# Review Role of RUNX in autoimmune diseases linking rheumatoid arthritis, psoriasis and lupus

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#### Abstract

Recent studies investigating the genetic susceptibility of systemic lupus erythematosus, rheumatoid arthritis and psoriasis have revealed a potential role for the RUNX proteins in the development of autoimmune disease. A new pathway of disease pathogenesis opens new avenues of research with thousands of questions that remain to be answered. In this review I attempt to propose how the RUNX proteins might be involved in these diseases and review current knowledge on this very interesting trio of transcription factors that was previously only suspected to be involved in cancer.

Keywords: autoimmunity, repression, runt-domain, susceptibility, transcription

# Introduction

The study of the genetics of complex diseases is now advancing rapidly as new genes are being discovered that are involved in susceptibility for a variety of diseases. However, more impressive is the fact that the identification of the genes and the polymorphisms involved in susceptibility is opening new avenues of study. The best example at hand is the recent identification of a polymorphism in the *PDCD1* (programmed cell death 1) gene as a susceptibility factor for systemic lupus erythematosus, coding for the immunoreceptor PD-1 [1]. The polymorphism identified, named PD1.3 and whose allele A is strongly associated with the disease, is so far the only polymorphism within the PDCD1 gene that can provide a functional explanation for the susceptibility related to this gene. Furthermore, the same allele A was associated to diabetes type 1 [2]. Association was also identified with rheumatoid arthritis [3].

The PD1.3 polymorphism is located in the fourth intron of the *PDCD1* gene [1]. Within the fourth intron there is a sequence of 160 base pairs enriched in binding sites for various transcription factors important in hematopoiesis, suggesting that this element might act as a regulatory enhancer. Importantly, the regulator element is not conserved in the mouse (ME Alarcón-Riquelme and L Prokunina, unpublished data), suggesting that the regulation of PD-1 is different in both species. The polymorphism associated with human lupus changed a common G nucleotide to an A nucleotide, thereby disrupting a binding site for what seemed to be the *RUNX1* transcription factor. The binding was tested on a simple band-shift assay (electrophoretic mobility-shift assay) with specific antibodies, experiments that supported the notion that the associated allelic variant did not allow binding of a protein complex and that the complex included, among other proteins, RUNX1 [1], thereby providing a functional explanation for the genetic association.

The potential role of RUNX1 was underscored by the recent finding by two groups describing polymorphisms strongly associated with psoriasis and rheumatoid arthritis [4,5], both of which, even if present in completely different genes, also disrupted binding sites for what seemed to be RUNX1. For rheumatoid arthritis [5], the authors investigated a complete 3-centimorgan genomic segment from human chromosome 5q31 that included the cytokine

TGF = transforming growth factor; Th = T helper; VEGF = vascular endothelial growth factor.

gene cluster, a cluster previously also linked to rheumatoid arthritis [6] and Crohn's disease [7] with a high-resolution single-nucleotide polymorphism genotyping. The search led to the pinning down of a single polymorphism disrupting the *RUNX1*-binding site within the organic cation transporter gene *SLC22A4* [5]. Furthermore, and providing a stronger case, the authors identified a preliminary association of rheumatoid arthritis with the *RUNX1* gene itself, with a SNP located in intron 6 of RUNX1 in chromosome 21q22. This is an interesting test and a first attempt to define disease pathways and identify susceptibility genes or susceptibility effects that might be epistatic, additive or independent.

Similarly, the psoriasis study analyzed a region previously identified by linkage in sibling pairs, and by thorough haplotype analysis narrowed it down to a single polymorphism (having excluded the remaining nine that showed association out of hundreds studied) that also disrupted a binding site for *RUNX1* [4]. This time the polymorphism was found in a non-coding intergenic region between *SLC9A3R1*, a solute carrier gene, and the *N*-acetyltransferase gene *NAT9*. It was impossible for the authors to determine which of the two genes was the target for the effects of the polymorphism, but *SLC9A3R1* was found expressed in skin and in T cells [4].

Thus genetics, in three studies, has led to the identification of at least four new genes potentially involved in autoimmunity. In the center, the runt-domain family of transcription factors seem to be potential major regulators.

In the studies described, the authors performed mobility assays and transfection experiments with which they could show the allelic effect of the polymorphisms on gene expression in reporter assays and their effect by cotransfection of RUNX1. In spite of these experiments, the possibility still remains that it is not RUNX1 the transcription factor that is binding to the altered sites, but that it might also be any of its sisters, RUNX2 or RUNX3. The reason for this is that the consensus sequence that is the binding site for the runt family of transcription factors is the same for all three members, so the artificial use of oligonucleotides or even co-transfection does not fully resolve the issue. At this point, only chromatin immunoprecipitation can directly provide an answer; with this technique we can analyze specifically which of the three transcription factors is binding in vivo to the target sequence in the gene of interest.

However, it is clear that the RUNX proteins have a role not yet understood in autoimmune diseases. What could this role be? The runt-domain family of transcription factors is involved in several diseases and acts on target genes in a variety of tissues [8]. The three members, RUNX1, RUNX2 and RUNX3, can be expressed in the same cell, but their binding to the consensus sequence is dependent on their relative levels and their affinity for the adaptor CBFB (core binding factor  $\beta$ ), with which all of the three can heterodimerize [9,10]. It is clear that each of the three RUNX proteins has different roles and that their tissue expression is different, but they might overlap in some of their functions. The runt domain is highly conserved down to Drosophila [11]. Indeed, the first member of the family of runt-domain transcription factors was the Drosophila regulatory gene runt, shown to determine segmentation patterns during embryogenesis and later found to have functions in sex determination and neurogenesis [12]. A second member, named lozenge, is required for cell patterning in the eye and for hematopoiesis. In humans the three genes are located in completely different chromosomes. RUNX1 is located in human chromosome 21, RUNX2 is located in chromosome 6, and RUNX3 is located in chromosome 1.

## The runt-domain family

Generally, the runt-domain transcription factors are considered to be repressors. Most of the studies performed so far in humans include the RUNX1 protein previously known as AML1a. AML1a was originally identified because it is frequently involved in mutations and translocations associated with acute myeloid leukemia [13]. The Aml1a-related translocations have provided an important source of study for the function of RUNX1 as a repressor as well as the proteins that have been found to be forming a fusion protein in various of the translocations. The t(8;21) translocation results in a fusion protein between RUNX1 and ETO, a zinc-finger protein that is most probably a transcription factor acting as a nuclear repressor [14-16]. Further translocations have been identified, including the t(12;21) translocation resulting in the fusion of RUNX1 with TEL [17-19], also a transcription factor, and a t(16;21) translocation in which RUNX1 fuses with MTG16 (myeloid transforming generelated protein 1) or the t(3;21) translocation involving the Evi-1 gene [20,21].

Thus, studies on the translocations and the resulting fusion proteins that disrupt RUNX1 or the fusion partner suggest a dominant-negative effect for RUNX1. Indeed, mice made deficient for RUNX1 lack development of their hematopoietic system in a dominant fashion [22]. In humans, haploinsufficiency due to structural mutations in RUNX1 leads to familial thrombocytopenia and a greatly increased risk for the development of acute myeloid leukemia [13,23,24]. As an observation, within a family described for RUNX1 haploinsufficiency, an individual with the mutation had rheumatoid arthritis [23].

Deficiency in RUNX2 (also called AML3) leads to bone malformation and boneless mice; RUNX2 is therefore of

major importance in skeletal development and in osteoblast and chondrocyte development [25,26], although recent evidence shows that RUNX1 might also be involved in skeletal development [27] and has been found expressed in the skin and other epithelial tissues [27]. Mice made deficient for RUNX3 develop gastric cancer, and these studies have also shown that RUNX3 is involved in the development of basal root ganglia [28,29].

However, there has never been any previous evidence that the RUNX proteins are involved in autoimmunity, either in mouse models or in human studies. The main reasons for this lack of evidence are that the recently produced deficiency models have strong dominant loss-of-function effects, and that RUNX1, the only one of the three to have been studied extensively in humans, has been related to leukemias.

This suggests that the effects of the RUNX proteins in autoimmunity are much more subtle and are possibly readable only at the level of specific cellular compartments; this is in line with what is expected for complex diseases.

## The RUNX proteins in immune development

Interestingly, conditional cellular models and the use of retroviral vectors have permitted the study of the RUNX proteins in more detail, although still in the mouse, and have provided evidence for the importance of the RUNX proteins in the immune system.

Both RUNX1 and RUNX3 are required in T cell development. It has recently been reported that RUNX1 is required for active repression in CD4<sup>-</sup>CD8<sup>-</sup> thymocytes, whereas RUNX3 is required for establishing epigenetic silencing in cytotoxic lineage thymocytes [30]. RUNX3deficient cytotoxic T cells, but not T helper (Th) cells, were reported to have defective responses to antigen, suggesting that RUNX proteins could have critical functions in lineage specification and in homeostasis of CD8-lineage T lymphocytes. In addition, RUNX1 and RUNX3 have been found to regulate the expression of CD4 during CD8 lineage commitment [31].

It has also been observed that RUNX1 inhibits the differentiation of naive CD4<sup>+</sup> T cells into the Th2 lineage [32]. This is done through direct influence on the main transcription factor regulating Th2 development, GATA-3.

Another interesting and recent finding is that the lack of RUNX3 in a mouse model results in eosinophilic airway inflammation. Interestingly, RUNX3 was found to be expressed in mouse mature dendritic cells and to mediate dendritic cell responses to transforming growth factor (TGF)- $\beta$  [33]. The authors observed that in the RUNX3 knockout mice, maturation of dendritic cells was accelerated when induced with lipopolysaccharide or

without induction, and showed an increased efficiency in stimulating T cells. It is also interesting that the skin epidermis of the RUNX3 knockout mice lacked epidermal Langerhans cells but not dendritic epidermal T cells.

RUNX3 is known to mediate lymphoid and myeloid activity of CD11a through direct interaction with its promoter, and the RUNX3 knockout mice showed aberrant expression of CD11a, CD11b and CD11c, the  $\beta_2$ -integrins.

The findings revealed by the RUNX3 knockout mouse might provide us with some ideas about how the involvement of the RUNX proteins could be explained in systemic lupus erythematosus, rheumatoid arthritis and psoriasis. It would be interesting to investigate the effect of the RUNX3 deficiency in another genetic backgound, to test whether a 'permissible' background would allow the development of an autoimmune phenotype.

#### **Regulation of targets of the RUNX proteins**

As mentioned previously, the RUNX proteins are transcription factors or repressors for various target genes, and their action might be modulated through many different signaling pathways exerting their affect at various cellular levels as well as at various developmental levels.

For example, RUNX2 is essential for skeletal development. It has been shown that RUNX2 is essential in osteoblast differentiation. RUNX2 regulates osteocalcin, osteoprotegerin, TGF- $\beta$  receptor 1, osteopontin and collagenase 3, among others, in osteoblasts [8,34,35]. Furthermore, RUNX2 is known to regulate the expression of osteopontin, collagenase 3 and vascular endothelial growth factor (VEGF) in chondrocytes [7,36–38].

A possibility exists that susceptibility to rheumatoid arthritis and part of the development of the disease might be related to the activity of RUNX2 in these tissues and its effect on some of the target genes, many of which, such as osteopontin [34,35], collagenase 3 [39] and VEGF [37], have been shown to have altered expression or have been otherwise implicated in rheumatoid arthritis. VEGF, a mediator of angiogenesis, has been correlated with disease severity and has also been found to be involved with psoriasis [40].

Both RUNX1 and RUNX3 have mainly been found to regulate genes expressed in lymphoid and myeloid cells. Among the targets of RUNX1 are the B cell-specific tyrosine kinase BLK, the T-cell antigen receptor  $\alpha$ ,  $\beta$ ,  $\gamma$  and  $\delta$  chains, CD3 and granulocyte/macrophage colony-stimulating factor in lymphoid cells. The genes encoding myeloperoxidase, complement receptor 1 and p21<sup>Waf1/Cip1</sup> have been shown to be among the target genes for RUNX1 in myeloid cells. Of these, p21 has been found to have a role in systemic lupus erythematosus [41] in animal

models, and there is extensive literature on the role of complement receptor 1 (previously known as the C3b receptor or CD35) in lupus and even in drug-induced systemic lupus erythematosus [42,43]. No targets have been thoroughly investigated for RUNX3. A more extensive list of target genes can be found in [8].

#### **Regulation of the RUNX proteins**

Little is known about the regulation of the RUNX proteins and the pathways in which they are controlled. Most of our knowledge comes from studies of RUNX2.

Structurally, the RUNX genes are very similar. In mammals, it seems that the gene encoding RUNX3 might have been the one from which the other two evolved [11]. Each of the RUNX genes is transcribed from two promoters [8]. For instance, RUNX2 is regulated distinctively in different tissues. Activator protein 1 regulates RUNX2 through binding to FosB in osteoblasts, whereas non-fimbrial adhesin (NFA)-1 regulates RUNX2 in non-osseous cells [44–46].

RUNX2 is also regulated by TGF- $\beta$ , and regulation by TGF- $\beta$  is dependent on the cellular compartment [47]. TGF- $\beta$  represses RUNX2 in an osteosarcoma cell line, whereas it induces RUNX2 in a myoblast precursor cell line. The effects of TGF- $\beta$  on RUNX2 seem to be mediated by the Smad factors [48]. Other proteins that regulate RUNX2 are the bone morphogenetic proteins, members of the TGF- $\beta$  superfamily [47]. These are also known to exert their effects through recruitment of the Smad proteins, in which case other Smads are involved. Tumor necrosis factor- $\alpha$  and FGF have also been shown to regulate RUNX2 [49]. In particular, tumor necrosis factor- $\alpha$  inhibits RUNX2.

It is interesting that retinoids bring about increased expression of the three RUNX proteins. Similarly, vitamin D3 also augmented the expression of the RUNX proteins in myeloid leukemia cells. It has recently been shown that estrogen (estradiol) enhances RUNX2 activity without changing RUNX2 expression or DNA binding affinity but through direct interaction with estrogen receptor  $\alpha$ . Glucocorticoids have been found to inhibit RUNX2 activity. All previous work suggests that RUNX2 might be very important in bone regeneration, bone formation and repair, and it is of particular interest when considering the susceptibility to response to treatment of patients with rheumatoid arthritis or to disease severity and damage.

Very little is known about the regulation of the other RUNX proteins, and it is evident that these have profound effects at numerous levels of cellular activities.

At present it is unclear how the RUNX proteins exert their effects and how their aberrant function leads to autoimmunity and inflammation. However, a new chapter of investigation has now been opened that might lead to many surprises [50].

# **Competing interests**

MEA-R is a shareholder or Everygene AB.

#### References

- Prokunina L, Castillejo-Lopez C, Oberg F, Gunnarsson I, Berg L, Magnusson V, Brookes AJ, Tentler D, Kristjansdottir H, Grondal G, et al.: A regulatory polymorphism in PDCD1 is associated with susceptibility to systemic lupus erythematosus in humans. Nat Genet 2002, 32:666-669.
- Nielsen C, Hansen D, Husby S, Jacobsen BB, Lillevang ST: Association of a putative regulatory polymorphism in the PD-1 gene with susceptibility to type 1 diabetes. *Tissue Antigens* 2003, 62:492-497.
- Prokunina L, Padyukov L, Bennet A, De Faire U, Wiman B, Prince J, Alfredsson L, Klareskog L, Alarcón-Riquelme ME: Association of the PD1.3 A allelle of the PDCD1 gene in patients with rheumatoid arthritis negative for rheumatoid factor and the shared epitope. *Arthritis Rheum* 2004, 50:1770-1773.
- Helms C, Cao L, Krueger JG, Wijsman EM, Chamian F, Gordon D, Heffernan M, Daw JA, Robarge J, Ott J, et al.: A putative RUNX1 binding site variant between SLC9A3R1 and NAT9 is associated with susceptibility to psoriasis. Nat Genet 2003.
- Tokuhiro S, Yamada R, Chang X, Suzuki A, Kochi Y, Sawada T, Suzuki M, Nagasaki M, Ohtsuki M, Ono M, et al.: An intronic SNP in a RUNX1 binding site of SLC22A4, encoding an organic cation transporter, is associated with rheumatoid arthritis. Nat Genet 2003, 35:341-348.
- John S, Eyre S, Myerscough A, Barrett J, Silman A, Ollier W, Worthington J: Linkage and association analysis of candidate genes in rheumatoid arthritis. J Rheumatol 2001, 28:1752-1755.
- Rioux JD, Daly MJ, Silverberg MS, Lindblad K, Steinhart H, Cohen Z, Delmonte T, Kocher K, Miller K, Guschwan S, et al.: Genetic variation in the 5q31 cytokine gene cluster confers susceptibility to Crohn disease. Nat Genet 2001, 29:223-228.
- 8. Otto F, Lubbert M, Stock M: Upstream and downstream targets of RUNX proteins. J Cell Biochem 2003, 89:9-18.
- Tahirov TH, Inoue-Bungo T, Morii H, Fujikawa A, Sasaki M, Kimura K, Shiina M, Sato K, Kumasaka T, Yamamoto M, et al.: Structural analyses of DNA recognition by the AML1/Runx-1 Runt domain and its allosteric control by CBFbeta. Cell 2001, 104: 755-767.
- Backstrom S, Huang SH, Wolf-Watz M, Xie XQ, Hard T, Grundstrom T, Sauer UH: Crystallization and preliminary studies of the DNA-binding runt domain of AML1. Acta Crystallogr D Biol Crystallogr 2001, 57:269-271.
- Rennert J, Coffman JA, Mushegian AR, Robertson AJ: The evolution of Runx genes I. A comparative study of sequences from phylogenetically diverse model organisms. *BMC Evol Biol* 2003, 3:4.
- 12. Coffman JA: Runx transcription factors and the developmental balance between cell proliferation and differentiation. *Cell Biol Int* 2003, **27**:315-324.
- 13. Barton K, Nucifora G: AML1 haploinsufficiency, gene dosage, and the predisposition to acute leukemia. *BioEssays* 2000, 22: 214-218.
- Erickson P, Gao J, Chang KS, Look T, Whisenant E, Raimondi S, Lasher R, Trujillo J, Rowley J, Drabkin H: Identification of breakpoints in t(8;21) acute myelogenous leukemia and isolation of a fusion transcript, AML1/ETO, with similarity to Drosophila segmentation gene, runt. Blood 1992, 80:1825-1831.
- Guerrasio A, Rosso C, Martinelli G, Lo Coco F, Pampinella M, Santoro A, Lanza C, Allione B, Resegotti L, Saglio G: Polyclonal haemopoieses associated with long-term persistence of the AML1-ETO transcript in patients with FAB M2 acute myeloid leukaemia in continous clinical remission. *Br J Haematol* 1995, 90:364-368.
- Kozu T, Miyoshi H, Shimizu K, Maseki N, Kaneko Y, Asou H, Kamada N, Ohki M: Junctions of the AML1/MTG8(ETO) fusion are constant in t(8;21) acute myeloid leukemia detected by

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reverse transcription polymerase chain reaction. Blood 1993, 82:1270-1276.

- Uchida H, Downing JR, Miyazaki Y, Frank R, Zhang J, Nimer SD: Three distinct domains in TEL-AML1 are required for transcriptional repression of the IL-3 promoter. Oncogene 1999, 18:1015-1022.
- Song H, Kim JH, Rho JK, Park SY, Kim CG, Choe SY: Functional characterization of TEL/AML1 fusion protein in the regulation of human CR1 gene promoter. *Mol Cells* 1999, 9:560-563.
- Romana SP, Mauchauffe M, Le Coniat M, Chumakov I, Le Paslier D, Berger R, Bernard OA: The t(12;21) of acute lymphoblastic leukemia results in a tel-AML1 gene fusion. *Blood* 1995, 85: 3662-3670.
- Kurokawa M, Ogawa S, Tanaka T, Mitani K, Yazaki Y, Witte ON, Hirai H: The AML1/Evi-1 fusion protein in the t(3;21) translocation exhibits transforming activity on Rat1 fibroblasts with dependence on the Evi-1 sequence. Oncogene 1995, 11:833-840.
- Lutterbach B, Hou Y, Durst KL, Hiebert SW: The inv(16) encodes an acute myeloid leukemia 1 transcriptional corepressor. Proc Natl Acad Sci USA 1999, 96:12822-12827.
- Okuda T, Nishimura M, Nakao M, Fujita Y: RUNX1/AML1: a central player in hematopoiesis. Int J Hematol 2001, 74:252-257.
- Song WJ, Sullivan MG, Legare RD, Hutchings S, Tan X, Kufrin D, Ratajczak J, Resende IC, Haworth C, Hock R, et al.: Haploinsufficiency of CBFA2 causes familial thrombocytopenia with propensity to develop acute myelogenous leukaemia. Nat Genet 1999, 23:166-175.
- Michaud J, Wu F, Osato M, Cottles GM, Yanagida M, Asou N, Shigesada K, Ito Y, Benson KF, Raskind WH, et al.: In vitro analyses of known and novel RUNX1/AML1 mutations in dominant familial platelet disorder with predisposition to acute myelogenous leukemia: implications for mechanisms of pathogenesis. Blood 2002, 99:1364-1372.
- Komori T: Requisite roles of Runx2 and Cbfb in skeletal development. J Bone Miner Metab 2003, 21:193-197.
- Kundu M, Javed A, Jeon JP, Horner A, Shum L, Eckhaus M, Muenke M, Lian JB, Yang Y, Nuckolls GH, et al.: Cbfβ interacts with Runx2 and has a critical role in bone development. Nat Genet 2002, 32:639-644.
- Lian JB, Balint E, Javed A, Drissi H, Vitti R, Quinlan EJ, Zhang L, Van Wijnen AJ, Stein JL, Speck N, *et al.*: Runx1/AML1 hematopoietic transcription factor contributes to skeletal development in vivo. J Cell Physiol 2003, 196:301-311.
  Inoue K, Ozaki S, Ito K, Iseda T, Kawaguchi S, Ogawa M, Bae SC,
- Inoue K, Ozaki S, Ito K, Iseda T, Kawaguchi S, Ogawa M, Bae SC, Yamashita N, Itohara S, Kudo N, et al.: Runx3 is essential for the target-specific axon pathfinding of trkc-expressing dorsal root ganglion neurons. Blood Cells Mol Dis 2003, 30:157-160.
- Moss SF: RUNX 3, apoptosis 0: a new gastric tumour suppressor. Gut 2003, 52:12-13.
- Woolf E, Xiao C, Fainaru O, Lotem J, Rosen D, Negreanu V, Bernstein Y, Goldenberg D, Brenner O, Berke G, et al.: Runx3 and Runx1 are required for CD8 T cell development during thymopoiesis. Proc Natl Acad Sci USA 2003, 100:7731-7736.
- Taniuchi I, Osato M, Egawa T, Sunshine MJ, Bae SC, Komori T, Ito Y, Littman DR: Differential requirements for Runx proteins in CD4 repression and epigenetic silencing during T lymphocyte development. *Cell* 2002, 111:621-633.
- 32. Komine O, Hayashi K, Natsume W, Watanabe T, Seki Y, Seki N, Yagi R, Sukzuki W, Tamauchi H, Hozumi K, et al.: The Runx1 transcription factor inhibits the differentiation of naive CD4+ T cells into the Th2 lineage by repressing GATA3 expression. J Exp Med 2003, 198:51-61.
- Fainaru O, Woolf E, Lotem J, Yarmus M, Brenner O, Goldenberg D, Negreanu V, Bernstein Y, Levanon D, Jung S, *et al.*: Runx3 regulates mouse TGFβ-mediated dendritic cell function and its absence results in airway inflammation. *EMBO J* 2004, 23: 969-979.
- Petrow PK, Hummel KM, Schedel J, Franz JK, Klein CL, Muller-Ladner U, Kriegsmann J, Chang PL, Prince CW, Gay RE, et al.: Expression of osteopontin messenger RNA and protein in rheumatoid arthritis: effects of osteopontin on the release of collagenase 1 from articular chondrocytes and synovial fibroblasts. Arthritis Rheum 2000, 43:1597-1605.
- Ohshima S, Yamaguchi N, Nishioka K, Mima T, Ishii T, Umeshita-Sasai M, Kobayashi H, Shimizu M, Katada Y, Wakitani S, et al.:

Enhanced local production of osteopontin in rheumatoid joints. *J Rheumatol* 2002, **29**:2061-2067.

- Stricker S, Fundele R, Vortkamp A, Mundlos S: Role of Runx genes in chondrocyte differentiation. *Dev Biol* 2002, 245:95-108.
- Ballara S, Taylor PC, Reusch P, Marme D, Feldmann M, Maini RN, Paleolog EM: Raised serum vascular endothelial growth factor levels are associated with destructive change in inflammatory arthritis. Arthritis Rheum 2001, 44:2055-2064.
- Paleolog EM, Young S, Stark AC, McCloskey RV, Feldmann M, Maini RN: Modulation of angiogenic vascular endothelial growth factor by tumor necrosis factor alpha and interleukin-1 in rheumatoid arthritis. Arthritis Rheum 1998, 41:1258-1265.
- Lindy O, Konttinen YT, Sorsa T, Ding Y, Santavirta S, Ceponis A, Lopez-Otin C: Matrix metalloproteinase 13 (collagenase 3) in human rheumatoid synovium. Arthritis Rheum 1997, 40:1391-1399.
- Detmar M: Evidence for vascular endothelial growth factor (VEGF) as a modifier gene in psoriasis. J Invest Dermatol 2004, 122:xiv-xv.
- Balomenos D, Martin-Caballero J, Garcia MI, Prieto I, Flores JM, Serrano M, Martinez AC: The cell cycle inhibitor p21 controls Tcell proliferation and sex-linked lupus development. *Nat Med* 2000, 6:171-176.
- Iida K, Mornaghi R, Nussenzweig V: Complement receptor (CR1) deficiency in erythrocytes from patients with systemic lupus erythematosus. J Exp Med 1982, 155:1427-1438.
- Moulds JM, Reveille JD, Arnett FC: Structural polymorphisms of complement receptor 1 (CR1) in systemic lupus erythematosus (SLE) patients and normal controls of three ethnic groups. *Clin Exp Immunol* 1996, 105:302-305.
- 44. Banerjee C, Hiebert SW, Stein JL, Lian JB, Stein GS: An AML-1 consensus sequence binds an osteoblast-specific complex and transcriptionally activates the osteocalcin gene. Proc Natl Acad Sci USA 1996, 93:4968-4973.
- 45. Hoffmann HM, Beumer TL, Rahman S, McCabe LR, Banerjee C, Aslam F, Tiro JA, van Wijnen AJ, Stein JL, Stein GS, et al.: Bone tissue-specific transcription of the osteocalcin gene: role of an activator osteoblast-specific complex and suppressor hox proteins that bind the OC box. J Cell Biochem 1996, 61:310-324.
- Prince M, Banerjee C, Javed A, Green J, Lian JB, Stein GS, Bodine PV, Komm BS: Expression and regulation of Runx2/Cbfa1 and osteoblast phenotypic markers during the growth and differentiation of human osteoblasts. J Cell Biochem 2001, 80:424-440.
- Bae SC, Lee KS, Zhang YW, Ito Y: Intimate relationship between TGF-beta/BMP signaling and runt domain transcription factor, PEBP2/CBF. J Bone Joint Surg Am 2001, 83-A Suppl 1:S48-S55.
- Leboy P, Grasso-Knight G, D'Angelo M, Volk SW, Lian JV, Drissi H, Stein GS, Adams SL: Smad-Runx interactions during chondrocyte maturation. J Bone Joint Surg Am 2001, 83-A Suppl 1: S15-S22.
- Ito Y, Miyazono K: RUNX transcription factors as key targets of TGF-beta superfamily signaling. Curr Opin Genet Dev 2003, 13:43-47.
- Alarcon-Riquelme ME: A RUNX trio with a taste for autoimmunity. Nat Genet 2003, 35:299-300.