

Review

Macrocycles and Supramolecules as Antioxidants: Excellent Scaffolds for Development of Potential Therapeutic Agents

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Abstract: Oxidative stress due to the high levels of reactive oxygen species (ROS) that damage biomolecules (lipids, proteins, DNA) results in acute inflammation. However, without proper intervention, acute inflammation progresses to chronic inflammation and then to several chronic diseases, including cancer, myocardial infarction, cardiovascular diseases, chronic inflammation, atherosclerosis, and more. There has been extensive research on the antioxidants of natural origin. However, there are myriad possibilities for the development of synthetic antioxidants for pharmacological applications. There is an increasing interest in the identification of novel synthetic antioxidants for the modulation of biochemical processes related to ROS. In this regard, derivatives of supramolecules, such as calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, porphyrin etc. are gaining attention for their abilities to scavenge the free radicals. Supramolecular chemistry offers excellent scaffolds for the development of novel antioxidants that can be used to modulate free radical reactions and to improve the disorders related to oxidative stress. This review focuses on the interdisciplinary approach for the design and development of novel synthetic antioxidants based on supramolecular scaffolds, with potentially protective effects against oxidative stress.

Keywords: supramolecules; calix[n]arene; resorcinarene; calixtyrosol; calixpyrrole; cucurbit[n]uril; porphyrin; antioxidant; radical scavenger; oxidative stress; reactive oxygen species; reactive nitrogen species

1. Introduction

Oxidative stress, due to the high levels of reactive oxygen species (ROS) that damage biomolecules (lipids, proteins, DNA), eventually leads to many chronic diseases, including cancer [1,2], myocardial infarction [3,4], cardiovascular diseases (CVD) [5–7], chronic inflammation [8], atherosclerosis [9] and more [10,11]. Free radicals such as hydroxyl (OH•), superoxide (O^{2-•}), and nitric monoxide (NO•) are the major ROS found in cells [12]. Antioxidants are the molecules that scavenge free radicals to combat cellular oxidative damage [13,14]. Antioxidants can slow or prevent the oxidation processes initiated by ROS that damages cells in the body [15]. Based on their activity, antioxidants fall into two categories: enzymatic and non-enzymatic antioxidants [16]. Enzymatic antioxidants break down and remove free radicals, while non-enzymatic antioxidants work by interrupting free radical chain reactions [17]. Antioxidants exert their effect via numerous mechanisms, including inhibition of the formation of free radicals by reducing hydroperoxides (ROO•) and hydrogen peroxides, the scavenging of active free radicals, sequestering metal ions and repairing and clearing oxidative



damages [18]. Antioxidants, therefore, have tremendous applications in the pharmaceutical industry due to their prophylactic and therapeutic activities [19,20].

There is increasing interest in the identification of novel antioxidants for the modulation of biochemical processes related to ROS [21,22]. The use of antioxidants, including enzymes, natural products, and synthetic compounds, is a logical therapeutic intervention for many ROS-related diseases, including cancer [23,24]. Considerable attention has been given to the design and development of potent antioxidants and the determination of their radical scavenging ability in complex samples, including cell culture studies and in vivo models. However, the development of novel antioxidants with both high pharmacological activity and few side effects is a challenging field [25]. In this regard, the use of supramolecular scaffolds, one of the many approaches that have been used for the development of novel antioxidants, can prove fruitful in the coming years.

Research on supramolecules and their applications has gained considerable attention in the last decade. Several classes of supramolecules have been developed for various applications, including chemosensors, biosensors, drug-delivery vehicles, and prodrugs. However, their application in the development of potent antioxidants has yet to be explored to its full capacity. Hence, supramolecular scaffolds such as calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, porphyrin etc. provide a novel platform to design and develop novel antioxidants with both high pharmacological activity and few side effects. The development of synthetic antioxidants is attractive, as it allows one to manipulate the physicochemical properties to improve the antioxidant properties for various applications. It is crucial to notice that there are several reports on the inclusion complexes of these macrocycles, including cyclodextrins with antioxidant activities [26–30]. However, this review is focused mainly on the direct chemical modifications of calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, and porphyrin scaffolds to synthesize the novel antioxidant compounds.

This review aims to explore the advances in the development of antioxidants, based on calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, and porphyrin (Figure 1).



Figure 1. Supramolecular scaffolds used in the development of antioxidants (**a**) calix[n]arene; (**b**) resorcinarene; (**c**) calixtyrosol; (**d**) calixpyrrole; (**e**) cucurbit[n]uril; (**f**) porphyrin.

2. Supramolecular Scaffolds Used for the Development of Antioxidants

2.1. Calix[n]arene

Calixarenes are cyclic oligomers obtained by the pyrolysis of a polymer produced from the condensation of formaldehyde with *p*-alkylphenols in the presence of a strong base [31,32]. Depending upon the number of monomers, they can be classified as calix[4]arene 1, calix[6]arene 2, and calix[8]arene **3** (Figure 2). Calixarenes can be substituted with a variety of functional groups on their upper and lower rims, resulting in molecules with a variety of biological applications [33]. Water-soluble calix[4]arene derivatives have been exploited for their molecular recognition properties [34–36]. Since the calixarenes can be tailored to improve their water solubility, they can be excellent platforms for the development of useful antioxidants. Calixarenes are also known to poses antioxidant features for over a couple of decades [37,38].



Figure 2. Calix[n]arene derivatives 1-6.

Gorghiu et al. reported the contribution of p-t-butylcalix[n]arenes (4, 5 and 6) (Figure 2) in the thermal stability of different types of low-density polyethylenes (LDPE) [39]. They found that the antioxidant feature of p-t-butylcalix[4]arene 4 is higher than the p-t-butylcalix[6]arene 5 and p-t-butylcalix[8]arene 6 for tested LDPE. The proposed mechanism is presented in Scheme 1.



Scheme 1. Mechanism of radical scavenging activity of t-butylcalix[n]arene derivatives.

Among the polyhydroxy phenolic compounds, the hydroxycinnamic acid derivatives are abundant in nature and are reported to possess strong antioxidant properties [40–42]. The amide derivatives of cinnamic acid are also reported to have significant radical scavenging ability [43]. Considering the potential of hydroxycinnamic acid (HA) as a potent antioxidant, Consoli et al. designed and synthesized the calixarene platform clustered with HA [44].

The radical scavenging and antioxidant activities determined by using 2,2-diphenyl-1picrylhydrazyl radical (DPPH[•]) and AIBN[•]–induced linoleic acid peroxidation test showed that the compounds 7 and 8 showed enhanced activity as compared to the corresponding monomers 9 and 10 (Figure 3). The rate constant values ($k_1 > 1.5 \times 10^4$) and the stoichiometric factors *n* (number of DPPH[•] quenched/antioxidant molecule) for 7 (n = 7.7) and 8 (n = 2.7) indicated their strong radical quenching ability.



Figure 3. Structures of calix[4]arene derivatives 7–19.

In 2012, Patel et al. reported the synthesis and evaluation of calix[4]arene based 1,3,4-oxadiazole and thiadiazole derivatives **11–19** (Figure 3) [45]. The conjugation of **4** (*tert*-butylcalix[4]arene) with

1,3,4-oxadiazole and 1,3,4-thiadiazole derivatives allowed them to synthesize novel antioxidant calixarene derivatives. All compounds showed significant DPPH[•] scavenging activity. Notably, the compounds containing the oxadiazole ring were more potent antioxidants than other compounds. The DPPH[•] scavenging activity of compound **18** (82.6%), containing oxadiazole ring and S bridge connecting to the calix[4]arene moiety, was comparable with the ascorbic acid (92.0%), which is used as a standard in the assays.

James et al. reported the synthesis and evaluation of antioxidant activities of amphiphilic phospholipid calix[4]arenes, that mimic the micellular delivery systems [46]. They evaluated the antioxidant activity compounds **20**, **21** and **22**, (Figure 4) using PC12 cells that were pre-treated with these compounds and then stressed with either H_2O_2 , menadione, or glutamate. The obtained results indicated that the PC12 cells treated compounds **20**, **21** and **22** showed significant inhibition in ROS generation than the untreated control groups. Furthermore, the loading of curcumin in the amphiphiles of these compounds not only increases the delivery of curcumin to the cytoplasm, but also enhances the antioxidant capacity. The results establish that calix[n]arenes are potentially useful synthetic antioxidants.



Figure 4. Structures of calix[4]arene derivatives 20-22.

There have been several reports on the methods to substitute the upper-rim and lower-rim of calix[4]arene [47,48]. Pur et al. came up with an idea to substitute the upper-rim of calix[4]arene with dihydropyrimidine moiety, as shown in Scheme 2 [49]. They devised a method to substitute four dihydropyrimidine moieties at the upper-rim of calix[4]arene by facial green Biginelli reaction. The resulting clusters of calix[4]arene-based dihydropyrimidines (24, 25) showed strong antiradical activity compared to the corresponding monomers.



Scheme 2. Synthetic pathway to substitute upper-rim of calix[4]arene with dihydropyrimidine [49].

Recently, Stephens et al. reported on the structural requirements for the antioxidant activity of calix[n]arenes and their associated antibacterial activity [50]. In this work, they studied ten calix[n]arene derivatives **26–35** for their antioxidant and antibacterial activities by linking them to Ag nanoparticles (Figure 5). In this study, they found that the calix[n]arenes bearing sulphonate groups on the upper-rim and/or lower-rim have intrinsic antioxidant capacity. The less sulfonated calix[n]arenes necessitate

the linkage to Ag nanoparticles in order to achieve similar efficacy. Therefore, these results indicate that the calix[n]arenes may be used as therapeutic agents in several diseases related to oxidative stress.



Figure 5. Structures of calix[4]arene derivatives 26–35.

Lu et al. reported the theoretical study that allowed one to find a way for improving the antioxidative activity of resveratrol by calix[4]arene-like tetramer [51]. In this study, they designed tetra-(3,5-dihydroxy)styryl-calix[4]arene (36, a calix[4]arene-like tetramer of resveratrol) and theoretically studied at the DFT-BP86/6-311+G(d,p) level of theory for antioxidant activity in comparison to resveratrol **37** (Figure 6). The spin density in 4'-phenoxyl radicals and cation radicals are found to delocalize over the whole molecular skeleton of **36**, which allows one to form more stable radicals than that of resveratrol. Further, it was revealed that calix[4]arene scaffold plays a crucial role in the delocalization of the unpaired electron. Therefore, the synthesis of analogues derived from the conjugation of calix[4]arene and resveratrol derivatives can lead to potentially strong antioxidant molecules.



Figure 6. Structures of calix[4]arene derivatives 36–39.

Recently, Ni et al. reported the antiradical and antioxidative activity of azocalix[4]arene derivatives **38**, **39** (Figure 6) [52]. In this work, they developed two azocalix[4]arene derivatives by a diazo coupling reaction between calix[4]arene and diazonium salts. Their antiradical and antioxidative activity of these compounds was evaluated by tow assays (hydroxyl radical scavenging assay and the pyrogallol autoxidation inhibition assay). The experimental and theoretical results identified both compounds **38** and **39** as potent antiradical and antioxidant agents. The synergism between the calix[4]arene moiety and the *para*-azo substituent groups at the upper rim contributed significantly towards the antiradical and antioxidant activity.

Iminecalix[4]arene derivatives are known to possess a deep cavity and hyperconjugation [53]. The water-soluble iminecalix[4]arene derivatives are reported for the molecular recognition of various organic molecules [54]. However, only recently, Silva et al. reported the novel iminecalix[4]arene derivatives (42–47) with anticandidal activity (Scheme 3) [55]. However, there are no reports on

the antioxidant activity of iminecalixarene derivatives. This class of calixarene has a long *pi*-aromatic conjugation system, which with proper substitutions, can be exploited for the antioxidant activities (Figure 7).



Scheme 3. Synthetic pathway for various derivatives of iminecalix[4]arenes [55].



Figure 7. Structures of iminecalix[4]arene derivatives 42-47.

Recently, Ozgu et al. reported on the iminecalix[4]arenes containing sulphonamide moieties [56]. The novel calix[4]azacrown sulphonamide Schiff bases were synthesized by the condensation of calix[4]azacrown aldehydes with various primary and secondary sulphonamides. The obtained compounds demonstrated relevant antioxidant activity compared with standard antioxidants used in the study.

2.2. Resorcinarene (calix[4]resorcinarene)

Resorcinarenes are a type of calixarene obtained by the condensation of 1,3-dihydroxybenzene (Resorcinol) with an aldehyde in an acidic environment (Scheme 4) [57,58]. Similar to calix[4]arenes, resorcinarenes also assume a cone shape, with eight hydroxyl groups on the upper-rim and four alkyl groups on the lower-rim [59,60]. Resorcinarene has been extensively studied for various applications along the lines of host-guest chemistry and redox-active macrocycles [61–63]. Since resorcinarene is a polyhydroxy-supramolecule, it offers a highly efficient scaffold for the development of antioxidants that can be tailored by varying the functional groups. Stable cyclic nitroxides are known to possess remarkable antioxidant activity due to their ability to scavenge superoxide, peroxide, and alkyl radicals [64–66].



Scheme 4. General scheme for the synthesis of resorcinarene derivatives.

In 2009, Vovk et al. reported on the antioxidant and antiradical activities of resorcinarene tetranitroxides (Figure 8) [67]. They synthesized the resorcinarene (48, 49) derivatives (50–53) by the aminomethylation of resorcinarene octols with 4-amino-TEMPO TEMPO ((4-Amino-2,2,6,6-tetramethylpiperidine-1-oxyl), and studied their antioxidant activities. They found that the Tetra- TEMPO resorcinarenes (50–53) are efficient scavengers of DPPH radicals and showed superoxide dismutase-like activity. These compounds were also found to have a high efficiency for inhibition of ABAP-induced peroxidation of linoleic acid. The macrocyclic structure of resorcinarene and intramolecular hydrogen bonding had a considerable contribution to the antiradical activity of these compounds. Recently, Ngurah reported the antioxidant activity of C-methoxyphenyl calix[4]resorcinarene 54 (Figure 8) [68]. The compound 56 showed reasonable antioxidant activity (IC50 = 79 ppm), but less than that of vitamin C (IC50 = 20.96 ppm), which was used as a standard in this study. Oliveira et al. reported the synthesis and comparative study on the antioxidant and anti-toxoplasma activities of vanillin 56 and its resorcinarene derivative 57 (Scheme 5a) [69].



Figure 8. Structures of resorcinarene derivatives 48-54.



Scheme 5. General scheme for the synthesis of resorcinarene derivative (a) compound 57 containing vanillin arms; (b) compound 60 containing 5-bromo-2-hydroxybenzene arms.

The in vivo acute toxicity assays of **56** and **57** demonstrated a substantial safety margin indicated by lack toxicity up to 300 mg/kg during 14 days after administration. The obtained compound **57** exhibited more significant antioxidant activity (84.9%) as compared to vanillin **56** (19.4%). These results indicate that the resorcinarene has excellent potential for the development of antioxidant and antiradical compounds. Recently, Abosadiya et al. reported the synthesis, characterization, x-ray structure, and biological activities of c-5-bromo-2-hydroxyphenylcalix[4]-2-methyl resorcinarene **60** (Scheme **5b**) [70]. The compound **60** was obtained by the condensation of 5-bromo-2-hydroxybenzaldehyde and 2-methylresorcinol in the presence of concentrated HCl. The compound **60** showed a strong activity (84.95%) in scavenging the DPPH free radicals. Since compound **60** is a polyphenolic compound, it inhibited the oxidation of DPPH, preferably using the hydrogen atoms to form the stable non-radical form of DPPH indicated by the formation of a pale-yellow color.

2.3. Calixtyrosol

Tyrosol is an essential antioxidant molecule with various biological activities. There has been an increasing interest in the development of strategies for the clustering of single drug units in order to design novel drugs. It is understood that the cluster effect can increase the pharmacological effect of a drug to a single drug unit [71].

Keeping this in mind, Pur et al. developed a novel calixtyrosol **64**, which is a novel cluster of tyrosol (Scheme 6) [72]. The free radical scavenging assay demonstrated that the calixtyrosol **64** has superior antioxidant activity (>5 fold) as compared to the single tyrosol unit. The enhanced antioxidant activity is attributed to the cluster of four impacted tyrosol units that show a synergistic effect in interactions with radicals.



Scheme 6. General scheme for the synthesis of calixtyrosol.

2.4. Calix[4]pyrrole

Calix[4]pyrroles have attracted substantial attention as supramolecular containers during the past two decades [73,74]. The calix[4]pyrroles are obtained by the condensation of pyrrole with acetone in the presence of an acid (Scheme 7a) [75]. Therefore, based on the use of ketone, the reaction with pyrrole, various derivatives of calix[4]pyrroles can be synthesized (Scheme 7b) by slight modifications in the reaction conditions [76].

Even though the calix[4]pyrrole offers an excellent scaffold that can be tailored for the development of potent antioxidants, it has been understudied in this regard. However, the complex of Au and calix[4]pyrrole has been studied for the antioxidant and radical scavenging efficiencies. Kongor et al. reported the synthesis and modeling of calix[4]pyrrole derivative **70** wrapped Au nanoprobes for the detection of Pb(II), antioxidant, and radical scavenging activities [77]. The calix[4]pyrrole wrapped Au nanoprobes were found to scavenge 76.4% of DPPH free radicles. They also found that the calix[4]pyrrole 70 had low antioxidant activity in comparison to its complex with the Au. More study on the development of antioxidants based on calix[4]pyrrole scaffold is required.



Scheme 7. (a) General scheme for the synthesis of calix[4]pyrrole 66; (b) Scheme for the synthesis of calix[4]pyrrole tetrahydrazide 70.

2.5. Cucurbituril (CB)

Cucurbituril, denoted as cucurbit[n]uril, is a supramolecule obtained by the condensation of glycoluril **73** with formaldehyde in the presence of strong acid [78]. The macrocycle was named cucurbituril, owing to its similarity to a pumpkin (*cucurbitaceae* family). The cucurbituril is often abbreviated as CB[n], highlighting the n glycoluril building blocks in the macrocycle [79,80] (Scheme 8).



Scheme 8. General scheme for the synthesis of cucurbituril (CB).

Depending on the number of glycoluril units, there are various derivatives of cucurbit[n]uril, including CB[5] 74, CB[6] 75, CB[7] 76, CB[8], CB[10], CB[13], CB[14], CB[15] [81–83] (Figure 9). Cucurbiturils have been studied for a variety of applications in chemistry, biology, and drug delivery [84].



Figure 9. Structures of cucurbit[n]uril derivatives 74–76.

Due to the excellent guest-binding ability of cucurbiturils in aqueous solutions, they play a significant role in the development of supramolecular nanostructures [85]. In 2013, Hou et al. constructed protein nanowires (Figure 10) through cucurbit[8]uril-based highly specific host-guest interaction [86]. The highly specific noncovalent interactions of CB[8] and a tripeptide FGG-tag, which was attached to the N-termini of dimeric glutathione S-transferase (GST), resulted in the formation of self-assembled protein nanowires. Then, the active site of selenoenzyme glutathione peroxidase (GPx) was introduced into GST for the construction of the functional protein nanowires. The functional protein nanowires that mimic the GPx-like activity showed high stability and exhibited substantial antioxidative properties to protect the mitochondria from oxidative stress. Therefore, the combination of cucurbituril host-guest interactions with proteins and enzymes presents a powerful tool for the construction of biologically active protein nanostructures.



GST nanowires

Figure 10. Formation of protein nanowires by CB[8]-based host-guest interactions.

Since cucurbituril has been extensively studied for its host-guest chemistry, it was used for the enhancement of the antioxidant activity of known antioxidants such as resveratrol. Recently, Lu et al. reported on the use of cucurbiturils to form inclusion complexes with resveratrol (Figure 11), in order to enhance the radical scavenging activity of the resveratrol [87]. Generally, the phenolic antioxidants resveratrol demonstrate their radical scavenging activity based on three mechanisms, such as the H atom transfer (HAT) mechanism, sequential proton loss electron transfer (SPLET) mechanism, and single electron transfer (SET) mechanism. Lu et al. found that the inclusion complexes res@CB[5] 77, res@CB[5] 78, and res@CB[5] 79 show the enhanced antioxidant capacity of resveratrol (Figure 11).

Recently, Kubota et al. developed a new class of artificial enzyme composed of cucurbit[10]uril, Mn-porphyrin, and imidazole [88]. They investigated a new class of artificial enzymes obtained by simply mixing the Mn-porphyrin, imidazole, and CB[10] in the aqueous solution. In this research study, they selected Mn(III)-5,10,15,20-tetrakis(1,3-dimethylimidazolium-2-yl)porphyrin (MndMIm4P) as a novel guest for CB[10]. Their simplified process has a great advantage over earlier reports that required a multi-step synthesis for the generation of artificial catalases. The preliminary in vitro study

in human cell lines suggested that the constructed artificial enzyme (Figure 12) catalytically eliminated the ROS, including H_2O_2 . The use of CB[10] in the construction of this artificial enzyme was crucial to increase its bioavailability. This study is one of a kind that demonstrates the meticulous use of supramolecules has a high potential to generate novel molecules as therapeutic antioxidants.



Figure 11. Inclusion complexes of resveratrol in cucurbiturils [87].



Figure 12. A new class of artificial enzyme (MndMIm4P@CB[10];Im) for the development of a therapeutic antioxidant [88].

2.6. Porphyrin

Porphyrins, metalloporphyrins, and related tetrapyrrolic macrocyclic compounds are a class of important heterocycles due to their wide applications in chemistry and biology [89,90]. They have been employed successfully in the areas of catalysis, medicine, and material science [91]. As shown in Scheme 9, a one-pot condensation of an aldehyde with pyrrole in acidic conditions can generate various derivatives of porphyrin **80**. Mackensen et al. reported the neuroprotection from delayed postischemic administration of a metalloporphyrin catalytic antioxidant [92]. They found that the administration of a metalloporphyrin had a significant neuroprotective effect, as it decreases postischemic superoxide mediated oxidative stress.



Scheme 9. Scheme for the synthesis of porphyrin.

Asayama et al. reported the manganese porphyrins (**81**, **82**) functionalized with the biomolecules (Figure 13) [93]. The developed porphyrin derivatives were studied for a potential role as antioxidants. The Mn-porphyrin (MnP) conjugated to catalase (**81**) was evaluated for its ability to catalyze the reduction of H_2O_2 to H_2O . This derivative was found to possess a dual function as SOD and catalase. The carbohydrate conjugated MnP (**82**) facilitated the anchoring of the conjugate on the cell surface by binding to a receptor. The results of this study suggest that the porphyrins can be modified to generate analogues with potent antioxidant activities.



Figure 13. Structures of Mn-porphyrin (MnP) conjugates 81, 82.

Fadda et al. reported the pharmacological activities of novel porphyrin derivatives [94,95]. They used a capping mechanism to produce a series of porphyrin derivatives **83–88** (Figure 14). All compounds were found to inhibit the peroxidation reactions in rat brain and kidney homogenates and rat erythrocyte hemolysis. The results of the ABTS assay indicated that the compounds **86** and **88** have a robust antioxidant activity, indicated by the 74.3% and 79.5% ABTS inhibition, respectively.



Figure 14. Structures of porphyrin derivatives 83-91.

In 2013, Tyurin et al. reported the redox characteristics and antioxidant activity of porphyrins with 2,6-dialkylphenol groups **89–91** (Figure 14) [96]. All compounds demonstrated significant DPPH scavenging activity. The antioxidant activity was attributed to the readily delocalized unpaired electron, resulting in the increased stability of a product after reaction with the radical.

Kubota et al. reported a synthesis of water-soluble MnP with antioxidant activities [97]. The Mn-porphyrin derivative **92** had a fair water-solubility and possessed SOD-like catalase-like activity. As shown in Scheme 10, the mechanistic study indicated the synergism of two Mn active sites gives a catalase-like activity to the compound. The restoration of the treadmill-running ability of SOD-deficient mouse indicates that the derivative **92** exhibited strong antioxidative activity in vivo.



Scheme 10. Multiple antioxidant activities of water-soluble di-nuclear Mn-Porphyrin.

Batini-Haberle et al. reviewed the SOD mimicking the activity of Mn porphyrins [98]. As shown in Figure 15, the dismutation process involves of two steps (i) SOD enzyme or MnP-based molecule acts as a (pro)oxidant in a first step, and as an antioxidant in the second step and closes the catalytic cycle, and (ii) acting as an oxidant, they produce H_2O_2 in the second step. Therefore, similar to SOD, its mimic MnPs can be considered to have applications in antioxidative defense in physiological conditions.

Tesakova et al. reported the tetraphenylporphyrene (93) and evaluation of its antioxidant activity by electrochemistry [99]. Abu-Melha reported the synthesis of *meso*-substituted porphyrins 94–101 and the evaluation of their antioxidant activity (Figure 16) [100]. Among derivatives 94–101, compounds 96, 98 and 101 exhibited the strongest antioxidant activity. The strong radical-scavenging activity of compounds 96 and 98 was credited to the presence of N and S atoms in their structures. It is well known that the presence of sulfur in compounds improves their radical-scavenging activity [101].

Porphyrins can be tailored to desired physicochemical properties by complexing with different metal ions, such as manganese, copper, zinc, silver, nickel etc. [102,103]. The *meso*-substituted porphyrins have gained considerable interest due to their ability to act as ligands for metal ions forming the complexes that have high applicability for therapeutic purposes [104,105]. Recently, Ahmed et al. synthesized and evaluated the antioxidant and anticancer activities of porphyrin derivatives **102–106** and their metalloporphyrin counterparts **107–114** (Figure 17) [106]. All compounds showed radical-scavenging activity. However, in particular, compounds **105** and **111** showed exceptional antioxidant activity as compared to ascorbic acid, which was used as a reference.



Figure 15. Mn porphyrins that mimic the SOD [98].



Figure 16. Structures of porphyrin derivatives 93–101.



Figure 17. Structures of porphyrin derivatives 102–114.

3. Discussion

ROS are highly reactive molecules that, in many cases, function as regulators of critical signaling pathways in living cells. The moderate levels of ROS are essential for various cellular processes, including gene expression. The elevated ROS levels alter the cell's microenvironment and damage the several biomolecules, including proteins, DNA leading to severe diseases like cancer and CVD. The generation of ROS and other free radicals in living cells is an inevitable consequence of aerobic metabolism. The levels of these highly reactive molecules is controlled by various antioxidant enzymatic and non-enzymatic pathways in the same cells. However, as evident from the disorders as a result of dysregulation of levels of ROS, the therapeutic use of external ROS modulators is inevitable.

In the last decade, the development of radical scavengers that mimic the activities of cellular enzymes such as SOD, catalase etc. has gained a tremendous interest of various scientific communities. The compounds that mimic enzyme activity function by one of the two methods, (i) by favoring a disproportionation process (SOD, catalase mimics) (ii) by undergoing reduction during the oxidation reactions (glutathione peroxidase mimics). There has been a tremendous amount of research on the identification of antioxidants from various natural sources. However, the development of synthetic antioxidants for therapeutic applications is a comparatively understudied field. The design, development, and synthesis of highly functional molecules with strong radical-scavenging activity warrants a suitable molecular scaffold that can be tailored for required physical, chemical, and biological properties.

Supramolecules such as calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, and porphyrin were used for the development of radical scavengers (Table 1). Calix[n]arens are the excellent scaffolds for the design of synthetic antioxidants, as the upper and lower rims of these molecules can be readily transformed by using various chemical reactions. The calix[n]arenes show a considerable amount of radical scavenging ability, owing to the aromatic hydroxyl groups that help in quenching the free radicals. Similar to the resveratrol derived analogues of calix[4]arenes, several other known antioxidant moieties can be substituted on the calixarene scaffold to afford stronger radical-scavengers. The resorcinarenes, as discussed earlier, can be used for the synthesis of functional compounds that may have significant applications in redox biology. The resorcinarene derivatives are efficient DPPH radical scavengers and were found to have SOD-like activity. The ability of resorcinarene to quench the free radicals is attributed to the intramolecular hydrogen bonding of these compounds. Calixtyrosol and calixpyrrole derived compounds were found to be excellent ROS modulators. However, these classes of macromolecules warrant further investigation in the development of therapeutically useful antioxidant compounds. The cucurbituril scaffold has been used for the development of molecular containers for various applications, including the enhancement of the radical-scavenger abilities of known antioxidants. The inclusion complexes of cucurbituril with molecules like resveratrol already proved beneficial to increase the reductive power of resveratrol and similar compounds.

Furthermore, the enormous study on the reaction mechanisms for the generation of novel derivatives, the cucurbituril scaffold, can be fine-tuned for the desired purposes. Hence, to see the novel antioxidants based on the cucurbituril scaffold in the near future is highly awaited. Among all supramolecules described in this review, porphyrins and their metal-complexed analogues are studied extensively; both in vitro and in vivo experiments for the radical-scavenging activities. As mentioned earlier, the porphyrin derivatives mimic the SOD and catalase like activities and thus have a great potential to be used therapeutically.

Supramolecular Scaffold	Compound	DPPH Radical Scavenging (%)	k_{inh}/k_p (M ⁻¹ S ⁻¹) ^a	k_{SOD} Activity (M ⁻¹ s ⁻¹)	Ref.
Calix[n]arene	7	-	7.25×10^{5}	-	F 4 4 1
	8	-	5.75×10^{5}	-	[44]
	18	82.6	-	-	[45]
	38	^b 71.7	-		[50]
	39	^b 63.2	-		[52]
Resorcinarene	50	$^{\rm c}$ 0.19 \pm 0.04	-	-	
	52	$^{\rm c}$ 0.18 ± 0.01	-	-	[67]
	54	86.3	-	-	
	57	84.9	-	-	[70]
	60	85.0	-	-	
Calixtyrosol	64	Significant	-	-	[72]
Calixpyrrole	70	76.4%	-	-	[77]
Cucurbit[n]uril	77–79	^d Significant	-	-	[87]
	MndMIm4P@CB[10];Im	- -	-	1×10^{7}	[88]
Porphyrin	80	-	-	5.4×10^{5}	[92]
	86	74.3	-	-	
	88	79.5	-	-	[20]
	90	77.36	-	-	[96]
	113	83.40	-	-	[106]

Table 1. Antioxidant activity of selected derivatives of calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, and porphyrin.

^a k_{inth} and k_p —rate constant of inhibition of free radical and propagation rate constant of the chain reaction, respectively. ^b [•OH] radical scavenging (%) activity. DPPH = 2,2-diphenyl-1-picrylhydrazyl radical. ^c ECR₅₀ = [scavenger]/[DPPH] producing 50% scavenging of DPPH after 5min reaction time. ^d antioxidant activity was higher than resveratrol.

4. Conclusions

In conclusion, supramolecular chemistry offers several classes of macrocycles, including calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, cucurbit[n]uril, and porphyrin. There are several reports on the modification reactions of these molecules to product functional compounds of desired properties. However, the application of these macromolecular scaffolds for the development of potent radical scavengers with pharmacological activities are not studied to its full potential. Therefore, this review elaborated on the derivatives of various supramolecules and their antioxidant activities. Porphyrin derivatives have a significant amount of research on their radical-scavenging abilities and reaction mechanisms supporting them. Similar to porphyrins, the calix[n]arene, resorcinarene, calixtyrosol, calixpyrrole, and cucurbit[n]uril derivatives offer great potential for the development of ROS modulators with significant pharmacological activities to prevent and treat the disorders resulting from oxidative stress.

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References

- 1. Reuter, S.; Gupta, S.C.; Chaturvedi, M.M.; Aggarwal, B.B. Oxidative stress, inflammation, and cancer: How are they linked? *Free Radic. Biol. Med.* **2010**, *49*, 1603–1616. [CrossRef]
- 2. Saha, S.K.; Lee, S.B.; Won, J.; Choi, H.Y.; Kim, K.; Yang, G.; Dayem, A.A.; Cho, S. Correlation between Oxidative Stress, Nutrition, and Cancer Initiation. *Int. J. Mol. Sci.* **2017**, *18*, 1544. [CrossRef] [PubMed]
- 3. Ramachandra, C.J.A.; Hernandez-Resendiz, S.; Crespo-Avilan, G.E.; Lin, Y.; Hausenloy, D.J. Mitochondria in acute myocardial infarction and cardioprotection. *EBioMedicine* **2020**, *57*, 102884. [CrossRef] [PubMed]

- Neuzil, J.; Rayner, B.S.; Lowe, H.C.; Witting, P.K. Oxidative stress in myocardial ischaemia reperfusion injury: A renewed focus on a long-standing area of heart research. *Redox Rep.* 2005, 10, 187–197. [CrossRef] [PubMed]
- Zhang, Y.; Murugesan, P.; Huang, K.; Cai, H. NADPH oxidases and oxidase crosstalk in cardiovascular diseases: Novel therapeutic targets. *Nat. Rev. Cardiol.* 2020, *17*, 170–194. [CrossRef]
- 6. Elahi, M.M.; Kong, Y.X.; Matata, B.M. Oxidative stress as a mediator of cardiovascular disease. *Oxid. Med. Cell Longev.* **2009**, *2*, 259–269. [CrossRef]
- 7. Peoples, J.N.; Saraf, A.; Ghazal, N.; Pham, T.T.; Kwong, J.Q. Mitochondrial dysfunction and oxidative stress in heart disease. *Exp. Mol. Med.* **2019**, *51*, 1–13. [CrossRef]
- 8. Federico, A.; Morgillo, F.; Tuccillo, C.; Ciardiello, F.; Loguercio, C. Chronic inflammation and oxidative stress in human carcinogenesis. *Int. J. Cancer* **2007**, *121*, 2381–2386. [CrossRef]
- 9. Yang, X.; Li, Y.; Li, Y.; Ren, X.; Zhang, X.; Hu, D.; Gao, Y.; Xing, Y.; Shang, H. Oxidative Stress-Mediated Atherosclerosis: Mechanisms and Therapies. *Front. Physiol.* **2017**, *8*, 600. [CrossRef]
- 10. Liguori, I.; Russo, G.; Curcio, F.; Bulli, G.; Aran, L.; Della-Morte, D.; Gargiulo, G.; Testa, G.; Cacciatore, F.; Bonaduce, D.; et al. Oxidative stress, aging, and diseases. *Clin. Interview Aging* **2018**, *13*, 757–772. [CrossRef]
- Pizzino, G.; Irrera, N.; Cucinotta, M.; Pallio, G.; Mannino, F.; Arcoraci, V.; Squadrito, F.; Altavilla, D.; Bitto, A. Oxidative Stress: Harms and Benefits for Human Health. *Oxid. Med. Cell Longev.* 2017, 2017, 8416763. [CrossRef] [PubMed]
- Valko, M.; Leibfritz, D.; Moncol, J.; Cronin, M.T.D.; Mazur, M.; Telser, J. Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* 2007, 39, 44–84. [CrossRef] [PubMed]
- 13. Nimse, S.B.; Pal, D. Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Adv.* **2015**, *5*, 27986–28006. [CrossRef]
- 14. Shi, D.; Yang, J.; Jiang, Y.; Wen, L.; Wang, Z.; Yang, B. The antioxidant activity and neuroprotective mechanism of isoliquiritigenin. *Free Radic. Biol. Med.* **2020**, *152*, 207–215. [CrossRef] [PubMed]
- 15. Warraich, U.; Hussain, F.; Kayani, H.U.R. Aging-Oxidative stress, antioxidants and computational modeling. *Heliyon* **2020**, *6*, e04107. [CrossRef] [PubMed]
- 16. Čolak, E.; Ignjatović, S.; Radosavljević, A.; Žorić, L. The association of enzymatic and non-enzymatic antioxidant defense parameters with inflammatory markers in patients with exudative form of age-related macular degeneration. *J. Clin. Biochem. Nutr.* **2017**, *60*, 100–107. [CrossRef]
- 17. Patlevič, P.; Vašková, J.; Švorc, P., Jr.; Vaško, L.; Švorc, P. Reactive oxygen species and antioxidant defense in human gastrointestinal diseases. *Integr. Med. Res.* **2016**, *5*, 250–258. [CrossRef]
- Hamley, I.W. Small Bioactive Peptides for Biomaterials Design and Therapeutics. *Chem. Rev.* 2017, 117, 14015–14041. [CrossRef]
- 19. Sindhi, V.; Gupta, V.; Sharma, K.; Bhatnagar, S.; Kumari, R.; Dhaka, N. Potential applications of antioxidants—A review. *J. Pharm. Res.* 2013, *7*, 828–835. [CrossRef]
- Panieri, E.; Buha, A.; Telkoparan-Akillilar, P.; Cevik, D.; Kouretas, D.; Veskoukis, A.; Skaperda, Z.; Tsatsakis, A.; Wallace, D.; Suzen, S. Luciano Saso 1Potential Applications of NRF2 Modulators in Cancer Therapy. *Antioxidants* 2020, *9*, 193. [CrossRef]
- 21. Yang, B.; Chen, Y.; Shi, J. Reactive Oxygen Species (ROS)-Based Nanomedicine. *Chem. Rev.* 2019, 119, 4881–4985. [CrossRef] [PubMed]
- 22. Ingold, K.U.; Pratt, D.A. Advances in Radical-Trapping Antioxidant Chemistry in the 21st Century: A Kinetics and Mechanisms Perspective. *Chem. Rev.* **2014**, *114*, 9022–9046. [CrossRef] [PubMed]
- 23. Morry, J.; Ngamcherdtrakul, W.; Yantasee, W. Oxidative stress in cancer and fibrosis: Opportunity for therapeutic intervention with antioxidant compounds, enzymes, and nanoparticles. *Redox Biol.* **2017**, *11*, 240–253. [CrossRef] [PubMed]
- Wang, J.; Luo, B.; Li, X.; Lu, W.; Yang, J.; Hu, Y.; Huang, P.; Wen, S. Inhibition of cancer growth in vitro and in vivo by a novel ROS-modulating agent with ability to eliminate stem-like cancer cells. *Cell Death Dis.* 2017, *8*, e2887. [CrossRef] [PubMed]
- 25. Hussain, H.H.; Babic, G.; Durst, T.; Wright, J.S.; Flueraru, M.; Chichirau, A.; Chepelev, L.L. Development of novel antioxidants: Design, synthesis, and reactivity. *J. Org. Chem.* **2003**, *68*, 7023–7032. [CrossRef] [PubMed]
- 26. López-Nicolás, J.M.; Rodríguez-Bonilla, P.; García-Carmona, F. Cyclodextrins and Antioxidants. *Crit. Rev. Food Sci. Nutr.* **2014**, *54*, 251–276. [CrossRef]

- 27. Wang, H.; Wang, S.; Zhu, H.; Wang, S.; Xing, J. Inclusion Complexes of Lycopene and β-Cyclodextrin: Preparation, Characterization, Stability and Antioxidant Activity. *Antioxidants* **2019**, *8*, 314. [CrossRef]
- Du, J.; Zhao, L.; Yang, S.; Huang, Y.; Chi, S.; Ruan, Q.; Zheng, P.; Hu, R.; Zhao, Y. Preparation, characterization, solubilization and antioxidant activity of polyamine modified β-cyclodextrins with baicalein inclusion complexes. *J. Incl. Phenom. Macrocycl. Chem.* **2019**, *93*, 203–213. [CrossRef]
- 29. Xu, Z.; Lian, X.; Li, M.; Zhang, X.; Wang, Y.; Tao, Z.; Zhang, Q. Effects of inclusion of chrysin in cucurbit [8] uril on its stability, solubility and antioxidant potential. *Chem. Res. Chin. Univ.* **2017**, *33*, 736–741. [CrossRef]
- Chao, J.; Wang, X.; Xu, M.; Zuo, Y. Characterization and enhanced antioxidant activity of the inclusion complexes of baicalin with p-sulfonatocalix [n] arenes. J. Incl. Phenom. Macrocycl. Chem. 2019, 93, 361–370. [CrossRef]
- 31. Gutsche, C.D. Calixarenes an Introduction, 2nd ed.; The Royal Society of Chemistry: Cambridge, UK, 2008.
- 32. Vicens, J.; Boehmer, V. Calixarenes: A Versatile Class of Macrocyclic Compounds; Springer: Dordrecht, The Netherlands, 1991.
- Nimse, S.B.; Kim, T. Biological Applications of Functionalized Calixarenes. *Chem. Soc. Rev.* 2013, 42, 366–386. [CrossRef] [PubMed]
- 34. Nimse, S.B.; Song, K.; Jung, C.; Eoum, W.; Kim, T. Molecular recognition of neutral substrates by new tetraaminocalix [4] arene derivative. *Bull. Korean Chem. Soc.* **2009**, *30*, 1247–1251.
- 35. Nimse, S.B.; Song, K.; Kim, J.; Kim, H.; Nguyen, V.; Eoum, W.; Jung, C.; Ta, V.; Kim, T. Aminocalix [4] arene: The effect of pH on the dynamics of gate and portals on the hydrophobic cavity. *Tetrahedron Lett.* **2010**, *51*, 6156–6160. [CrossRef]
- Nimse, S.B.; Kim, J.; Song, K.; Kim, J.; Lee, J.T.; Nguyen, V.; Ta, V.; Kim, T. Selective recognition of the ditopic trimethylammonium cations by water-soluble aminocalix [4] arene. *Tetrahedron Lett.* 2011, 52, 3751–3755. [CrossRef]
- 37. Seiffarth, K.; Schulz, M.; Goermar, G.; Bachmann, J. Calix [n] arenes—New light stabilizers for polyolefins. *Polym. Degrad. Stab.* **1989**, *24*, 73–80. [CrossRef]
- 38. Feng, W.; Yuan, L.H.; Zheng, S.Y.; Huang, G.L.; Qiao, J.L.; Zhou, Y. The effect of p-tert-butylcalix [n] arene on γ-radiation degradation of polypropylene. *Rad. Phys. Chem.* **2000**, *57*, 425–429. [CrossRef]
- 39. Gorghiu, L.M.; Dumitrescu, C.; Olteanu, R.L.; Bumbac, M.; Jipa, S. Comparative Study of Some Calix[n] arenes and other Phenolic Compounds Influence on the thermal Stability of LDPE. *Rom. Rep.* **2004**, *56*, 466–472.
- 40. Kikuzaki, H.; Hisamoto, M.; Hirose, K.; Akiyama, K.; Taniguchi, H. Antioxidant properties of ferulic acid and its related compounds. *J. Agric. Food Chem.* **2002**, *50*, 2161–2168. [CrossRef]
- 41. Chen, J.; Yang, J.; Ma, L.; Li, J.; Shahzad, N.; Kim, C.K. Structure-antioxidant activity relationship of methoxy, phenolic hydroxyl, and carboxylic acid groups of phenolic acids. *Sci. Rep.* **2020**, *10*, 5666. [CrossRef]
- 42. Wrigstedt, P.; Kylli, P.; Pitkänen, L.; Nousiainen, P.; Tenkanen, M.; Sipilä, J. Synthesis and Antioxidant Activity of Hydroxycinnamic Acid Xylan Esters. *J. Agric. Food Chem.* **2010**, *58*, 6937–6943. [CrossRef]
- 43. Nimse, S.B.; Pal, D.; Mazumder, A.; Mazumder, R. Synthesis of Cinnamanilide Derivatives and their Antioxidant and Antimicrobial Activity. *J. Chem.* **2015**, 2015, 208910. [CrossRef]
- 44. Consoli, G.M.L.; Galante, E.; Daquino, C.; Granata, G.; Cunsolo, F.; Geraci, C. Hydroxycinnamic acid clustered by a calixarene platform: Radical scavenging and antioxidant activity. *Tetrahedron Lett.* **2006**, *47*, 6611–6614. [CrossRef]
- Patel, M.B.; Modi, N.R.; Raval, J.P.; Menon, S.K. Calix [4] arene based 1,3,4-oxadiazole and thiadiazole derivatives: Design, synthesis, and biological evaluation. *Org. Biomol. Chem.* 2012, 10, 1785–1794. [CrossRef] [PubMed]
- James, E.; Eggers, P.K.; Harvey, A.R.; Dunlop, S.A.; Fitzgerald, M.; Stubbs, K.A.; Raston, C.L. Antioxidant phospholipid calix [4] arene mimics as micellular delivery systems. *Org. Biomol. Chem.* 2013, *11*, 6108–6112. [CrossRef]
- Kumar, R.; Sharma, A.; Singh, H.; Suating, P.; Kim, H.S.; Sunwoo, K.; Shim, I.; Gibb, B.C.; Kim, J.S. Revisiting Fluorescent Calixarenes: From Molecular Sensors to Smart Materials. *Chem. Rev.* 2019, 119, 9657–9721. [CrossRef]
- Kumar, R.; Lee, Y.O.; Bhalla, V.; Kumar, M.; Kim, J.S. Recent developments of thiacalixarene based molecular motifs. *Chem. Soc. Rev.* 2014, 43, 4824–4870. [CrossRef]

- 49. Pur, F.N.; Dilmaghani, K.A. New Antiradical Clusters Synthesized Using the First Green Biginelli Reactions of Calix [4] arene. *Pharm. Chem. J.* 2016, *50*, 80–82. [CrossRef]
- 50. Stephens, E.K.; Tauran, Y.; Coleman, A.W.; Fitzgerald, M. Structural requirements for anti-oxidant activity of calix [n] arenes and their associated anti-bacterial activity. *Chem. Commun.* **2015**, *51*, 851–854. [CrossRef]
- 51. Lu, L.; Ruan, Z.; Ni, J.; Chen, J.; Shu, H.; Wang, Y.; Liu, Y. Improvement of antioxidative activity of resveratrol by calix [4] arene-like tetramer: A theoretical study. *Comput. Theor. Chem.* **2019**, *1148*, 1–7. [CrossRef]
- 52. Ni, J.; Lu, L.; Liu, Y. Antiradical and Antioxidative Activity of Azocalix [4] arene Derivatives: Combined Experimental and Theoretical Study. *Molecules* **2019**, *24*, 485. [CrossRef]
- 53. Nimse, S.B.; Song, K.; Kim, J.; Ta, V.; Nguyen, V.; Kim, T. Synthesis and Modification of Novel Iminecalix [4] arene Derivatives. *Bull. Korean Chem. Soc.* **2011**, *32*, 1143–1145. [CrossRef]
- 54. Nimse, S.B.; Kim, J.; Ta, V.; Kim, H.; Song, K.; Jung, C.; Nguyen, V.; Kim, T. New water-soluble iminecalix [4] arene with a deep hydrophobic cavity. *Tetrahedron Lett.* **2009**, *52*, 7346–7350. [CrossRef]
- 55. Silva, C.M.; da Silva, D.L.; Magalhães, T.F.F.; Alves, R.B.; de Resende-Stoianoff, M.A.; Martins, F.T.; Fátima, A. Iminecalix [4] arenes: Microwave-assisted synthesis, X-ray crystal structures, and anticandidal activity. *Arabian J. Chem.* 2019, 12, 4365–4376. [CrossRef]
- Oguz, M.; Kalay, E.; Akocak, S.; Nocentini, A.; Lolak, N.; Boga, M.; Yilmaz, M.; Supuran, C.T. Synthesis of calix [4] azacrown substituted sulphonamides with antioxidant, acetylcholinesterase, butyrylcholinesterase, tyrosinase and carbonic anhydrase inhibitory action. *J. Enz. Inhib. Med. Chem.* 2020, *35*, 1215–1223. [CrossRef] [PubMed]
- 57. Aoyama, Y.; Tanaka, Y.; Toi, H.; Ogoshi, H. Polar host–guest interaction. Binding of nonionic polar compounds with a resorcinol-aldehyde cyclooligomer as a lipophilic polar host. *J. Am. Chem. Soc.* **1988**, *110*, 634–635. [CrossRef]
- Gropp, C.; Quigley, B.L.; Diederich, F. Molecular Recognition with Resorcin [4] arene Cavitands: Switching, Halogen-Bonded Capsules, and Enantioselective Complexation. J. Am. Chem. Soc. 2018, 140, 2705–2717. [CrossRef]
- 59. MacGillivray, L.R.; Atwood, J.L. A chiral spherical molecular assembly held together by 60 hydrogen bonds. *Nature* **1997**, *389*, 469–472. [CrossRef]
- 60. Shivanyuk, A.; Rebek, J. Assembly of Resorcinarene Capsules in Wet Solvents. *J. Am. Chem. Soc.* **2003**, 125, 3432–3433. [CrossRef]
- 61. Zhang, Q.; Catti, L.; Tiefenbacher, K. Catalysis inside the Hexameric Resorcinarene Capsule. *Acc. Chem. Res.* **2018**, *51*, 2107–2114. [CrossRef]
- 62. Avram, L.; Cohen, Y.; Rebek, J. Recent advances in hydrogen-bonded hexameric encapsulation complexes. *Chem. Commun.* **2011**, 47, 5368–5375. [CrossRef]
- 63. Payne, D.T.; Webre, W.A.; Matsushita, Y.; Zhu, N.; Futera, Z.; Labuta, J.; Jevasuwan, W.; Fukata, N.; Fossey, J.S.; D'Souza, F.; et al. Multimodal switching of a redox-active macrocycle. *Nat. Commun.* **2019**, *10*, 1007. [CrossRef] [PubMed]
- 64. Soule, B.P.; Hyodo, F.; Matsumoto, K.; Simone, N.L.; Cook, J.A.; Krishna, M.C.; Mitchell, J.B. The chemistry and biology of nitroxide compounds. *Free Radic. Biol. Med.* **2007**, *42*, 1632–1650. [CrossRef] [PubMed]
- 65. Samuni, A.M.; Barenholz, Y. Site-activity relationship of nitroxide radical's antioxidative effect. *Free Radic. Biol. Med.* **2003**, *34*, 177–185. [CrossRef]
- 66. Damiani, E.; Castagna, R.; Astolfi, P.; Greci, L. Aromatic and aliphatic mono- and bis-nitroxides: A study on their radical scavenging abilities. *Free Radic. Res.* **2005**, *39*, 325–336. [CrossRef]
- 67. Vovk, A.I.; Shivanyuk, A.M.; Bugas, R.V.; Muzychka, O.V.; Melnyk, A.K. Antioxidant and antiradical activities of resorcinarene tetranitroxides. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 1314–1317. [CrossRef] [PubMed]
- 68. Ngurah, B.I.G.M. Synthetic C-Methoxyphenyl Calix [4] Resorcinarene and Its Antioxidant Activity. J. Appl. Chem. Sci. 2018, 5, 403–408. [CrossRef]
- Oliveira, C.B.S.; Meurer, Y.S.R.; Oliveira, M.G.; Medeiros, W.M.T.Q.; Silva, F.O.N.; Brito, A.C.F.; Pontes, D.L.; Andrade-Neto, V.F. Comparative Study on the Antioxidant and Anti-Toxoplasma Activities of Vanillin and Its Resorcinarene Derivative. *Molecules* 2014, *19*, 5898–5912. [CrossRef]
- Abosadiya, H.M.; Hasbullah, S.A.; Mackeen, M.M.; Low, S.C.; Ibrahim, N.; Koketsu, M.; Yamin, B.M. Synthesis, Characterization, X-ray Structure and Biological Activities of C-5-Bromo-2-hydroxyphenylcalix [4]-2-methyl Resorcinarene. *Molecules* 2013, *18*, 13369–13384. [CrossRef]

- 71. Pur, N.F. Calixdrugs: Calixarene-based clusters of established therapeutic drug agents. *Mol. Divers.* **2016**, *20*, 781–787.
- 72. Pur, N.F.; Dilmaghani, K.A. Calixtyrosol: A Novel Calixarene Based Potent Radical Scavenger. *Iranian J. Pharma. Res.* 2015, 14, 1181–1187.
- 73. Pinalli, R.; Pedrini, A.; Dalcanale, A. Biochemical sensing with macrocyclic receptors. *Chem. Soc. Rev.* 2018, 47, 7006–7026. [CrossRef] [PubMed]
- 74. Kim, D.S.; Sessler, J.L. Calix [4] pyrroles: Versatile molecular containers with ion transport, recognition, and molecular switching functions. *Chem. Soc. Rev.* **2015**, *44*, 532–546. [CrossRef] [PubMed]
- 75. Gale, P.A.; Anzenbacher, P.; Sessler, J.L. Calixpyrroles II. Coord. Chem. Rev. 2001, 222, 57–102. [CrossRef]
- 76. Kongor, A.; Panchal, M.; Mehta, V.; Bhatt, K.; Bhagat, D.; Tipre, D.; Jain, V.K. Basketing nanopalladium into calix [4] pyrrole as an efficient catalyst for Mizoroki-Heck reaction. *Arab. J. Chem.* 2017, 10, 1125–1135. [CrossRef]
- 77. Kongor, A.; Panchal, M.; Athar, M.; Makwana, B.; Sindhav, G.; Jha, P.C.; Jain, V. Synthesis and modeling of calix [4] pyrrole wrapped Au nanoprobe for specific detection of Pb(II): Antioxidant and radical scavenging efficiencies. *J. Photochem. Photobiol. A Chem.* **2018**, *364*, 801–810. [CrossRef]
- 78. Freeman, W.A.; Mock, W.L.; Shih, N.Y. Cucurbituril. J. Am. Chem. Soc. 1981, 103, 7367–7368. [CrossRef]
- 79. Kim, J.; Jung, I.-S.; Kim, S.-Y.; Lee, E.; Kang, J.-K.; Sakamoto, S.; Yamaguchi, K.; Kim, K. New Cucurbituril Homologues: Syntheses, Isolation, Characterization, and X-ray Crystal Structures of Cucurbit [n] uril (n = 5, 7, and 8). *J. Am. Chem. Soc.* **2000**, *122*, 540–541. [CrossRef]
- 80. Day, A.I.; Blanch, R.J.; Arnold, A.P.; Lorenzo, S.; Lewis, G.R.; Dance, I. A Cucurbituril-Based Gyroscane: A New Supramolecular Form. *Angew. Chem., Int. Ed.* **2002**, *41*, 275–277. [CrossRef]
- 81. Barrow, S.J.; Kasera, S.; Rowland, M.J.; Barrio, J.; Scherman, O.A. Cucurbituril-Based Molecular Recognition. *Chem. Rev.* **2015**, *115*, 12320–12406. [CrossRef]
- 82. Murray, J.; Kim, K.; Ogoshi, T.; Yao, W.; Gibb, B.C. The aqueous supramolecular chemistry of cucurbit [n] urils, pillar [n] arenes and deep-cavity cavitands. *Chem. Soc. Rev.* **2017**, *46*, 2479–2496. [CrossRef]
- 83. Day, A.; Arnold, A.P.; Blanch, R.J.; Snushall, B. Controlling Factors in the Synthesis of Cucurbituril and Its Homologues. *J. Org. Chem.* **2001**, *66*, 8094–8100. [CrossRef] [PubMed]
- 84. Lee, J.W.; Samal, S.; Selvapalam, N.; Kim, H.; Kim, K. Cucurbituril Homologues and Derivatives: New Opportunities in Supramolecular Chemistry. *Acc. Chem. Res.* **2003**, *36*, 621–630. [CrossRef] [PubMed]
- 85. Lagona, J.; Mukhopadhyay, P.; Chakrabarti, S.; Isaacs, L. The Cucurbit [n] uril Family. *Angew. Chem.* 2005, 117, 4922–4949. [CrossRef]
- Hou, C.; Li, J.; Zhao, L.; Zhang, W.; Luo, Q.; Dong, Z.; Xu, J.; Liu, J. Construction of Protein Nanowires through Cucurbit [8] uril-based Highly Specific Host–Guest Interactions: An Approach to the Assembly of Functional Proteins. *Angew. Chem. Int. Ed. Engl.* 2013, *52*, 5590–5593. [CrossRef]
- 87. Lu, L.; Zhu, S.; Zhang, H.; Li, F.; Zhang, S. Theoretical study of complexation of resveratrol with cyclodextrins and cucurbiturils: Structure and antioxidative activity. *RSC Adv.* **2015**, *5*, 14114–14122. [CrossRef]
- Kubota, R.; Takabe, T.; Arima, K.; Taniguchi, H.; Asayama, S.; Kawakami, H. New class of artificial enzyme composed of Mn-porphyrin, imidazole, and cucurbit [10] uril toward use as a therapeutic antioxidant. *J. Mater. Chem. B* 2018, *6*, 7050–7059. [CrossRef]
- 89. Tian, J.; Huang, B.; Nawaz, M.H.; Zhang, W. Recent advances of multi-dimensional porphyrin-based functional materials in photodynamic therapy. *Coord. Chem. Rev.* **2020**, *420*, 213410. [CrossRef]
- 90. Lee, H.; Hong, K.I.; Jang, W.D. Design and applications of molecular probes containing porphyrin derivatives. *Coord. Chem. Rev.* **2018**, 354, 46–73. [CrossRef]
- 91. Singh, S.; Aggarwal, A.; Bhupathiraju, N.V.S.D.K.; Arianna, G.; Tiwari, K.; Drain, C.M. Glycosylated Porphyrins, Phthalocyanines, and Other Porphyrinoids for Diagnostics and Therapeutics. *Chem. Rev.* **2015**, *115*, 10261–10306. [CrossRef]
- Mackensen, G.B.; Patel, M.; Sheng, H.; Calvi, C.L.; Batinic'-Haberle, I.; Day, B.J.; Liang, L.P.; Fridovich, I.; Crapo, J.D.; Pearlstein, R.D.; et al. Neuroprotection from Delayed Postischemic Administration of a Metalloporphyrin Catalytic Antioxidant. *J. Neurosci.* 2001, *21*, 4582–4592. [CrossRef]
- Asayama, S.; Mori, T.; Nagaoka, S.; Kawakami, H. Chemical modification of manganese porphyrins with biomolecules for new functional antioxidants. *J. Biomater. Sci. Polym. Ed.* 2003, 14, 1169–1179. [CrossRef] [PubMed]

- 94. Fadda, A.A.; El-Mekawy, R.E.; El-Shafei, A.; Freeman, H.S.; Hinks, D.; El-Fedawy, M. Design, Synthesis, and Pharmacological Screening of Novel Porphyrin Derivatives. *J. Chem.* **2013**, 340230. [CrossRef]
- 95. Fadda, A.A.; El-Mekawy, R.E.; El-Shafei, A.I.; Freeman, H. Synthesis and pharmacological screening of novel meso-substituted porphyrin analogs. *Arch. Pharm. (Weinheim)* **2013**, *346*, 53–61. [CrossRef] [PubMed]
- 96. Tyurin, V.Y.; Zhang, J.; Moiseeva, A.A.; Milaeva, E.R.; Belykh, D.V.; Buravlev, E.V.; Rocheva, T.K.; Chukicheva, I.Y.; Kuchin, A.V. Comparative study of redox characteristics and antioxidant activity of porphyrins with 2,6-dialkylphenol groups. *Dokl. Chem.* **2013**, *450*, 152–155. [CrossRef]
- 97. Kubota, R.; Imamura, S.; Shimizu, T.; Asayama, S.; Kawakami, H. Synthesis of Water-Soluble Dinuclear Mn-Porphyrin with Multiple Antioxidative Activities. *ACS Med. Chem. Lett.* **2014**, *5*, 639–643. [CrossRef]
- Batinic-Haberle, I.; Tovmasyan, A.; Spasojevic, I. An educational overview of the chemistry, biochemistry and therapeutic aspects of Mn porphyrins—From superoxide dismutation to H₂O₂-driven pathways. *Redox Biol.* 2015, *5*, 43–65. [CrossRef]
- 99. Tesakova, M.V.; Parfenyuk, V.I. The Electrochemical Evaluation of the Antioxidant Activity of Substituted Tetraphenylporphyrins. *Russ. J. Electrochem.* **2017**, *53*, 1281–1285. [CrossRef]
- 100. Abu-Melha, S. Efficient synthesis of meso-substituted porphyrins and molecular docking as potential new antioxidant and cytotoxicity agents. *Arch. Pharm. Chem. Life Sci.* **2018**, 352, 1800221. [CrossRef]
- Shaaban, S.; Negm, A.; Sobh, M.A.; Wessjohann, L.A. Expeditious Entry to Functionalized Pseudo-peptidic Organoselenide Redox Modulators via Sequential Ugi/SN Methodology. *Anti-Cancer Agents Med. Chem.* 2016, 16, 621–632. [CrossRef]
- 102. Zhang, J.; Wang, A.; Zhao, W.; Li, C.; Chen, X.; Wang, Y.; Zhu, W.; Zhong, Q. Influence of metal-porphyrins on the photocatalysis of graphitic carbon nitride. *Dyes Pigment.* **2018**, *153*, 241–247. [CrossRef]
- 103. Shabangu, S.M.; Babu, B.; Soy, R.C.; Managa, M.; Sekhosana, K.E.; Nyokong, T. Photodynamic antimicrobial chemotherapy of asymmetric porphyrin-silver conjugates towards photoinactivation of Staphylococcus aureus. J. Coord. Chem. 2020, 73, 593–608. [CrossRef]
- 104. Munechika, R.; Biju, V.; Takano, Y.; Harashima, H.; Yamada, Y. The optimization of cancer photodynamic therapy by utilization of a pi-extended porphyrin-type photosensitizer in combination with MITO-Porter. *Chem. Commun.* **2020**, *56*, 1145–1148.
- 105. Zhang, Q.; He, J.; Yu, W.; Li, Y.; Liu, Z.; Zhou, B.; Liu, Y. A promising anticancer drug: A photosensitizer based on the porphyrin skeleton. *RSC Med. Chem.* **2020**, *11*, 427–437. [CrossRef]
- 106. Ahmed, A.; Omar, W.A.E.; El-Asmy, H.A.; Abou-Zeid, L.; Fadda, A.A. Docking studies, antitumor and antioxidant evaluation of newly synthesized porphyrin and metalloporphyrin derivatives. *Dyes Pigment*. 2020, 108728. [CrossRef]



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