The fine line between lifespan extension and shortening in response to caloric restriction

Kirk Szafranski¹ and Karim Mekhail^{1,2,*}

¹Department of Laboratory Medicine and Pathobiology; Faculty of Medicine; University of Toronto; Toronto, ON Canada; ²Canada Research Chairs Program; Faculty of Medicine; University of Toronto; Toronto, ON Canada

Keywords: caloric restriction, sirtuins, TOR, genome stability, oxidative stress, autophagy

Abbreviations: AL, Ad libitum; BER, base excision repair; CR, calorie restriction; DSB, double stranded break; eNoSC, energydependent nucleolar silencing complex; ETC, electron transport chain; Idh2, isocitrate dehydrogenase 2; NER, nucleotide excision repair; NHEJ, non-homologous end joining; rDNA, ribosomal DNA; ROS, reactive oxygen species; Sir2, silent information regulator 2; SIRT1, Sirtuin 1; SIRT3, Sirtuin 3; SIRT6, Sirtuin 6; Sod2, superoxide dismutase 2; TOR, target of rapamycin; Vma, vacuolar membrane ATPase

Caloric restriction (CR) is generally linked to lifespan extension in various organisms and may limit age-associated diseases. Processes through which caloric restriction promotes lifespan include obesity-countering weight loss, increased DNA repair, control of ribosomal and telomeric DNA repeats, mitochondrial regulation, activation of antioxidants, and protective autophagy. Several of these protective cellular processes are linked to the suppression of TOR (target of rapamycin) or the activation of sirtuins. In stark contrast CR fails to extend or even shortens lifespan in certain settings. CR-dependent lifespan shortening is linked to weight loss in the non-obese, mitochondrial hyperactivity, genomic inflexibility, and several other processes. Deciphering the balance between positive and negative effects of CR is critical to understanding its ultimate impact on aging. This knowledge is especially needed in order to fulfill the promise of using CR or its mimetic drugs to counteract age-associated diseases and unhealthy aging.

Introduction

The existence of conserved longevity pathways may seem counterintuitive from an evolutionary perspective. This is because evolution selects for reproductive success rather than long life. In fact, a leading theory of the evolution of aging, antagonistic pleiotropy, even stipulates that genotypes promoting reproduction earlier in life actually accelerate the aging process later in life (Fig. 1).¹ Consistent with this notion, several species experience a decline in early reproductive rate under CR conditions.¹ In contrast, a recent report indicates that the same genes can confer high early-life fitness and long life.^{2,3} While future work is needed to improve our understanding of the evolution of aging and longevity, it is clear that various genetic and environmental

conditions can alter lifespan. One of the promises of the study of conserved aging and/or longevity pathways is that it will lead to applications that help us reduce the morbidity associated with age-related diseases as well as increase overall human lifespan.

It has long been appreciated that caloric restriction (CR) is a wide-ranging and potent anti-aging intervention. As early as 1935, it was observed that reduction of caloric intake relative to ad libitum (AL, or unrestricted feeding) results in an extension in the average and maximal lifespan of laboratory mice.⁴ Since then, similar findings have been reported for a diverse range of organisms including yeast, nematodes, fruit flies, fish, rats, mice, and dogs, among others.^{5,6} The discovery of such a potent antiaging intervention has set the stage for research into the biology of aging and its modulation by caloric restriction.⁵

However, contrary to the well-documented positive effects of caloric restriction, several studies reported caloric restriction to be neutral or even detrimental to lifespan. For example, studies have found that caloric restriction regimens fail to impact lifespan in rhesus monkeys,7 wild mice,8 medflies,9 an isolate of the nematode C. Remanai,10 the spider L. Hasselti,11 and some yeast strains.^{12,13} Even more striking, CR actually shortened lifespan in several models including houseflies,14 male butterflies,15 the rotifer Cephalodella sp.,16 ILSXISS mice strains,^{17,18} and some yeast strains (Table 1).¹³ The work done with ILSXISS mice is particularly poignant. A meta-analysis of all mice studies excluding the ILSXISS strains reveals an average CR-dependent increase in lifespan of 15%.19 When the ILSXISS strains are included in the meta-analysis the average increase in lifespan drops to 2.9%.19 The ILSXISS studies set CR at 60% of AL intake in agreement with common standards in the field but it remains possible that more or less substantial restrictions may promote the lifespan of both ILSXISS and other mice strains. Taken together, these studies indicate that standard caloric restriction regimens do not universally promote longevity in various organisms. In other words, several more variables may exist within the equation determining the impact of CR on lifespan than originally anticipated.

Caloric restriction has been proposed to impact lifespan by affecting genomic stability, autophagy, oxidative stress, nutrient

^{*}Correspondence to: Karim Mekhail; Email: karim.mekhail@utoronto.ca Submitted: 12/10/2013; Revised: 01/17/2014; Accepted: 01/21/2014; Published Online: 01/27/2014 http://dx.doi.org/10.4161/nucl.27929

intake, and body weight, among other processes. Here we briefly highlight some of these processes and assess how they may be differentially impacted by CR to either positively or negatively affect lifespan. For reviews more focused on fully deciphering aging processes, we refer the reader elsewhere.^{5,20-28}

Reduction of Body Weight under CR Can Positively or Negatively Affect Lifespan

A CR diet will tend to promote the loss of body mass. How this impacts lifespan depends on which tissues are catabolized and the starting weight of the individual.

As they start following CR regimens, obese individuals typically increase their lifespan as they lose fat mass. Simply put, this is because obesity is correlated with a number of age-associated pathologies such as cardiovascular disease and diabetes.^{29,30} Consistent with this rationale, in obese human males, CR reduces body fat while also significantly decreasing obesity-related pathologies such as high blood pressure and chronic inflammation.³¹ In addition, in obese mice, the combination of CR and omega-3-polyunsaturated fatty acid intake simultaneously counteracts adiposity and chronic inflammation.³² Thus, within obesity settings, CR-mediated weight loss is generally beneficial.

In contrast, fat loss under CR is linked to lifespan shortening in the non-obese. For example, in non-obese mice, CR-induced fat loss is inversely correlated with lifespan.³³ In addition, ageassociated pathologies such as respiratory disease correlated with body weight loss in non-obese humans although CR can lower biomarkers for cardiovascular disease in this segment of the population.³⁴⁻³⁶ Although losing body weight appears to lower lifespan in non-obese humans aged 50–70 even when health status is considered, it is unclear if this applies to other age cohorts.³⁴⁻³⁶ Together, these studies and rationales suggest that the starting weight of an individual may dictate whether CR-induced fat loss positively or negatively impacts lifespan.

In addition to fat modulation, CR can also decrease muscle mass. Increased muscle mass has been shown to have an important protective effect in several age-related diseases and losing muscle mass can therefore be deleterious.³⁷ Indeed, even in ILSXISS mice, lean/muscle body mass correlated positively with lifespan under CR conditions.³³ Taken together, these studies suggest that CR-triggered losses in muscle mass can shorten lifespan.

Therefore, the impact of CR-mediated weight loss on lifespan may be bidirectional dependent on the starting weight and tissue(s) catabolized. This indicates that there may be an optimal body mass density for lifespan. In humans, recent evidence has pointed toward this as being at the high end of what is typically considered a healthy weight, 22.5–25 kg/m².³⁸ Alternatively, it is possible that weight loss in and of itself is a stressor whose deleterious side effects on lifespan are only mitigated if the starting weight is significantly above the ideal. Either way, these findings clearly show that CR does not universally promote lifespan.

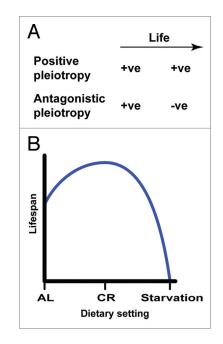


Figure 1. The evolution of aging and dietary effects on lifespan. (**A**) Positive and antagonistic pleiotropy theories suggest that alterations conferring advantages in early life respectively trigger beneficial and deleterious effects at the post-reproductive age. (**B**) Generalized relationships between diets and lifespan.

Nutrient Balance Affects the Response to CR

Interestingly, the magnitude and quality of weight loss may also depend on the nutritional composition of the CR diet itself. In fact, the very idea that it is solely the actual decrease in caloric content that accounts for CR-dependent lifespan modulation has been questioned. Instead, it may be that the varying restriction of nutrients in different diets, which may be commonly referred to as a CR diet, can positively or negatively impact lifespan depending on the particular nutrients affected.

Studies conducted in both flies and mice lend support to this notion. The fruit fly D. melanogaster can be subjected to a yeastbased or sugar-based diet. Decreased intake of yeast or sugar increases lifespan.³⁹ Interestingly, the positive effect on lifespan per calorie decreased was much more substantial under the yeast-based diet relative to the sugar-based counterpart.³⁹ This suggests that it is not simply the decreased caloric content per se that solely impacts lifespan. One possibility is that restriction of yeast-based diets also limits the amount of other nutrients within this diet. Consistent with this reasoning, methionine restriction significantly increases murine lifespan independently of caloric content.^{40,41} It is conceivable that the restriction of several aminoacids or nutrients would have similar effects. Therefore, in addition to altering overall caloric intake, certain CR diets may also extend lifespan via restriction of various lifespan-limiting dietary components or nutrients.

Specific nutritional composition may also explain contradictory results on the impact of different CR diets on lifespan. For example, one of many possible explanations for

Species Name	Common Name	Effect of CR on Lifespan	Reference
Saccharomyces cerevisiae	Budding yeast	Positive in majority of strains, negative in sod2 mutants, others	12–13
Caenorhabditis elegans	Worm	Positive	95–96
Caenorhabditis remanai	Worm	Neutral	10
Cephalodella sp.	Worm	Negative	16
Drosophila melanogaster	Fruitfly	Positive	39
Ceratitis capitata	Medfly	Neutral	9
Musca domestica	Housefly	Negative	14
Latrodectus hasseltii	Redback spider	Neutral	11
Frontinella pyramitela	Bowl and doily spider	Positive	125
Euphaedra sp.	Butterfly	Negative in males	15
Charaxes sp.	Butterfly	Negative in males	15
Rattus norvegicus	Brown rat	Positive	19
Mus musculus	House mouse	Positive in most strains, neutral in wild mice, negative in most ILSXISS strains	8, 17, 19
Canis lupus familiaris	Domesticated dog	Positive	126
Macaca mulatta	Rhesus monkey	Conflicting reports—appears to be neutral with good nutrition	7, 42

Table 1. The effect of CR on various species

these apparently contradictory results on the effect of CR on rhesus monkey lifespan may be the dietary composition; the study in which the monkeys responded to CR had AL diets with higher sucrose and lower antioxidants and omega 3 polyunsaturated acids.^{7,42} Not surprisingly, these data imply that different nutrients may promote or limit lifespan. Thus, different CR regimens with multiple variations in overall nutrient balance are very likely to trigger a wide range of responses with respect to lifespan. For example, a CR regimen that restricts methionine while maintaining antioxidants might extend lifespan while a regimen that limits omega-3-polyunsaturated acids while maintaining sucrose may shorten lifespan.

Overall, it is not surprising that the underlying nutritional composition of a diet influences the ultimate impact of CR regimens on lifespan. Future research in organisms with complex diets should carefully control for the nutritional composition of diets in order to accurately distinguish calorie-dependent from nutrient-dependent effects on lifespan. In addition, one should revisit some of the previously published CR studies to eliminate any potential confounding factors that may be linked to background nutrient deprivation or starvation. More specifically, experiments showing a decline in longevity in response to CR have not been generally performed across a range of nutrient levels and it remains possible that a number of these reports would have revealed enhanced longevity under similar caloric but different nutrient conditions.

Crosstalk between CR and Genome Stability

Beyond confounding dietary designs, CR certainly influences multiple processes operating at the cellular level.43-45 For example, CR can promote genome stability by sustaining DNA repair processes and also protecting repetitive DNA loci such as telomeres and rDNA. However, pre-existing conditions in these endogenous pathways can occur in certain genetic settings, in which case the effect of CR on lifespan may rapidly turn from positive to deleterious.

Connections between CR and several DNA repair processes do exist. One of the DNA repair pathways influenced by CR is base excision repair (BER), which repairs small non-helical distorting lesions in DNA. BER is the most commonly used DNA repair pathway in mammals.⁴⁶ BER declines with age but caloric restriction prevents such age-associated declines in mice. 47,48 This is likely linked to the ability of CR to promote rate limiting factors in the BER pathway. For example, CR increases the enzymatic activity of apyramidine/apurinic endonuclease as well as the expression of DNA polymerase β.47,49 Caloric restriction is also capable of activating nucleotide excision repair (NER), which repairs helical-distorting DNA damage caused by large bulky adducts.50 In mice, as with BER, NER rates decline with age but this decline is prevented by CR.⁵⁰ In addition, NER dysfunction is linked to skin cancer development and the premature aging disease xeroderma pigmentosum.⁵¹ Another DNA repair pathway in which CR may be implicated is non-homologous end joining (NHEJ), which repairs the majority of DNA double-stranded breaks (DSBs) in mammalian cells.⁴³ The autoantigen Ku is a DNA binding protein in the NHEJ pathway.⁵² Interestingly, Ku expression declines with age in rats but CR can counteract this phenomenon.53 Future work will show if CR impacts actual NHEJ rates and if this in turn directly affects lifespan. Taken together, CR may promote lifespan by promoting the lifelong maintenance of several DNA repair pathways.

Importantly, the effects of CR are not limited to DNA repair. In fact, CR also modulates processes that help prevent DNA damage from occurring in the first place. One area of intense investigation has been to understand the impact of CR on conserved repetitive DNA loci known to significantly affect

cellular lifespan from yeast to human.54 In particular, budding yeast has served as a highly valuable tool to decipher many conserved cellular aging mechanisms.⁵⁵⁻⁵⁸ Yeast lifespan can be analyzed both in terms of replicative lifespan (number of daughter cells produced by a new mother cell) and chronological lifespan (survival of non-dividing cells). Importantly, CR extends both types of yeast lifespan.⁵⁹ Lifespan of the budding yeast S. cerevisiae is highly dependent on the stability of repetitive DNA loci, in particular the rDNA (rDNA) repeats as well as the telomeres.^{57,60-63} Due to their highly repetitive nature, rDNA repeats are prone to recombination. While this can be protective under extreme stress conditions, aberrant or hyperactive recombination within the repeats generally leads to chromosome instability and shortens cellular lifespan.55,57,64 CR typically decreases recombination within the rDNA repeats via a form of rDNA silencing that represses intergenic RNA Pol II-dependent transcription and this extends lifespan.65-67 CR has been proposed to suppress recombination within rDNA repeats through multiple

mechanisms including the repression of the nutrient-sensing target of rapamycin (TOR) complex as well as activation of the conserved NAD+-dependent histone deacetylase Sir2 (silent information regulator 2).62,65,66 Interestingly, Sir2 is required for CR-dependent extension of replicative but not chronological lifespan.^{68,69} More recently, CR has also been proposed to increase lifespan by suppressing the activity of rDNA origins of replication preventing them from deleteriously competing with weaker replication origins elsewhere in the genome.⁷⁰ In human cells, the Sir2 homolog SIRT1 (Sirtuin 1) acts as a subunit of the eNoSC (energy-dependent nucleolar silencing complex) to ensure a form of rDNA silencing that represses RNA Pol I-dependent transcription in a glucose-dependent manner.⁷¹ SIRT1, which is also activated by CR, is linked to cell survival in human cells.⁷² It is likely that SIRT1-dependent rDNA silencing increases cellular lifespan by decreasing deleterious recombination, similarly to yeast.

On another front, telomeres are linear DNA sequences that are located at the ends of linear chromosomes and are amplified to prevent excessive chromosome shortening and subsequent genome destabilization during cell division.⁷³⁻⁷⁶ Telomeres also help propagate Sir2-dependent silent chromosome structures along nearby regions on chromosome arms.^{55,77,78} Importantly, it is clear that telomere length maintenance as well as downstream sirtuin-dependent silent chromatin assembly are both critical to lifespan maintenance.^{60,61,73,79} CR can promote subtelomeric silencing in yeast through Sir2 and this translates to a longer cellular lifespan.⁶² SIRT6 (Sirtuin 6), which is another human Sir2 homolog, also promotes telomeric silencing in human cells.⁸⁰ As CR increases SIRT6 levels, this suggests that CR may promote mammalian lifespan in part by increasing telomeric silencing.⁸¹ Furthermore, in mice and rats, a CR diet helps maintain the

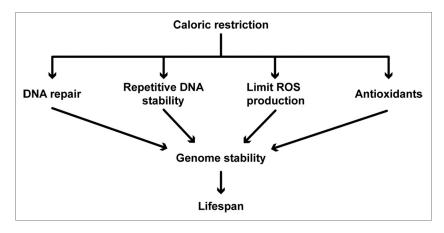


Figure 2. Calorie restriction (CR) increases lifespan by increasing genomic stability. Stability of DNA is maintained by increasing DNA repair pathways and controlling repetitive DNA loci. DNA repair pathways controlled by CR include base excision repair (BER), nucleotide excision repair (NER), and non-homologous end-joining (NHEJ). At the repetitive DNA loci, CR prevents deleterious recombination at the rDNA repeats, increases telomeric silencing, and maintains telomere length to increase lifespan. CR can also increase genomic stability by lowering the production of reactive oxygen species (ROS) as well as promoting the function of antioxidants (e.g., superoxide dismutase enzymes). CR may decrease ROS production by increasing mitochondrial efficiency or by decreasing mitochondrial membrane potential.

length of telomeres over the lifetime of the animal.^{82,83} Telomere length maintenance is a strong predictor of lifespan and it is thus likely that CR-dependent telomere length maintenance promotes lifespan.^{73,79} SIRT1 may also regulate telomere length and attenuate age-associated telomere shortening, suggesting that CR acts upstream of SIRT1 to regulate telomere length and promote lifespan.^{84,85} However, TOR also appears to be important for telomere length maintenance and lifespan in yeast.^{86,87} Therefore, it is likely that CR acts through both sirtuins and TOR modulation to in order to optimize lifespan-sustaining telomeric functions. It is important to note that additional CR-dependent processes maintaining rDNA/telomere stability may still exist as currently identified pathways only partly account for the beneficial effects of CR at these repetitive genomic loci. Together, current literature clearly indicates that CR activates processes operating at least at the repetitive DNA loci, rDNA, and telomeres, in order to prevent genome instability (Fig. 2). We also note that the dysregulation of other types of repetitive DNA sequences such as transposable elements have been linked to genome instability and aging.⁸⁸ It is therefore possible that CR may somehow control these elements in order to promote lifespan. Overall, CR is a potent genome maintenance intervention that both prevents DNA damage and promotes DNA repair.

Although these genome-stabilizing effects of CR can generally be viewed as beneficial, it is possible to imagine various settings in which they may ultimately shorten lifespan. For example, by decreasing DNA recombination capacity and genomic flexibility, cells often lose the ability to efficiently adapt to variable environmental conditions. In addition, cellular aging can be beneficial in multicellular organisms that need to balance new cell generation and old cell clearance within tissues and organs. Consistent with this rationale, it was recently discovered that senescence itself is a normal part of development.⁸⁹⁻⁹¹ This may eventually help explain how gestational CR is linked to accelerated aging in rats.⁹² It is also possible that hyperactivation of DNA repair processes too early in life may disrupt the fine line between telomere maintenance and DNA repair processes. This could in turn lead to the erroneous recognition of telomeres as broken DNA ends. Future studies possibly employing systems biology tools to assess relationships between various CR-dependent genome maintenance processes at different stages of life could help clarify these points.

CR-Autophagy Connections

Calorie restriction also increases autophagy, which is the mechanism responsible for catabolizing cellular components such as organelles by targeting them for lysosome-dependent degradation.^{93,94} While eliminating old cellular components that may be malfunctioning and/or cytotoxic, autophagy thereby also mobilizes energy reserves in times of stress.⁹³ Several studies suggest that CR may promote lifespan by operating in part through autophagy.

Consistent with this notion, CR mimetics fail to extend the lifespan of autophagy-deficient *C. Elegans.*⁹⁴⁻⁹⁶ In addition, *Arabidopsis* requires autophagy genes for lifespan extension under light restriction, which is the autotrophic analog of CR.⁹⁷ These data indicate that autophagy can mediate CR-dependent lifespan extension within various settings.

CR may promote autophagy through several avenues. Interestingly, these may implicate Sirtuin activation and TOR suppression.^{94,98} For example, absence of the essential autophagic modulator Beclin-1 abolishes the lifespan extension that has been observed in *C. elegans* upon overexpression of the sirtuin Sir2.1.⁹⁴ We note that these effects may reflect changes to processes that are independent of Sir2.1 itself, whose overexpression may not actually underlie the extended lifespan phenotypes initially reported.^{99,100} CR-dependent suppression of TOR also promotes autophagy in a variety of species and does so independently of sirtuins in *C. elegans.*^{94,98} This may be linked to the ability of TOR to suppress adenosine monophosphate-activated protein kinase, which is a potent activator of autophagy.¹⁰¹⁻¹⁰³ Thus, sirtuin activation and TOR suppression may partly underlie autophagy-dependent lifespan extension by CR.

Healthy aging is also thought to depend on the proper maintenance of adult stem cells.²⁶ Interestingly, autophagy is essential for the lifelong maintenance of hematopoietic stem cells and for supporting an old blood system.¹⁰⁴ This phenomenon appears to implicate a FOXO3A-dependent gene expression program and is activated by CR.

Overall, it is reasonable to conclude that autophagy underlies at least some of the beneficial effects of CR on lifespan. If specific autophagy-related processes can also be linked to the negative effects of CR on lifespan in certain settings remains unclear. One possibility may be that activating autophagy when autophagic vesicles cannot fuse with lysosomes such as in Danon disease would be deleterious as this leads to an accumulation of non-functional autophagic vesicles.¹⁰⁵ In fact, aberrant autophagy genes are also linked to several other diseases including cancer (ovarian, breast, prostate, and colon), autoimmune diseases (lupus), asthma, Crohn disease, and others.²⁵ This greatly increases the clinical settings in which CR-dependent activation of autophagy may simply exacerbate phenotypes.

Links between CR, Oxidative Stress, and Energy Metabolism

In addition to promoting the autophagy of organelles including mitochondria, CR can decrease oxidative stress through several distinct pathways. Oxidative stress in an organism is largely due to the accumulation of reactive oxygen species (ROS). By damaging nucleic acids, proteins, and other molecules, ROS decrease the lifespan of many organisms.^{106,107} CR can promote lifespan by both lowering the production of ROS as well as promoting the function of antioxidants that can repair ROS-induced damage. However, CR also promotes mitochondrial activity, which inevitably increases the chance of ROS production. Thus, a delicate balance must be maintained for CR to actually decrease ROS-induced damage and extend lifespan.

Antioxidants can scavenge ROS and generally maintain a reducing environment that promotes lifespan.¹⁰⁸ Importantly, CR promotes the function of several antioxidants. In mice, CR activates SIRT3 (Sirtuin 3), which in turn promotes the deacetylation and consequent activation of the antioxidant enzyme Sod2 (superoxide dismutase 2).108 SIRT3 activation also promotes the glutathione antioxidant Idh2 (isocitrate dehydrogenase 2), which decreases age-associated hearing loss in mice.¹⁰⁹ Therefore, CR may operate in part through sirtuins to promote the function of antioxidants and extend lifespan. As antioxidant activity counteracts the deleterious effects of ROS, CR-dependent upregulation of antioxidants can promote lifespan extension. Of note, several reports have suggested that the antioxidant-dependent impact of CR on lifespan may reflect tissue specific processes that can also differ between organisms.110-112

CR may also be capable of promoting lifespan by increasing mitochondrial efficiency and energy production.^{23,113} Indeed, CR increases mitochondrial respiration rates as well as the overall number of mitochondria in mouse cells.¹¹⁴ CR can also increase the number of mitochondria in human cells.¹¹⁵ While increased mitochondrial bioenergetics can be beneficial, mitochondria are in fact the major site of ROS production. So how can it be beneficial to increase mitochondrial function under CR? One explanation is that CR increases overall mitochondrial efficiency, thereby decreasing the number of electrons stalling in the electron transport chain (ETC) and preventing excessive ROS generation. Electrons stall in the ETC when their rate of entry exceeds their rate of transit.23 This then creates an environment that is prone to generate ROS.¹¹⁶ It was proposed that CR-dependent improvement of mitochondrial bioenergetics may prevent electron stalling by permitting mitochondria to simultaneously process a greater number of electrons.^{23,117}

Consistent with this possibility, CR-treated rats have decreased electron leaks within the ETC complex I.¹¹⁸ Thus, by increasing mitochondrial efficiency, CR may limit ROS-generating electron leaks and promote lifespan.

CR can reduce electron leak by maintaining low mitochondrial membrane potential.^{119,120} CR maintains a low membrane potential at least in part by regulating vacuolar pH. The latter increases with age and this lowers vacuolar storage capabilities.¹²¹ In turn, this increases the concentration of free cytoplasmic amino acids.¹²¹ It is thought that mitochondrial catabolism of cytoplasmic amino acids places a burden on mitochondrial carrier proteins and this in turn may overwhelm and increase mitochondrial membrane potential.^{121,122} CR helps maintain a low vacuolar pH over lifespan, possibly through the lifespan-modulating usual suspect TOR.¹²¹ Therefore, by maintaining a high vacuolar pH, CR can lower mitochondrial membrane potential, thereby decreasing ROS production and increasing lifespan. Taken together, these studies point to CR as a suppressor of ROS production as well as an activator of antioxidant processes.

However, activation of the mitochondria is not always beneficial. In yeast Sod2 knockout cells, the switch toward respiration under CR causes a massive decrease in lifespan.¹³ Similarly, CR shortens lifespan in mice with malfunctioning Sod1.¹²³ This suggests that the increase in antioxidant activity is critical to maintain lifespan under CR. Without it, the increase in mitochondrial activity will increase oxidative damage. This seems to occur even if CR can lower ROS generation during respiration in an antioxidant-independent fashion. Therefore, in these settings, the magnitude of oxidative damage caused by CR-mediated increases in respiration must be greater than the amount of oxidative damage decreased due to elevated mitochondrial efficiency and the maintenance of a low mitochondrial membrane potential.

Furthermore, CR shortens the lifespan of yeast cells lacking Vma (vacuolar membrane ATPase).^{13,121} Vma proteins are responsible for vacuolar H+-ATPase function and therefore maintain vacuolar acidity by proton transport. Thus, CR fails to acidify vacuoles and increase amino acid storage in Vmadeficient cells. This would then limit the ability of CR to lower mitochondrial membrane potential. The fact that CR shortens lifespan when its ability to lower mitochondrial membrane potential is impaired suggests that a low mitochondrial membrane potential is also required to compensate for the increased chance of ROS production in the presence of CR-driven mitochondrial activation (Fig. 3). Taken together, these data show that CR must maintain a low mitochondrial membrane potential and promote antioxidant functions in order to compensate for the elevation in ROS levels that typically occurs upon CR-dependent increases in mitochondrial activity.

Overall, these findings paint the picture of a very delicate balance between lifespan extension and suppression through CR's effect on mitochondria. Cells will increase respiration and mitochondrial number, likely to compensate for decreased energy intake. This then increases the probability of ROS production. CR compensates for this via activation of antioxidant proteins and increasing mitochondrial efficiency via modulation of

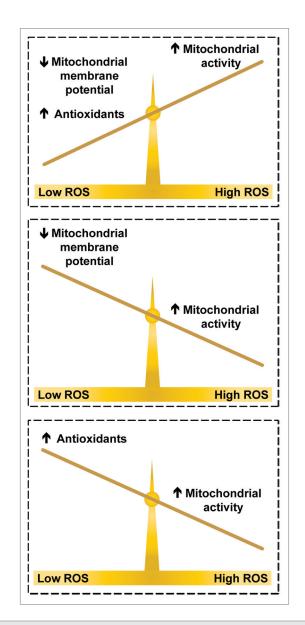


Figure 3. Calorie restriction (CR) influences reactive oxygen species (ROS) production through a delicate balance. When CR decreases mitochondrial membrane potential and increases antioxidant expression, ROS production is reduced relative to ad libitum, and lifespan may be increased (top scale). If either of these factors is absent, ROS production will be increased relative to ad libitum and lifespan will be decreased (bottom scales).

mitochondrial membrane potential. Overall, the combination of these changes allows CR to actually limit ROS-dependent damage. However, when CR is unable to affect antioxidants or mitochondrial membrane potential, ROS production is higher than in AL. Conditions that alter the ability of CR to positively affect mitochondrial membrane potential can thus switch the effect of CR on lifespan from positive to negative. Lifespan outcomes may also reflect the notion that the effect of CR on antioxidants can display tissue and organism specificities. Overall, this delicate balance further highlights how CR is a broad acting intervention that may just be the key to unlocking the mysteries of aging.

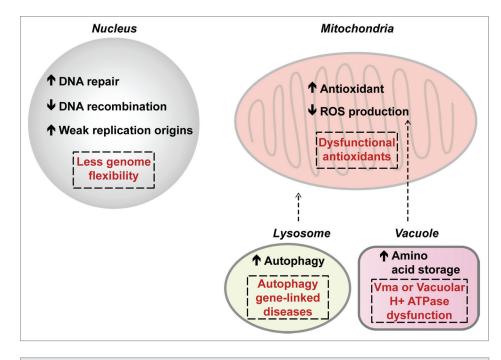


Figure 4. Summary of the effects of calorie restriction (CR) on various cellular components. CR increases DNA repair, promotes telomere length, decreases recombination particularly within repetitive DNA loci, and may also ensure the function of weak DNA replication origins. Taken together, the net effect of these changes is a decreased genomic flexibility, and this may in turn prevent cells from efficiently adapting to various stress conditions (nuclear dashed box). CR-dependent hyperactivation of DNA repair processes early in life may disrupt the balance between DNA repair and telomere maintenance. With regard to mitochondrial processes, CR increases antioxidant function and lowers membrane potential to lower ROS production even in the presence of CR-dependent mitochondrial hyperactivity. However, in settings where CR-dependent increases in mitochondrial activity are not mitigated by other processes such as in superoxide dismutase mutant, CR increases ROS-dependent damage (mitochondrial dashed box). In the vacuole, CR lowers pH, which promotes amino acid storage and may help lower mitochondrial membrane potential and overall ROS levels. Vacuolar defects can cause CR to have a net negative effect on lifespan (dashed box inside vacuole). In the lysosome, CR promotes an increase in autophagy, which can help eliminate old organelles, including dysfunctional mitochondria. This helps mobilize cellular energy stores and limit toxicity caused by defective organelles. Mutations within autophagy genes are linked to a large number of clinical settings and this can in turn cause partial CR-dependent activation of autophagic processes triggering toxicity and lowering lifespan (dashed box within lysosome).

Perspective

Calorie restriction has been proposed to impact lifespan by a great number of mechanisms, some of which are discussed here (Fig. 4). It is just emerging that various seemingly subtle changes in a genetic background can cause CR to have dramatically different consequences on lifespan. A recent study conducted in budding yeast indicates that the number of pathways the disruption of which causes the effects of CR on lifespan to change from neutral or positive to negative is substantial.¹³ These pathways include oxidative stress response, vacuolar function, and protein catabolism among others. It is important to note that different mutations impacting even the

same pathway or organelle may cause CR to have different effects depending on the specific mutation. For example, although CR lowers the lifespan of superoxide dismutase mutants, it is mainly mitochondrial mutants that were found to positively respond to CR.¹³ However, several of the mutants whose lifespan responds positively to CR may reflect the ability of this broad acting dietary intervention to activate processes that correct or counteract defects triggered by the initial mutation. For example, TOR inhibition, which is also achieved by CR, can alleviate mitochondrial disease in a mouse model of Leigh syndrome.124 In contrast, within the setting of other mitochondrial diseases or even clinical conditions linked to dysfunctional autophagy genes, CR may actually shorten lifespan. This is likely to be only the tip of the iceberg as one can also imagine that CR will also exert unpredictable effects on cells and/or organisms carrying multiple genetic alterations or polymorphisms. Thus, the task ahead of the CR and/ or aging field is really enormous. Luckily, the fact that CR is expected to have a wide range of consequences on lifespan also implies that this dietary intervention will be beneficial within large fractions of the human population, be they healthy or suffering from various diseases. As we often strive to decrease our caloric intake in today's health-conscious society, it will

also be just as important to identify those of us who may actually be harmed rather than helped by caloric restriction.

Disclosure of Potential Conflicts of Interest

No potential conflict of interest was disclosed.

Acknowledgments

We thank members of the Mekhail lab for discussions. In particular, we thank Jayesh S Salvi, Janet NY Chan, and Daniel Chung for comments. K.S. is supported by a University of Toronto Graduate Entrance Scholarship. The Mekhail laboratory is supported by the Canadian Institutes of Health Research as well as the Canada Research Chairs program.

References

- Partridge L, Piper MD, Mair W. Dietary restriction in Drosophila. Mech Ageing Dev 2005; 126:938-50; PMID:15935441; http://dx.doi.org/10.1016/j. mad.2005.03.023
- Maklakov AA. Aging: why do organisms live too long? Curr Biol 2013; 23:R1003-5; PMID:24262824; http://dx.doi.org/10.1016/j.cub.2013.10.002
- Kimber CM, Chippindale AK. Mutation, condition, and the maintenance of extended lifespan in Drosophila. Curr Biol 2013; 23:2283-7; PMID:24210612; http://dx.doi.org/10.1016/j. cub.2013.09.049
- McCay CM, Crowell MF, Maynard LA. The effect of retarded growth upon the length of life span and upon the ultimate body size. J Nutr 1935; 10:63-79
- Kaeberlein M. Lessons on longevity from budding yeast. Nature 2010; 464:513-9; PMID:20336133; http://dx.doi.org/10.1038/nature08981
- Weindruch R, Walford RL. The retardation of aging and disease by dietary restriction. Springfield, Ill., U.S.A.: C.C. Thomas, 1988.
- Mattison JA, Roth GS, Beasley TM, Tilmont EM, Handy AM, Herbert RL, Longo DL, Allison DB, Young JE, Bryant M, et al. Impact of caloric restriction on health and survival in rhesus monkeys from the NIA study. Nature 2012; 489:318-21; PMID:22932268; http://dx.doi.org/10.1038/ nature11432
- Harper JM. Wild-derived mouse stocks: an underappreciated tool for aging research. Age (Dordr) 2008; 30:135-45; PMID:19424863; http://dx.doi. org/10.1007/s11357-008-9057-0
- Carey JR, Liedo P, Harshman L, Zhang Y, Müller HG, Partridge L, Wang JL. Life history response of Mediterranean fruit files to dietary restriction. Aging Cell 2002; 1:140-8; PMID:12882344; http://dx.doi. org/10.1046/j.1474-9728.2002.00019.x
- Sutphin GL, Kaeberlein M. Dietary restriction by bacterial deprivation increases life span in wildderived nematodes. Exp Gerontol 2008; 43:130-5; PMID:18083317; http://dx.doi.org/10.1016/j. exger.2007.10.019
- Kasumovic MM, Brooks RC, Andrade MC. Body condition but not dietary restriction prolongs lifespan in a semelparous capital breeder. Biol Lett 2009; 5:636-8; PMID:19515652; http://dx.doi. org/10.1098/rsbl.2009.0335
- Kaeberlein M, Steffen KK, Hu D, Dang N, Kerr EO, Tsuchiya M, Fields S, Kennedy BK. Comment on "HST2 mediates SIR2-independent lifespan extension by calorie restriction". [author reply]. Science 2006; 312:1312, author reply 1312; PMID:16741098; http://dx.doi.org/10.1126/ science.1124608
- Schleit J, Johnson SC, Bennett CF, Simko M, Trongtham N, Castanza A, Hsieh EJ, Moller RM, Wasko BM, Delaney JR, et al. Molecular mechanisms underlying genotype-dependent responses to dietary restriction. Aging Cell 2013; 12:1050-61; PMID:23837470; http://dx.doi.org/10.1111/ acel.12130
- Cooper TM, Mockett RJ, Sohal BH, Sohal RS, Orr WC. Effect of caloric restriction on life span of the housefly, Musca domestica. FASEB J 2004; 18:1591-3; PMID:15319362
- Molleman F, Ding J, Boggs CL, Carey JR, Arlet ME. Does dietary restriction reduce life span in male fruitfeeding butterflies? Exp Gerontol 2009; 44:601-6; PMID:19580860; http://dx.doi.org/10.1016/j. exger.2009.06.008
- Weithoff G. Dietary restriction in two rotifer species: the effect of the length of food deprivation on life span and reproduction. Oecologia 2007; 153:303-8; PMID:17453249; http://dx.doi.org/10.1007/ s00442-007-0739-6

- Liao CY, Rikke BA, Johnson TE, Diaz V, Nelson JF. Genetic variation in the murine lifespan response to dietary restriction: from life extension to life shortening. Aging Cell 2010; 9:92-5; PMID:19878144; http://dx.doi. org/10.1111/j.1474-9726.2009.00533.x
- Rikke BA, Liao CY, McQueen MB, Nelson JF, Johnson TE. Genetic dissection of dietary restriction in mice supports the metabolic efficiency model of life extension. Exp Gerontol 2010; 45:691-701; PMID:20452416; http://dx.doi.org/10.1016/j. exger.2010.04.008
- Swindell WR. Dietary restriction in rats and mice: a meta-analysis and review of the evidence for genotype-dependent effects on lifespan. Ageing Res Rev 2012; 11:254-70; PMID:22210149; http:// dx.doi.org/10.1016/j.arr.2011.12.006
- Mostoslavsky R, Esteller M, Vaquero A. At the crossroad of lifespan, calorie restriction, chromatin and disease: meeting on sirtuins. Cell Cycle 2010; 9:1907-12; PMID:20458180; http://dx.doi. org/10.4161/cc.9.10.11481
- van Diepeningen AD, Maas MF, Huberts DH, Goedbloed DJ, Engelmoer DJ, Slakhorst SM, Koopmanschap AB, Krause F, Dencher NA, Sellem CH, et al. Calorie restriction causes healthy life span extension in the filamentous fungus Podospora anserina. Mech Ageing Dev 2010; 131:60-8; PMID:20026344; http://dx.doi.org/10.1016/j. mad.2009.12.002
- Fusco S, Maulucci G, Pani G. Sirt1: def-eating senescence? Cell Cycle 2012; 11:4135-46; PMID:22983125; http://dx.doi.org/10.4161/ cc.22074
- Guarente L. Mitochondria--a nexus for aging, calorie restriction, and sirtuins? Cell 2008; 132:171-6; PMID:18243090; http://dx.doi.org/10.1016/j. cell.2008.01.007
- Cornu M, Albert V, Hall MN. mTOR in aging, metabolism, and cancer. Curr Opin Genet Dev 2013; 23:53-62; PMID:23317514; http://dx.doi. org/10.1016/j.gde.2012.12.005
- Choi AM, Ryter SW, Levine B. Autophagy in human health and disease. N Engl J Med 2013; 368:1845-6; PMID:23656658; http://dx.doi.org/10.1056/ NEJMra1205406
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell 2013; 153:1194-217; PMID:23746838; http://dx.doi. org/10.1016/j.cell.2013.05.039
- Schreiber KH, Kennedy BK. When lamins go bad: nuclear structure and disease. Cell 2013; 152:1365-75; PMID:23498943; http://dx.doi.org/10.1016/j. cell.2013.02.015
- Liu J, Kim J, Oberdoerffer P. Metabolic modulation of chromatin: implications for DNA repair and genomic integrity. Front Genet 2013; 4:182; PMID:24065984; http://dx.doi.org/10.3389/ fgene.2013.00182
- Després JP. Health consequences of visceral obesity. Ann Med 2001; 33:534-41; PMID:11730160; http:// dx.doi.org/10.3109/07853890108995963
- Reilly JJ, Methven E, McDowell ZC, Hacking B, Alexander D, Stewart L, Kelnar CJ. Health consequences of obesity. Arch Dis Child 2003; 88:748-52; PMID:12937090; http://dx.doi. org/10.1136/adc.88.9.748
- 31. Kitada M, Kume S, Takeda-Watanabe A, Tsuda S, Kanasaki K, Koya D. Calorie restriction in overweight males ameliorates obesity-related metabolic alterations and cellular adaptations through antiaging effects, possibly including AMPK and SIRT1 activation. Biochim Biophys Acta 2013; 1830:4820-7; PMID:23800577; http://dx.doi.org/10.1016/j. bbagen.2013.06.014

- 32. Flachs P, Rühl R, Hensler M, Janovska P, Zouhar P, Kus V, Macek Jilkova Z, Papp E, Kuda O, Svobodova M, et al. Synergistic induction of lipid catabolism and anti-inflammatory lipids in white fat of dietary obese mice in response to calorie restriction and n-3 fatty acids. Diabetologia 2011; 54:2626-38; PMID:21779874; http://dx.doi.org/10.1007/ s00125-011-2233-2
- Liao CY, Rikke BA, Johnson TE, Gelfond JA, Diaz V, Nelson JF. Fat maintenance is a predictor of the murine lifespan response to dietary restriction. Aging Cell 2011; 10:629-39; PMID:21388497; http:// dx.doi.org/10.1111/j.1474-9726.2011.00702.x
- Fontana L, Meyer TE, Klein S, Holloszy JO. Longterm calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. Proc Natl Acad Sci U S A 2004; 101:6659-63; PMID:15096581; http://dx.doi.org/10.1073/pnas.0308291101
- Whitlock G, Lewington S, Sherliker P, Clarke R, Emberson J, Halsey J, Qizilbash N, Collins R, Peto R; Prospective Studies Collaboration. Body-mass index and cause-specific mortality in 900 000 adults: collaborative analyses of 57 prospective studies. Lancet 2009; 373:1083-96; PMID:19299006; http://dx.doi.org/10.1016/S0140-6736(09)60318-4
- Holloszy JO, Fontana L. Caloric restriction in humans. Exp Gerontol 2007; 42:709-12; PMID:17482403; http://dx.doi.org/10.1016/j. exger.2007.03.009
- Wisløff U, Najjar SM, Ellingsen O, Haram PM, Swoap S, Al-Share Q, Fernström M, Rezaei K, Lee SJ, Koch LG, et al. Cardiovascular risk factors emerge after artificial selection for low aerobic capacity. Science 2005; 307:418-20; PMID:15662013; http:// dx.doi.org/10.1126/science.1108177
- Whitlock G, Lewington S, Sherliker P, Clarke R, Emberson J, Halsey J, Qizilbash N, Collins R, Peto R; Prospective Studies Collaboration. Body-mass index and cause-specific mortality in 900 000 adults: collaborative analyses of 57 prospective studies. Lancet 2009; 373:1083-96; PMID:19299006; http://dx.doi.org/10.1016/S0140-6736(09)60318-4
- Mair W, Piper MD, Partridge L. Calories do not explain extension of life span by dietary restriction in Drosophila. PLoS Biol 2005; 3:e223; PMID:16000018; http://dx.doi.org/10.1371/ journal.pbio.0030223
- Miller RA, Buehner G, Chang Y, Harper JM, Sigler R, Smith-Wheelock M. Methionine-deficient diet extends mouse lifespan, slows immune and lens aging, alters glucose, T4, IGF-I and insulin levels, and increases hepatocyte MIF levels and stress resistance. Aging Cell 2005; 4:119-25; PMID:15924568; http:// dx.doi.org/10.1111/j.1474-9726.2005.00152.x
- Zimmerman JA, Malloy V, Krajcik R, Orentreich N. Nutritional control of aging. Exp Gerontol 2003; 38:47-52; PMID:12543260; http://dx.doi. org/10.1016/S0531-5565(02)00149-3
- Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB, Cruzen C, Simmons HA, Kemnitz JW, et al. Caloric restriction delays disease onset and mortality in rhesus monkeys. Science 2009; 325:201-4; PMID:19590001; http:// dx.doi.org/10.1126/science.1173635
- Heydari AR, Unnikrishnan A, Lucente LV, Richardson A. Caloric restriction and genomic stability. Nucleic Acids Res 2007; 35:7485-96; PMID:17942423; http://dx.doi.org/10.1093/nar/ gkm860
- Lombard DB, Chua KF, Mostoslavsky R, Franco S, Gostissa M, Alt FW. DNA repair, genome stability, and aging. Cell 2005; 120:497-512; PMID:15734682; http://dx.doi.org/10.1016/j. cell.2005.01.028
- Oberdoerffer P, Sinclair DA. The role of nuclear architecture in genomic instability and ageing. Nat Rev Mol Cell Biol 2007; 8:692-702; PMID:17700626; http://dx.doi.org/10.1038/ nrm2238

- 46. Maynard S, Schurman SH, Harboe C, de Souza-Pinto NC, Bohr VA. Base excision repair of oxidative DNA damage and association with cancer and aging. Carcinogenesis 2009; 30:2-10; PMID:18978338; http://dx.doi.org/10.1093/carcin/bgn250
- Cabelof DC, Yanamadala S, Raffoul JJ, Guo Z, Soofi A, Heydari AR. Caloric restriction promotes genomic stability by induction of base excision repair and reversal of its age-related decline. DNA Repair (Amst) 2003; 2:295-307; PMID:12547392; http:// dx.doi.org/10.1016/S1568-7864(02)00219-7
- Stuart JA, Karahalil B, Hogue BA, Souza-Pinto NC, Bohr VA. Mitochondrial and nuclear DNA base excision repair are affected differently by caloric restriction. FASEB J 2004; 18:595-7; PMID:14734635
- 49. Kisby GE, Kohama SG, Olivas A, Churchwell M, Doerge D, Spangler E, de Cabo R, Ingram DK, Imhof B, Bao G, et al. Effect of caloric restriction on base-excision repair (BER) in the aging rat brain. Exp Gerontol 2010; 45:208-16; PMID:20005284; http:// dx.doi.org/10.1016/j.exger.2009.12.003
- Guo Z, Heydari A, Richardson A. Nucleotide excision repair of actively transcribed versus nontranscribed DNA in rat hepatocytes: effect of age and dietary restriction. Exp Cell Res 1998; 245:228-38; PMID:9828120; http://dx.doi.org/10.1006/ excr.1998.4269
- Lehmann AR, McGibbon D, Stefanini M. Xeroderma pigmentosum. Orphanet J Rare Dis 2011; 6:70; PMID:22044607; http://dx.doi. org/10.1186/1750-1172-6-70
- Jeggo PA. DNA-PK: at the cross-roads of biochemistry and genetics. Mutat Res 1997; 384:1-14; PMID:9201268; http://dx.doi.org/10.1016/ S0921-8777(97)00009-8
- Um JH, Kim SJ, Kim DW, Ha MY, Jang JH, Kim DW, Chung BS, Kang CD, Kim SH. Tissuespecific changes of DNA repair protein Ku and mtHSP70 in aging rats and their retardation by caloric restriction. Mech Ageing Dev 2003; 124:967-75; PMID:14499502; http://dx.doi.org/10.1016/ S0047-6374(03)00169-6
- Steffen KK, Kennedy BK, Kaeberlein M. Measuring replicative life span in the budding yeast. J Vis Exp 2009; PMID:19556967; http://dx.doi. org/10.3791/1209
- Gottlieb S, Esposito RE. A new role for a yeast transcriptional silencer gene, SIR2, in regulation of recombination in ribosomal DNA. Cell 1989; 56:771-6; PMID:2647300; http://dx.doi. org/10.1016/0092-8674(89)90681-8
- Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, Zipkin RE, Chung P, Kisielewski A, Zhang LL, et al. Small molecule activators of sirtuins extend Saccharomyces cerevisiae lifespan. Nature 2003; 425:191-6; PMID:12939617; http://dx.doi.org/10.1038/nature01960
- Kaeberlein M, McVey M, Guarente L. The SIR2/3/4 complex and SIR2 alone promote longevity in Saccharomyces cerevisiae by two different mechanisms. Genes Dev 1999; 13:2570-80; PMID:10521401; http://dx.doi.org/10.1101/ gad.13.19.2570
- Kobayashi T, Heck DJ, Nomura M, Horiuchi T. Expansion and contraction of ribosomal DNA repeats in Saccharomyces cerevisiae: requirement of replication fork blocking (Fob1) protein and the role of RNA polymerase I. Genes Dev 1998; 12:3821-30; PMID:9869636; http://dx.doi.org/10.1101/ gad.12.24.3821
- Fontana L, Partridge L, Longo VD. Extending healthy life span--from yeast to humans. Science 2010; 328:321-6; PMID:20395504; http://dx.doi. org/10.1126/science.1172539

- Chan JN, Poon BP, Salvi J, Olsen JB, Emili A, Mekhail K. Perinuclear cohibin complexes maintain replicative life span via roles at distinct silent chromatin domains. Dev Cell 2011; 20:867-79; PMID:21664583; http://dx.doi.org/10.1016/j. devcel.2011.05.014
- Dang W, Steffen KK, Perry R, Dorsey JA, Johnson FB, Shilatifard A, Kaeberlein M, Kennedy BK, Berger SL. Histone H4 lysine 16 acetylation regulates cellular lifespan. Nature 2009; 459:802-7; PMID:19516333; http://dx.doi.org/10.1038/nature08085
- Salvi JS, Chan JN, Pettigrew C, Liu TT, Wu JD, Mekhail K. Enforcement of a lifespan-sustaining distribution of Sir2 between telomeres, matingtype loci, and rDNA repeats by Rif1. Aging Cell 2013; 12:67-75; PMID:23082874; http://dx.doi. org/10.1111/acel.12020
- Takeuchi Y, Horiuchi T, Kobayashi T. Transcriptiondependent recombination and the role of fork collision in yeast rDNA. Genes Dev 2003; 17:1497-506; PMID:12783853; http://dx.doi.org/10.1101/ gad.1085403
- Mekhail K, Moazed D. The nuclear envelope in genome organization, expression and stability. Nat Rev Mol Cell Biol 2010; 11:317-28; PMID:20414256; http://dx.doi.org/10.1038/nrm2894
- Riesen M, Morgan A. Calorie restriction reduces rDNA recombination independently of rDNA silencing. Aging Cell 2009; 8:624-32; PMID:19732046; http://dx.doi.org/10.1111/j.1474-9726.2009.00514.x
- Smith DL Jr., Li C, Matecic M, Maqani N, Bryk M, Smith JS. Calorie restriction effects on silencing and recombination at the yeast rDNA. Aging Cell 2009; 8:633-42; PMID:19732044; http://dx.doi. org/10.1111/j.1474-9726.2009.00516.x
- Smith JS, Boeke JD. An unusual form of transcriptional silencing in yeast ribosomal DNA. Genes Dev 1997; 11:241-54; PMID:9009206; http://dx.doi.org/10.1101/gad.11.2.241
- Lin SJ, Defossez PA, Guarente L. Requirement of NAD and SIR2 for life-span extension by calorie restriction in Saccharomyces cerevisiae. Science 2000; 289:2126-8; PMID:11000115; http://dx.doi. org/10.1126/science.289.5487.2126
- Smith DL Jr., McClure JM, Matecic M, Smith JS. Calorie restriction extends the chronological lifespan of Saccharomyces cerevisiae independently of the Sirtuins. Aging Cell 2007; 6:649-62; PMID:17711561; http://dx.doi. org/10.1111/j.1474-9726.2007.00326.x
- Kwan EX, Foss EJ, Tsuchiyama S, Alvino GM, Kruglyak L, Kaeberlein M, Raghuraman MK, Brewer BJ, Kennedy BK, Bedalov A. A natural polymorphism in rDNA replication origins links origin activation with calorie restriction and lifespan. PLoS Genet 2013; 9:e1003329; PMID:23505383; http://dx.doi. org/10.1371/journal.pgen.1003329
- Murayama A, Ohmori K, Fujimura A, Minami H, Yasuzawa-Tanaka K, Kuroda T, Oie S, Daitoku H, Okuwaki M, Nagata K, et al. Epigenetic control of rDNA loci in response to intracellular energy status. Cell 2008; 133:627-39; PMID:18485871; http:// dx.doi.org/10.1016/j.cell.2008.03.030
- Cohen HY, Miller C, Bitterman KJ, Wall NR, Hekking B, Kessler B, Howitz KT, Gorospe M, de Cabo R, Sinclair DA. Calorie restriction promotes mammalian cell survival by inducing the SIRT1 deacetylase. Science 2004; 305:390-2; PMID:15205477; http://dx.doi.org/10.1126/ science.1099196
- Allsopp RC, Vaziri H, Patterson C, Goldstein S, Younglai EV, Futcher AB, Greider CW, Harley CB. Telomere length predicts replicative capacity of human fibroblasts. Proc Natl Acad Sci U S A 1992; 89:10114-8; PMID:1438199; http://dx.doi. org/10.1073/pnas.89.21.10114

- Blackburn EH, Gall JG. A tandemly repeated sequence at the termini of the extrachromosomal ribosomal RNA genes in Tetrahymena. J Mol Biol 1978; 120:33-53; PMID:642006; http://dx.doi. org/10.1016/0022-2836(78)90294-2
- Chan SW, Blackburn EH. New ways not to make ends meet: telomerase, DNA damage proteins and heterochromatin. Oncogene 2002; 21:553-63; PMID:11850780; http://dx.doi.org/10.1038/ sj.onc.1205082
- Collado M, Blasco MA, Serrano M. Cellular senescence in cancer and aging. Cell 2007; 130:223-33; PMID:17662938; http://dx.doi.org/10.1016/j. cell.2007.07.003
- Hoppe GJ, Tanny JC, Rudner AD, Gerber SA, Danaie S, Gygi SP, Moazed D. Steps in assembly of silent chromatin in yeast: Sir3-independent binding of a Sir2/Sir4 complex to silencers and role for Sir2-dependent deacetylation. Mol Cell Biol 2002; 22:4167-80; PMID:12024030; http://dx.doi. org/10.1128/MCB.22.12.4167-4180.2002
- Wellinger RJ, Zakian VA. Everything you ever wanted to know about Saccharomyces cerevisiae telomeres: beginning to end. Genetics 2012; 191:1073-105; PMID:22879408; http://dx.doi.org/10.1534/ genetics.111.137851
- 79. Heidinger BJ, Blount JD, Boner W, Griffiths K, Metcalfe NB, Monaghan P. Telomere length in early life predicts lifespan. Proc Natl Acad Sci U S A 2012; 109:1743-8; PMID:22232671; http://dx.doi. org/10.1073/pnas.1113306109
- Tennen RI, Bua DJ, Wright WE, Chua KF. SIRT6 is required for maintenance of telomere position effect in human cells. Nat Commun 2011; 2:433; PMID:21847107; http://dx.doi.org/10.1038/ ncomms1443
- Kanfi Y, Shalman R, Peshti V, Pilosof SN, Gozlan YM, Pearson KJ, Lerrer B, Moazed D, Marine JC, de Cabo R, et al. Regulation of SIRT6 protein levels by nutrient availability. FEBS Lett 2008; 582:543-8; PMID:18242175; http://dx.doi.org/10.1016/j. febslet.2008.01.019
- Pendergrass WR, Penn PE, Li J, Wolf NS. Age-related telomere shortening occurs in lens epithelium from old rats and is slowed by caloric restriction. Exp Eye Res 2001; 73:221-8; PMID:11446772; http://dx.doi. org/10.1006/exer.2001.1033
- Vera E, Bernardes de Jesus B, Foronda M, Flores JM, Blasco MA. Telomerase reverse transcriptase synergizes with calorie restriction to increase health span and extend mouse longevity. PLoS One 2013; 8:e53760; PMID:23349740; http://dx.doi. org/10.1371/journal.pone.0053760
- 84. Kim S, Bi X, Czarny-Ratajczak M, Dai J, Welsh DA, Myers L, Welsch MA, Cherry KE, Arnold J, Poon LW, et al. Telomere maintenance genes SIRT1 and XRCC6 impact age-related decline in telomere length but only SIRT1 is associated with human longevity. Biogerontology 2012; 13:119-31; PMID:21972126; http://dx.doi.org/10.1007/s10522-011-9360-5
- Palacios JA, Herranz D, De Bonis ML, Velasco S, Serrano M, Blasco MA. SIRT1 contributes to telomere maintenance and augments global homologous recombination. J Cell Biol 2010; 191:1299-313; PMID:21187328; http://dx.doi. org/10.1083/jcb.201005160
- Kwan EX, Foss E, Kruglyak L, Bedalov A. Natural polymorphism in BUL2 links cellular amino acid availability with chronological aging and telomere maintenance in yeast. PLoS Genet 2011; 7:e1002250; PMID:21901113; http://dx.doi.org/10.1371/journal. pgen.1002250
- Ungar L, Harari Y, Toren A, Kupiec M. Tor complex 1 controls telomere length by affecting the level of Ku. Curr Biol 2011; 21:2115-20; PMID:22169538; http://dx.doi.org/10.1016/j.cub.2011.11.024

- Wood JG, Helfand SL. Chromatin structure and transposable elements in organismal aging. Front Genet 2013; 4:274; PMID:24363663; http://dx.doi. org/10.3389/fgene.2013.00274
- Banito A, Lowe SW. A new development in senescence. Cell 2013; 155:977-8; PMID:24267881; http://dx.doi.org/10.1016/j.cell.2013.10.050
- Muñoz-Espín D, Cañamero M, Maraver A, Gómez-López G, Contreras J, Murillo-Cuesta S, Rodríguez-Baeza A, Varela-Nieto I, Ruberte J, Collado M, et al. Programmed cell senescence during mammalian embryonic development. Cell 2013; 155:1104-18; PMID:24238962; http://dx.doi.org/10.1016/j. cell.2013.10.019
- Storer M, Mas A, Robert-Moreno A, Pecoraro M, Ortells MC, Di Giacomo V, Yosef R, Pilpel N, Krizhanovsky V, Sharpe J, et al. Senescence is a developmental mechanism that contributes to embryonic growth and patterning. Cell 2013; 155:1119-30; PMID:24238961; http://dx.doi. org/10.1016/j.cell.2013.10.041
- Tarry-Adkins JL, Martin-Gronert MS, Chen JH, Cripps RL, Ozanne SE. Maternal diet influences DNA damage, aortic telomere length, oxidative stress, and antioxidant defense capacity in rats. FASEB J 2008; 22:2037-44; PMID:18230683; http://dx.doi. org/10.1096/fj.07-099523
- Eisenberg T, Knauer H, Schauer A, Büttner S, Ruckenstuhl C, Carmona-Gutierrez D, Ring J, Schroeder S, Magnes C, Antonacci L, et al. Induction of autophagy by spermidine promotes longevity. Nat Cell Biol 2009; 11:1305-14; PMID:19801973; http://dx.doi.org/10.1038/ncb1975
- 94. Morselli E, Maiuri MC, Markaki M, Megalou E, Pasparaki A, Palikaras K, Criollo A, Galluzzi L, Malik SA, Vitale I, et al. Caloric restriction and resveratrol promote longevity through the Sirtuin-1-dependent induction of autophagy. Cell Death Dis 2010; 1:e10; PMID:21364612; http://dx.doi. org/10.1038/cddis.2009.8
- Hansen M, Chandra A, Mitic LL, Onken B, Driscoll M, Kenyon C. A role for autophagy in the extension of lifespan by dietary restriction in C. elegans. PLoS Genet 2008; 4:e24; PMID:18282106; http://dx.doi. org/10.1371/journal.pgen.0040024
- Jia K, Levine B. Autophagy is required for dietary restriction-mediated life span extension in C. elegans. Autophagy 2007; 3:597-9; PMID:17912023
- Minina EA, Sanchez-Vera V, Moschou PN, Suarez MF, Sundberg E, Weih M, Bozhkov PV. Autophagy mediates caloric restriction-induced lifespan extension in Arabidopsis. Aging Cell 2013; 12:327-9; PMID:23331488; http://dx.doi.org/10.1111/ acel.12048
- Díaz-Troya S, Pérez-Pérez ME, Florencio FJ, Crespo JL. The role of TOR in autophagy regulation from yeast to plants and mammals. Autophagy 2008; 4:851-65; PMID:18670193
- Burnett C, Valentini S, Cabreiro F, Goss M, Somogyvári M, Piper MD, Hoddinott M, Sutphin GL, Leko V, McElwee JJ, et al. Absence of effects of Sir2 overexpression on lifespan in C. elegans and Drosophila. Nature 2011; 477:482-5; PMID:21938067; http://dx.doi.org/10.1038/ nature10296
- 100. Viswanathan M, Guarente L. Regulation of Caenorhabditis elegans lifespan by sir-2.1 transgenes. Nature 2011; 477:E1-2; PMID:21938026; http:// dx.doi.org/10.1038/nature10440
- Kaeberlein M, Kapahi P. Cell signaling. Aging is RSKy business. Science 2009; 326:55-6; PMID:19797648; http://dx.doi.org/10.1126/science.1181034

- 102. Salminen A, Kaarniranta K. AMP-activated protein kinase (AMPK) controls the aging process via an integrated signaling network. Ageing Res Rev 2012; 11:230-41; PMID:22186033; http://dx.doi. org/10.1016/j.art.2011.12.005
- 103. Selman C, Tullet JM, Wieser D, Irvine E, Lingard SJ, Choudhury AI, Claret M, Al-Qassab H, Carmignac D, Ramadani F, et al. Ribosomal protein S6 kinase 1 signaling regulates mammalian life span. Science 2009; 326:140-4; PMID:19797661; http://dx.doi. org/10.1126/science.1177221
- 104. Warr MR, Binnewies M, Flach J, Reynaud D, Garg T, Malhotra R, Debnath J, Passegué E. FOXO3A directs a protective autophagy program in haematopoietic stem cells. Nature 2013; 494:323-7; PMID:23389440; http://dx.doi.org/10.1038/ nature11895
- 105. Nishino I, Fu J, Tanji K, Yamada T, Shimojo S, Koori T, Mora M, Riggs JE, Oh SJ, Koga Y, et al. Primary LAMP-2 deficiency causes X-linked vacuolar cardiomyopathy and myopathy (Danon disease). Nature 2000; 406:906-10; PMID:10972294; http:// dx.doi.org/10.1038/35022604
- Balaban RS, Nemoto S, Finkel T. Mitochondria, oxidants, and aging. Cell 2005; 120:483-95; PMID:15734681; http://dx.doi.org/10.1016/j. cell.2005.02.001
- 107. Du C, Anderson A, Lortie M, Parsons R, Bodnar A. Oxidative damage and cellular defense mechanisms in sea urchin models of aging. Free Radic Biol Med 2013; 63:254-63; PMID:23707327; http://dx.doi. org/10.1016/j.freeradbiomed.2013.05.023
- Qiu X, Brown K, Hirschey MD, Verdin E, Chen D. Calorie restriction reduces oxidative stress by SIRT3-mediated SOD2 activation. Cell Metab 2010; 12:662-7; PMID:21109198; http://dx.doi. org/10.1016/j.cmet.2010.11.015
- 109. Someya S, Yu W, Hallows WC, Xu J, Vann JM, Leeuwenburgh C, Tanokura M, Denu JM, Prolla TA. Sirt3 mediates reduction of oxidative damage and prevention of age-related hearing loss under caloric restriction. Cell 2010; 143:802-12; PMID:21094524; http://dx.doi.org/10.1016/j.cell.2010.10.002
- 110. Parkes TL, Elia AJ, Dickinson D, Hilliker AJ, Phillips JP, Boulianne GL. Extension of Drosophila lifespan by overexpression of human SOD1 in motorneurons. Nat Genet 1998; 19:171-4; PMID:9620775; http:// dx.doi.org/10.1038/534
- 111. Martin I, Jones MA, Grotewiel M. Manipulation of Sod1 expression ubiquitously, but not in the nervous system or muscle, impacts age-related parameters in Drosophila. FEBS Lett 2009; 583:2308-14; PMID:19540235; http://dx.doi.org/10.1016/j. febslet.2009.06.023
- 112. Li J, Li T, Zhang X, Tang Y, Yang J, Le W. Human superoxide dismutase 1 overexpression in motor neurons of Caenorhabditis elegans causes axon guidance defect and neurodegeneration. Neurobiol Aging 2014; 35:837-46; PMID:24126158; http:// dx.doi.org/10.1016/j.neurobiolaging.2013.09.003
- Merry BJ. Molecular mechanisms linking calorie restriction and longevity. Int J Biochem Cell Biol 2002; 34:1340-54; PMID:12200030; http://dx.doi. org/10.1016/S1357-2725(02)00038-9
- 114. Nisoli E, Tonello C, Cardile A, Cozzi V, Bracale R, Tedesco L, Falcone S, Valerio A, Cantoni O, Clementi E, et al. Calorie restriction promotes mitochondrial biogenesis by inducing the expression of eNOS. Science 2005; 310:314-7; PMID:16224023; http:// dx.doi.org/10.1126/science.1117728

- 115. Civitarese AE, Carling S, Heilbronn LK, Hulver MH, Ukropcova B, Deutsch WA, Smith SR, Ravussin E; CALERIE Pennington Team. Calorie restriction increases muscle mitochondrial biogenesis in healthy humans. PLoS Med 2007; 4:e76; PMID:17341128; http://dx.doi.org/10.1371/journal.pmed.0040076
- 116. Barros MH, Bandy B, Tahara EB, Kowaltowski AJ. Higher respiratory activity decreases mitochondrial reactive oxygen release and increases life span in Saccharomyces cerevisiae. J Biol Chem 2004; 279:49883-8; PMID:15383542; http://dx.doi. org/10.1074/jbc.M408918200
- 117. Lin SJ, Kaeberlein M, Andalis AA, Sturtz LA, Defossez PA, Culotta VC, Fink GR, Guarente L. Calorie restriction extends Saccharomyces cerevisiae lifespan by increasing respiration. Nature 2002; 418:344-8; PMID:12124627; http://dx.doi. org/10.1038/nature00829
- 118. Sanz A, Caro P, Ibañez J, Gómez J, Gredilla R, Barja G. Dietary restriction at old age lowers mitochondrial oxygen radical production and leak at complex I and oxidative DNA damage in rat brain. J Bioenerg Biomembr 2005; 37:83-90; PMID:15906153; http:// dx.doi.org/10.1007/s10863-005-4131-0
- 119. Korshunov SS, Skulachev VP, Starkov AA. High protonic potential actuates a mechanism of production of reactive oxygen species in mitochondria. FEBS Lett 1997; 416:15-8; PMID:9369223; http://dx.doi. org/10.1016/S0014-5793(97)01159-9
- 120. López-Lluch G, Hunt N, Jones B, Zhu M, Jamieson H, Hilmer S, Cascajo MV, Allard J, Ingram DK, Navas P, et al. Calorie restriction induces mitochondrial biogenesis and bioenergetic efficiency. Proc Natl Acad Sci U S A 2006; 103:1768-73; PMID:16446459; http://dx.doi.org/10.1073/pnas.0510452103
- 121. Hughes AL, Gottschling DE. An early age increase in vacuolar pH limits mitochondrial function and lifespan in yeast. Nature 2012; 492:261-5; PMID:23172144; http://dx.doi.org/10.1038/ nature11654
- 122. Palmieri F, Agrimi G, Blanco E, Castegna A, Di Noia MA, Iacobazzi V, Lasorsa FM, Marobbio CM, Palmieri L, Scarcia P, et al. Identification of mitochondrial carriers in Saccharomyces cerevisiae by transport assay of reconstituted recombinant proteins. Biochim Biophys Acta 2006; 1757:1249-62; PMID:16844075; http://dx.doi.org/10.1016/j. bbabio.2006.05.023
- 123. Patel BP, Safdar A, Raha S, Tarnopolsky MA, Hamadeh MJ. Caloric restriction shortens lifespan through an increase in lipid peroxidation, inflammation and apoptosis in the G93A mouse, an animal model of ALS. PLoS One 2010; 5:e9386; PMID:20195368; http://dx.doi.org/10.1371/journal. pone.0009386
- 124. Johnson SC, Yanos ME, Kayser EB, Quintana A, Sangesland M, Castanza A, Uhde L, Hui J, Wall VZ, Gagnidze A, et al. mTOR inhibition alleviates mitochondrial disease in a mouse model of Leigh syndrome. Science 2013; 342:1524-8; PMID:24231806; http://dx.doi.org/10.1126/ science.1244360
- 125. Austad SN. Life extension by dietary restriction in the bowl and doily spider, Frontinella pyramitela. Exp Gerontol 1989; 24:83-92; PMID:2707314; http:// dx.doi.org/10.1016/0531-5565(89)90037-5
- 126. Kealy RD, Lawler DF, Ballam JM, Mantz SL, Biery DN, Greeley EH, Lust G, Segre M, Smith GK, Stowe HD. Effects of diet restriction on life span and agerelated changes in dogs. J Am Vet Med Assoc 2002; 220:1315-20; PMID:11991408; http://dx.doi. org/10.2460/javma.2002.220.1315