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ORIGINAL RESEARCH ARTICLE

Ecological approaches to informing public health policy and risk assessments on emerging vector-borne zoonoses

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Received 25 March 2009 Revised 27 November 2009 Accepted 1 December 2009 Pathogens associated with vector-borne zoonoses occur in enzootic cycles within nature. They are driven by a combination of vertebrate host and invertebrate vector population dynamics, which in turn respond to changes in environmental stimuli. Human involvement in these cycles, and hence the occurrence of human disease, is often to act as incidental host. From a public health perspective our ability to better predict human outbreaks of these diseases and prepare intervention and mitigation strategies relies on understanding the natural cycle of pathogen transmission. This requires consideration of, for example, invertebrate and vertebrate ecology and biology, climatology, land use and habitat change. Collectively, these can be referred to as medical entomology and medical ecology. This article reviews the importance for inclusion of such disciplines when assessing the public health risk from vector-borne zoonoses and summarizes the possible future challenges and driving forces for changes in vector status and vector-borne zoonoses emergence, with a particular focus on a UK and European context.

Introduction

UK public health risk assessment process

When preparing for and responding to emerging diseases, government officials and scientists are expected to make rapid assessments of risk and inform policy decisions through expert steering groups. Infections such as influenza, SARS, chikungunya and West Nile virus (WNV) have tested our abilities to predict epidemics and provide timely, well-informed advice to policy decision makers.

Since 2003, the Human Animal Infections Risk Surveillance (HAIRS) group, coordinated by the Health Protection Agency (HPA), has provided expert advice on emerging infections to the UK Chief Medical Officer's National Expert Panel on New and Emerging Infections. This group draws in experts largely from the medical and veterinary sectors across the UK¹ and assesses the risks posed to UK public health by potential emerging infectious diseases threats using a published methodology.² The Medical Entomology and Zoonoses Ecology (MEZE) group at HPA are members of HAIRS, providing input to discussions primarily on vector-borne zoonoses (VBZ). When considering the possible risks of infections that essentially occur within nature (whether it be wildlife, or an invertebrate vector), various considerations

are required that may be crucial to the final assessment of public health risk.

Role of medical ecology and medical entomology in informing risk assessments and policy

A large proportion of emerging human pathogens have a zoonotic source and many of these have wildlife origins or are transmitted via arthropod vectors.^{3–5} Given the prospects of climate change, coupled with large-scale ecological and agricultural change, the dynamics of these pathogens will ebb and flow and we are already beginning to see signs of this in Europe.^{6–14} With increasing global travel and trade, the geographic distributions of key vectors and their pathogens are equally dynamic.^{15–19}

Such pathogens occur in enzootic cycles within nature, with humans often acting as incidental hosts. To fully understand and to better predict the dynamics of associated human disease outbreaks, we need to have an intimate knowledge of their ecology and transmission cycle. Without such an understanding, the ability to prepare for, react to, and mitigate the effect of these pathogens is severely hindered. An ecological perspective to the risk assessment process can quickly shed light on what appears initially as an

unpredictable complex disease dynamic. In contrast, it can also complicate a seemingly simple disease formula. It may also provide greater opportunities to disrupt the transmission cycle at its weakest point, which in turn can reduce or eliminate the public health burden.

This review does not aim to cover all components of disease emergence such as clinical, virological, bacteriological or immunological changes; although the authors agree such drivers have equal importance. This article will purely review where an ecological/entomological input has been required by MEZE in assessing VBZ risk and, using examples elsewhere in Europe, provide an overview of other ecologically driven disease transmission scenarios that have relevance to informing policy decisions and risk assessment on VBZ; concluding with some discussion of what challenges might arise in the UK over the next 50 years.

No ethical approval for this study was required; an in-depth review of the literature was performed using all relevant VBZ keywords in PubMed. Keywords included: vector-borne, arboviruses, zoonoses, wildlife zoonoses, mosquito, tick, *Phlebotomus*, Europe, Hantavirus, *Dirofilaria*, West Nile, Sindbis, Tahyna, Tick-borne encephalitis, Crimean-Congo haemorrhagic fever, chikungunya, *Aedes albopictus*, invasive mosquitoes, *Ixodes, Dermacentor, Hyalomma, Echinococcus multilocularis, Aedes vexans, Culex pipiens, Rhipicephalus sanguineus*, malaria, disease ecology, mosquitoes and wetlands.

Vector-borne diseases in twenty-first century Europe

Since the turn of the century, Europe has witnessed unexpected outbreaks of bluetongue virus (BTV) in northern countries, 20 the continued spread and establishment of the exotic mosquito A. albopictus to most of the Mediterranean countries,²¹ and the emergence of chikungunya virus (CHIKV) in Italy.^{22,23} Crimean-Congo haemorrhagic fever virus (CCHFV) and its Hyalomma tick vector have also emerged in parts of Turkey, and for the first time caused clinical disease in humans in both Turkey and Greece.^{24,25} New foci of Tick-borne encephalitis virus (TBEV) have been reported further north in Scandinavia.²⁶ Usutu virus (USUV) has emerged in Austria, 27 Hungary 28 and Spain, 29 and outbreaks of WNV continue to appear in France, 30 Hungary,³¹ and Romania,³² and emerge in Italy.³³ One of our commonest vector-borne diseases, Lyme borreliosis, has also seen an increase in incidence rates between 2001 and 2005 in many European countries, with some rates increasing two- or threefold.³⁴

The arrival and expansion of BTV, CHIKV and USUV raises questions as to which pathogens and/or vectors will next appear in Europe. Will we see the movement of Rift Valley fever virus (RVFV) from Africa and the Arabian Peninsula? Will CCHFV expand to match the distribution of its vector? Will Aedes aegypti, the principal vector of dengue (DENV), spread from Madeira to mainland Portugal and beyond? Looking further ahead perhaps, will we see the re-emergence

of malaria and DENV to previously endemic zones in the Mediterranean basin (for example malaria occurred in Italy and large outbreaks of DENV occurred in Greece, although the latter will be dependant on the spread of invasive *Aedes*)? The latter few might seem fanciful, but we should not be complacent. If we learn any lesson from the past, it is time to expect, and prepare, for the unexpected.

Developing a baseline distribution of invertebrate vectors

A crucial component to any risk assessment on VBZ is knowledge of the distribution and abundance of potential vector species. Without this baseline data it is impossible to assess whether a 'new finding' of an unusual or exotic species is significant. It is not generally considered to be costeffective to have long-term nationwide surveillance programmes for all potential vectors, particularly when they do not currently transmit disease. Occasionally, there are funded programmes that monitor species abundance at a number of key sites across a country and these data are important; however it tells us very little about the nationwide distribution of vector species. It is also of limited use in the development of predictive distribution maps, unless the survey locations are both numerous and reflect the sufficient diversity of ecozones. Recently, in response to the outbreak of CHIKV in Italy, ECDC funded a project to map the European distribution of A. albopictus. In less than 3 months, and following concerted European-wide collaboration between entomologists, a map now exists for this exotic mosquito species across more than 20 countries. This has facilitated predictive mapping of their current distribution and the potential consequences of climate change.35

Without substantive and ongoing funded programmes to map vector species, the sustainable and affordable option is to capitalize on and engage the volunteer recording network. In the UK, naturalists and scientists have been recording species occurrence since the eighteenth century, with 15 million records of 12,000 species currently held by the Biological Records Centre.³⁶ Of course much of the data relate to non-vector species of biodiversity interest, but it illustrates the potential of thousands of invertebrate recorders willing to submit data. In 2005, MEZE established a tick recording scheme,³⁷ and has now received 1000 records and >4000 ticks from across the UK.³⁸ Added to 4500 historical records, this provides our best data set for ticks, and is now being used to facilitate geo-spatial mapping. A similar initiative for mosquitoes has also been running since 2005.³⁹ Although this kind of data are often regarded as 'ad hoc' and unsystematic, they inform public and veterinary health authorities where nuisance-biting vectors occur, provide an early warning system for exotic or expanding species and provide a bank of samples for pathogen analysis. Furthermore, it is very cost-effective and requires the time of the scheme coordinators to perform and validate identifications.

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Understanding vector behaviour and classification

For non-medical entomologists, a list of potential vector species and their distribution can be misleading without specific knowledge on their biology, ecology, classification and vector competence. Recent examples of this relate to potential mosquito vectors of WNV and RVFV. During the outbreak of WNV in North America, C. pipiens was quickly implicated as a key vector in the eastern states. 40 In the UK it exists as two separate forms, an exclusive ornithophagic pipiens form, and predominantly mammalophilic (also partly ornithophilic) molestus form. 41 These two forms adopt hugely different behaviour and aquatic habitats and do not, as far as we are aware, hybridize. In the USA, however they do and some studies have found that 40% of C. pipiens are hybrids and consequently feed on both birds and humans, thus driving the transmission cycle. 42 In the UK then, although our pipiens form might be an important enzootic vector, it is unlikely to be a human vector despite its synanthropic container breeding habits and its propensity for over-wintering in the cool recesses of houses. 43 The status of the British forms of this mosquito is still a topic of much debate in DNA barcoding circles.

Similarly, one of the primary vectors of RVFV is *A. vexans*, which also exists as a species complex including subspecies *vexans* and *arabiensis*; for example, *A. vexans arabiensis* is an important vector in Senegal and Saudi Arabia. *A. vexans* occurs also in parts of western Europe^{44,45} and in a recent study involving MEZE in Epping Forest was found to cause nuisance biting close to the London Olympics village site.⁴⁶ Should this be a concern, and should we assess its taxonomy and vector competence?

A number of studies, including those by MEZE on WNV and SINV, have taken an entomological perspective to risk assessment, by reviewing the status, behaviour and distribution of putative vector species. 43,47–49 This rapidly identifies which species could function as vectors, their likely aquatic breeding sites (and hence synanthropic/sylvatic issues) and, if known, their biting behaviour. Often important data are lacking, particularly information on the host preferences (for example bird- or horse biting) of certain vector species, which can be significant in understanding transmission cycles and disease risk. Such reviews also help to target research gaps for field entomology and ecology and to identify key species for vector competence studies and viral isolations.

Vertebrate host dynamics and vector-borne infections

Knowledge of the involvement of vertebrate species as reservoirs or amplifying species can rapidly affect the level of risk attributed to an emerging disease. Even with an abundant vector population, the absence of key vertebrate hosts for all or part of the year is significant. Studies from Sweden and Finland on Sindbis virus (SINV; the cause of Ockelbo and Pogosta diseases in humans) illustrate this point very well. 50,51 SINV exists within a bird–mosquito

cycle. In Sweden, it cycles between passerines and *Culex* mosquitoes, with *Aedes* providing the link to humans, ^{52–54} and in Finland vertebrate hosts are tetraonid birds. ⁵⁵ Following reports of antibodies to this virus in the UK⁵⁶ where all key putative mosquito vectors occur, ⁴³ concerns were raised over possible endemic transmission, and an ecological assessment was performed by MEZE. ⁴⁸

This work concluded that particular tetraonid bird species (Tetrao urogallus, Tetrao tetrao) that amplify the virus in Finland⁵⁴ also occur in Britain, but only in a few Scottish forest and moorland sites. Both species are rare and consequently unlikely to be important in amplifying the virus or contributing significantly to virus cycles in birds. Given the ubiquity of passerines in the UK however, and their general role as reservoirs in Sweden, it is suggested that perhaps a lack of reservoir hosts might not be a limiting factor in transmission. However, Swedish studies found that human outbreaks of Ockelbo disease appeared to be linked to the occurrence of certain migratory thrush species. 55,57 Redwing (Turdus iliacus), fieldfare (T. pilaris) and song thrush (T. philomelos) were found to have high prevalence rates for virus and were implicated as key amplifying hosts. These species breed in Sweden in the summer and have large numbers of fledgelings during the main peak in mosquito activity. In autumn, redwing and fieldfare fly south to spend the winter in Britain, usually after the peak in mosquitoes, and hence are unlikely to contribute significantly to virus amplification. An understanding of the role of different hosts and bird life histories can significantly change our perception of disease risk.

Impact of weather and climate on vector, host and pathogen dynamics

Temperature-dependent distribution and seasonal activity of vectors

Many aspects of ecological systems are affected by climatic changes, particularly the effect of temperature. The expected change in summer and winter temperature as a consequence of climate change will change vector behaviour and seasonality, and often this requires a longer-term assessment of risk. For example, phlebotomine sandflies are acutely responsive to temperature changes and new reports of their survival further north (that is, north of the Mediterranean) have been reported in Germany.⁵⁸ Phlebotomus sandflies are vectors of canine and human leishmaniasis, and there is a genuine concern that travelling dogs that are returning to the UK with Leishmania parasites could create a reservoir of infection, which in the event of sandflies establishing could lead to local cycles.⁵⁹ In the absence of current surveillance for sandflies, MEZE are working with expert dipterists to develop an early warning system.

Mosquitoes use temperature as a seasonal cue, and along with photoperiodic cues, these have been used to model their seasonal activity. For temperate strains of exotic species, like *A. albopictus*, models, developed by MEZE and others, have assessed areas for establishment and seasonal

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activity in the event of an importation,^{60,61} thus providing an insight into possible areas for nuisance biting and to target surveillance.

Changes in climate also affect the seasonality and/or distribution of ticks, with evidence of a gradual northerly spread of I. ricinus in response to consecutive mild winters in Scandinavia. For R. sanguineus, it has even been suggested that host preference is altered in response to temperature, with the affinity of immature R. sanguineus for humans apparently increased during short periods of warmer weather (>25°C) in France. Given the recent discussion over permitting dogs with tick treatment to travel to the UK, the influence of climate is significant when advising on the potential for establishment of R. sanguineus in the UK.

Effects of temperature on extrinsic incubation of pathogens

Many arthropod-borne pathogens undergo a period of development through the vector, known as the extrinsic incubation period. This period, particularly for parasites like Plasmodium and Dirofilaria, is temperature dependent, with parasite development enhanced above a set temperature threshold. Laboratory and field studies on these two parasites in the USA on Dirofilaria^{65–68} and in Russia on Plasmodium⁶⁹ have aided MEZE and others in our predictions of the likely period of incubation of their larval stages through a hypothetical mosquito.^{70–72} This enables us to predict currently (that is based on current weather patterns) how many days the development of the parasite through to the infective third larval stage might be permissible. This information is making us re-assess our opinions on whether northern Europe is too cold to prevent or limit transmission of seemingly tropical or Mediterranean pathogens. However, other factors, such as availability of infected hosts, abundance of receptive vectors and the impact of treatments, need to be considered.

Effects of sequential climate events on reservoir host dynamics

Studies on the impact of weather patterns on non-arthropod wildlife zoonotic infections, for example, those caused by Puumala virus, have shown that interpreting sequential climate events can allow prediction to be made about changes in the abundance of reservoir hosts (that is, Myodes glareolus voles)73 and outbreaks of human disease (for example Nephropathia epidemica). 74,75 The essence of this approach taken by MEZE is to predict the peak years in tree nut production that drive the peaks of vole cycles. In the case of beech (Fagus sylvatica), a sequence of climate events over 2-3 years stimulates a synchronous increase in seed/nut production; known as a 'masting event'. The vole population consequently peaks the following year and stimulates an increased circulation of virus, which leads to outbreaks of human disease. Equally this has implications for other pathogens like B. burgdorferi that circulates in rodent populations.

Societal change masking climate change

These relatively simple uses of weather and climate data are providing important insights into the effects of climate change and possible changes in disease risk. However, climate should not be taken in isolation. Recent studies on TBEV in eastern Europe have shown that climate changes may be masking underlying societal changes, suggesting that economic- and climate-influenced factors are combining to impact on the transmission of TBEV. For example, in the Baltic states there has been an increase in outdoor activities as a consequence of increased wealth since the decline of Communism; a decline in industry has changed the amount of local solar radiation, adapting the local climate, thus favouring co-incident seasonality of tick stages, thus increasing co-feeding transmission, and hence increased exposure to virus. ^{76–79} Indeed, the concept of co-feeding of immature I. ricinus in determining TBEV endemic foci in central Europe was one of the first studies to show the complexities of tick seasonality and behaviour of disease transmission cycles.^{80,81}

Human impacts: trade and travel

The impact of global trade and human travel on vector-borne disease is well established. The best examples are the worldwide spread of exotic mosquitoes on used tyres (for example *A. albopictus, Ochlerotatus japonicus, A. aegypti*), and the importation of CHIKV by a human traveller from India to Italy.⁸² Movement of animals and potentially infected or infested animals often goes unnoticed and can have significant implications. There is still much debate about causes of the emergence of WNV in North America with introduction of the virus through imported infected birds being one theory.^{83,84} The involvement of livestock movement in pathogen spread was suggested as one of the factors involved in the emergence of RVFV in Saudi Arabia; imported from Africa.⁸⁵

A recent example studied by MEZE has identified a different implication to the movement of wildlife zoonoses through pet travel. There is currently much debate in the UK over the relaxation of tapeworm control in dogs returning to the UK from continental Europe. 86–88 Current controls are in place to maintain the UK free of E. multilocularis, 89 which is an emerging urban zoonosis in continental Europe⁹⁰ exacerbated by the urban fox population.⁹¹ This parasite occurs within enzootic cycles between voles (including the water vole, Arvicola terrestris) and the fox (Vulpes vulpes). A recent assessment by MEZE⁹² has highlighted that in addition to the public health issues, the importation of this parasite by dogs into the UK could undermine current UK wildlife policy⁹³ and legislation⁹⁴ that promote the protection of water voles and enhance both their habitats and the grassland habitats of another potential reservoir host, the field vole (Microtus agrestis). Coupled with an established urban fox population there is the prospect that an inadvertent importation of this parasite on health risk assessment grounds could conflict with environmental impact assessment, thus undermining green initiatives and establishing

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enzootic cycles that would be difficult to eradicate. MEZE findings⁹² also question whether such an act could be at odds with EU legislation^{95,96} on non-native species and their effect on native fauna.

Multiple drivers for emergence

Although not discussed in depth here, it is now recognized that viral mutation and a change in vector specificity were important drivers in the large-scale epidemics of CHIKV in the Indian Ocean region.⁹⁷ The subsequent outbreak of CHIKV in Italy further showed how a combination of factors such as global trade in tyres and global travel of infected individuals can act in synergy to drive the emergence of an exotic virus vectored by an exotic mosquito.

Current unpublished MEZE work in Kosova is showing that multiple drivers may be driving cycles of CCHFV and *Hyalomma* ticks. It appears that a combination of the decline in agriculture (hence the encroachment of scrub and grassland close to the village fringe), the dramatic increase in reservoir host numbers as a possible consequence of predator decline, coupled with the importation of virus-naive livestock (following the recent conflict), could all be contributory factors to increasing the abundance and public exposure of *Hyalomma* and enhancing the circulation of CCHFV in ticks in parts of rural Kosova.

Possible driving forces for future change in VBZ in the UK

A key component of 'horizon scanning' is looking for future issues. Arthropod disease vectors such as mosquitoes and ticks are, like all invertebrates, acutely responsive to changes in their environment, with an ability to evolve to local climate rapidly. There has been much discussion over the role of weather and climate on infectious diseases, with some models able to predict future epidemics and new foci. Notable examples for RVFV, 98 meningitis, 99 malaria 100 and TBEV 101 have included environmental parameters such as rainfall, wind direction, temperature and seasonal climate. Much less discussion has focussed on non-climate-related environmental changes, or environmental changes as a consequence of adaptation to climate change. Some examples from a UK context are briefly discussed here.

Biodiversity-enriching strategies are likely to have some impact on vector survival, activity and spread. Recent UK biodiversity initiatives promote habitat connectivity by encouraging wildlife corridors on a national 102,103 and a local 104 scale. These strategies aim to de-fragment habitats, allowing more vulnerable species of nature conservation concern to move between habitats and hence respond to climatic changes. Often this landscape ecology approach encourages the parcelling of the landscape into habitat patches that form mosaics of biotypes linked by an ecological matrix connecting the fragments of species-rich habitats.

It is not known what effect this will have on disease vectors. Undoubtedly it provides a more robust network for movement of vector hosts (for example, deer, small mammals) and an increase of ecotonal habitat (that is, habitat at the interface of two habitats, for example grassland and woodland) so often favoured by *I. ricinus* ticks and their hosts (for example pheasant). Some authors argue that habitat de-fragmentation can increase the transmission of zoonotic and anthropozoonotic diseases by bringing humans, livestock, wildlife and their pathogens ever closer together. MEZE studies have shown that the spatial heterogeneity of ticks within a varied landscape is predictable, and more recent MEZE work, looking at the impact of woodland ride management on *I. ricinus*, has found that hot spots of questing tick activity are predictable within lowland woodlands, and that ride and bracken management can be planned to mitigate the public's exposure to ticks and limit the tick's survival in the mat layer.

In the UK, part of enhancing habitat connectivity has involved paying environmental subsides to encourage farmers to create field margins and re-instate hedgerows, ponds and ditches. 110 Furthermore, on the fragments of relict calcareous grassland, which remain among the farmland, traditional grazing strategies are being re-established to ensure that a diverse sward creates a diverse flora supporting specialist invertebrates. There is little known about the impact of these strategies on tick survival and movement. Ongoing MEZE work suggests that field margins can act as *I. ricinus* habitats, and there are suggestions (anecdotal and published) that the movement of livestock can lead to the establishment of ticks in new geographical foci. 111

Allied to terrestrial UK initiatives are nationwide plans to return large parts of the UK's drained agricultural land to wetland, fen and salt marsh (for example UK Wetland Vision). 112 This is likely to have huge biodiversity benefits by making the wildlife of existing wetlands more sustainable, increasing the general available resource for wildlife as well as providing better management of flood waters in low lying areas, river valleys and flood alleviation schemes in urban areas. It would seem prudent therefore that habitat creation plans include a health risk assessment component to the management plan, focusing on the possible effects on mosquito habitats and nuisance biting/disease transmission. MEZE are currently working on such an approach with the Great Fen Project; an environmental project that plans to restore >3000 hectares of arable farmland in Cambridgeshire back to fen, wetland and wet woodland. Some question whether new wetland means more mosquitoes, 113 others argue that focus should be given to managing habitats for the vectors' natural predators as they may have some role in keeping vector populations at manageable levels. 114 Wetland initiatives along the coast have other motives, such as creating salt marshes to help defend an eroding coastline (for example, in eastern England) or by inadvertently adapting coastal habitats to harness the power of tides, such as the Severn Tidal Barrage. 115 Undoubtedly, the creation of new salt marshes could provide new habitats for our brackish water Anopheles sp., which lost their habitats, and the malaria they transmitted, following wide-scale drainage in

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the nineteenth century. $^{116-119}$ However, the arguments here for the return of malaria are more complex and have been discussed elsewhere. 120

In urban areas, changes are also occurring, whether it is an extension of urban areas into the peri-urban fringe, the rewilding and greening of urban habitats, restoration of industrialized river corridors or wildlife gardening. MEZE have received several reports to its tick recording scheme regarding high levels of tick activity in residential gardens, mostly attributable to the encouragement of wildlife to feed and shelter close to human habitation.³⁸ Often we cannot pick and choose the wildlife we attract. Although we may wish to see feeding birds and browsing deer, we also have to accept that we may also attract rats and ticks.

Furthermore, given the recent spread of the container habitat mosquito *A. albopictus* in Europe, and its involvement in CHIKV cycles, ^{35,121} there is good reason to be cautious about the management of water in domestic settings. Currently water butts provide a suitable habitat for mosquito species such as *C. pipiens* and *Culiseta annulata*, which are not currently involved as disease vectors in the UK. Following the introduction of an exotic vector species that can exploit these habitats, a re-assessment of our domestic husbandry regarding conserving water will be required, as occurs in Italy. ¹²²

In conclusion, possible environmental change may have significant impacts on the future changes in VBZ. It is paramount therefore that medical entomologists and epidemiologists work with ecologists, environmental scientists and land managers to pre-empt, prepare for and mitigate the possible changes in VBZ through environmental management. Achieving this preparedness and establishing routes of communication take time, but it should not be left to hastily prepared responses to nuisance biting or outbreaks of disease. A balanced evidence-based opinion is required whereby public health matters are considered, but not in an alarmist way that undermines biodiversity and water management objectives. We have the opportunity as public health practitioners to work with the environment sector to ensure that an increase in biodiversity does not lead to emergence or re-emergence of VBZ.

Disclaimer

The opinions expressed in this article are those of the authors and not necessarily those of the Health Protection Agency.

References

- 1 Health Protection Agency [HPA]. *The Human Animal Infections and Risk Surveillance (HAIRS) Group: First Report 2004–2007* 2008; 1–30, www.hpa.org.uk.
- 2 Walsh AL, Morgan D. Identifying hazards, assessing the risks. Vet Rec 2005;157:684–7.
- 3 Taylor LH, Latham SM, Woolhouse MEJ. Risk factors for human disease emergence. *Philos Trans R Soc London B Biol Sci* 2001;356:983–9.

- 4 Simpson VR. Wild animals as reservoirs of infectious diseases in the UK. *Vet I* 2002:163:128–46.
- 5 Kruse H, Kirkemo AM, Handeland K. Wildlife as a source of zoonotic infections. *Emerg Infect Dis* 2004;**10**:2067–72.
- 6 Mattingly PF. The Biology of Mosquito-Borne Disease. The Science of Biology Series No 1. George Allen and Unwin: London, 1969.
- 7 Kovats RS, Campbell-Lendrum DH, McMichael AJ, Woodward A, Cox JH. Early effects of climate changes: do they include changes in vector-borne disease? *Philos Trans R Soc London B Biol Sci* 2001;356:1057–68.
- 8 Parola P, Raoult D. Tick-borne bacterial diseases emerging in Europe. *Clin Microbiol Infect* 2001;7:80–3.
- 9 Greger M. The human/animal interface: emergence and resurgence of zoonotic infectious diseases. Crit Rev Microbiol 2007; 33:243–99.
- 10 Pugliese A, Beltramo T, Torre D. Emerging and re-emerging viral infections in Europe. *Cell Biochem Funct* 2007;**25**:1–13.
- 11 Vorou RM, Papavassiliou VG, Tsiodras S. Emerging zoonoses and vector-borne infections affecting humans in Europe. *Epidemiol Infect* 2007;135:1231–47.
- 12 Gage KL, Burkot TR, Eisen RJ, Hayes EB. Climate and vector-borne diseases. *Am J Prev Med* 2008;35:436–50.
- 13 Patz JA, Olson SH, Uejio CK, Gibbs HK. Disease emergence from global climate and land use change. *Med Clin North Am* 2008;92:1473–91.
- 14 Gould EA, Higgs S. Impact of climate change and other factors on emerging arbovirus diseases. *Trans R Soc Trop Med Hyg* 2009;103:109–21.
- 15 Romi R. History and updating on the spread of *Aedes albopictus* in Italy. *Parassitologia* 1995;37:99–103.
- 16 Chomel BB, Belotto A, Meslin FX. Wildlife, exotic pets and emerging zoonoses. *Emerg Infect Dis* 2007;13:6–11.
- 17 Scholte EJ, Jacobs F, Linton YM, Dijkstra E, Fransen J, Takken W. First record of *Aedes (Stegomyia) albopictus* in the Netherlands. *Euro Mosq Bull* 2007;22:5–9.
- 18 Enserink M. A mosquito goes global. Science 2008;320:864-6.
- 19 Pluskota B, Storch V, Braunbeck T, Beck M, Becker N. First record of *Stegomyia albopicta* (Skuse) in Germany. *Euro Mosq Bull* 2008:26:1–5.
- 20 Purse BV, Brown HE, Harrup L, Mertens PPC, Rogers DJ. Invasion of bluetongue and other orbivirus infections into Europe; the role of biological and climatic processes. *Rev Sci Tech* 2008;27:427–42.
- 21 Eritja R, Escosa R, Lucientes J, Marques E, Molina R, Roiz D, *et al.*Worldwide invasion of vector mosquitoes: present European distribution and challenges for Spain. *Biol Invasions* 2005;7: 87–97.
- 22 Chretien J-P, Linthicum KJ. Chikungunya in Europe; what's next? *Lancet* 2007;370:1805–6.
- 23 Pavan WO. Chikununya—Italy (Emilia Romagna): confirmed. ProMed. 3 Sep 2007 [cited 2009 12 February] archive no. 20070903.2899. Available from URL http://www.promedmail.org.
- 24 Karti SS, Odabasi Z, Korten V, Yilmaz M, Sonmez M, Caylan R. Crimean-Congo haemorrhagic fever in Turkey. *Emerg Infect Dis* 2004;10:1379–84.
- 25 Papa A, Maltezou HC, Tsiodras S, Dalla VG, Papadimitriou T, Pierroutsakos I, *et al.* A case of Crimean-Congo haemorrhagic fever in Greece, June 2008. *Euro Surveill* 2008;13:7–9.
- 26 Haglund M. Occurrence of TBE in areas previously considered being non-endemic. *Int J Med Microbiol* 2002;**291** (Suppl 33): 50–4.
- 27 Weissenböck H, Kolodziejek J, Url A, Lussy H, Rebel-Bauder B, Nowotny N. Emergence of Usutu virus, an African mosquitoborne *flavivirus* of the Japanese encephalitis virus group, Central Europe. *Emerg Infect Dis* 2002;8:652–6.
- 28 Bakonyi T, Erdélyi K, Ursu K, Ferenczi E, Csörgő T, Lussy H, et al. Emergence of Usutu virus in Hungary. J Clin Microbiol 2007;45:3870–4.

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- 29 Busquets N, Alba A, Allepuz A, Aranda C, Núòez JI. Usutu virus sequences in *Culex pipiens* (Diptera: Culicidae), Spain. *Emerg Infect Dis* 2008;14:861–3.
- 30 Giudice PD, Schuffenecker I, Vandenbos F, Counillon E, Zeller H. Human West Nile virus, France. *Emerg Infect Dis* 2004;10:1885–6.
- 31 Krisztalovics K, Molnár Z, Csohán Á, Bán E, Zöldi V, Kaszás K. West Nile virus infections in Hungary, August-September 2008. *Euro Surveill* 2008;**13**: pii: 19030.
- 32 Hubalek Z, Halouzka J. West Nile fever—a remerging mosquitoborne viral disease in Europe. *Emerg Infect Dis* 1999;5:643–50.
- 33 Savini G, Monaco F, Calistri P, Lelli R. Phylogenetic analysis of West Nile virus isolated in Italy in 2008. *Euro Surveill* 2008;13: pii: 19048.
- 34 Smith R, Takkinen J, Editorial Team. Lyme borreliosis: Europewide coordinated surveillance and action needed? *Euro Surveill* 2006:11:2977.
- 35 Schaffner F, Hendrickx G, Scholte EJ, Medlock JM, Angelini P, Ducheyne E. Development of *Aedes albopictus* risk maps. TigerMaps project report. European Centre for Disease Prevention and Control: Stockholm, 2009.
- 36 Biological Records Centre [Online]. 2009 [cited 2009 27 November]. Available from: www.brc.ac.uk.
- 37 Pietzsch ME, Medlock JM, Jones L, Avenell D, Abbott J, Harding P, *et al.* Distribution of *Ixodes ricinus* in the British Isles: investigation of historical records. *Med Vet Entomol* 2005;**19**: 306–14
- 38 Jameson LJ, Medlock JM. Spatial distribution of tick species parasitizing humans and animals in Great Britain, 2005–2009. In preparation.
- 39 Medlock JM, Snow KR. British mosquitoes. *Br Wildl* 2008;**19**: 338–46.
- 40 Higgs S, Snow KR, Gould E. The potential for West Nile virus to establish outside of its natural range: a consideration of potential mosquito vectors in the United Kingdom. *Trans R Soc Trop Med Hyg* 2004;**98**:82–7.
- 41 Cranston PS, Ramsdale CD, Snow KR, White GB. Adults, Larvae and Pupae of British Mosquitoes (Culicidae). Scientific Publication, no. 48. Freshwater Biological Association: Ambleside, Cumbria, 1987.
- 42 Fonseca DM, Keyghobadi N, Malcolm CA, Mehmet C, Schaffner F, Mogi M, *et al.* Emerging vectors in the *Culex pipiens* complex. *Science* 2004;303:1535–8.
- 43 Medlock JM, Snow KR, Leach S. Potential transmission of West Nile virus in the British Isles: an ecological review of candidate mosquito bridge vectors. *Med Vet Entomol* 2005;19:2–1.
- 44 Snow KR, Rees AT, Bulbeck SJ. A Provisional Atlas of the Mosquitoes of Britain. University of East London: London, 1998.
- 45 Snow KR, Ramsdale CD. Distribution chart for European mosquitoes. *Euro Mosq Bull* 1999;3:14–31.
- 46 Snow KR, Medlock JM. The mosquitoes of Epping Forest, Essex, UK. Euro Mosq Bull 2008;26:9–17.
- 47 Romi R, Pontuale G, CIufolini MG, Fiorentini G, Marchi A, Nicoletti L, *et al.* Potential vectors of West Nile virus following an equine disease outbreak in Italy. *Med Vet Entomol* 2004;18: 14–19.
- 48 Medlock JM, Snow KR, Leach S. Possible ecology and epidemiology of medically important mosquito-borne arboviruses in Great Britain. *Epidemiol Infect* 2007;135:466–82.
- 49 Moutailler S, Krida G, Schaffner F, Vazeille M, Failloux A. Potential vectors of Rift Valley fever virus in the Mediterranean region. *Vector Borne Zoonotic Dis* 2008;8:749–53.
- 50 Lundstrom JO. Mosquito-borne viruses in Western Europe: a review. *J Vector Ecol* 1999;24:1–39.
- 51 Brummer-Korvenkontio M, Vapalahti O, Kuusisto P, Saikku P, Manni T, Koskela P, *et al.* Epidemiology of Sindbis virus infections in Finland 1981–96: possible factors explaining a peculiar disease pattern. *Epidemiol Infect* 2002;129:335–45.
- 52 Francy DB, Jaenson TG, Lundström JO, Schildt EB, Espmark A, Henriksson B, *et al.* Ecologic studies of mosquitoes and birds as

- hosts of Ockelbo virus in Sweden and isolation of Inkoo and Batai viruses from mosquitoes. *Am J Trop Med Hyg* 1989;**41**: 355–63
- 53 Lundstrom JO, Niklasson B, Francy DB. Swedish *Culex torrentium* and *Cx.pipiens* (Diptera: Culicidae) as experimental vectors of Ockelbo virus. *J Med Entomol* 1990;27:561–3.
- 54 Turell MJ, Lundstrom JO, Niklasson B. Transmission of Ockelbo virus by Aedes cinereus, Ae. communis and Ae. excrucians (Diptera: Culicidae) collected in an enzootic area in central Sweden. J Med Entomol 1990;27:266–8.
- 55 Lundstrom JO, Turell MJ, Niklasson B. Antibodies to Ockelbo virus in three orders of birds (Anseriformes, Galliformes and Passeriformes) in Sweden. *J Wildl Dis* 1992;28:144–7.
- 56 Buckley A, Dawson A, Moss SR, Hinsley SA, Bellamy PE, Gould EA. Serological evidence of West Nile virus, Usutu virus and Sindbis virus infection of birds in the UK. *J Gen Virol* 2003;84:2819–28.
- 57 Lundstrom JO, Lindström KM, Olsen B, Dufva R, Krakower DS. Prevalence of Sindbis virus neutralizing antibodies among Swedish passerines indicates that thrushes are the main amplifying hosts. *J Med Entomol* 2001;38:289–97.
- 58 Naucke TJ, Pesson B. Presence of *Phlebotomus mascittii* in Germany. *Parasitol Res* 2000;86:335–6.
- 59 Shaw SE, Langton DA, Hillman TJ. Letter-canine leishmaniasis in the UK. *Vet Rec* 2008;**163**:253–4.
- 60 Medlock JM, Avenell D, Barrass I, Leach S. Analysis of the potential for survival and seasonal activity of *Aedes albopictus* (Diptera: Culicidae) in the United Kingdom. *J Vector Ecol* 2006;31:292–304.
- 61 Takumi K, Scholte E-J, Braks M, Reusken C, Avenell D, Medlock JM. Introduction, establishment and seasonal activity of *Aedes albopictus* in the Netherlands. *Vector Borne Zoonotic Dis* 2009;9:191–6.
- 62 Lindgren E, Tälleklint L, Polfeldt T. Impact of climate change on the northern latitude limit and population density of the disease-transmitting European tick *Ixodes ricinus*. *Environ Health Perspect* 2000;**108**:119–23.
- 63 Parola P, Socolovschi C, Jeanjean L, Bitam I, Fournier P, Sotto A, et al. Warmer weather linked to tick attack and emergence of severe rickettsioses. PLoS Negl Trop Dis 2008;2:e338.
- 64 Jameson LJ, Phipps LP, Medlock JM. Surveillance for exotic ticks on companion animals in the United Kingdom. *Vet Rec* 166:202–3.
- 65 Fortin JF, Slocombe JOD. Temperature requirements for the development of *Dirofilaria immitis* in *Aedes triseriatus* and *Ae.* vexans. Mosq News 1981;41:625–33.
- 66 Knight DH, Lok JB. Seasonality of heartworm infection and implications for chemoprophylaxis. Clin Tech Small Anim Prac 1998:13:77–82.
- 67 Lok JB, Knight DH. Laboratory verification of a seasonal heartworm model. *Proceedings of the Heartworm Advances in Heartworm Disease Symposium 1998*. American Heartworm Society: Batavia IL, 1998, pp 15–20.
- 68 Slocombe JOD, Surgeoner GA, Srivastava B. Determination of heartworm transmission period and its use in diagnosis and control. *Proceedings of the Heartworm Symposium 1989*. American Heartworm Society: Washington, DC, 1989. pp 19–26.
- 69 Detinova T. Age-grouping methods in Diptera of medical importance. *Monograph Series*. World Health Organization: Geneva, 1962, pp 13.
- 70 Genchi C, Rinaldi L, Cascone C, Mortarino M, Cringoli G. Is heartworm disease really spreading in Europe? Vet Parasitol 2005;133:137–48.
- 71 Svobodova V, Misonova P. The potential risk of *Dirofilaria immitis* becoming established in the Czech Republic by imported dogs. *Vet Parasitol* 2005;128:137–40.
- 72 Medlock JM, Barrass I, Kerrod E, Taylor MA, Leach S. Analysis of climatic predictions for extrinsic incubation of *Dirofilaria* nematodes in the UK. *Vector Borne Zoonotic Dis* 2007;7:4–14.

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- 73 Bennett E, Clement J, Sansom P, Hall I, Leach S, Medlock JM. Environmental and ecological potential for enzootic cycles of Puumala hantavirus in Great Britain. Epidemiol Infect 138:91–8.
- 74 Clement J, Vercauteren J, Verstraeten WW, Ducoffre G, Barrios JM, Vandamme A-M, *et al.* Relating increasing hantavirus incidences to the changing climate: the mast connection. *Int J Health Geogr* 2009;**8**:1.
- 75 Tersago K, Verhagen R, Servais A, Heyman P, Ducoffre G, Leirs H. Hantavirus disease (*Nephropathia epidemica*) in Belgium: effects of tree seed production and climate. *Epidemiol Infect* 2009;137: 250–6.
- 76 Sumilo D, Bormane A, Asokliene L, Lucenko I, Vasilenko V, Randolph S. Tick-borne encephalitis in the Baltic states: identifying risk factors in space and time. *Int J Med Microbiol* 2006;**296** (Suppl 40): 76–9.
- 77 Sumilo D, Asokliene L, Bormane A, Vasilenko V, Golovljova I, Randolph SE. Climate change cannot explain the upsurge of tick-borne encephalitis in the Baltics. *PLoS ONE* 2007;2:e500.
- 78 Sumilo D, Bormane A, Asokliene L, Vasilenko V, Golovljova I, Avsic-Zupanc T, *et al.* Socio-economic factors in the differential upsurge of tick-borne encephalitis in central and Eastern Europe. *Rev Med Virol* 2008;18:81–95.
- 79 Sumilo D, Bormane A, Vasilenko V, Golovljova I, Asokliene L, Zygutiene M, et al. Upsurge of tick-borne encephalitis in the Baltic states at the time of political transition, independent of changes in public health practices. Clin Microbiol Infect 2009;15:75–80.
- 80 Labuda M, Nuttall PA, Kozuch O, Eleckova E, Williams T, Zuffova E, *et al.* Non-viraemic transmission of tick-borne encephalitis virus: a mechanism for arbovirus survival in nature. *Experientia* 1993;**49**:802–5.
- 81 Labuda M, Jones LD, Williams T, Danielova V, Nuttall PA. Efficient transmission of tick-borne encephalitis virus between co-feeding ticks. *J Med Entomol* 1993;30:295–9.
- 82 Angelini R, Finarelli AC, Angelini P, Po C, Petropulacos K, Macini P, *et al.* An outbreak of chikungunya fever in the province of Ravenna, Italy. *Euro Surveill* 2007;12:E070906.1.
- 83 Rappole JH, Derrickson SR, Hubalek Z. Migratory birds and spread of West Nile virus in the western hemisphere. *Emerg Infect Dis* 2000;6:319–28.
- 84 Reed KD, Meece JK, Henkel JS, Shukla SK. Birds, migration and emerging zoonoses: West Nile virus, Lyme disease, Influenza A and Enteropathogens. *Clin Med Res* 2003;1:5–12.
- 85 Gerdes GH. Rift Valley fever. Rev Sci Tech 2004;23:613-23.
- 86 Comment: pet travel: DEFRA considers relaxing the rules. *Vet Rec* 2007:32:534–5.
- 87 Comment: PETS under pressure. Vet Rec 2008;162:425.
- 88 Trees AJ. Disease threats from travelling pets. *Vet Rec* 2009;**164**: 28–9.
- 89 Smith GC, Gangadharan B, Taylor Z, Laurenson MK, Bradshaw H, Hide G, *et al.* Prevalence of zoonotic important parasites in the red fox (*Vulpes vulpes*) in Great Britain. *Vet Parasitol* 2003;118:133–42.
- 90 Eckert J, Deplazes P. Alveolar echinococcosis in humans: the current situation in central Europe and the need for countermeasures. *Parasitol Today* 1999;15:315–19.
- 91 Deplazes P, Hegglin D, Gloor S, Romig T. Wilderness in the city: the urbanisation of *Echinococcus multilocularis*. *Trends Parasitol* 2004:20:77–84.
- 92 Medlock JM, Leach S. *Echinococcus multilocularis* and possible cycles in UK wildlife. *Vet Rec* 2009;**164**:789–90.
- 93 Planning and Policy Statement 9 (2005): Biodiversity and Geological Conservation. 2005 [cited 11 March 2009]: Available from URL: http://www.communities.gov.uk/publications/planningandbuilding/pps.
- 94 Defra. Extra protection for the water vole [online], 2008 [cited 11 March 2009]. Available from URL: http://www.defra.gov.uk/news/2008/080226a.htm.

- 95 EU Habitats Directive. (1992) [cited 11 March 2009]: Available from URL: http://www.ec.europa.eu/environment/nature/legislation/habitatsdirective/index_en.htm.
- 96 Convention on Conservation of European Wildlife and Habitats. [cited 11 March 2009]: Available from URL: http://www.coe.int/t/dg4/cultureheritage/conventions/bern/default_en.asp.
- 97 Tsetsarkin AK, Vanlandingham DL, McGee CE, Higgs S. A single mutation in chikungunya virus affects vector specificity and epidemic potential. *PLoS Pathog* 2007;3:1895–906.
- 98 Bicout DJ, Sabatier P. Mapping Rift Valley fever vectors and prevalence using rainfall variations. *Vector Borne Zoonotic Dis* 2004;4:33–42.
- 99 Thomson MC, Molesworth AM, Djingarey MH, Yameogo KR, Belanger F, Cuevas LE. Potential of environmental models to predict meningitis epidemics in Africa. *Trop Med Int Health* 2006;11:781–8.
- 100 Thomson MC, Doblas-Reyes FJ, Mason SJ, Hagedorn R, Connor SJ, Phindela T, *et al.* Malaria early warnings based on seasonally climate forecasts from multi-model ensembles. *Nature* 2006;**439**: 576–9.
- 101 Randolph SE. The shifting landscape of tick-borne zoonoses: tick-borne encephalitis and Lyme borreliosis in Europe. *Phil Trans R Soc B* 2001;356:1045–56.
- 102 Heller NE, Zavaleta ES. Biodiversity management in the face of climate change: a review of 22 years of recommendations. *Biol Conserv* 2009;142:14–32.
- 103 Wildlife Trusts: A Living Landscape. [cited 15 March]: Available at URL: http://www.wildlifetrusts.org/?section = environment: livinglandscapes.
- 104 Natural England: Environmental Stewardship. [cited 15 March]: Available at URL: http://www.naturalengland.org.uk/ourwork/farming/funding/es/default.aspx.
- 105 Despommier D, Ellis BR, Wilcox BA. The role of ecotones in emerging infectious diseases. *EcoHealth* 2007;3:281–9.
- 106 Woodford MH. Veterinary aspects of ecological monitoring: the natural history of emerging infectious diseases of human, domestic animals and wildlife. *Trop Anim Health Prod* 2009;41: 1023–33.
- 107 Merckx T, Feber RE, Riordan P, Townsend MC, Bourn NAD, Parsons MS, *et al.* Optimizing the biodiversity gain from agri-environment schemes. *Agric Ecosyst Environ* 2009;**130**: 177–82.
- 108 Medlock JM, Pietzsch ME, Rice NV, Jones L, Kerrod E, Avenell D, et al. Investigation of ecological and environmental determinants for the presence of questing Ixodes ricinus on Gower, South Wales. J Med Entomol 2008;45:314–25.
- 109 Medlock JM. Spatial distribution of questing deer ticks, *Ixodes ricinus*, along woodland rides and glades in south Wiltshire: implications for woodland management and public exposure. University of Bristol MSc dissertation 2009; 1–85.
- 110 Radley G, O'Reilly J, Jowitt A. Environmental Stewardship. *Br Wildl* 2005;**16**:257–63.
- 111 Jameson LJ, Medlock JM. Results of HPA tick surveillance in Great Britain. *Vet Rec* 2009;**165**:154.
- 112 Wetland Vision Project. Accessed online 1 March 2009. Available from URL: http://www.wetlandvision.org.uk.
- 113 Fuller M. Allaying public fears of health issues on wetlands. *Br Wildl* 2005;**16**:332–8.
- 114 Medlock JM, Snow KR. Natural predators and parasites of British mosquitoes—a review. *Eur Mosq Bull* 2008;25:1–11.
- 115 SDC. *Tidal Power in the UK*. Severn Barrage Proposals: UK. Research Report 3; 2007.
- 116 Snow K. Malaria and mosquitoes in Britain: the effect of global climate change. *Euro Mosq Bull* 1999;4:17–24.
- 117 Kuhn KG, Campbell-Lendrum DH, Armstrong B, Davies CR. Malaria in Britain: past, present, and future. *Proc Natl Acad Sci USA* 2003;100:9997–10001.

www.eht-journal.org page 8/9

Emerging Health Threats Journal 2010, 3:e1

- 118 Ramsdale CD, Gunn N. History and prospects for mosquito-borne disease in Britain. *Euro Mosq Bull* 2005;**20**:15–30.
- 119 Snow KR, Medlock JM. The potential impact of climate change on the distribution and prevalence of mosquitoes in Britain. *Euro Mosq Bull* 2006;21:1–10.
- 120 Lindsay ŚW, Willis SG. Predicting future areas suitable for vivax malaria in the United Kingdom, commissioned as part of the UK Government's Foresight project: Infectious Diseases: Preparing
- for the Future. Foresight Directorate, Office of Science and Innovation, UK, 2006; 1–15.
- 121 Chretien J-P, Anyamba A, Bedno SA, Breiman RF, Sang R, Sergon K, *et al.* Drought-associated chikungunya emergence along coastal East Africa. *Am J Trop Med Hyg* 2007;**76**: 405–407.
- 122 ECDC Mission Report 2007. Chikungunya in Italy: joint ECDC/WHO visit for a European risk assessment. 2007.

www.eht-journal.org page 9/9