Investigating White Matter Perfusion Using Optimal Sampling Strategy Arterial Spin Labeling at 7 Tesla

Alexander G. Gardener and Peter Jezzard*

Purpose: Cerebral blood flow (CBF) is an informative physiological marker for tissue health. Arterial spin labeling (ASL) is a noninvasive MRI method of measuring this parameter, but it has proven difficult to measure white matter (WM) CBF due to low intrinsic contrast-to-noise ratio compared with gray matter (GM). Here we combine ultra-high field and optimal sampling strategy (OSS) ASL to investigate WM CBF in reasonable scan times.

Methods: A FAIR-based ASL sequence at 7T was combined with a real-time-feedback OSS technique, to iteratively improve post-label image acquisition times (TIs) on a tissueand subject-specific basis to obtain WM CBF quantification.

Results: It was found 77% of WM voxels gave a reasonable CBF fit. Averaged WM CBF for these voxels was found to be 16.3 ± 1.5 mL/100 g/min (discarding partial-volumed voxels). The generated TI schedule was also significantly different when altering the OSS weighted-tissue-mask, favoring longer TIs in WM.

Conclusion: WM CBF could be reasonably quantified in over 75% of identified voxels, from a total preparation and scan time of 15 min. OSS results suggest longer TIs should be used versus general GM ASL settings; this may become more important in WM disease studies. **Magn Reson Med 73:2243–2248, 2015.** © 2014 The Authors. Magnetic Resonance in Medicine published by Wiley Periodicals, Inc. on behalf of International Society for Magnetic Resonance in Medicine. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

Key words: ASL; white matter; CBF, perfusion; ultra-high field

INTRODUCTION

MRI arterial spin labeling (ASL) is a noninvasive method of labeling arterial blood to investigate cerebral blood flow (CBF) (1–3). Tissue perfusion is an important physiological marker and ASL has applications in studying function and pathology. Following the labeling RF pulse (pulsed ASL, PASL) or labeling period (continuous ASL, CASL), there is a time delay (TI) before acquisition of images superior to labeled blood vessels, to allow perfusion into the tissue. An otherwise-matched control image (without labeling) is acquired on alternate acquisitions. On comparing the labeled (tag) and nonlabeled (control) images the difference is found to depend on voxel perfusion. Using calibration scans, acquiring data at multiple TIs and postprocess fitting to a suitable model, voxel-by-voxel quantification of CBF and blood arterial transit time (ATT) can be achieved (4). However, most ASL studies focus on the investigation of gray matter (GM) tissue (5–7).

The study of white matter (WM) CBF by ASL has proven challenging, mainly due to low intrinsic difference signal (Δ M) between control and tag images, which limits the contrast-to-noise ratio (CNR). From literature the WM Δ M signal is reported to be 2–4 times smaller than GM Δ M, with arterial arrival times also longer, leading to increased relaxation (and signal loss) of labeled blood before arrival and perfusion into WM tissue (8). There are also concerns that partial voluming of GM within tissue voxels leads to over-estimation of the WM CBF. Nevertheless, there is increasing interest in using ASL for white matter disease studies and so robust quantification methods are highly desirable, for instance in WM leukoaraiosis and Alzheimer's disease (9–11).

Recent studies have investigated developments in labeling sequences, such as the pseudo-Continuous ASL (pCASL) technique (12). In one study, pCASL was used to investigate the number of acquisitions of single postlabel delay time (PLD) images and effect on WM CNR (13); in another, multiple-PLD pCASL acquisition was combined with fine detail ROI analysis (14). Furthermore, PASL acquisition, combined with single-voxel spectroscopic-like localization, has been suggested to overcome CNR and arrival time limitations (15). With the availability of ultrahigh field (UHF) human scanners an alternative approach is available, to take advantage of higher SNR and longer T₁ blood relaxation times at these field strengths, which benefit ASL (16,17). Coupled with an optimal sampling strategy (OSS) approach, to optimize the selection of TI times for WM CBF measurement using real-time feedback (18), the aim of this study was to demonstrate WM CBF quantification and acquire data in a timeframe suitable for future multi-parameter white matter disease investigations.

METHODS

Scanning was performed using a 7 Tesla (T) Siemens whole-body scanner (Siemens Healthcare, Erlangen, Germany), equipped with a combined head-only 32-channel receive and single-channel transmit coil (Nova Medical, Wilmington, MA). The physical dimensions of this coil

FMRIB Centre, Nuffield Department of Clinical Neurosciences, University of Oxford, John Radcliffe Hospital, Oxford, United Kingdom.

^{*}Correspondence to: Peter Jezzard, Ph.D., FMRIB Centre, Nuffield Department of Clinical Neurosciences, John Radcliffe Hospital, Oxford, OX3 9DU UK. E-mail: peterj@fmrib.ox.ac.uk

Additional Supporting Information may be found in the online version of this article.

Received 23 December 2013; revised 2 June 2014; accepted 3 June 2014 DOI 10.1002/mrm.25333

Published online 20 June 2014 in Wiley Online Library (wileyonlinelibrary.co-m).

^{© 2014} The Authors. Magnetic Resonance in Medicine published by Wiley Periodicals, Inc. on behalf of International Society for Magnetic Resonance in Medicine. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.



to the subject's neck, with an inside diameter of 290 mm and length 280 mm; the receive coil had diameter 180 mm and height 240 mm. Due to ultra-high field $B_0/$ B_1 profile and efficiency issues (16,19), a multiple-TI Flow-sensitive Alternating Inversion Recovery (FAIR) Quantitative Imaging of Perfusion Using a Single Subtraction (QUIPSS2) PASL sequence was used for the study (3,17,20). In FAIR a nonselective inversion pulse is centered on the image volume as the tag condition; a selective inversion covering the image slab is the control, so inflowing blood is not inverted. Subtraction of the two leaves a perfusion-weighted signal difference, as static tissue in the image is the same in both. Frequency-Offset Corrected Inversion (FOCI) label pulses, and Shinnar-Le Roux (SLR)-optimized saturation pulses (for image pre- and postlabel saturation and QUIPSS2), were used in this implementation (21-23). Pulse efficiencies were assessed using a Polydimethylsiloxane (PDMS) oil phantom (T_1 of 1400 ms at 7T). These were found to be better than 95% of desired flip-angle across spatial profile, at up to ± 10 cm from B₀ isocenter. The sharpness of the inversion and saturation pulse edges was less than 1 voxel (3 mm) width for ramp, for up to 20 cm pulse thickness (nonselective inversion thickness).

For this study an OSS approach was used with FAIR, to iteratively improve TI_2 values as the scan progressed using

real-time feedback, optimizing delay time choice (18,24). In OSS-ASL, N pairs of control-tag images (where N equals number of TIs) are acquired, with an improved schedule of N TI_2s generated from included voxels (24). This is fed back to the scanner control, updating sequence timings, and the new TI_2 set acquired. This process repeats to generate an ever-improving TI_2 set. All generation and feedback functions ran on the image-processing pipeline on the reconstruction computer, running in the background whilst data were acquired as per normal operation.

Eight healthy adult male subjects were scanned (ages 33 ± 5 years), giving informed consent under an institutional development protocol. Unless noted all imaging was performed at a resolution of $3 \times 3 \times 5 \text{ mm}^3$ with a 64^2 matrix, with six axial slices (2 mm slice gap) acquired in ascending order. Gradient-echo EPI acquisition (6/8 partial k-space coverage) was used, with flowcrushing gradients in the slice-direction (critical velocity of 5 cm/s) to suppress unwanted large vessel signal. Double inversion recovery (DIR) images were acquired before the OSS-ASL scans to generate voxel masks for the OSS process. These were thresholded to binary masks and stored in scanner memory until used by OSS. Three approaches were used; (A) WM voxel mask (TR 30 s; TI values of 0, 4300 ms, acquire at 5400 ms); (B) GM voxel mask (TR 30 s; TI values of 0, 3560 ms, acquire at 4220 ms); and (C) a WM-GM mask constructed from the WM-

Table 1

Subject and	Averaged CBF	Values from BA	ASIL Fits for:	GM, All	(WM-GM);	Significant	(z-stat >2) (WM-GM); a	and Lastly	All (z	z-stat >	>2)
WM Voxels, v	which Includes	Some GM Par	tial-Voluming	Effects								

Subject	1	2	3	4	5	6	7	8	Mean CBF (mL/100 g/min)
GM CBF	43.3	33.6	51.2	48.8	69.3	57.6	58.2	43.4	50.7 ± 11.0
(WM-GM) CBF	13.2	14.0	13.9	13.6	14.5	12.8	14.2	12.2	13.6 ± 0.8
(WM-GM) z>2 CBF	16.3	15.4	16.5	16.6	19.6	15.0	16.3	14.7	16.3 ± 1.5
WM z>2 CBF	17.9	16.6	17.4	18.5	20.1	16.1	17.4	17.5	17.7 ± 1.2

mask minus overlapping GM voxels, to minimize partial volume effects. Representative DIR images and masks are presented in Figure 1. DIR timings were generated from T₁-based simulations, and were adjusted at the scanner for the best tissue contrast (25). For FAIR scans a QUIPSS2 pulse was applied at 0.7 s postlabel inversion (TI_1) . A long TI_1 would limit the minimum achievable TI_2 value, whilst a shorter TI_1 might reduce the amount of tagged blood delivered to the brain; values of 0.6 to 1.2 s have been suggested (20). Pre/postinversion saturation pulses were used to reduce potential $B_1\ {\rm profile}$ errors. TE was set as short as possible at 18 ms, including time for flow-crushers. The TR was 4 s, which gave 95% SAR at 7T; this limited the maximum number of slices. The selective inversion was wider than the imaging slab to prevent interference from poor pulse profile; effective gap between bolus edge and the inferior slice was 20 mm. The OSS TI₂ search space was set to 0.8 s to 2.55 s, to fill the TR period and allow OSS flexibility in optimization fits for either tissue. Eight initial TI₂ values were acquired in a block at the start of each OSS-ASL, evenly spaced every 0.25 s; for subsequent blocks OSSgenerated TI2 values were used. For each of the OSS scans 160 images (10 blocks) were acquired, taking 11 min in total per ASL scan. An unprepared M₀ image was acquired for CBF quantification, with a TR of 20 s but matching ASL scan parameters. With the preparation (DIR), calibration (M_0) and tissue-weighted OSS scans, total duration per subject was approximately 45 min.

Analysis

Data were processed off-line using FSL tools (26) and Matlab (Mathworks, Natick, MA) to prepare ΔM timeseries. Motion-correction and registration to M₀ calibration image, brain extraction and multi-channel coil correction were performed before pair-wise control-tag subtraction. The "Bayesian Inference for Arterial Spin Labeling" (BASIL) tool was used to fit ΔM data to a single-compartment ASL kinetic model (voxel-by-voxel) using all TI₂ values from each scan (4,27). The effects of acquisition delay on the TI₂ value for each slice and the WM and GM T₁ relaxation times at 7T were accounted for in fitting (tissue T₁ 1.4 and 1.9 s, respectively) (28,29). The M_{0-blood} term in the standard model was approximated from M_{0-csf}, measured in the ventricles (corrected for T₂^{*} decay and blood:water partition coefficient) (30).

Fitted parameters were investigated using ROIs generated from thresholded, brain-extracted DIR images registered to M_0 images. Voxels found to be in GM and WMthresholded ROIs were discarded to avoid partial volume contamination. BASIL-generated z-statistics for the CBF fit were used as a marker of goodness-of-fit to the model parameters (27,31). It was found that the third slice acquired contained the highest number of WM-voxels. For consistency this slice was used to investigate a representative ATT for labeled blood. The OSS-generated TI_2 sets were also compared for the different tissue weightings.

RESULTS

Table 1 presents mean CBF values for tissue ROIs, averaged over all slices for subjects, then across subjects. The 'GM' row indicates gray matter voxels without overlapping WM voxels; "(WM-GM)" includes all voxels identified from the WM minus the GM mask; "(WM-GM) z>2" used the same base ROI, but only includes voxels with a fitted CBF z-stat greater than 2 (approximately 95% confidence in fitted value, P = 0.05). Across subjects this was found to be $77 \pm 8\%$ of "(WM-GM)" voxels; these are referred to as "significant (WM-GM)" voxels. The mean CBF of WM increased by 2.7 mL/100 g/min when poorly fitted voxels were excluded (second versus third row of Table 1). The increase in subject 5 CBF after discarding poorly fitting voxels accounts for the increased standard deviation in the significant (WM-GM) voxels. The bottom row presents average CBF for the whole WM mask (z-stat > 2), including any overlapping GM voxels. This is significantly higher (P < 0.01) across all subjects than the significant (WM-GM) CBF, likely due to partial-volume GM contamination. DIR images show good demarcation between GM and WM, checked against MNI-152 library segmentations. These results emphasize the importance of voxel consideration in ROI analysis. It would be worthwhile investigating WM CBF across a wider sample set to improve the precision of these values (32). There was greater variation across subject GM CBF values, with two subjects having particularly high and low outliers. Excluding these outliers in the average GM CBF gave a similar mean but reduced standard deviation (to $50.4 \pm 6.0 \text{ mL}/100 \text{ g/min}$).

Figure 2 presents representative CBF maps for 3 slices, and Figure 3 presents the same images but with perfusion scaled specifically to a WM CBF range for clarity (with any value above 35 mL/100 g/min shown as white). The two tissue regions exhibit fairly even CBF distributions. Figure 4 presents representative fitted ATT maps, to match Figure 2; slice 3 is the right-most image. Longer ATT values are seen in WM regions, as expected. Representative fitted ATT values are presented in the Supporting Information (Supporting Table S1, which is available online) for a single slice (matching slice across subjects). As expected, significant (WM-GM) ATT values are longer than those found in GM by an average 0.19 s, reflecting later arrival of labeled blood to WM tissue.



FIG. 2. Fitted representative CBF maps for a GM-like perfusion range of 0 to 120 mL/ 100 g/min.

This would be difficult to observe in a single-TI ASL experiment; it may also vary in WM-disease studies from aberrant (faster or slower) blood flow (33).

Supporting Figure S1 plots the real-time generated OSS TI_2 distributions for the final OSS TI_2 block, averaged across subjects for each OSS (tissue) type. It shows longer TI_2 values are preferred for WM OSS, and longer still for (WM-GM) OSS, likely reflecting the later arrival of labeled blood. This causes a shift of the kinetic model peak; the OSS algorithm then favors a later acquisition window versus GM OSS. Using a paired two-tailed Student's t-test there was found to be significant difference between the (WM-GM) and GM OSS TI_2 distributions (P < 0.01), suggesting OSS was masking-tissue dependent and showing sensitivity to blood delivery.

DISCUSSION

This study shows it is possible to obtain CBF and ATT values for WM at 7T using an OSS FAIR-QUIPSS2 PASL sequence, in 15 min of acquisition including preparation, calibration and 11 min of ASL scanning. Prior work had attempted to investigate WM perfusion using PASL at 3T, but it was found that WM CBF fits were poor even with 20 min of data acquisition; over 80% of voxels were not fitted or had low z-stat, suggesting uncertainty in the CBF values obtained. The combination of ultrahigh field and OSS proved a powerful tool in this initial study, with over three quarters of identified WM voxels having what were considered reasonable CBF fits. The parameter values found are comparable to literature, with a significant (WM-GM) CBF value that is slightly

lower than in previous studies, reflecting care in selecting ROI voxels (avoiding partial volume effects) and discarding 23% of poor CBF-fitted voxels. The ratio of GM to significant (WM-GM) CBF values was found to be 3.1:1, falling within a range from previous reports [ratios: 2.5:1 to 5:1 (8,13–15,33–35)]. ATT values showed a difference of approximately 0.2 s between GM and significant (WM-GM), similar to previously reported; due to specific ASL settings this difference is a more useful marker than absolute ATT value (14,33,35,36).

Using OSS gave information on suitable TI₂ ranges for WM ASL compared with GM ASL. What may be optimal acquisition times for one tissue may not be optimal for another. There is little currently in the literature exploring this, especially at ultra-high field-strength where T₁ values are longer. Often, protocols are based on lowerfield studies and used with little modification. Furthermore, there is an argument that CBF fits from ΔM benefits from having a large set of delay times, against repeat acquisitions at a few TI₂ values (27,37). OSS intrinsically provides such data, with subject- and tissue-specific TI₂ values generated during acquisition. The results suggest for healthy adults a range of 1.1 to 2.3 s is suitable for WM, which is a longer than is typically used in lower field / GM-tissue studies. A range of 1.0 to 2.0 s is suggested for GM at 7T; theory suggests a peak around 1.8 s to match tissue/blood T1 values. This study used a relatively high number of TI2 values (eight) to give the OSS a large data set for its optimization. It would be worth investigating if a smaller number of TIs and acquisition blocks resulted in similar TIs and WM CBF values; this would allow a reduction in ASL scan time, to move



FIG. 3. The same fitted CBF images as Figure 2, but rescaled to a more WM-like perfusion range of 0 to 35 mL/100 g/min to highlight WM tissue regions.



FIG. 4. Fitted representative ATT maps, slices to match Figures 2 and 3; displayed on a range of 0 to 1.2 s.

toward a clinically useful total time (ideally less than 10 min for everything).

The resolution of images acquired in this study kept EPI acquisitions short with lower TE, and provided more signal per voxel. We assessed CNR from mean difference signal divided by background region standard deviation; at max signal (3rd/4th TI typically) the ratio of GM:WM CNR was 3.1:1; by the 8th TI this was 2.4:1, showing a greater proportional loss in GM CNR (and signal), but still remaining above WM CNR. The limit of viable CNR for longer TIs, such as for reduced or delayed perfusion disease studies, is an area of interest in future studies. Higher resolution requires additional data acquisition due to lower CNR, and results in longer TEs, introducing more BOLD signal contamination. At 7T the optimal TE for BOLD shortens to 25-30 ms from 30-50 ms at 3T, depending on region. Parallel imaging was not used in this study, as it provided no reduction in TE and complicated real-time processing; however, it may reduce TE in a high-resolution study. One advantage of highresolution acquisition would be to further improve ROI selection, limiting partial volume contamination and allowing regional or structural ROI analysis.

The potential of OSS-ASL in studying WM perfusion in ageing and/or disease is an area of particular interest. Such studies may benefit from OSS adapting to unusual blood dynamics in subjects, with optimal TI₂s varying from healthy subjects; this information may not be known before scanning. Using a wide TI₂ search space and allowing OSS sufficient optimization runs should provide subject- and/or pathological-optimized TI₂ values. It is worth noting that OSS ΔM images retain all ASL perfusion information compared with standard ASL, whilst offering additional information for little additional cost. Postprocessing is similar to standard ASL analysis, substituting individual TI₂ values for each ΔM image rather than repeated blocks of TI₂ values.

The current lack of a scanner body-transmit coil, or a dedicated neck-labeling coil, prevented implementation of a higher CNR OSS-pCASL technique in this study; the latter coil is under development. This would potentially allow another method of obtaining high-resolution ASL imaging; alternatively, pCASL could reduce the number of acquisitions needed per scan, making the shorter pCASL acquisition more clinically useful, though this risks reducing the optimization power of OSS if too few blocks are generated. A problem that remains with ultrahigh field systems is SAR; this is not limited to ASL. Parallel transmit methods may offer future solutions to this issue, as well as offering improved label pulses, or novel methods of vessel suppression.

ACKNOWLEDGMENTS

We acknowledge the UK Medical Research Council (MRC) for grant funding, and colleague James Meakin for assistance with sequence coding. We acknowledge the Dunhill Medical Trust for salary support (P.J.).

REFERENCES

- Detre JA, Zhang WG, Roberts DA, Silva AC, Williams DS, Grandis DJ, Koretsky AP, Leigh JS. Tissue-specific perfusion imaging using arterial spin-labeling. NMR Biomed 1994;7:75–82.
- Edelman RR, Siewert B, Darby DG, Thangaraj V, Nobre AC, Mesulam MM, Warach S. Qualitative mapping of cerebral blood-flow and functional localization with echo-planar MR-imaging and signal targeting with alternating radio-frequency. Radiology 1994;192:513– 520.
- 3. Kim SG. Quantification of relative cerebral blood-flow change by flow-sensitive alternating inversion-recovery (fair) technique application to functional mapping. Magn Reson Med 1995;34:293–301.
- Buxton RB, Frank LR, Wong EC, Siewert B, Warach S, Edelman RR. A general kinetic model for quantitative perfusion imaging with arterial spin labeling. Magn Reson Med 1998;40:383–396.
- Deibler AR, Pollock JM, Kraft RA, Tan H, Burdette JH, Maldjian JA. Arterial spin-labeling in routine clinical practice, part 1: technique and artifacts. AJNR Am J Neuroradiol 2008;29:1228–1234.
- Petersen ET, Zimine I, Ho YCL, Golay X. Non-invasive measurement of perfusion: a critical review of arterial spin labelling techniques. Br J Radiol 2006;79:688–701.
- Wong EC. Quantifying CBF with pulsed ASL: technical and pulse sequence factors. J Magn Reson Imaging 2005;22:727–731.
- van Gelderen P, de Zwart JA, Duyn JH. Pittfalls of MRI measurement of white matter perfusion based on arterial spin labeling. Magn Reson Med 2008;59:788–795.
- Mandell DM, Han JS, Poublanc J, Crawley AP, Kassner A, Fisher JA, Mikulis DJ. Selective reduction of blood flow to white matter during hypercapnia corresponds with leukoaraiosis. Stroke 2008;39:1993– 1998.
- Kimura N, Nakama H, Nakamura K, Aso Y, Kumamoto T. Effect of white matter lesions on brain perfusion in Alzheimer's disease. Dement Geriatr Cogn Disord 2012;34:256–261.
- Huynh TJ, Murphy B, Pettersen JA, Tu H, Sahlas DJ, Zhang L, Symons SP, Black S, Lee TY, Aviv RI. CT perfusion quantification of small-vessel ischemic severity. AJNR Am J Neuroradiol 2008;29: 1831–1836.
- Dai W, Garcia D, de Bazelaire C, Alsop D. Continuous flow-driven inversion for arterial spin labeling using pulsed radio frequency and gradient fields. Magn Reson Med 2008;60:1488–1497.

- van Osch MJ, Teeuwisse WM, van Walderveen MA, Hendrikse J, Kies DA, van Buchem MA. Can arterial spin labeling detect white matter perfusion signal? Magn Reson Med 2009;62:165–173.
- 14. Lu K, Liu TT, Wong EC, Jung Y. Regional white matter perfusion measurement using an optimized pseudo-continuous ASL MRI. In Proceedings of the 17th Annual Meeting of ISMRM, Honolulu, Hawaii, USA, 2009. Abstract 1521.
- Pohmann R. Accurate, localized quantification of white matter perfusion with single-voxel ASL. Magn Reson Med 2010;64:1109–1113.
- Teeuwisse WM, Webb AG, van Osch MJR. Arterial spin labeling at ultra-high field: all that glitters is not gold. Int J Imaging Syst Technol 2010;20:62–70.
- Gardener AG, Gowland PA, Francis ST. Implementation of quantitative perfusion imaging using pulsed arterial spin labeling at ultrahigh field. Magn Reson Med 2009;61:874–882.
- Xie J, Gallichan D, Gunn RN, Jezzard P. Optimal design of pulsed arterial spin labeling MRI experiments. Magn Reson Med 2008;59: 826–834.
- Schepers J, van Osch MJP, Bartels LW, Heukels SN, Viergever MA, Nicolay K. The effect of B1 field inhomogeneity and the nonselective inversion profile on the kinetics of FAIR-based perfusion MRI. Magn Reson Med 2005;53:1355–1362.
- Wong EC, Buxton RB, Frank LR. Quantitative imaging of perfusion using a single subtraction (QUIPSS and QUIPSS II). Magn Reson Med 1998;39:702-708.
- Payne GS, Leach MO. Implementation and evaluation of frequency offset corrected inversion (FOCI) pulses on a clinical MR system. Magn Reson Med 1997;38:828-833.
- Holm DA, Sidaros K. Slice profile optimization in arterial spin labeling using presaturation and optimized RE pulses. Magn Reson Med 2006;24:1229–1240.
- Yongbi MN, Branch CA, Helpern JA. Perfusion imaging using FOCI RF pulses. Magn Reson Med 1998;40:938–943.
- 24. Xie J, Clare S, Gallichan D, Gunn RN, Jezzard P. Real-time adaptive sequential design for optimal acquisition of arterial spin labeling MRI data. Magn Reson Med 2010;64:203–210.
- Redpath TW, Smith FW. Use of a double inversion recovery pulse sequence to image selectively grey or white brain matter. Br J Radiol 1994;67:1258–1263.

- Smith SM, Jenkinson M, Woolrich MW, et al. Advances in functional and structural MR image analysis and implementation as FSL. Neuroimage 2004;23:S208–S219.
- Chappell MA, Groves AR, Whitcher B, Woolrich MW. Variational Bayesian inference for a nonlinear forward model. IEEE Trans Signal Process 2009;57:223–236.
- Bluestein KT, Pitt D, Knopp MV, Schmalbrock P. T1 and proton density at 7 T in patients with multiple sclerosis: an initial study. Magn Reson Imaging 2012;30:19–25.
- 29. Ikonomidou VN, van Gelderen P, de Zwart JA, Duyn JH. T1 measurements at 7T with application to tissue specific imaging. In Proceedings of the 14th Scientific Meeting of ISMRM, Seattle, Washington, USA, 2006. Abstract 920.
- Herscovitch P, Raichle ME. What is the correct value for the brainñblood partition coefficient for water. J Cereb Blood Flow Metab 1985; 5:65–69.
- MacIntosh BJ, Filippini N, Chappell MA, Woolrich MW, Mackay CE, Jezzard P. Assessment of arterial arrival times derived from multiple inversion time pulsed arterial spin labeling MRI. Magn Reson Med 2010;63:641–647.
- 32. Liu Y, Zhu X, Feinberg D, Guenther M, Gregori J, Weiner MW, Schuff N. Arterial spin labeling MRI study of age and gender effects on brain perfusion hemodynamics. Magn Reson Med 2012;68:912–922.
- 33. Paling D, Petersen ET, Tozer DJ, Altmann DR, Wheeler-Kingshott CA, Kapoor R, Miller DH, Golay X. Cerebral arterial bolus arrival time is prolonged in multiple sclerosis and associated with disability. J Cereb Blood Flow Metab 2014;34:34–42.
- 34. Parkes LM, Rashid W, Chard DT, Tofts PS. Normal cerebral perfusion measurements using arterial spin labeling: reproducibility, stability, and age and gender effects. Magn Reson Med 2004;51:736–743.
- Gunther M, Oshio K, Feinberg DA. Single-shot 3D imaging techniques improve arterial spin labeling perfusion measurements. Magn Reson Med 2005;54:491–498.
- Chen Y, Wang DJ, Detre J. Comparison of arterial transit times estimated using arterial spin labeling. Magn Reson Mater Phys Biol Med 2012;25:135–144.
- 37. Lee W, Janik R, Scouten A, Stefanovic B, Sled JG. Efficient sampling of early signal arrival for estimation of perfusion and transit time in whole-brain arterial spin labeling. Magn Reson Med 2012;68:179–187.