





Draft Genome Sequences of Eight *Mycobacterium montefiorensis* Strains Isolated from Salamanders in Captivity

 Takeshi Komine,^a Hyogo Ihara,^a  Hanako Fukano,^b Yoshihiko Hoshino,^b Osamu Kurata,^a Shinpei Wada^a

^aLaboratory of Aquatic Medicine, School of Veterinary Medicine, Nippon Veterinary and Life Science University, Musashino, Tokyo, Japan

^bDepartment of Mycobacteriology, Leprosy Research Center, National Institute of Infectious Diseases, Higashi-Murayama, Tokyo, Japan

ABSTRACT *Mycobacterium montefiorensis* is a nontuberculous mycobacterium that causes infections in fish and salamanders. Here, we report annotated draft genome sequences of eight strains that were isolated in 2014 and 2018 from salamanders reared in an aquarium in Japan.

Mycobacterium montefiorensis is a nontuberculous mycobacterium (NTM) that causes mycobacteriosis in fish and salamanders (1–3). *Mycobacterium montefiorensis* has also been isolated from soil and pond water (4). We sequenced the genomes of eight *M. montefiorensis* strains that were isolated in 2014 and 2018 from salamanders reared in Niigata City Aquarium (Niigata, Japan).

Several dead salamanders in the aquarium were collected (Table 1) and routinely dissected. The liver tissues were sampled, homogenized, and decontaminated with 1 mL of *N*-acetyl-L-cysteine-sodium citrate-NaOH for no more than 15 min. After neutralization with 6 mL of phosphate buffer (pH 6.8), the samples were centrifuged; the pellets obtained were then inoculated on Middlebrook 7H10 agar supplemented with 10% BBL Middlebrook oleic acid-albumin-dextrose-catalase (OADC) enrichment (Becton, Dickinson and Company, USA) and on 2% Ogawa egg slants (Kyokuto Pharmaceutical Industrial Co., Ltd., Japan). Isolates obtained were identified as *M. montefiorensis* based on the Runyon classification system (5) and phylogenetic analysis of the 401-bp 65-kDa heat shock protein gene (*hsp65*) with the Tb11/Tb12 primer set (6).

Frozen stocks (–80°C in 20% glycerol) of *M. montefiorensis* strains were streaked on 2% Ogawa slants, and single colonies were grown at 25°C for approximately 4 weeks. The collected colonies were boiled at 95°C for 15 min, frozen at –20°C overnight, and disrupted twice (4,500 rpm for 1 min) with approximately 0.5-mm-diameter zirconia/silica beads (BioSpec Products, Inc., USA) using a Micro Smash MS-100 disrupter (Tomy Digital Biology Co., Ltd., Japan). Genomic DNA was extracted using the NucleoSpin Plant II kit (Macherey-Nagel GmbH & Co. KG, Germany) in accordance with the manufacturer's instructions.

Sequencing libraries were prepared using the Nextera XT DNA library preparation kit (Illumina, USA) and sequenced on the Illumina HiSeq X platform (2 × 150 bp). The quality of the raw reads was assessed with FastQC v0.11.9 (7). The sequence reads were trimmed for quality using fastp v0.20.1 (8) and assembled *de novo* using Platanus_B v1.1.0 (9). We also assembled the genome of the type strain (*M. montefiorensis* ATCC BAA-256) using the raw data (accession number [DRR361296](https://ncbi.nlm.nih.gov/nucl/DRR361296)) from the National Center for Biotechnology Information (NCBI) GenBank database using the same method as described above. Automated annotation was conducted with the DNA Data Bank of Japan (DDBJ) Fast Annotation and Submission Tool (DFAST) (<https://dfast.ddbj.nig.ac.jp>). The annotated assemblies for eight strains and the type strain were deposited in the DDBJ. All genomic statistics are given in Table 1. Default parameters were used for all software unless otherwise noted.

Average nucleotide identity (ANI) analysis was conducted to determine relationships

Editor Frank J. Stewart, Montana State University

Copyright © 2022 Komine et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Takeshi Komine, fishvet.tk@gmail.com.

The authors declare no conflict of interest.

Received 24 July 2022

Accepted 12 October 2022

Published 31 October 2022

TABLE 1 Strain information and assembly statistics

Strain	Isolate source	Year of isolation	No. of raw reads	Genome size (bp)	No. of contigs	N_{50} (bp)	Coverage (x)	Total no. of CDSs ^a	G+C content (%)	SRA accession no.	Contig accession no.
NJB14191	Hakuba salamander (<i>Hynobius hidamontanus</i>)	2014	3,646,896	5,744,673	123	171,591	85	5,348	65.1	DRR357474	BQYA000000000
NJB14192	Hakuba salamander (<i>H. hidamontanus</i>)	2014	1,841,220	5,776,754	234	157,068	43	5,366	65.1	DRR357475	BQYB000000000
NJB14194	Hakuba salamander (<i>H. hidamontanus</i>)	2014	1,946,204	5,753,077	155	169,431	46	5,382	65.1	DRR357476	BQYC000000000
NJB14195	Hakuba salamander (<i>H. hidamontanus</i>)	2014	2,424,346	5,749,641	116	201,183	58	5,381	65.1	DRR357477	BQYD000000000
NJB14197	Hakuba salamander (<i>H. hidamontanus</i>)	2014	3,047,344	5,745,062	108	201,635	72	5,364	65.1	DRR357478	BQYE000000000
NJB18182	Tohoku hynobiid salamander (<i>Hynobius lichenatus</i>)	2018	2,308,180	5,764,439	210	136,848	52	5,352	65.1	DRR357479	BQYF000000000
NJB18183	Tohoku hynobiid salamander (<i>H. lichenatus</i>)	2018	2,811,918	5,749,386	127	207,711	64	5,376	65.1	DRR357480	BQYG000000000
NJB18185	Tohoku hynobiid salamander (<i>H. lichenatus</i>)	2018	4,174,608	5,739,217	88	240,008	99	5,381	65.1	DRR357481	BQYH000000000
ATCC BAA-256	Green moray (<i>Gymnothorax funebris</i>)	NC ^b	8,308,120	5,226,877	735	16,108	396	4,149	65.2	DRR361296	BSAJ000000000

^a CDSs, coding sequences.^b NC, not clear.

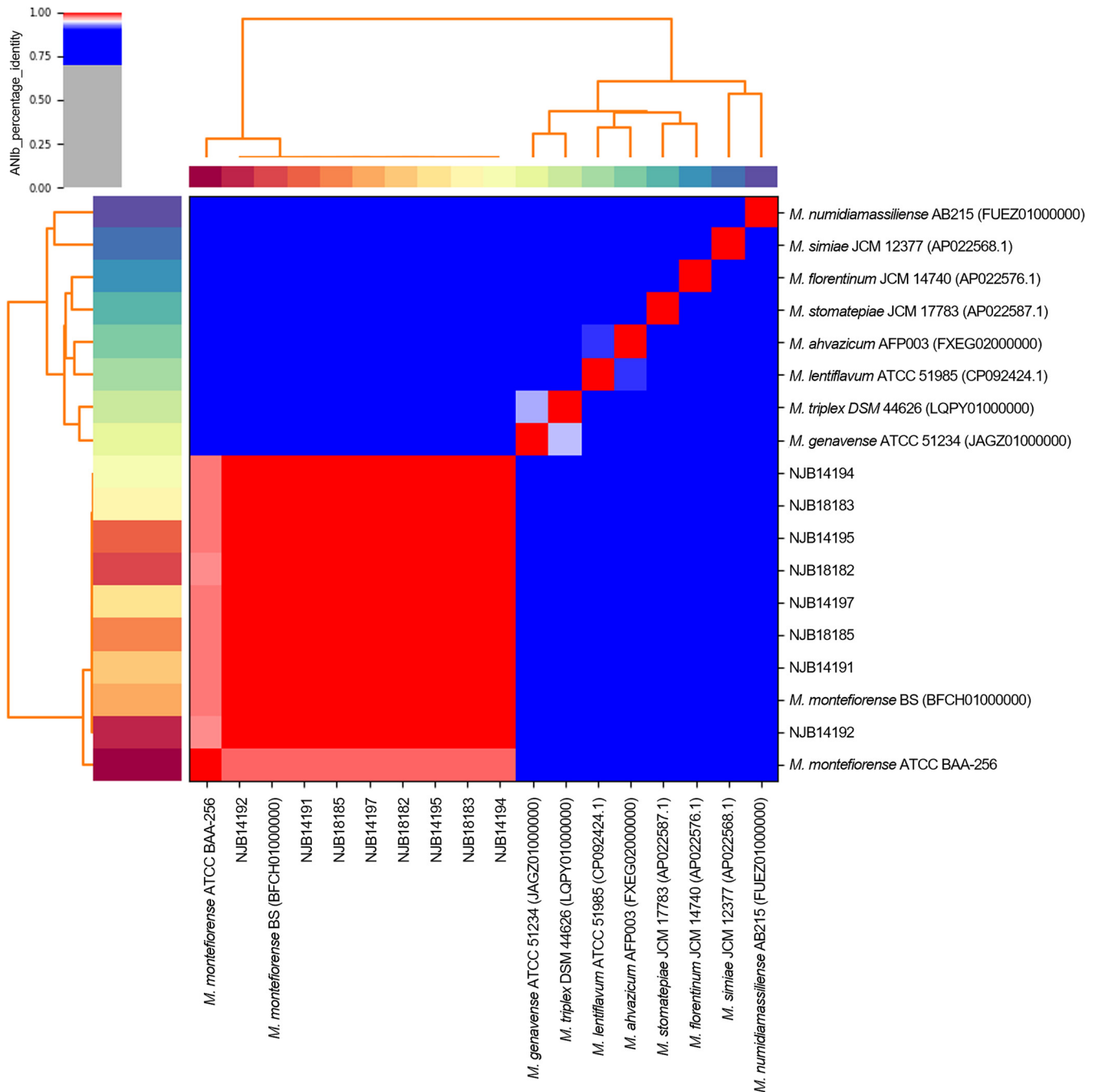


FIG 1 Heatmap of ANI values for 18 *Mycobacterium* strains. The heatmap was generated using pyani v0.2.11 and a BLAST-based approach (ANiB).

among *Mycobacterium* species. ANI values were determined for the whole-genome sequences using pyani v0.2.11 and a BLAST-based approach (ANiB) (10). The ANI heatmap is shown in Fig. 1, and the ANI values for comparisons between the eight strains and *M. montefiorensis* ATCC BAA-256 were >97.2%.

We report the draft genome sequences of eight *M. montefiorensis* strains that were isolated from salamanders in captivity. These sequences will improve our understanding of the pathogenicity and evolution of this mycobacterial species.

Data availability. The genome sequencing and assembly projects have been deposited in the DDBJ under BioProject accession number [PRJDB13312](https://www.ncbi.nlm.nih.gov/bioproject/PRJDB13312). See Table 1 for the DDBJ Sequence Read Archive (DRA) and DDBJ accession numbers.

ACKNOWLEDGMENTS

We thank Mallory Eckstut from Edanz (<https://jp.edanz.com/ac>) for editing a draft of the manuscript.

This study was supported in part by grants from the Japan Agency for Medical Research and Development/Japan International Cooperation Agency (AMED) to Y.H. (grants JP20fk0108064, JP20fk0108075, JP21jm0510004, JP22fk0108093, JP22fk0108129, JP22fk0108608, JP22gm1610003, JP22wm0125007, JP22wm0225004, JP22wm0225022, JP22wm0325003, and JP22wm0325054) and H.F. (grant JP22wm0325054); by a Grant-in-Aid for Fostering Joint International Research (B) to Y.H. (grant JP19KK0217) and Grants-in-Aid for Early-Career Scientists to H.F. (grants JP18K15966 and JP22K16382) from the Japan Society for the Promotion of Science (JSPS); and by a Grant-in-Aid for Scientific Research (B) to Y.H. (grant JP20H02282) from JSPS. The funders had no role in the study design, data collection, data analysis, the decision to publish, or preparation of the manuscript.

REFERENCES

1. Herbst LH, Costa SF, Weiss LM, Johnson LK, Bartell J, Davis R, Walsh M, Levi M. 2001. Granulomatous skin lesions in moray eels caused by a novel *Mycobacterium* species related to *Mycobacterium triplex*. *Infect Immun* 69:4639–4646. <https://doi.org/10.1128/IAI.69.7.4639-4646.2001>.
2. Levi MH, Bartell J, Gandolfo L, Smole SC, Costa SF, Weiss LM, Johnson LK, Osterhout G, Herbst LH. 2003. Characterization of *Mycobacterium montefiorensis* sp. nov., a novel pathogenic mycobacterium from moray eels that is related to *Mycobacterium triplex*. *J Clin Microbiol* 41:2147–2152. <https://doi.org/10.1128/JCM.41.5.2147-2152.2003>.
3. Fukano H, Yoshida M, Shimizu A, Iwao H, Katayama Y, Omatsu T, Mizutani T, Kurata O, Wada S, Hoshino Y. 2018. Draft genome sequence of *Mycobacterium montefiorensis* isolated from Japanese black salamander (*Hynobius nigrescens*). *Genome Announc* 6:e00448-18. <https://doi.org/10.1128/genomeA.00448-18>.
4. Makovcova J, Slany M, Babak V, Slana I, Kralik P. 2014. The water environment as a source of potentially pathogenic mycobacteria. *J Water Health* 12:254–263. <https://doi.org/10.2166/wh.2013.102>.
5. Runyon EH. 1959. Anonymous mycobacteria in pulmonary disease. *Med Clin North Am* 43:273–290. [https://doi.org/10.1016/S0025-7125\(16\)34193-1](https://doi.org/10.1016/S0025-7125(16)34193-1).
6. Telenti A, Marchesi F, Balz M, Bally F, Böttger EC, Bodmer T. 1993. Rapid identification of mycobacteria to the species level by polymerase chain reaction and restriction enzyme analysis. *J Clin Microbiol* 31:175–178. <https://doi.org/10.1128/jcm.31.2.175-178.1993>.
7. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
8. Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>.
9. Kajitani R, Yoshimura D, Ogura Y, Gotoh Y, Hayashi T, Itoh T. 2020. Platanus_B: an accurate *de novo* assembler for bacterial genomes using an iterative error-removal process. *DNA Res* 27:dsaa014. <https://doi.org/10.1093/dnares/dsaa014>.
10. Pritchard L, Glover RH, Humphris S, Elphinstone JG, Toth IK. 2016. Genomics and taxonomy in diagnostics for food security: soft-rotting enterobacterial plant pathogens. *Anal Methods* 8:12–24. <https://doi.org/10.1039/C5AY02550H>.