



Genome Sequence of *Burkholderia plantarii* ZJ171, a Tropolone-Producing Bacterial Pathogen Responsible for Rice Seedling Blight

Yuan Qian,ª Haruna Matsumoto,ª Wenzhuo Li,ª Guonian Zhu,ª Yasuyuki Hashidoko,^b Yang Hu,^c Mengcen Wang^a

Institute of Pesticide and Environmental Toxicology, Zhejiang University, Hangzhou, China^a; Research Faculty of Agriculture, Hokkaido University, Sapporo, Japan^b; Department of Forest Mycology and Plant Pathology, Swedish University of Agricultural Science, Uppsala, Sweden^c

Burkholderia plantarii is the causal agent of rice seedling blight. Here, we report the draft genome sequence of *B. plantarii*, which contains 8,020,831 bp, with a G+C content of 68.66% and a predicted 7,688 coding sequences. The annotated genome sequence will provide further insight into its pathogenicity.

Received 29 September 2016 Accepted 14 October 2016 Published 8 December 2016

Citation Qian Y, Matsumoto H, Li W, Zhu G, Hashidoko Y, Hu Y, Wang M. 2016. Genome sequence of *Burkholderia plantarii* ZJ171, a tropolone-producing bacterial pathogen responsible for rice seedling blight. Genome Announc 4(6):e01318-16. doi:10.1128/genomeA.01318-16.

Copyright © 2016 Qian et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license

Address correspondence to Yang Hu, yang.hu@slu.se, or Mengcen Wang, wmctz@zju.edu.cn.

B*witholderia plantarii* is a rice-pathogenic bacterium causing seedling blight by the production of tropolone, a phytotoxin, as the virulence factor (1, 2). Rice seedlings showed typical symptoms, such as chlorosis, stunting, and root growth inhibition (3), when infested by *B. plantarii* or treated with tropolone only (4, 5). This disease was first reported in a rice nursery in Chiba Prefecture, Japan, in 1985 and was found to be widespread in other major rice-growing regions of Asia, including China (6). Particularly during recent years, the greenhouse has been adopted as an important measure for transplantation and production of rice seedlings in Asia, which fostered outbreaks of seedling blight and caused severe damages to rice cultivation (6). In this report, the genome sequence of the *B. plantarii* strain ZJ171 was determined, assembled, and annotated.

B. plantarii ZJ171 (16S rRNA gene under accession no. LC020026) was isolated from a rice paddy in Zhejiang Province, China (29°48.467'N, 120°17.867'E). The whole genome of *B. plantarii* ZJ171 was sequenced by Illumina HiSeq 2500 pairedend sequencing technology at Biomarker (Beijing, China). The reads were assembled using Velvet (7), and gene annotation was carried out using GLIMMER for contigs equal or longer than 1,000 bp (8). The assembled genome consisted of 150 contigs, 8,020,831 bp (G+C content of 68.66%), and an N_{50} of 98,282 bp. It comprised 7,688 coding sequences of genes and 58 predicted RNA genes (46 tRNA genes, 1 rRNA gene, and 11 microRNA genes).

Analysis of the genome of *B. plantarii* ZJ171 revealed critical genes involved in biofilm formation, tropolone production, and the regulatory, biosynthetic, and secretory systems related to tropolone production. Also, quorum sensing (QS) is known to regulate *B. plantarii* virulence (2). Previously, a pair of *N*-acyl homoserine lactone (AHL)-QS genes, *pla-plaR*, were found to be involved in activation of tropolone production (2). In the assembled genome of *B. plantarii* ZJ171, 2 additional paired AHL-QS genes and 21 orphan AHLs regulator-coding genes without the AHL synthase-coding genes (LuxR family transcriptional regulators) were found, indicating the more sophisticated regulatory

patterns for the diverse responses to interspecies communication with the host plant and various competitors. Interestingly, it was first noticed that the gene coding for enoyl coenzyme A (CoA) hydratase (*rpfF*) was existed in *B. plantarii* ZJ171, which functions as the key component of the BDSF (*cis*-2-dodecenoic acid) signaling pathway controlling *Burkholderia cepacia* complex (Bcc) virulence (9, 10), suggesting the possible involvement of the *Burkholderia* diffusible signal factor (BDSF) pathway in the regulation of *B. plantarii* virulence. Similar to *Burkholderia glumae*, genes coding for virulence-related enzymes, such as lipase LipA (11) and LysR-type transcriptional activator (12), have also been detected in *B. plantarii* ZJ171.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession no. MKGK00000000. The version described in this paper is version MKGK01000000. The BioProject designation for this project is PRJNA323430. The BioSample accession no. is SAMN05178893.

ACKNOWLEDGMENTS

This work was supported by the National Natural Science Foundation of China (grant no. 31501684) and by the Zhejiang Provincial Natural Science Foundation of China (grant no. LQ16C14001).

FUNDING INFORMATION

This work, including the efforts of Mengcen Wang, was funded by National Natural Science Foundation of China (NSFC) (31501684). This work, including the efforts of Mengcen Wang, was funded by Zhejiang Provincial Natural Science Foundation of China (LQ16C14001).

REFERENCES

- Azegami K, Nishiyama K, Watanabe Y, Suzuki T, Yoshida M, Nose K, Toda S. 1985. Tropolone as a root growth-inhibitor produced by a plant pathogenic *Pseudomonas* sp. causing seedling blight of rice. Ann Phytopathol Soc Japn 51:315–317.
- 2. Wang M, Hashimoto M, Hashidoko Y. 2013. Repression of tropolone production and induction of a *Burkholderia plantarii* pseudo-biofilm by carot-4-en-9,10-diol, a cell-to-cell signaling disrupter produced by

Trichoderma virens. PLoS One 8:e78024. http://dx.doi.org/10.1371/journal.pone.0078024.

- 3. Azegami K, Nishiyama K, Kato H. 1988. Effect of iron limitation on *"Pseudomonas plantarii"* growth and tropolone and protein production. Appl Environ Microbiol 54:844–847.
- Wang M, Hashimoto M, Hashidoko Y. 2013. Carot-4-en-9,10-diol, a conidiation-inducing sesquiterpene diol produced by *Trichoderma virens* PS1-7 upon exposure to chemical stress from highly active iron chelators. Appl Environ Microbiol 79:1906–1914. http://dx.doi.org/10.1128/AEM.03531-12.
- Wang M, Tachibana S, Murai Y, Li L, Lau SY, Cao M, Zhu G, Hashimoto M, Hashidoko Y. 2016. Indole-3-acetic acid produced by *Burkholderia heleia* acts as a phenylacetic acid antagonist to disrupt tropolone biosynthesis in *Burkholderia plantarii*. Sci Rep 6:22596. http:// dx.doi.org/10.1038/srep22596.
- 6. Wang M, Wei P, Cao M, Zhu L, Lu Y. 2015. First report of rice seedling blight caused by *Burkholderia plantarii* in North and Southeast China. Plant Dis 100:645.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. http:// dx.doi.org/10.1101/gr.074492.107.

- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. http://dx.doi.org/10.1093/bioinformatics/btm009.
- Ryan RP, An SQ, Allan JH, McCarthy Y, Dow JM. 2015. The DSF family of cell-cell signals: an expanding class of bacterial virulence regulators. PLoS Pathog 11:e1004986. http://dx.doi.org/10.1371/journal.ppat.1004986.
- Schmid N, Pessi G, Deng Y, Aguilar C, Carlier AL, Grunau A, Omasits U, Zhang L-H, Ahrens CH, Eberl L. 2012. The AHL- and BDSFdependent quorum sensing systems control specific and overlapping sets of genes in *Burkholderia cenocepacia* H111. PLoS One 7:e49966. http:// dx.doi.org/10.1371/journal.pone.0049966.
- Ham JH, Melanson RA, Rush MC. 2011. Burkholderia glumae: next major pathogen of rice? Mol Plant Pathol 12:329–339. http://dx.doi.org/ 10.1111/j.1364-3703.2010.00676.x.
- Kim J, Kim JG, Kang Y, Jang JY, Jog GJ, Lim JY, Kim S, Suga H, Nagamatsu T, Hwang I. 2004. Quorum sensing and the LysR-type transcriptional activator ToxR regulate toxoflavin biosynthesis and transport in *Burkholderia glumae*. Mol Microbiol 54:921–934. http://dx.doi.org/ 10.1111/j.1365-2958.2004.04338.x.