: a case control study of infertile women. Hum Reprod. 2001;16:2050-5. PMID: 11574490 DOI: 10.1093/humrep/16.10.2050

Peres RM, Sanseverino, MTV, Schüler-Faccini L. Exposure to environmental contaminants during pregnancy and its effects in the fetal well-being: a review. Rev Hosp Clin P Alegre. 2001;21:368-78. PMID: 18308050 DOI: 10.1016/j. fertnstert.2007.12.041

Peretz J, Flaws JA. Bisphenol A down-regulates rate-limiting Cyp11a1 to acutely inhibit steroidogenesis in cultured mouse antral follicles. Toxicol Appl Pharmacol. 2013;271:249-56. PMID: 23707772 DOI: 10.1016/j. taap.2013.04.028

Petro EM, Leroy JL, Covaci A, Fransen E, De Neubourg D, Dirtu AC, De Pauw I, Bols PE. Endocrine-disrupting chemicals in human folicular fluid impair in vitro oocyte developmental competence. Hum Reprod. 2012;27:1025-33. PMID: 22267834 DOI: 10.1093/humrep/der448

Polyzos AS, Kountouras J, Deretzi G, Zavos C, Mantzoros CS. The emerging role of endocrine disruptors in pathogenesis of insulin resistance: a concept implicating nonalcoholic fatty liver disease. Curr Mol Med. 2012;12:68-82. PMID: 22082482 DOI: 10.2174/156652412798376161

Porpora MG, Medda E, Abballe A, Bolli S, De Angelis I, di Domenico A, Ferro A, Ingelido AM, Maggi A, Panici PB, De Felip E. Endometriosis and organochlorinated environmental pollutants: a case-control study on italian women of reproductive age. Environ Health Perspect. 2009;117:1070-5. PMID: 19654915 DOI: 10.1289/ehp.0800273

Quignot N, Arnaud M, Robidel F, Lecomte A, Tournier M, Cren-Olivé C, Barouki R, Lemazurier E. Characterization of endocrine-disrupting chemicals based on hormonal balance disruption in male and female adul rats. Reprod Toxicol. 2012;33:339-52. PMID: 22285353 DOI: 10.1016/j. reprotox.2012.01.004

Rier SE, Turner WE, Martin DC, Morris R, Lucier GW, Clark GC. Serum levels of TCDD and dioxine-like chemicals in Rhesus monkeys chronically exposed to dioxin: correlation of increased serum PCB levels with endometriosis. Toxicol Sci. 2001;59:147-59. PMID: 11134554 DOI: 10.1093/tox-sci/59.1.147

Rivera OE, Varayoud J, Rodríguez HA, Muñoz-de-Toro M, Luque EH. Neonatal exposure to bisphenol A or diethylstylbestrol alters the ovarian folicular dynamics in the lamb. Reprod Toxicol. 2011;32:304-12. PMID: 21722727 DOI: 10.1016/j.reprotox.2011.06.118 Rochester JR, Bolden AL. Bisphenol S and F: A Systematic Review and Comparison of the Hormonal Activity of Bisphenol A Substitutes. Environ Health Perspect. 2015;123:643-50. PMID: 25775505 DOI: 10.1289/ehp.1408989

Rodríguez HA, Santambrosio N, Santamaría CG, Muñozde-Toro M, Luque EH. Neonatal exposute to bisphenol A reduces the pool of primordial follicles in the rat ovary. Reprod Toxicol. 2010;30:550-7. PMID: 20692330 DOI: 10.1016/j.reprotox.2010.07.008

Safe SH, Zacharewski T. Organochlorine exposure and risk for breast cancer. Prog Clin Biol Res. 1997;396:133-45. PMID: 9108595

Sanseverino MTV, Spritzer DT, Schuler-Faccini L, eds. Manual de Teratogenese. Porto Alegre: Universidade Federal do Rio Grande do Sul; 2001.

Sartain CV, Hunt PA. An old culprit but a new story: Bisphenol A and "NextGen" bisphenols. Fertil Steril. 2016;106:820-6. PMID: 27504789 DOI: 10.1016/j.fertnstert.2016.07.1114

Shen Y, Xu Q, Ren M, Feng X, Cai Y, Gao Y. Measurement of phenolic environmental estrogens in women with uterine leiomyoma. PLoS One. 2013;8:e79838. PMID: 24255718 DOI: 10.1371/journal.pone.0079838

Signorile PG, Spugnini EP, Mita L, Mellone P, D'Avino A, Bianco M, Diano N, Caputo L, Rea F, Viceconte R, Portaccio M, Viggiano E, Citro G, Pierantoni R, Sica V, Vincenzi B, Mita DG, Baldi F, Baldi A. Pre-natal exposure of mice to bisphenol A elicits an endometriosis-like phenotype in female offspring. Gen Comp Endocrinol. 2010;168:318-25. PMID: 20350546 DOI: 10.1016/j.ygcen.2010.03.030

Signorile PG, Spugnini EP, Citro G, Viceconte R, Vincenzi B, Baldi F, Baldi A. Endocrine-disruptors in utero cause ovarian damages linked to endometriosis. Front Biosci (Elite Ed). 2012;4:1724-30. PMID: 22201988 DOI: 10.2741/ e493

Simsa P, Mihalyi A, Schoeters G, Koppen G, Kyama CM, Den Hond EM, Fülöp V, D'Hooghe TM. Increased exposure to dioxine-likecompounds is associated with endometriosis in a case-control study in women. Reprod Biomed Online. 2010;20:681-8. PMID: 20211585 DOI: 10.1016/j. rbmo.2010.01.018

Solomon KR, Giesy JP, LaPoint TW, Giddings JM, Richards RP. Ecological risk assessment of atrazine in North American surface waters. Environ Toxicol Chem. 2013;32:10-1. PMID: 23147529 DOI: 10.1002/etc.2050

Soto AM, Sonnenscheim C, Chung KL, Fernandez MF, Olea N, Serrano FO. The E-Screen assay as a tool to identify estrogens: an update on estrogenic environmental pollutants. Environ Health Perspect. 1995;103:113-22. PMID: 8593856 DOI: 10.1289/ehp.95103s7113

Souter I, Smith KW, Dimitriadis I, Ehrlich S, Williams PL, Calafat AM, Hauser R. The association of bisphenol A urinary concentrations with antral follicle counts and other measures of ovarian reserve in women undergoing infertility treatment. Reprod Toxicol. 2013;42:224-31. PMID: 24100206 DOI: 10.1016/j.reprotox.2013.09.008

Su PH, Huang PC, Lin CY, Ying TH, Chen JY, Wang SL. The effect of in utero exposure to dioxins and polychlorinated biphenyls on reproductive development in eight year-old children. Environ Int. 2012;39:181-7. PMID: 22208758 DOI: 10.1016/j.envint.2011.09.009

Svechnikova I, Svechnikov K, Söder O. The influence of di-(2-ethylhexyl) phtalate on steroidogenesis by the ovarian granulosa cells of immature female rats. J Endocrinol. 2007;194:603-9. PMID: 17761899 DOI: 10.1677/JOE-07-0238

Takayanagi S, Tokunaga T, Liu X, Okada H, Matsushima A, Shimohigashi Y. Endocrine disruptor bisphenol A strongly binds to human estrogen-related receptor gamma (ERRgamma) with high constitutive activity. Toxicol Lett. 2006;167:95-105. PMID: 17049190 DOI: 10.1016/j. toxlet.2006.08.012

Taketa Y, Yoshida M, Inoue K, Takahashi M, Sakamoto Y, Watanabe G, Taya K, Yamate J, Nishikawa A. Differential stimulation pathway of progesterone secretion from newly formed corpora lutea in rats treated with ethylene glycol monomethyl ether, sulpiride or antrazine. Toxicol Sci . 2011;121:267-78. PMID: 21427058 DOI: 10.1093/toxsci/ kfr062

Tinfo NS, Hotchkiss MG, Buckalew AR, Zorrilla LM, Cooper RL, Laws SC. Understanding the effects of atrazine on steroidogenesis in rat granulosa and H295R adrenal cortical carcinoma cells. Reprod Toxicol. 2011;31:184-93. PMID: 21126571 DOI: 10.1016/j.reprotox.2010.11.005

Troisi R, Hyer M, Hatch EE, Titus-Ernstoff L, Palmer JR, Strohsnitter WC, Herbst AL, Adam E, Hoover RN. Medical conditions among adults offspring prenataly exposed to diethylstilbestrol. Epidemiology. 2013;24:430-8. PMID: 23474687 DOI: 10.1097/EDE.0b013e318289bdf7

Upson K, Sathyanarayana S, De Roos AJ, Thompson ML, Scholes D, Dills R, Holt VL. Phtalates and risk of endometriosis. Environ Res. 2013;126:91-7. PMID: 23890968 DOI: 10.1016/j.envres.2013.07.003

Veiga-Lopez A, Luense LJ, Christenson LK, Padmanabhan V. Developmental programming: gestational bisphenol-A treatment alters trajectory of fetal ovarian gene expression. Endocrinology. 2013;154:1873-84. PMID: 23525218 DOI: 10.1210/en.2012-2129

Vélez MP, Arbuckle TE, Fraser WD. Female exposure to phenols and phthalates and time to pregnancy: the Maternal-Infant Research on Environmental Chemicals (MIREC) Study. Fertil Steril. 2015;103:1011-20.e2. PMID: 25681860 DOI: 10.1016/j.fertnstert.2015.01.005

Watanabe M, Ohno S, Nakajin S. Effects of Bisphenol A on the expression of cytochrome P450 aromatase (CYP19) in human fetal osteoblastic and granulosa cell-like lines. Toxicol Lett. 2012;210:95-9. PMID: 22327052 DOI: 10.1016/j. toxlet.2012.01.020

Weuve J, Hauser R, Calafat AM, Missmer AS, Wise LA. Association of exposure to phthalates with endometriosis and uterine leiomyomata: findings from NHANES, 1999-2004. Environ Health Perspect. 2010;118:825-32. PMID: 20185384 DOI: 10.1289/ehp.0901543 Xi W, Lee CK, Yeung WS, Giesy JP, Wong MH, Zhang X, Hecker M, Wong CK. Effect of perinatal and postnatal bisphenol A exposure to the regulation circuits at the hypothalamus-pituitary-gonadal axis of CD-1 mice. Reprod Toxicol. 2011;31:409-17. PMID: 21182934 DOI: 10.1016/j. reprotox.2010.12.002

Xu C, Chen JA, Qiu Z, Zhao Q, Luo J, Yang L, Zeng H, Huang Y, Zhang L, Cao J, Shu W. Ovotoxicity and PPAR-mediated aromatase downregulation in female Sprague-Dawley rats following combined oral exposure to benzo[a]pyrene and di-(2-ethylhexyl) phthalate. Toxicol Lett. 2010;199:323-32. PMID: 20920559 DOI: 10.1016/j.toxlet.2010.09.015

Xu N, Kwon S, Abbott DH, Geller DH, Dumesic DA, Azziz R, Guo X, Goodarzi MO. Epigenetic mechanism underlying the development of polycystic ovary syndrome (PCOS)-like phenotypes in prenatally androgenized rhesus monkeys. PLoS One. 2011;6:e27286. PMID: 22076147 DOI: 10.1371/journal.pone.0027286 Yoshida M, Takahashi M, Inoue K, Hayashi S, Maekawa A, Nishikawa A. Delayed adverse effects of neonatal exposure to diethylstilbestrol and their dose dependency in female rats. Toxicol Pathol. 2011;39:823-34. PMID: 21747122 DOI: 10.1177/0192623311413785

Zhou F, Zhang L, Liu A, Shen Y, Yuan J, Yu X, Feng X, Xu Q, Cheng C. Measurement of phenolic environmental estrogens in human urine samples by HPLC-MS/MS and primary discussion the possible link-age with uterine leiomyoma. J Chromatogr B Analyt Technol Biomed Life Sci. 2013;938:80-5. PMID: 24060595 DOI: 10.1016/j. jchromb.2013.08.032

Zota AR, Park JS, Wang Y, Petreas M, Zoeller RT, Woodruff TJ. Polibrominated diphenyl ethers, hidroxilated polibrominated diphenyl ethers and measures of thyroid function in second trimester pregnant women in California. Environ Sci Technol. 2011;45:7896-905. PMID: 21830753 DOI: 10.1021/es200422b