



Review

Opportunities, Challenges and Pitfalls of Using Cannabidiol as an Adjuvant Drug in COVID-19 †

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† To the memory of our friend Professor Daniel Moura (Department of Pharmacology, University of Porto, Portugal), who was scientifically active in the field of ACE2 and angiotensin II.

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Abstract: Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection may lead to coronavirus disease 2019 (COVID-19) which, in turn, may be associated with multiple organ dysfunction. In this review, we present advantages and disadvantages of cannabidiol (CBD), a non-intoxicating phytocannabinoid from the cannabis plant, as a potential agent for the treatment of COVID-19. CBD has been shown to downregulate proteins responsible for viral entry and to inhibit SARS-CoV-2 replication. Preclinical studies have demonstrated its effectiveness against diseases of the respiratory system as well as its cardioprotective, nephroprotective, hepatoprotective, neuroprotective and anti-convulsant properties, that is, effects that may be beneficial for COVID-19. Only the latter two properties have been demonstrated in clinical studies, which also revealed anxiolytic and antinociceptive effects of CBD (given alone or together with Δ^9 -tetrahydrocannabinol), which may be important for an adjuvant treatment to improve the quality of life in patients with COVID-19 and to limit post-traumatic stress symptoms. However, one should be aware of side effects of CBD (which are rarely serious), drug interactions (also extending to drugs acting against COVID-19) and the proper route of its administration (vaping may be dangerous). Clearly, further clinical studies are necessary to prove the suitability of CBD for the treatment of COVID-19.

Keywords: ACE2; cannabidiol; COVID-19; SARS-CoV-2; respiratory disease



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1. Introduction

Coronavirus disease 2019 (COVID-19) is caused by a new virus entity, the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and has grown to be one of the most dangerous pandemics in the history of mankind [1–3]. Its death toll of more than 2.2 million (1 February 2021) people has a similar magnitude like the Asian flu (1957–1958; 1–4 million) and Hong Kong flu (1968–1970; 1–4 million) and is already higher than that of the swine flu (2009–2010; up to 0.6 million; [4]). It is responsible for significantly more fatalities than the severe acute respiratory syndrome (SARS, 2002–2004; 774 deaths) and Middle East respiratory syndrome (MERS, 2012–present; 935 deaths) pandemics caused by the SARS-CoV and MERS-CoV viruses, respectively [4]; the latter two, like SARS-CoV-2, belong to the *Coronaviridae* family [5].

Vaccines protecting against SARS-CoV-2 have become available within a year of its emergence and they appear to be effective and safe [6,7]. Although general vaccination began in December 2020 in many countries, it will take months until the number of vaccinated people is high enough to provide herd immunity [6]. Therapeutic approaches are largely symptomatic and supportive. Many drugs have been examined in clinical

studies, mostly with disappointing results [2,3,8,9]. Nonetheless, the antiviral drug remdesivir shortens hospitalization time and the glucocorticoid dexamethasone even reduces mortality [10,11]. In addition, it is suggested that alternative compounds, including phytochemicals and natural agents targeting coronavirus development directly or as a result of their immunomodulatory effects, could be applied as potential therapies and for the prevention of COVID-19 [12–15].

The two senior authors of the present article (B.M. and E.S.) serve as editors of a Special Issue of *Int. J. Mol. Sci.* dedicated to research on cannabidiol (CBD), which has been suggested as a putative drug against COVID-19 but the views of different investigators on this compound have been found to vary greatly. Thus, we decided to conduct a review on the advantages and disadvantages of using CBD as a potential agent for the prevention and treatment of COVID-19 based on scientific reports on its influence on selected disease models and in clinical studies. We also summarize current knowledge about the effects of this compound on SARS-CoV-2 infection. The aim of the present review is to examine the suitability of CBD (Section 3) as an antiviral drug against SARS-CoV-2 (Section 4) and as an agent for the prevention and treatment of disease states in the preclinical (Section 5) and clinical (Section 6) settings. The review ends with a synopsis of the pros and cons of using CBD as a potential drug to treat COVID-19 (Section 7).

2. Mode of Infection and Symptoms of COVID-19

Infection with SARS-CoV-2 occurs mainly by aerosol/droplet transmission through direct contact with an infected person. The virus enters the body through the epithelial cells of the tongue, bronchi and lungs after attaching to angiotensin-converting enzyme 2 (ACE2). An important function of membrane-bound and soluble ACE2 is degradation of angiotensin II (Ang II) to angiotensin 1-7 (Ang 1-7), which exerts a beneficial influence, as opposed to numerous detrimental effects of high concentrations of Ang II (for details, see Figure 1 and Supplementary Table S1). The affinity of SARS-CoV-2 to ACE2 is determined by glycoprotein S1 localized on the characteristic viral “spikes.” S1 binds to the enzyme via the receptor-binding domain. Transmembrane serine protease 2 (TMPRSS2), which allows the entry of the virus into the cytoplasm of host cells [1,2,5,16], also plays a significant role in COVID-19 infection. Moreover, quite recently, it was shown that the membrane protein neuropilin-1 (NRP1) promotes SARS-CoV-2 entry [17].

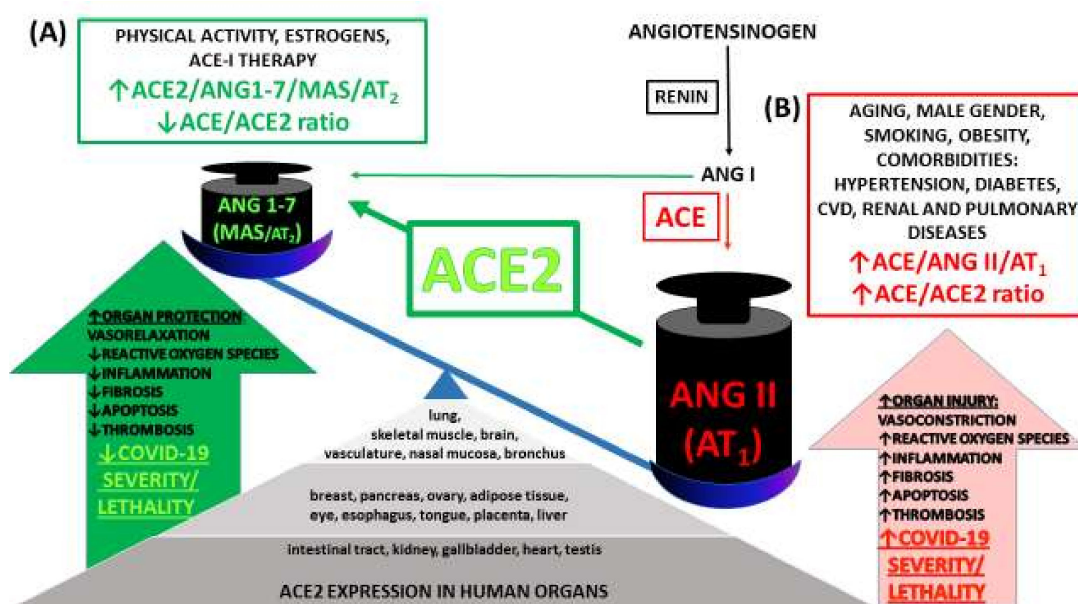


Figure 1. Expression of angiotensin-converting enzyme 2 (ACE2) in human tissues and organs, its counter-regulatory effects on the ACE → Ang II → AT₁ axis and interaction with coronavirus disease 2019 (COVID-19). ACE2 is ubiquitous and widely expressed in many organs targeted and damaged by COVID-19 caused by severe acute respiratory syndrome coronavirus 2

(SARS-CoV-2). It is a membrane-bound enzyme and an endogenous counter-regulator of the renin-angiotensin hormonal cascade. It degrades angiotensin II (Ang II) to angiotensin 1-7 (Ang 1-7) that exerts beneficial effects opposed to those of Ang II. Ang 1-7 acts through the G protein-coupled receptor MAS and, to a lesser extent, Ang II type 2 receptors (AT₂). ACE and ACE2 and their major products, Ang II and Ang 1-7, respectively, are linked in almost a ying/yang process, that is, when one decreases, the other increases and vice versa [18]. Thus, reduced activity of the deleterious ACE → Ang II → Ang II receptor type 1 (AT₁) axis (red) is coupled with increased activity of the protective ACE2 → Ang 1-7 → MAS receptor axis (green). A lower ACE/ACE2 ratio (A) (occurring in women, in exercise-trained individuals and patients well-treated with ACE inhibitors (ACE-I)) leads to beneficial effects such as vasorelaxation, anti-inflammatory, anti-oxidative, anti-fibrotic and anti-thrombotic effects that predispose towards a lower risk of cardiovascular disease (CVD) and better COVID-19 outcomes. By contrast, a high ACE/ACE2 ratio (B) that is increased in males, elderly and many pathologies (especially CVD, pulmonary and renal diseases and obesity) may aggravate COVID-19 infection [19–22].

COVID-19 is usually asymptomatic. In most symptomatic patients, SARS-CoV-2 infection is mild with symptoms including fever, shortness of breath, coughing, fatigue, anosmia, ageusia and muscle pain. Less common symptoms are nausea, vomiting and diarrhea [1,2,16,23]. The risk of developing a severe form of the disease increases with male gender, age and smoking; a further aggravation occurs in people with comorbidities such as hypertension, diabetes, obesity, cardiovascular or chronic respiratory system diseases, in which the concentration of the soluble form of ACE2 is substantially higher (Figure 1). By contrast, in healthy individuals ACE2 activity is much lower or not detected [24,25].

Because ACE2 is ubiquitous and widely expressed in the heart, blood vessels, gut, lungs (particularly in type 2 pneumocytes and macrophages) and in different types of cells (for details, see Figure 1 and Supplementary Table S1), COVID-19 is a multiple-organ disease, as summarized in Figure 2. One of the most common complications is an acute respiratory distress syndrome (ARDS) resulting from the cytokine storm phenomenon, a violent and uncontrolled inflammatory reaction in response to the presence of the virus in the host organism [1,2,16,23]. However, the negative consequences of COVID-19 are also connected with other disorders of the respiratory system [1,16,26,27], with the cardiovascular system (for review, see [1,16,22,26,28–30]) and with dangerous hematological complications, particularly thromboembolism [1,16,31–36]. In addition, pathological changes in the renal [1,16,27,33,37,38], gastrointestinal, hepatic, pancreatic [1,16,27,31,39,40] and nervous systems [1,27,41–43] have been described. Disturbances of the eyes [1,44], endocrine [45,46] and reproductive functions [31,45–47], skeletal muscles [1,33,48] and skin [1,49,50] occur as well (for details, see Figure 2).

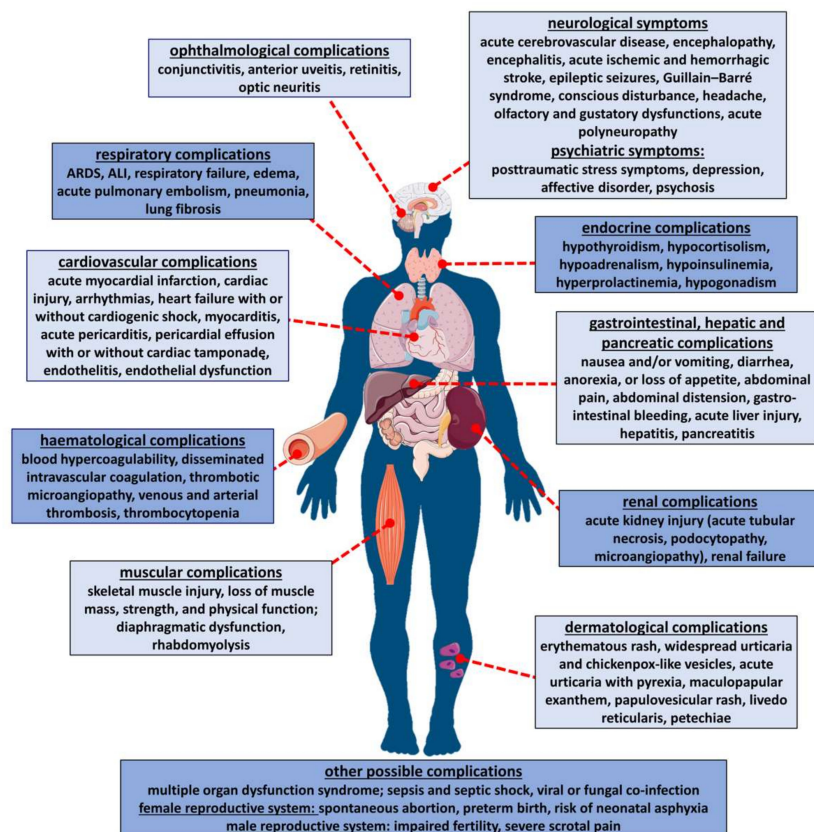


Figure 2. Systemic manifestation of COVID-19 infection. For respective literature, see Section 2. ALL, acute lung injury; ARDS, acute respiratory distress syndrome.

3. Cannabidiol—Pharmacological Potential and Mechanism of Action

Non-intoxicating CBD is a phytocannabinoid isolated from the *Cannabis sativa* plant [13,14] and, apart from the psychoactive Δ^9 -tetrahydrocannabinol (THC), represents the best-studied compound in this group.

Numerous studies have demonstrated a range of beneficial properties associated with CBD, including anti-inflammatory, antioxidant, antiarthritic, cardio- and neuro-protective, anticonvulsant, procognitive and analgesic effects (for detail, see reviews, for example, [51–53]). At the moment, there are two indications. CBD (Epidiolex®) has been approved for the treatment of intractable childhood-onset epilepsy (Dravet and Lennox-Gastaut syndrome). Its combination with THC (nabiximols (Sativex®); 100 μ L of oromucosal spray contains 2.5 mg CBD and 2.7 mg THC) has been approved for the therapy of spasticity in multiple sclerosis. Moreover, a potential therapeutic effect of CBD is suggested for inflammatory and autoimmune diseases, anxiety disorders, schizophrenia, depression, Alzheimer’s disease, Parkinson’s disease, chronic pain, cancer and diabetic complications [54,55].

With respect to the potential use of CBD in COVID-19, sixteen publications have so far appeared. Four publications are experimental (see Section 4) whereas another twelve discuss data from the literature.

Within the group of papers dealing with the literature, seven publications consider the potential use of CBD for the treatment of COVID-19 mainly due to its potent anti-inflammatory activity [14,15,56–60]. However, as suggested by another three authors, there is a lack of high-quality studies dedicated to the anti-inflammatory effects of CBD [61,62] or to its effects on the central nervous system (e.g., anxiety or neurological complications) [63]. Two authors warn that the anti-inflammatory action of CBD might exert a potential detrimental effect on the immune system even leading to enhancement of viral infections [64,65].

One should also consider that vaping CBD was associated with severe COVID-19 problems [57] and not overlook the unclear impact of “over-the-counter” CBD on the immunity of the SARS-CoV-2 infection [62]. Due to the lack of rigorous legal regulations, CBD is indeed commonly used as over-the-counter product, often of unapproved and unknown composition [51,54,55].

The multi-directional properties of CBD mentioned above arise from its complex mechanism of action. CBD has a low affinity for cannabinoid receptors (CB-Rs); it acts as a negative allosteric modulator of cannabinoid receptor type 1 (CB₁-R) and as an inverse agonist of cannabinoid receptor type 2 (CB₂-R). In addition, CBD acts via many other molecular targets including G-protein-coupled receptors (GPCRs; for example, activation of the peroxisome proliferator-activated γ (PPAR γ) receptor and serotonin 5-HT_{1A} and 5-HT_{2A} receptors) and ionotropic receptors (e.g., activation of vanilloid TRPV1 but inhibition of serotonin 5-HT₃ receptors). Moreover, it inhibits various transporters (e.g., adenosine uptake) and enzyme activities (e.g., fatty acid amide hydrolase (FAAH), an enzyme responsible for the degradation of the endocannabinoid anandamide) (for details and other molecular targets, see reviews [51,66]). In order to explain its effect against oxidative/nitrative stress, direct effects on the mitochondria and nuclei have been taken into consideration as additional molecular mechanisms [66].

4. Potential Antiviral Activity of CBD

Various effects and molecular mechanisms of CBD have been described in the previous section and some of them may be helpful in the context of the COVID-19 infection. Before discussing them in preclinical studies (Section 5) and in the clinical setting (Section 6), we would like to draw the attention of the reader to four experimental papers on the basis of which the potential usefulness of CBD against the SARS-CoV-2 virus was suggested.

In the first paper, Wang et al. [67] examined whether the gateways for the entry of the virus into cells, ACE2 and TMPRSS2, are affected by CBD. They found that high-CBD *Cannabis sativa* extracts decreased ACE2 and TMPRSS2 protein levels in artificial human 3D models of oral, airway or intestinal tissues primed by tumor necrosis factor α (TNF- α) plus interferon γ (IFN- γ). Extracts had different cannabinoid and terpene profiles and not all extracts under study were equally effective. In particular, pure CBD failed to affect ACE2 and TMPRSS2 protein levels in the airways, suggesting an entourage effect of the components of the extracts; the possibility that a component in the extracts other than CBD is the active principal component can so far not be excluded. Some extracts produced undesired molecular effects, that is, upregulated the levels of the ACE2 gene and protein. The idea of using high-CBD products (administered, for example, via mouth wash) to limit the entry of SARS-CoV-2 into susceptible hosts seems to be attractive but requires unambiguous scientific confirmation [56].

In the second paper (Raj et al. [68]), a direct antiviral effect of CBD was identified. The authors first screened several cannabinoids *in silico* and then examined CBD and THC, which appeared to have particularly promising effects in cultured Vero cells infected with SARS-CoV-2. CBD exhibited an IC₅₀ value of 8 μ M for its inhibitory effect on SARS-CoV-2 replication and was at least as potent, in this respect, as the antiviral drugs remdesivir, chloroquine and lopinavir, which are already used for the treatment of COVID-19 [2,3,8,9]. Again, these interesting data await unambiguous confirmation. By the way, CBD also inhibits the replication of hepatitis C virus *in vitro* [69] but was not active against the hepatitis B virus [69] or the Kaposi sarcoma-associated herpesvirus [70].

In the third and fourth experimental study suggesting the potential usefulness of CBD for treatment of COVID-19 [71,72] an acute respiratory distress syndrome was induced in mice by poly(I:C), a synthetic analogue of viral double-stranded RNA. Table 1 shows that CBD indeed had a beneficial effect in this condition. Chronic CBD application was also effective against the negative consequences of the infection in mice suffering from Theiler’s murine encephalomyelitis virus, which induces demyelinating disease [73,74]. However, in the aforementioned *in vivo* experiments, the beneficial effects of chronic

CBD administration were due to its anti-inflammatory properties and not direct antiviral activities (Table 1).

5. Preclinical Studies on the Use of CBD for COVID-19 Treatment

As discussed in the previous section, CBD has a direct as well as an indirect antiviral effect by reducing the number of ACE2 molecules via which the SARS-CoV-2 virus enters cells. Table 1 shows that, in preclinical models, CBD has a beneficial effect in many disease states also occurring in COVID-19. We mainly concentrated on the in vivo effects of chronic CBD administration. Acute in vivo or in vitro effects were considered if they were of interest with respect to COVID-19. Table 1 differentiates between prophylactic (preventive) and therapeutic CBD administration, that is, whether CBD was given before (or simultaneously with) the stimulus leading to the disease or after the pathological state had fully developed, respectively.

First of all, we would like to underline the effectiveness of CBD found in experimental models of respiratory failure. As already mentioned in the previous section, chronic CBD administration reduced the acute respiratory distress syndrome and the cytokine storm induced by polyriboinosinic:polyribocytidylic acid (poly(I:C)), a synthetic analogue of viral RNA [71,72]. Interestingly, it increased the expression of blood apelin [72], which serves as a catalytic substrate for ACE2 [24]. Moreover, the administration (mainly intraperitoneally (i.p.)) of CBD improved lung function and reduced inflammation in experimental acute lung injury (ALI) [75,76], pulmonary hypertension [77], lung injury induced by brain hypoxic/ischemic damage [78] and asthma [79,80] (Table 1). The beneficial influence of CBD resulted mainly from its significant anti-inflammatory properties [75,76,78–80]. Importantly, CBD has been shown to improve lung function [76], gas exchange [77], blood oxygen saturation [77] and to reduce allergen-induced airway obstruction [81]. Undoubtedly, a favorable effect of CBD is also the strong relaxation of the human pulmonary artery determined under in vitro conditions [82]. The unequivocally beneficial action profile of CBD is, however, contrasted by the results obtained by Karmaus et al. [83], who described a proinflammatory effect of prophylactically administered CBD (once daily for 3 days) in lipopolysaccharide-induced lung inflammation in mice. Moreover, CBD does not have antitussive properties and does not affect trachea contraction [84]. In most studies, CBD (i.p. or per os (p.o.)) in doses of 5–10 mg/kg per day) was administered prophylactically or therapeutically for 2–4 days only (Table 1). Only in the rat model of monocrotaline-induced pulmonary hypertension CBD was given prophylactically for 3 weeks [77].

A cardioprotective influence of CBD has been mainly shown in experiments in which prophylactic administration (predominantly in one dose given before occlusion or reperfusion) prevented the negative consequences of experimental myocardial infarction by decreasing the infarct size or arrhythmia ([85–88]; Table 1). Beneficial therapeutic effects (including improvement of cardiac systolic and diastolic dysfunction, reduction in coronary vasoconstriction, enhancement of mesenteric artery vasorelaxation and improvement of metabolic parameters) of chronic CBD administration (1 to 11 weeks) have been demonstrated in autoimmune myocarditis [89], diabetic cardiomyopathy [90], primary and secondary hypertension [91,92] and in Zucker diabetic fatty rats [93]. As in the case of respiratory failure, the favorable influence of CBD on the cardiovascular system is connected mainly with its anti-inflammatory properties and, in addition, with its antioxidative, antinitrative and antifibrotic effects (for details, see Table 1).

There is a significant association between severe COVID-19 and the occurrence of thromboembolism (for review, see [1,16,32–36]). Unfortunately, only a few publications have addressed the influence of CBD on the components or parameters of hemostasis. Thus, the chronic administration of CBD normalized the plasma tissue plasminogen activator and plasminogen activator inhibitor-1 enhanced by monocrotaline-induced pulmonary hypertension in rats [77]. A decrease in platelet aggregation occurred after a single CBD dose given before (but not after) myocardial infarction [87]. CBD (injected prophylactically,

acutely or chronically) failed to reduce vessel thrombogenesis and did not modify human platelet aggregation when given *in vitro* [94].

Nephroprotective properties of CBD have been described only for its acute administration (one or maximally two doses) in experimental models of ischemic acute kidney injury [95–97]. They were connected with its anti-inflammatory, antioxidative and anti-inflammatory properties (Table 1).

With respect to disorders of the gastrointestinal tract, cases of acute and chronic prophylactic CBD administration have been associated with anti-nausea and antiemetic effects in experimental nausea and vomiting induced by lithium chloride via the activation of serotonin 5-HT_{1A} receptors [98]. Since CBD antagonizes 5-HT₃ receptors under *in vivo* conditions [99], its antiemetic effect might also result from the antagonism of 5-HT₃ receptors. Hepatoprotective properties of CBD have been described in models of liver injury induced by hepatic artery and portal vein occlusion [100], chronic ethanol administration [101], thioacetamide [102] or cocaine [103]. Again, the beneficial influence of CBD is connected mainly with its anti-inflammatory and antioxidative properties. Unfortunately, with the exception of chronic ethanol administration [101], CBD was only given once. Prophylactic administration of CBD had a beneficial influence in experimental acute pancreatitis, based on its anti-inflammatory properties [104]. Moreover, its therapeutic administration at one [105] or three [106] doses was shown to reduce intestinal hypermotility (Table 1). Interestingly, cannabis extract with a high CBD content reduced inflammatory changes in the colon more strongly than CBD did alone [106].

The most common symptoms of COVID-19 include anosmia, ageusia and fever. A PubMed-based search did not identify any publications showing that CBD is useful in the case of loss of smell or taste. With respect to the nervous system, one might mention its prohedonic activity, which occurred upon chronic administration in rats that were exposed to chronic unpredictable mild stress [107]. Moreover, CBD is approved for the treatment of Dravet syndrome, a condition that features recurrent seizures triggered by fever [108]. Its effective anticonvulsant activity has been confirmed both in human (for review, see [108]) and different experimental models (for example [109,110], see Table 1). In addition, a neuroprotective influence of CBD (administered once or twice) has been determined in hepatic encephalopathy [102], perinatal hypoxia/ischemia encephalopathy [111,112], sepsis-related encephalitis (including increase in integrity of blood-brain barrier; [113]) or cerebral ischemia [114,115]. CBD (given for up to 10 days) had also a beneficial effect on encephalomyelitis and multiple sclerosis induced by Theiler's murine encephalomyelitis virus [73,74].

Tears and the eyes in general, create portals for coronavirus entry. Although we did not find any publications regarding the potential application of CBD for conjunctivitis (the most common ocular manifestation of COVID-19), one should remember that CBD has been suggested as a putative novel therapy for diabetic retinopathy [116] and retinal inflammation ([117]; Table 1). Both protective effects are associated with the anti-inflammatory and antioxidative actions of CBD which are also beneficial in the reduction of the negative consequences of perinatal hypoxia/ischemia [111,112]. In this context, one may consider the risk of neonatal asphyxia in children from mothers suffering from COVID-19 (Figure 2).

Unfortunately, there are limited publications suggesting potential beneficial effects of CBD in endocrine, muscular and dermatological disorders, that also are listed among those related to COVID-19 (Table 1). Thus, chronic CBD administration has been shown to reduce hyperglycemia and to improve metabolic dysfunction [118,119] in experimental models of obesity/diabetes, which is important if one considers that high glucose plasma levels and diabetes are risk factors for COVID-19 [120]. Moreover, CBD was found to prevent losses in functionality due to skeletal muscle degeneration [121]. Beneficial anti-inflammatory properties of chronic and acute administration of CBD on skin function have been shown in nude rats [122] and *in vitro* in human sebocytes, keratinocytes and skin organ culture [123,124].

Table 1. Potential use of cannabidiol (CBD) for COVID-19 treatment as suggested by preclinical studies.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Respiratory	<u>ARDS</u> induced by poly(I:C)	mouse	5 mg/kg, i.p. therapeutic, once a day for 3 days	cytokine storm and ARDS symptoms totally or partially improved (blood oxygen saturation, perivascular and peribronchiolar interstitial inflammatory infiltrate, lung fibrosis, hypertrophy and pulmonary edema) ↓IL-6 expression and ↓neutrophil frequency in the lung	<u>anti-inflammatory</u>	[71,72]
			5 mg/kg, i.p. therapeutic, once a day for 3 days	improvement of lung structure ↓T cells and ↑neutrophils returned towards the normal level, ↑expression of apelin in the blood	<u>anti-inflammatory</u> regulation of apelin level	[72]
	<u>ALI</u> induced by LPS	mouse	1–80 mg/kg, i.p. prophylactic; one dose before ALI induction (effects determined 1, 2 and 4 days after LPS)	<u>lungs</u> : ↓leukocyte count; ↓leukocyte migration into lungs; ↓MPO activity; ↓vascular permeability; <u>BALF</u> : ↓pro-inflammatory cytokines (TNF- α , IL-6) and chemokines (MCP-1, MIP-2)	<u>anti-inflammatory</u> ; partially dependent on adenosine A _{2A} -Rs	[75]
			20 or 80 mg/kg, i.p. therapeutic; one dose 6 h after ALI induction (effects determined 24 h after LPS)	<u>lungs</u> : function improved: ↓resistance; ↓tissue damping and stiffness; ↓leukocyte migration into lungs; ↓MPO activity; ↓vascular permeability; <u>BALF</u> : ↓pro-inflammatory cytokines (TNF- α , IL-6) and chemokines (MCP-1, MIP-2)	<u>anti-inflammatory</u> ; antagonists not used	[76]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Respiratory	lung inflammation induced by LPS	mouse	75 mg/kg, p.o. prophylactic; once a day for 3 days; LPS 1 h before the last dose of CBD	lungs: ↑inflammatory changes in tissue; BALF: ↑pro-inflammatory TNF- α , IL-6, IL-23, GCSF; slight ↑inflammatory cells	pro-inflammatory; antagonists not used	[83]
	pulmonary hypertension due to monocrotaline	rat	10 mg/kg, i.p. prophylactic; once a day for 21 days	heart: ↓right ventricular systolic pressure; ↔ right hypertrophy and lung edema; ↑blood oxygen saturation; plasma: ↓leukocytes	↑blood oxygen saturation; antagonists not used	[77]
	lung injury induced by brain hypoxic–ischemic damage	newborn piglets	1 mg/kg, i.v. therapeutic; one dose 30 min after lung injury	improvement of gas exchange; ↑TLC, lungs: ↓ histological damage and edema; ↓leukocyte migration into lungs, ↓inflammatory changes; ↓vascular permeability; BALF: ↓pro-inflammatory cytokines (IL-1)	anti-inflammatory 5-HT _{1A} -Rs in all parameters, except for improvement of gas exchange; site of CBD action (brain and/or lungs) unclear	[78]
		rat	5 mg/kg, i.p. therapeutic; once a day for 2 days	serum: ↓ IL-4, IL-5, IL-6, IL-13 and TNF- α ; ↔ IL-10	anti-inflammatory; antagonists not used	[79]
	asthma induced by ovalbumin	mouse	5 or 10 mg/kg, i.p. therapeutic; once a day for 3 days	↓airway resistance; ↓alveolar collapse areas; ↓collagen in airways and alveolar septa; lung and BALF: ↓pro-inflammatory cytokines (IL-4, IL-5, IL-13)	anti-inflammatory; anti-fibrotic; airway resistance: CB ₁ -Rs other effects: CB _{1/2} -Rs	[80]
	airway obstruction induced by ovalbumin	guinea-pig	1 mg/kg, i.v. prophylactic; one dose	↓airway obstruction induced by ovalbumin	bronchoprotective; reduction of the antigen-induced contractile responses	[81]
	cough induced by aerosolized citric acid	guinea-pig	prophylactic; aerosolized solution of 10 mg/mL for 20 minutes using a nebulizer	cough inhibition only in three out of eight animals	antagonists not used	[84]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Respiratory	isolated trachea	guinea-pig	10 μ M	\leftrightarrow resting basal tension; \leftrightarrow contractions induced electrically or by methacholine	antagonists not used	[84]
	isolated human pulmonary artery	human	0.1–30 μ M	almost full concentration-dependent vasorelaxation	endothelium-dependent vasodilatation mediated via K ⁺ channels, IP, EP ₄ , TRPV1 and PPAR γ receptors	[82]
Cardiovascular		rabbit	0.1 mg/kg, i.v. prophylactic; one dose before occlusion and one before reperfusion	<u>heart</u> : \downarrow infarct area; \uparrow left ventricular function; \uparrow blood supply to perfusion-defective region; \downarrow neutrophil infiltration; \downarrow MPO activity <u>plasma</u> : \downarrow cardiac troponin I	<u>anti-inflammatory;</u> <u>cardioprotective;</u> <u>potentially anti-ischemic;</u> antagonists not used	[85]
	<u>myocardial infarction induced by left coronary artery occlusion</u>		5 mg/kg, i.p. prophylactic before occlusion and once a day thereafter for 7 days	<u>heart</u> : \downarrow infarct size; \leftrightarrow HR; \downarrow leukocyte infiltration; <u>serum</u> : \downarrow IL-6 CRP, TNF- α	<u>anti-inflammatory;</u> <u>cardioprotective;</u> <u>potentially anti-ischemic;</u> antagonists not used	[86]
		rat	50 μ g/kg, i.v. prophylactic; one dose before occlusion	<u>heart</u> : \downarrow infarct area; \downarrow arrhythmias; \leftrightarrow HR	<u>antiarrhythmic;</u> <u>cardioprotective;</u> antagonists not used	[87]
			50 μ g/kg, i.v. prophylactic; one dose before reperfusion	<u>heart</u> : \downarrow infarct area; \leftrightarrow arrhythmias	antagonists not used	
			50 μ g/kg, i.v. prophylactic; one dose before occlusion	<u>heart</u> : \downarrow arrhythmias; \leftrightarrow HR	antiarrhythmic; potential involvement of A ₁ -Rs	[88]
	<u>autoimmune myocarditis</u>	mouse	10 mg/kg, i.p. therapeutic; once a day for 46 days	<u>heart</u> : improved systolic and diastolic dysfunction and myocardial stiffness; \downarrow left ventricular inflammatory changes; \downarrow necrosis; oxidative stress; \downarrow fibrosis	<u>anti-inflammatory;</u> <u>anti-oxidative;</u> <u>anti-fibrotic;</u> <u>cardioprotective</u> antagonists not used	[89]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Cardiovascular	diabetic cardiomyopathy induced by streptozotocin	mouse	1, 10 or 20 mg/kg, i.p. therapeutic; once a day for 4 or 11 weeks	heart: ↑diastolic and systolic left ventricular function; ↓oxidative and nitrative stress; ↓inflammation and NF-κB activation; ↓ fibrosis; ↓expression of pro-fibrotic genes	anti-inflammatory; antioxidative; anti-nitrative; anti-fibrotic; cardioprotective; antagonists not used	[90]
	changes in vascular endothelium function in Zucker diabetic fatty	rat	10 mg/kg, i.p. therapeutic for 7 days	mesenteric arteries: ↑endothelium-dependent vasorelaxation due to COX- or NO-mediated mechanisms; serum: ↓cardiovascular biomarkers (C-peptide, insulin and intracellular adhesion molecule-1); ↔glucose, body weight	vasoprotective; improvement in the profile of cardiovascular and metabolic parameters	[93]
	hypertension: primary (SHR); secondary (DOCA-salt)	rat	10 mg/kg, i.p. therapeutic; once a day for 2 weeks	↔blood pressure, HR; heart: ↓oxidative stress; ↓carbachol-induced coronary constriction; ↓left ventricular cardiomyocyte width; ↔left ventricular hypertrophy	anti-oxidative; antagonists not used	[91,92]
Hematological	myocardial infarction induced by left coronary artery occlusion	rat	- 50 µg/kg, i.v. before occlusion - 50 µg/kg, i.v. before reperfusion	↓platelet aggregation ↔platelet aggregation	antagonists not used	[87]
	pulmonary hypertension due to monocrotaline	rat	10 mg/kg, i.p. prophylactic; once a day for 21 days	plasma: ↓ t-PA and PAI-1	antagonists not used	[77]
	pharmacologically induced thrombus in ear venules	mouse	5 mg/kg, i.p. prophylactic; acute: one dose 30 min before thrombus induction; chronic for 3 days	acute: ↔vessel thrombogenesis chronic: ↔vessel thrombogenesis	antagonists not used	[94]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Hematological	<u>platelet aggregation</u>	human	in vitro, 0.1–10 μ M	\leftrightarrow resting platelets; \leftrightarrow activation induced by thrombin receptor activating peptide		[94]
	Renal	acute kidney injury induced by renal ischemia/reperfusion	rat	5 mg/kg, i.v. two doses before occlusion and after reperfusion	kidney: \downarrow tubular necrosis and dilatation; \downarrow inflammatory changes; \downarrow NF- κ B, COX-2, TNF- α and iNOS; serum: \downarrow oxidative and nitrative stress	anti-inflammatory; anti-oxidative; anti-nitrative; nephroprotective; antagonists not used
mouse			5 mg/kg, i.a. one dose after occlusion	kidney: \downarrow MPO activity; \downarrow IL-1, TNF- α and NO levels; \downarrow lipid and protein oxidative damage; \leftrightarrow nitrite/nitrate levels	anti-inflammatory; anti-oxidative; nephroprotective; CB ₁ -Rs and CB ₂ -Rs expression unaltered	[96]
mouse		10 mg/kg i.p. one dose before reperfusion	kidney: pattern of innate lymphoid cells restored to control values	nephroprotective; antagonists not used	[97]	
Gastrointestinal	<u>nausea or vomiting induced by lithium chloride</u>	rat shrew	prophylactic; acute: 5 and 20 mg/kg, s.c. chronic: 5 mg/kg s.c. for 7 days	acute and chronic: \downarrow nausea and/or vomiting	anti-nausea, antiemetic; 5-HT _{1A} -Rs	[98]
	<u>hepatic ischemia/reperfusion injury induced by hepatic artery and portal vein occlusion</u>	mouse	3 or 10 mg/kg, i.p. prophylactic; one dose before reocclusion or 90 min after	serum: \downarrow AST and ALT; liver: \downarrow inflammatory changes; \downarrow cell apoptosis (10 mg/kg only); \downarrow pro-inflammatory cytokines; \downarrow oxidative and nitrative stress (10 mg/kg only); \downarrow neutrophil migration to liver tissue	anti-inflammatory; antioxidant; anti-nitrative; hepatoprotective; independent of CB ₂ -Rs	[100]
	<u>liver injury and steatosis induced by chronic ethanol administration</u>	mouse	5 or 10 mg/kg, i.p. prophylactic for 11 days during the ethanol exposure	serum: \downarrow AST and ALT; liver: \downarrow inflammatory changes; \downarrow pro-inflammatory chemokines; \downarrow neutrophil accumulation; \downarrow oxidative burst of neutrophils; \downarrow oxidative and nitrative stress	anti-inflammatory; anti-oxidative; hepatoprotective; antagonists not used	[101]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Gastrointestinal	<u>hepatic encephalopathy due to thioacetamide</u>	mouse	5 mg/kg, i.p. <u>therapeutic</u> ; one dose after thioacetamide injection	<u>liver</u> : ↔necrosis; <u>plasma</u> : ↓ALT and AST, ammonia and bilirubin	<u>partly hepatoprotective</u> ; antagonists not used	[102]
	<u>acute hepatic toxicity induced by cocaine</u>	mouse	30, 60 and 90 mg/kg, i.p. <u>prophylactic</u> ; acute 30 min before cocaine injection	<u>liver</u> : ↓acute inflammation and damage (↓histological changes) <u>serum</u> : ↓ALT; ↓acute behavioral seizure	<u>anti-inflammatory</u> ; FAAH inhibitor did not modify cocaine-induced changes in liver	[103]
	<u>acute pancreatitis induced by cerulein</u>	mouse	0.5 mg/kg, i.p. <u>prophylactic</u> ; 8 doses (2 before and 6 simultaneously with cerulein)	<u>pancreas</u> : ↓pathological changes, ↓MPO activity in pancreas tissue; <u>plasma</u> : ↓amylase and lipase; ↓IL-6 and TNF-α	<u>anti-inflammatory</u> ; <u>pancreas-protective</u> ; possibly via GPR55 (presence in pancreas)	[104]
	<u>inflammation and intestinal hypermotility induced by croton oil</u>	mouse	5 and 10 mg/kg, i.p. <u>therapeutic</u> ; one dose to mice with inflammation	<u>intestine</u> : ↓hypermotility	involvement of CB ₁ -Rs (but not CB ₂ -Rs) and FAAH	[105]
	<u>colitis induced by intracolonic dinitrobenzensulfonic acid</u>	mouse	5–30 mg/kg, i.p. and 10–60 mg/kg, p.o. <u>therapeutic</u> for 3 days after colitis induction	<u>intestine</u> : ↓hypermotility; ↔colitis; ↓colon weight and MPO activity	<u>anti-inflammatory</u> ; antagonists not used	[106]
Neurological	<u>chronic unpredictable mild stress model of depression</u>	rat	10 mg/kg, i.p. <u>prophylactic</u> for 28 days	higher rate of body weight gain and sucrose preference compared to controls	<u>prohedonic</u> ; antagonists not used	[107]
	<u>various acute seizure models</u>	rat mouse	one different i.v. dose dependent on the model	acute antiseizure activity	<u>antiseizure activity</u>	[109]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Neurological	status epilepticus— spontaneous recurrent seizures (RISESRS) model	rat	200 mg/kg for 7 weeks	↓seizure burden and motor comorbidities; reversal of the epilepsy-induced cognitive deficits		[109]
	seizure induced by pentylene-tetrazole	mouse	60 mg/kg, i.p. prophylactic; once before induction of epileptic attack	↓seizure duration; ↓EEG changes	anti-convulsant; CB ₁ , CB ₂ and TRPV1 receptors	[110]
	hepatic encephalopathy induced by thioacetamide	mouse	5 mg/kg, i.p. therapeutic; one dose after thioacetamide injection	↑neurological and cognitive functions; ↑activity; ↓activated astrocytes	procognitive; neuroprotective; antagonists not used	[102]
	sepsis-related encephalitis induced by LPS	mouse	3 mg/kg, i.v. one dose simultaneously with LPS	↑integrity of blood–brain barrier; ↓leukocyte margination in brain vessels; ↔level of oxidative stress; ↓TNF-α and COX-2	anti-inflammatory; neuroprotective	[113]
	perinatal hypoxic- ischemic encephalopathy induced by occlusion of carotid arteries	newborn piglet	1 mg/kg, i.v. therapeutic; one dose 30 min after induction of brain injury	brain: ↓EEG changes; ↓neuronal mortality; ↓excitotoxicity; ↓IL-1; ↓oxidative stress	anti-inflammatory; anti-oxidative; neuroprotective partially dependent on 5-HT _{1A} -Rs and CB ₂ -Rs ↔brain endocannabinoid levels	[111]
	1 mg/kg, i.v. therapeutic; one dose after induction of brain injury		brain: ↑activity (EEG); ↓neuronal mortality; ↓excitotoxicity; ↓oxidative stress; ↓TNF-α; effects on excitotoxicity, oxidative stress and TNF-α additive to those of hypothermia	anti-inflammatory; anti-oxidative; neuroprotective; antagonists not used	[112]	

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Neurological	<u>cerebral ischemia</u> (stroke model) induced by MCA occlusion	mouse	0.1; 1; 3 mg/kg, i.p. two doses (before and after occlusion)	<u>brain</u> : ↑neurological function and motor coordination; dose-dependent ↓infarct area ↓MPO activity and ↑CBF (tested only at 3 mg/kg); <u>blood</u> : ↔pCO ₂ , pO ₂	<u>anti-inflammatory</u> ; <u>neuroprotective</u> ; partially dependent on 5-HT _{1A} -Rs independent of CB ₁ , CB ₂ and TRPV1 receptors	[114,115]
	<u>encephalomyelitis</u> induced by TMEV	mouse	180 mg/kg, i.p. twice daily starting 2 days before (prophylactic) or 3 days after infection (therapeutic)	↓acute behavioral seizures from 5 days (prophylactic) and 6 days after infection onward (therapeutic)	<u>anti-inflammatory</u> , <u>anti-oxidative</u> (not confirmed experimentally)	[73]
	<u>multiple sclerosis</u> induced by TMEV	mouse	5 mg/kg, i.p. therapeutic, once a day for 7 days	sub chronic effects (after 8 days): ↓transmigration of leukocytes to the nervous parenchyma by downregulating the expression of VCAM-1, CCL2 and CCL5 and the proinflammatory cytokine IL-1β and by attenuating the activation of microglia	<u>anti-inflammatory</u> partial involvement of adenosine A _{2A} -Rs (experiments with an appropriate antagonist)	[74]
			same treatment for 10 days	chronic effects (after 80 days): improvement of motor deficits ↓microglial activation and pro-inflammatory cytokine production		
Eye	<u>retinal inflammation</u> due to LPS	rat	1 mg/kg, i.p. prophylactic before LPS treatment	↓retinal TNF-α levels	<u>anti-inflammatory</u> ; due to A ₂ -Rs but not A ₁ -Rs	[117]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Important for COVID-19 Endocrine	<u>high-fat diet-induced obesity</u>	rat	10 mg/kg, i.p. <u>prophylactic</u> for 2 weeks	plasma: ↓insulin; skeletal muscle: improved insulin signal transduction and glycogen recovery	↓lipotoxicity, leading to insulin-sensitization in myocytes; ↓expression of CB ₁ , CB ₂ , TRPV1 and 5-HT _{1A} receptors	[118]
	<u>type 1 diabetes by streptozotocin submitted to chronic cerebral hypoperfusion</u>	rat	10 mg/kg, i.p. 30 min before and for 30 days after cerebral hypoperfusion surgery	↓body weight; plasma: ↓hyperglycemia; ↑insulinemia; ↓AGEs and fructosamine; ↓dyslipidemia (LDL, HDL, TGs and total cholesterol levels); ↓AST and ALT; ↑memory performance	improvement of metabolic dysfunction; <u>hepatoprotective;</u> <u>neuroprotective;</u> <u>anti-inflammatory</u>	[119]
Muscular	<u>Duchenne muscular dystrophy</u> caused by dystrophin deficiency	mouse	60 mg/kg, i.p., <u>therapeutic;</u> three times a week for 2 weeks	<u>muscle:</u> prevention of the functionality loss and tissue degeneration; restoration of locomotor activity; ↓inflammation (IL-6, TNF-α); muscle strength and autophagy restored	anti-inflammatory; (involvement of TRP channels—based on in vitro experiments)	[121]
Skin	skin irradiated with UVA/UVB	nude rat	2.5 g in 100 g petrolatum applied to the back of rats every 12 h for 4 weeks	↓UV-induced changes in inflammation; apoptosis and oxidative stress	prevention of UV-induced metabolic changes in epidermal keratinocytes	[122]
	cultured human sebocytes and human skin organ culture	human	10 μM	↓lipogenic actions of arachidonic acid and a combination of linoleic acid and testosterone; suppression of sebocyte proliferation	anti-inflammatory (adenosine A _{2A} Rs); <u>sebostatic;</u> lipostatic; antiproliferative (TRP4)	[123]
	experimental model of allergic contact dermatitis in keratinocytes	human	5, 10, 20 μM	inhibition of polyinosinic-polycytidylic acid-induced release of MCP-2, IL-6, IL-8 and TNF-α; no cytotoxic effect	anti-inflammatory via CB ₂ and TRPV1 receptors	[124]

Table 1. Cont.

	Model	Species	Dose and Route of Administration of CBD	Effects	CBD Properties Important for COVID-19; Mechanisms	References
Other	sepsis induced by cecal ligation and puncture	rat	10 mg/kg, i.p. Acute—therapeutic; one dose (after sepsis induction)	↓lipid peroxidation in lung, heart and kidney; ↓oxidative protein damage in spleen, liver and heart; ↓oxidative damage of proteins in striatum, cortex and hippocampus; ↑oxidative damage of proteins in lung	anti-oxidative; organo-protective; <u>pro-oxidative in lungs</u>	[125]
			10 mg/kg, i.p. Chronic—therapeutic; once daily for 9 days; first dose after sepsis induction	↓mortality; improvement of memory-related processes; ↓lipid peroxidation in kidney; ↓oxidative protein damage in spleen, liver, heart	anti-oxidative; organo-protective; <u>procognitive</u>	

The following antagonists were used to describe the multimodal mechanism of cannabidiol (CBD), namely A₂-Rs due to blockade by ZM241385 [75,117]; A₁-Rs by DPCPX [88]; 5-HT_{1A}-Rs by WAY100635 [78,98,111,114,115]; CB₁-Rs by AM251 [80,110] and by rimonabant [105,114,115], CB₂-Rs by AM630 [80,110,111,114,115,124] or by SR144528 [105]; TRPV1 by capsaizepine [82,114,115], SB36679 [110] or 5'-iodoresiniferatoxin [124] and IP, EP4 and PPAR (antagonism via L161982, Cay10441 and GW9662, respectively [82]). Moreover, some experiments were performed on knockout mice, that is, CB₂^{-/-} [100]. 5-HT_{1A}-Rs, serotonin receptor type 1A; A_{1A}-R, A_{2A}-R, adenosine receptor type A_{1A} and A_{2A}; AGEs, advanced glycation end-products; ALI, acute lung injury; ALT, alanine transaminase; ARDS, acute respiratory distress syndrome; AST, aspartate transaminase; BALF, bronchoalveolar lavage fluid; CB-R, cannabinoid receptor; CB₁-R, CB-R type 1; CB₂-R, CB-R type 2; CBD, cannabidiol; CBF, cerebral blood flow; CCL2, CCL5, C-C motif chemokine ligand 2 and 5; COX-2, cyclooxygenase 2; CRP, C-reactive protein; DOCA, deoxycorticosterone acetate; DPCPX, 8-cyclopentyl-1,3-dipropylxanthine; EEG, electroencephalography; EP₄, prostanoid EP₄ receptor; FAAH, fatty acid amide hydrolase; GCSF, granulocyte colony stimulating factor; GPR55, G protein-coupled receptor, resembling to some extent the CB-Rs; HDL, high density lipoprotein; HR, heart rate; i.a. intraarterially; IL-n, interleukin n, for example, IL-1, interleukin 1; iNOS, inducible nitric oxide synthase; IP, prostacyclin receptor; i.p. intraperitoneally; i.v. intravenously; LDL, low-density lipoprotein; LPS, lipopolysaccharide; MCA, middle cerebral artery; MCP-1, monocyte chemoattractant protein-1; MIP-2, macrophage inflammatory protein-2; MPO, myeloperoxidase; NF-κB, nuclear factor κB; NO, nitric oxide; PAI-1, plasminogen activator inhibitor-1; pCO₂, partial pressure of carbon dioxide; p.o. per os, orally; pO₂, partial pressure of oxygen; poly(I:C), polyriboinosinic:polyribocytidylic acid, synthetic analogue of viral double-stranded RNA; PPAR, peroxisome proliferator-activated receptor type gamma; SHR, spontaneously hypertensive rats; STZ, streptozotocin; TGs, triglycerides; TLC, total lung capacity; TMEV, Theiler's murine encephalomyelitis virus-induced demyelinating disease; TNF-α, tumor necrosis factor α; t-PA, tissue plasminogen activator; TRP, transient receptor potential; TRPVn, transient receptor potential vanilloid subfamily member n; UVA and UVB, ultraviolet A and B; VCAM-1, vascular cell adhesion molecule-1. ↑, increase; ↓, decrease; ↔, no change.

Importantly, the effectiveness of CBD has been shown in multiple organ dysfunction. Acute and chronic administration in an experimental model of sepsis reduced mortality, lipid peroxidation and oxidative damage of proteins in many vital organs [125]. However, one should keep in mind that CBD, given acutely, had a pro-oxidative effect and increased oxidative damage of proteins in the lungs ([125]; Table 1).

In summary, preclinical studies show that acute and chronic administration of CBD through prophylactic and/or therapeutical interventions has numerous beneficial effects in organs that are also targeted by the coronavirus; they mainly result from CBD's anti-inflammatory and antioxidative actions. The precise mechanism(s) is/are still unknown although in a few studies, the involvement of cannabinoid CB₁, CB₂, GPR55, vanilloid TRPV1, adenosine A_{2A} and serotonin 5-HT_{1A} receptors has been described (Table 1).

6. Use of CBD for COVID-19 Treatment? Clinical Studies

In the [ClinicalTrials.gov](https://clinicaltrials.gov) database (accessed on 1 February 2021), we found six clinical trials (two active, not recruiting; two not yet recruiting; and two recruiting) in which the use of CBD is being addressed in the context of COVID-19. Two studies are dedicated to the use of CBD in patients with mild to moderate symptoms, including the study entitled "Cannabidiol for COVID-19 patients with mild to moderate symptoms" (daily doses of 300 mg for 14 days) and the study entitled "Synthetic CBD as a therapy for COVID-19" (dose and duration of administration not specified). Another two studies are dedicated to patients with higher risk, including the study entitled "Cannabidiol treatment for severe and critical coronavirus (COVID-19) pulmonary infection" (daily doses of 300 mg for 14–28 days or until discharge) and the study entitled "Cannabidiol in patients with COVID-19 and cardiovascular disease or risk factors" (daily doses of 525 mg/70 kg for 28 days). The fifth study, entitled "Outcomes mandate national integration with Cannabis as medicine for prevention and treatment of COVID-19 (OMNI-Can)" will examine the efficacy and safety of using medical cannabis for chronic medical conditions, including COVID-19 (dose and duration of administration not specified). As suggested by the title "Burnout and distress prevention with cannabidiol in front-line health care workers dealing with COVID-19," the final study is dedicated to the hospital staff rather than to patients (daily doses of 175 mg/70 kg for 28 days).

Since it will take some time until the latter studies have been completed, the question arises as to whether there are other clinical studies based on CBD that may point to its suitability for the prevention/treatment of COVID-19 (Table 2). In the [ClinicalTrials.gov](https://clinicaltrials.gov) database (accessed on 1 February 2021), there are 186 items regarding formulations containing CBD (59 studies were signed as completed; the results of 13 of the latter trials are presented in the database and 9 of them have been published). In contrast to the results of preclinical studies (Table 1), so far, no clinical studies have demonstrated promising effects of CBD on patients with respiratory failure (Table 2). There are two publications based on a few patients with chronic obstructive pulmonary disease (COPD). CBD given acutely together with THC in vaporized form [126] or as a sublingual spray [127] had no or only minimal beneficial effects on airway function, exertional breathlessness at rest and during exercise and simulated breathlessness (Table 2). Clinical trials indicating a potential usefulness of CBD in cardiovascular, hematological and renal symptoms associated with COVID-19 could not be found.

Table 2. Efficacy and safety of cannabidiol in clinical studies.

	Disease	<i>n</i>	* Study Design; Dose of CBD and THC	Application	Final Results and/or Conclusions, Properties Important for COVID-19	Profile Safety/Side Effects	References
Respiratory	COPD	16	* Cannabis 35 mg (THC, 18.2%, CBD, 0.1%); acute	vaporized	no effect on airway function, exertional breathlessness at rest and exercise		[126]
		9	* THC: CBD 2.7:2.5 mg/spray; maximum single dose of 4 sprays	oromucosal spray	no effect on simulated breathlessness in COPD subjects; <u>↓unpleasantness of breathlessness as judged by descriptors</u>		[127]
Gastrointestinal	chemotherapy-induced nausea and vomiting (CINV)	7	* THC: CBD 2.7:2.5 mg/spray, ≤3 sprays within 2 h after chemotherapy plus ≤8 sprays each at days 2, 3 and 4	oromucosal spray	<u>better protection against delayed CINV compared to standard antiemetic therapy alone</u>	well tolerated	[128]
	cancer-related anorexia-cachexia syndrome	99	* THC 2.5 mg and CBD 1 mg; twice daily 1 h before meals for 6 weeks	p.o.	no effect on patients' appetite or quality of life	well tolerated	[129]
	aspirin-induced increased gut permeability	10	* CBD:600 mg	p.o.	<u>↓increased gut permeability</u>		[130]
	moderately active Crohn's disease	20	* CBD 10 mg/kg twice daily for 8 weeks	p.o.	no clinical improvement	excellent tolerability and safety profile	[131]
	ulcerative colitis	60	* 250 mg CBD-rich extract (up to 4.7% THC) twice daily before meals for 10 weeks	p.o.	no effect on ulcerative colitis but <u>↑quality of life outcomes</u>	mild/moderate, mainly dizziness and somnolence	[132]
Psychiatric	anxiety in healthy volunteers	40	* CBD:300 mg acute	p.o.	<u>↓anxiety to simulated public speaking</u>		[133]
		10	* CBD:400 mg acute	p.o.	<u>↓subjective anxiety to a simulated public speaking test, ↑mental sedation</u>		[134]

Table 2. Cont.

	Disease	n	* Study Design; Dose of CBD and THC	Application	Final Results and/or Conclusions, Properties Important for COVID-19	Profile Safety/Side Effects	References
Psychiatric	anxiety in healthy volunteers	60	* CBD:100, 300 and 900 mg acute	p.o.	↓subjective anxiety with a dose-dependent bell-shaped curve (effective dose:300 mg only)	CBD 300 mg has a lower sedation level than clonazepam	[135]
	naïve social anxiety disorder	24	* CBD:600 mg acute	p.o.	↓subjective anxiety, ↓cognitive impairment and discomfort in speech performance; no changes in blood pressure, heart rate and skin flow	absence of psychoactive or cognitive effects	[136]
		10	* CBD:400 mg acute	p.o.	↓subjective anxiety, changes in regional cerebral flow		[137]
	stress-related disorders	11	** CBD: flexible doses, starting from 25 to 49 mg/d for 8 weeks	p.o.	↓stress-related disorders (including ↓nightmares)	well tolerated, no patients discontinued treatment due to side effects; mild side effects: fatigue, reduced concentration; gastrointestinal bloating or pain	[138]
	psychiatric patients with anxiety or poor sleep	103	CBD:25 mg/d to 50–75 mg/d; for 1–3 months	p.o.	↓anxiety in a sustained manner, ↓sleep disturbances	well tolerated, fatigue (may be related to dosing), mild sedation, dry eyes	[139]
	patients at high risk for psychosis	32	* CBD:600 mg/day for 1 week	p.o.	cortisol reaction: tended to be better; anxiety: tended to be better; ↓negative self-statement		[140]
		33	* CBD:600 mg acute	p.o.	putative antipsychotic effect by normalizing motivational salience and moderating motor response		[141]
	cannabis use disorder	128	* THC:CBD 2.7:2.5 mg/spray for 6 days, up to 32 sprays/d	oromucosal spray	↓anxiety, ↓depression, ↓craving	no differences in adverse effects between THC:CBD and placebo group	[142]

Table 2. Cont.

Disease	<i>n</i>	* Study Design; Dose of CBD and THC	Application	Final Results and/or Conclusions, Properties Important for COVID-19	Profile Safety/Side Effects	References
regular cannabis users	20	** CBD:200 mg for 10 weeks	p.o.	<u>↓depressive symptoms,</u> <u>↓psychotic symptoms,</u> <u>↑attentional switching,</u> <u>↑verbal learning, ↑memory</u>		[143]
drug-abstinent patients with history of heroin abuse	42	* CBD 400 or 800 mg/d for 3 days	p.o.	<u>↓anxiety,</u> <u>↓craving,</u> <u>↓HR,</u> <u>↓salivary cortisol levels</u>	no serious adverse events; mild side effects: diarrhea, headache, tiredness, fatigue	[144]
volunteers pre-selected for high paranoid traits	32	* CBD:600 mg acute	p.o.	<u>no benefits on anxiety</u> <u>or persecutory ideation</u>		[145]
schizophrenia	42	*** CBD; week 1: gradual increase to 800 mg/day; weeks 2–4:800 mg/day	p.o.	<u>↓positive psychotic symptoms (no</u> <u>difference compared to amisulpride),</u> <u>mitigation of psychotic symptoms</u>	marked tolerability / safety (with respect to weight, prolactin, hepatic or cardiac functions) compared with current medications	[146]
schizophrenia	88	* CBD:1000 mg/d for 6 weeks on top of antipsychotic medication	p.o.	<u>↓positive psychotic symptoms,</u> <u>↓impressions and severity of illness,</u> <u>cognitive performance and overall</u> <u>functioning tended to be improved</u>	good tolerance; diarrhea, nausea, headache, infections, insomnia; mild ↓blood pressure and moderate chest pain; no significant changes in prolactin, weight, liver function, inflammatory markers or HDL cholesterol levels	[147]
	36	* CBD:600 mg/day for 6 weeks	p.o.	<u>no improvement of cognitive</u> <u>impairments and</u> <u>psychotic symptoms in stable</u> <u>antipsychotic-treated outpatients</u>	well tolerated with no worsening of mood, suicidality or movement side effects; sedation	[148]

Table 2. Cont.

	Disease	n	* Study Design; Dose of CBD and THC	Application	Final Results and/or Conclusions, Properties Important for COVID-19	Profile Safety/Side Effects	References
Pain	chronic pain	94	CBD-rich hemp extract ¹ for 8 weeks	soft gels	↓chronic pain and ↑life quality (↓opioid use, ↑sleep quality)		[149]
	chronic pain in kidney transplant patients	7	CBD increasing from 50 to 150 mg twice a day for 3 weeks	p.o.	2 patients, total improvement of pain 4 patients, <u>partial pain improvement</u>	no serious adverse effects; dizziness, nausea, dry mouth, drowsiness, intermittent episodes of heat	[150]
	chronic pain in patients with fibromyalgia	20	* THC:CBD (mg) 1. 22.4: < 1 2. 13.4:17.8 3. < 1:18.4	single vapor inhalation	<u>small analgesic responses</u>	limited, such as dizziness and nausea	[151]
	neuropathic pain in patients with MS	20	THC:CBD 2.7:2.5 mg/spray for 4 weeks, 8 sprays/d	sublin. spray	↓pain rating, ↑life quality	few side effects: dizziness, nausea, dry mouth and weakness	[152]
	peripheral neuropathic pain associated with diabetes or allodynia	380	THC:CBD 2.7:2.5 mg/spray for 38 weeks, ≤8 sprays per 3 h and ≤24 sprays every 24 h	oromucosal spray	↓pain in the majority of patients	safe and well tolerated; patients did not seek to increase their dose with time but 23% of patients ceased medication due to adverse effects (2% infections)	[153]
	painful diabetic neuropathy	30	* THC:CBD 2.7:2.5 mg/spray; dose was titrated over 2 weeks, followed by a 10-week maintenance phase (4 sprays/d)	sublin. spray	no significant improvements in pain rating and life quality		[154]
	advanced cancer patients with chronic pain	199	* THC:CBD 2.7:2.5 mg/spray: initially 4 up to 7 sprays/d	oromucosal spray	average pain score not superior to placebo		[155]

Table 2. Cont.

	Disease	<i>n</i>	* Study Design; Dose of CBD and THC	Application	Final Results and/or Conclusions, Properties Important for COVID-19	Profile Safety/Side Effects	References
Neurological	Dravet syndrome in children	198	* CBD: 10 and 20 mg/kg/d for 14 weeks	p.o.	↓frequency of seizures, improvement of patients' condition; similar effectiveness of both doses	↓appetite, diarrhea, somnolence, pyrexia, and fatigue, ↑aminotransferases in patients also taking valproate sodium; at 10 mg/kg/d better tolerance and safety profile	[156]
		108	* CBD:20 mg/kg/d for 14 weeks	p.o.	↓frequency of drug-resistant seizures	somnolence, ↓appetite, diarrhea, ↑aminotransferases in patients also taking valproate	[157]
	Lennox–Gastaut syndrome in children and adults	225	* CBD:10 or 20 mg/kg/d for 28 days	p.o.	↓epileptic seizures in some patients, ↓total seizure frequency, improvement of patients' condition	somnolence, ↓appetite, diarrhea (at 20 mg/kg/d), threefold ↑aminotransferases	[158]
		171	* CBD:20 mg/kg/d for 14 weeks	p.o.	↓frequency of drug-resistant seizures	in general good tolerance of CBD as add-on therapy; mild or moderate side effects: ↓appetite, vomiting, diarrhea, somnolence, fever	[159]
	multiple sclerosis	160	* THC:CBD 2.7:2.5 mg/spray up to a maximum of 120 mg THC and 120 mg CBD/day with no more than 20 mg of each in any 3-h period for 6 weeks	oromucosal spray	↓spasticity associated with MS	good tolerance, no effects on cognition or mood; dizziness, disturbance in attention, headache, fatigue, somnolence, disorientation, feeling drunk, vertigo, application site discomfort, nausea, diarrhea, mouth ulceration	[160]

Table 2. Cont.

	Disease	<i>n</i>	* Study Design; Dose of CBD and THC	Application	Final Results and/or Conclusions, Properties Important for COVID-19	Profile Safety/Side Effects	References
Neurological	multiple sclerosis	137	THC:CBD 2.7:2.5 mg/spray up to 48 sprays/day for 21–814 days (extension study of Wade et al. [160])	oromucosal spray	↓spasticity associated with MS, clinical effect maintained after a long treatment period	serious adverse effects: seizures, fall, aspiration pneumonia, gastroenteritis; mild adverse effects: sore mouth, oromucosal disorder, oral pain, altered attention, dizziness, diarrhea, nausea	[161]
		12	THC:CBD 5.4:5.0 mg (two sublin. sprays at a 15-min interval)	oromucosal spray	↓spasticity postural and motor tests unchanged	no differences in adverse events between THC:CBD and placebo groups	[162]
		460	* THC:CBD 2.7:2.5 mg/spray; maximally 12 sprays/day for 12 weeks	oromucosal spray	↓spasticity (resistant to previous antispasticity treatment) and associated symptoms in patients with moderate to severe MS		[163]
Skin	psoriasis	5	CBD-enriched ointment, twice daily, for 3 months	ointment	in both diseases, <u>improved skin parameters</u>	no irritant or allergic reactions	[164]
	atopic dermatitis	5					

* double-blind, placebo-controlled, randomized clinical trial; ** open-label study; *** double-blind, randomized clinical trial with active control group (amisulpride) CBD, cannabidiol; CINV, chemotherapy-induced nausea and vomiting; COPD, chronic obstructive pulmonary disease; d, day; HDL, high-density lipoprotein; HR, heart rate; MS, multiple sclerosis; *n*, number of patients; p.o. per os; sublin., sublingual; THC, Δ⁹-tetrahydrocannabinol; ↑, increase, ↓, decrease, ↔, no effect. ¹ 15.7 mg CBD, 0.5 mg THC, 0.3 mg cannabidivarin, 0.9 mg cannabidiolic acid, 0.8 mg cannabichrome and >1% botanical terpene blend.

With respect to the gastrointestinal complications related to COVID-19 (see Figure 2), the effect found for Sativex[®] (combination of 2.7 mg THC and 2.5 mg CBD), given on top of the standard antiemetic therapy against delayed chemotherapy-induced nausea and vomiting, is remarkable but is based on 7 patients only [128]. CBD given orally with THC did not increase appetite or raise the quality of life in patients with cancer-related anorexia-cachexia syndrome [129]. Regarding inflammatory-related disorders, a single dose of CBD reduced aspirin-induced increased gut permeability [130], suggesting its effectiveness in disorders such as inflammatory bowel disease. On the other hand, CBD given chronically, alone or with THC, did not lead to any clinical improvement in patients with moderately active Crohn's disease [131] or ulcerative colitis [132] (Table 2).

Although the effect of CBD on organ function has been considered only in a few clinical trials (as opposed to preclinical studies), its suitability for the treatment of some psychiatric disorders including post-traumatic stress, generalized anxiety, panic disorder and social anxiety, which may also occur in the context of COVID-19, is suggested by numerous clinical studies (for review, see [108]) (Figure 2, Table 2). CBD (given acutely or chronically) has been shown to reduce subjective anxiety and/or other reactions induced by stress (e.g., a simulated public speaking test) in healthy volunteers [133–135], people with naïve social anxiety disorder [136,137], patients with post-traumatic stress disorders [138], some psychiatric patients [139], persons at high risk for psychosis [140,141] and present cannabis [142,143] and past heroin users [144]. Only in a study on volunteers preselected for high paranoid traits [145] did CBD fail to attenuate anxiety. The possibility that CBD may be an effective treatment for schizophrenia has also been considered. CBD was shown to reduce positive psychotic symptoms of schizophrenia in studies by Leweke et al. [146] and McGuire et al. [147] but not in the trial by Boggs et al. [148] (Table 2).

COVID-19 is associated with painful symptoms, including myalgia, headache and abdominal or chest pain (Figure 2). Antinociceptive activity of CBD has been found in patients suffering from chronic pain [149], including pain in kidney transplant recipients [150] and in individuals with fibromyalgia [151], multiple sclerosis [152], diabetes and allodynia [153,154] but not in patients with advanced cancer [155]. In the latter studies, CBD was mainly administered in combination with THC as an oromucosal or sublingual spray (Table 2).

SARS-CoV-2 patients may develop encephalopathic symptoms ranging from alteration in consciousness to delirium, seizures and muscular damage (Figure 2). On the other hand, individuals with epilepsy and multiple sclerosis develop changes that not only increase their risk of morbidity from COVID-19 but may also mask the presentation of acute respiratory symptoms which can potentially delay the diagnosis of COVID-19. Published trials (Table 2) refer to the treatment of Dravet syndrome in children [156,157] and Lennox–Gastaut syndrome in children and adults [158,159]. Cannabidiol was administered at a dose of 10 or 20 mg/kg/day for 14 weeks and was found to reduce seizure frequency. Sativex[®] was used as an oromucosal spray in patients with multiple sclerosis and was found to reduce spasticity [160–163]. As described above, Epidiolex[®] (which contains a 100 mg/mL solution of CBD for oral administration) and Sativex[®] have been approved for the treatment of intractable childhood-onset epilepsy (Dravet and Lennox–Gastaut syndromes) and as a therapy for spasticity in multiple sclerosis, respectively.

In one study [164], chronic use of CBD-enriched ointment improved skin parameters in inflammatory skin diseases (Table 2). However, Epidiolex[®] induced a delayed skin rash in one patient with medically refractory epilepsy [165].

Importantly, as shown in Table 2, CBD, given alone or together with THC, is generally well tolerated, usually with no severe adverse events or clinical worsening. The most common side effects are nausea and vomiting, loss of appetite, diarrhea, fever and an increased concentration of aminotransferases. Even orally administered spray can produce mild to moderate unwanted effects including dizziness, nausea, diarrhea, oral pain and oromucosal disorder. Unfortunately, these side effects, to some extent, resemble the symptoms of COVID-19.

In summary, considering the effects of COVID-19 on multiple organs (Figure 2), a CBD-based pharmacotherapy that has been approved by the health authorities is limited to the treatment of some rare types of seizures and the spasticity associated with multiple sclerosis. In addition to these neuroprotective properties, anxiolytic and antinociceptive effects of CBD have been shown in clinical trials. Thus, CBD, given alone or together with THC, may be important as an adjuvant treatment to improve the well-being and quality of life of patients with COVID-19 and may even be used after recovery to limit post-traumatic stress symptoms. Further clinical studies are necessary to clarify beneficial observations obtained in small groups of patients or in cases where conflicting results have been found. Unfortunately, clinical studies (Table 2) have often failed to confirm the promising observations found in preclinical experiments (Table 1).

7. Opportunities, Challenges and Pitfalls of Cannabidiol Use as a COVID-19 Therapy

Cannabidiol is an interesting medicine with various pharmacological properties. Our main question is whether it is justified to recommend CBD as a therapy for COVID-19. Unfortunately, despite the identification in preclinical studies of some beneficial properties that are important for COVID-19 treatment, there are still numerous questions that need to be addressed. Preclinical and clinical effects (including their sites of action), the quality of CBD preparations, the route of administration, dosing, side effects and drug interactions are discussed in detail below.

First, the potential effects of CBD against COVID-19 may comprise the following four mechanisms.

- a. Numerous preclinical findings (Table 1) and reviews regarding the potential use of CBD in COVID-19 treatment [14,15,56–60] suggest that CBD has beneficial anti-inflammatory and antioxidative effects, which can be expected to improve the systemic symptoms that are characteristic of SARS-CoV-2 infection. Unfortunately, a comparison of the results of preclinical (Table 1) and clinical (Table 2) studies demonstrates that the favorable preclinical properties may not translate into the clinical setting (or that the appropriate clinical studies have not been conducted; see also reviews [58,61,62]). Above all, there is no confirmation of such beneficial effects of CBD with regard to its effects on respiratory failure. In addition, careful examination of whether the influence of CBD on the immune system could exacerbate viral infection is required (reviews [64,65]). Indeed, viral, fungal infections and pneumonia infections are listed among the side effects of CBD [166–169]. Moreover, it has to be considered that CBD, which was found to have a pro-oxidative effect in one preclinical study ([83]; Table 1), might even aggravate the feared cytokine storm.
- b. Anxiolytic and antinociceptive properties of CBD, given alone or together with THC, which have been identified in clinical studies (Table 2), suggest that it may be used as an adjuvant treatment to improve the quality of life of patients with COVID-19 and, even after recovery, may limit post-traumatic stress symptoms. However, well-designed double-blind, placebo-controlled clinical trials regarding the efficacy of CBD against COVID-19-associated panic, anxiety, depression and neurological complications are so far missing [63].
- c. The decreases in ACE2 and TMPRSS2 protein expression in a human tissue model are extremely interesting but so far this effect has only been shown in an in vitro study by Wang et al. [67]. Even if this mechanism was also found to occur in vivo, certain issues would have to be considered, for example, the mechanism was found for some CBD-rich extracts but did not occur when pure CBD was used. The consequence would be that an extract would need to be administered instead of a pure substance and oral administration would not be possible (for problems associated with topical administration, see below). Next, the question arises about the extent to which ACE2 and TMPRSS2 have to be decreased in order to obtain a robust antiviral effect. Moreover, it is unclear whether a reduction in ACE2 (the importance of which

is highlighted in Figure 1) will lead to problems other than the struggle against the virus.

- d. Inhibition of the replication of SARS-CoV-2 by CBD in a cell line also represents an interesting mechanism, although this effect has only been shown in a study by Raj et al. [68] and transfer to the in vivo situation of the human body is unclear.

Second, a wide range of over-the-counter CBD-based products is now available, for example, capsules, sprays, oil droppers, gummies and plant materials to be used by smoking, dry vaporizers and e-liquid vaporizers [54]. Unfortunately, these products may be of questionable quality, are not subject to appropriate safety controls and are characterized by unknown CBD content. There is little evidence of the pharmacological activity or health benefits of non-commercial CBD preparations [54,167]. Importantly, people taking non-commercial preparations on their own should be very careful due to the possibility of dangerous consequences, such as respiratory depression following, for example, CBD gummy ingestion [170] or CBD oil overdose [171]. One should also consider that the impact of “over-the-counter” CBD on the immunity of the SARS-CoV-2 infection is unclear [62]. The use of CBD as a medication in products like Epidiolex[®] and Sativex[®] (see Section 3) is approved, that is, these products fulfil the essential criteria related to efficacy, safety and pharmaceutical quality.

Third, one of the most severe COVID-19 symptoms is connected with disorders of the respiratory system (i.e. ARDS). Therefore, inhalation might be a particularly appropriate route of CBD administration. The relatively high bioavailability level of ~30% (whereas oral administration leads to a value of 6% only [172]) is another argument in favor of this route of administration. However, the vaping of cannabis products (including CBD) may lead to acute pulmonary toxicity [57,173], casting some doubt on the suitability of this method of administration; it is unclear as to whether similar problems would occur if pure CBD was used instead. Since Epidiolex[®] has been administered effectively as an oromucosal spray (Table 2), the use of a mouthwash with CBD-rich extracts might represent an interesting strategy to lower ACE2 topically [67]. It is of interest that the bioavailability of oromucosal administration of CBD is not higher than that of its oral application [172].

Fourth, with respect to dosing, most data refer to neurological disorders and this is in line with the approved indications (reviewed by Britch et al. [54]; Millar et al. [174]). Unfortunately, clinical trials dedicated to the use of CBD to treat inflammation are virtually nonexistent and clinically meaningful conclusions can therefore not be drawn [54]. A special note is necessary for Sativex[®] (2.7 mg THC plus 2.5 mg CBD per spray). If one uses 48 sprays per day (the maximum dose used in the study of Wade et al. [160] for the treatment of multiple sclerosis), the dose of CBD will be 120 mg, corresponding to ~1.7 mg/kg. This value is much lower than that planned for use in the three CBD trials listed on [ClinicalTrials.gov](https://clinicaltrials.gov) (2.5–7.5 mg/kg; see Section 6) or that recommended for use to treat seizures in children (10–20 mg/kg). CBD may increase the positive effects of THC and simultaneously attenuate the negative ones. A closer look, however, shows that both assumptions frequently do not hold true [175–179].

Fifth, CBD is generally and also according to the Critical Review Report 2018 of the World Health Organization [180], regarded as a safe compound (for review, see [54,181]). However, the authors of the latter two reviews postulated the need to conduct additional clinical trials. Indeed, various side effects of CBD have recently been described [166–169]. In addition to the increased risk of infection mentioned above, the increased tendency for respiratory depression and aspiration to occur, that is, symptoms also occurring in COVID-19, should be considered.

Sixth, CBD interacts with drug-metabolizing enzymes both of phase I (CYP3A4, CYP2C9 and CYP2C19) and phase II (uridine-5'-diphosphoglucosyltransferase) [181–183]. If CBD is used as an anticonvulsant, its combination with other anticonvulsants like clobazam or valproate might increase the risk of side effects [182,184], for example, thrombocytopenia was identified in one-third of 87 pediatric patients treated concurrently with cannabidiol and valproate [184]. Before CBD is used for the treatment of COVID-19, its pos-

sible interaction with the frontline therapy against COVID-19 should be carefully checked. The combination of CBD and glucocorticoids might lead to an increased plasma concentration of dexamethasone, since some glucocorticoids are substrates for CYP3A4 [185]. Due to the high therapeutic index of acutely administered glucocorticoids, this drug interaction may be tolerated. The situation is, however, entirely different for the antithrombin warfarin, the plasma level of which is also increased by CBD [183,186]. In this case, the interaction may lead to life-threatening bleeding.

8. Conclusions

COVID-19 is associated with multiple organ dysfunction/failure and a high mortality rate. The COVID-19 pandemic has made everyday life difficult and vaccination against SARS-CoV-2 has only just begun (and its final success is still unknown). New mutations of SARS-CoV-2 are appearing [187], so new promising therapies against COVID-19 are constantly being suggested. These include natural products, for example, CBD, a non-intoxicating phytocannabinoid from the cannabis plant with valuable pharmacological properties including strong anti-inflammatory, antioxidant, antiemetic, anticonvulsant, antipsychotic and anxiolytic properties. The broad pharmacological effectiveness and potential sites of action of CBD are shown in Figure 3. Besides its well-known antioxidant properties, downregulation of ACE2 and TMPRSS2 proteins (which are responsible for the entry of the SARS-CoV-2 virus into host cells) [67] and inhibition of SARS-CoV-2 replication [68] have only been shown very recently. Results require confirmation by independent groups and have to be demonstrated in humans *in vivo*. Numerous preclinical studies have shown the effectiveness of CBD in treating diseases of the respiratory system (including ARDS, one of the most dangerous symptoms of COVID-19) and its cardioprotective, nephroprotective, hepatoprotective, neuroprotective and anticonvulsant properties, that is, properties that could be beneficial for the treatment of COVID-19 (Section 5, Table 1). The beneficial influence of CBD results mainly from its significant anti-inflammatory and antioxidant properties. The anti-inflammatory properties of CBD are also responsible for a reduction in the short- and long-term consequences of viral infection, as suggested by a few *in vitro* and *in vivo* experiments (Section 4).

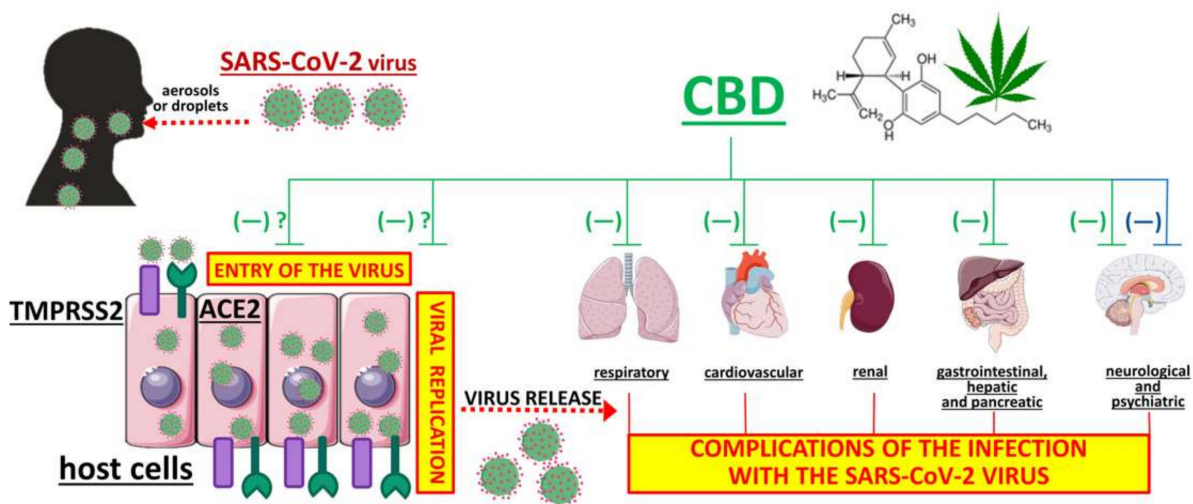


Figure 3. Potential therapeutic effect of cannabidiol against the SARS-CoV-2 virus infection. Three mechanisms have to be considered. The first and second mechanisms are the inhibitory effects on virus entry [67] and replication [68], respectively. These mechanisms have been described only recently and it is unclear whether they also occur in the human body. The third mechanism is a beneficial effect against complications which may also occur under a SARS-CoV-2 virus infection. In animal studies such a beneficial effect has been shown in five organ systems (green lines; Table 1) whereas in humans an effect on neurological and psychiatric disorders has been shown only (blue line; Table 2). ACE2, angiotensin-converting enzyme 2; CBD, cannabidiol; TMPRSS2, transmembrane serine protease 2.

Unfortunately, so far, clinical studies have not confirmed the beneficial anti-inflammatory properties of CBD but it is suggested that the anxiolytic and antinociceptive properties of CBD (given alone or together with THC) may be important regarding its use as an adjuvant treatment to improve the quality of life of patients with COVID-19 and, after recovery, to limit post-traumatic stress symptoms (Section 6, Table 2). When using CBD, one should be aware of its side effects (which are rarely serious), its frequent drug interactions (which also extend to drugs used for COVID-19 treatment) and the most appropriate administration route (vaping may be effective but sometimes also dangerous). Clearly, further clinical studies are necessary to confirm the beneficial observations made for small numbers of patients, to clarify conflicting results and to broaden our understanding of the true therapeutic potential of CBD against COVID-19.

Supplementary Materials: The following are available online at <https://www.mdpi.com/1422-0067/22/4/1986/s1> [188–279].

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Abbreviations

5-HT _{1A} -R	serotonin receptor type 1A
A _{1A} -R	adenosine receptor type A _{1A}
A _{2A} -R	adenosine receptor type A _{2A}
ACE2	angiotensin-converting enzyme 2
ACE-I	angiotensin-converting-enzyme inhibitors
AGEs	advanced glycation endproducts
ALI	acute lung injury
ALT	alanine transaminase
Ang II	angiotensin II
Ang 1–7	angiotensin 1–7
AST	aspartate transaminase
ARDS	acute respiratory distress syndrome
BALF	bronchoalveolar lavage fluid
CB-R	cannabinoid receptor
CB ₁ -R	cannabinoid CB ₁ receptor
CB ₂ -R	cannabinoid CB ₂ receptor
CBD	cannabidiol
CBF	cerebral blood flow
CCL2, CCL5	C-C motif chemokine ligand 2 and 5
CINV	chemotherapy-induced nausea and vomiting
COPD	chronic obstructive pulmonary disease
COVID-19	coronavirus disease 2019
COX-2	cyclooxygenase 2
CRP	C-reactive protein
CVD	cardiovascular disease

d	day(s)
DOCA	deoxycorticosterone acetate
DPCPX	8-cyclopentyl-1,3-dipropylxanthine
EEG	electroencephalography
EP ₄	prostanoid EP ₄ receptor
FAAH	fatty acid amide hydrolase
GCSF	granulocyte colony stimulating factor
HDL	high density lipoprotein
HR	heart rate
i.a.	intra-arterially
IFN- γ	interferon γ
IL- <i>n</i>	interleukin <i>n</i> , e.g., IL-1, interleukin-1
IL-1 β	interleukin-1 β
iNOS	inducible nitric oxide synthase
IP	prostacyclin receptor
i.p.	intraperitoneally
i.v.	intravenously
LAD	left anterior descending artery
LCx	left circumflex coronary artery
LDL	low-density lipoprotein
LPS	lipopolysaccharide
MCA	middle cerebral artery
MCP-1	monocyte chemoattractant protein-1
MERS	middle east respiratory syndrome
MIP-2	macrophage inflammatory protein-2
MPO	myeloperoxidase
MS	multiple sclerosis
NF- κ B	nuclear factor κ B
NO	nitric oxide
NRP1	protein neuropilin-1
PAI-1	plasminogen activator inhibitor-1
pCO ₂	partial pressure of carbon dioxide
p.o.	per os, orally
pO ₂	partial pressure of oxygen
poly(I:C)	polyriboinosinic:polyribocytidylic acid
PPAR γ	peroxisome proliferator-activated receptor type γ
SARS	severe acute respiratory syndrome
SARS-CoV-2	severe acute respiratory syndrome coronavirus 2
SHR	spontaneously hypertensive rats
STZ	streptozotocin
TGs	triglycerides
THC	Δ^9 -tetrahydrocannabinol
TLC	total lung capacity
TMEV	Theiler's murine encephalomyelitis virus-induced demyelinating disease
TMPRSS2	transmembrane serine protease 2
TNF- α	tumor necrosis factor α
t-PA	tissue plasminogen activator
TRP	transient receptor potential
TRPV1	transient receptor potential vanilloid subfamily member 1
UVA	ultraviolet A
VCAM-1	vascular cell adhesion molecule-1

References

1. Baj, J.; Karakuła-Juchnowicz, H.; Teresiński, G.; Buszewicz, G.; Ciesielka, M.; Sitarz, E.; Forma, A.; Karakuła, K.; Flieger, W.; Portincasa, P.; et al. Specific and non-specific clinical manifestations and symptoms: The current state of knowledge. *J. Clin. Med.* **2020**, *9*, 1753. [[CrossRef](#)] [[PubMed](#)]

2. Tu, Y.F.; Chien, C.S.; Yarmishyn, A.A.; Lin, Y.Y.; Luo, Y.H.; Lin, Y.T.; Lai, W.Y.; Yang, D.M.; Chou, S.J.; Yang, Y.P.; et al. A review of SARS-CoV-2 and the ongoing clinical trials. *Int. J. Mol. Sci.* **2020**, *21*, 2657. [CrossRef]
3. Peng, M. Outbreak of COVID-19: An emerging global pandemic threat. *Biomed. Pharmacother.* **2020**, *129*, 110499. [CrossRef] [PubMed]
4. List of Epidemics. Available online: https://en.wikipedia.org/wiki/List_of_epidemics (accessed on 1 February 2021).
5. Cannalire, R.; Stefanelli, I.; Cerchia, C.; Beccari, A.R.; Pelliccia, S.; Summa, V. Entry inhibitors: Small molecules and peptides targeting virus or host cells. *Int. J. Mol. Sci.* **2020**, *21*, 5707. [CrossRef]
6. World Health Organisation. Draft Landscape of COVID-19 Candidate Vaccines. Available online: <https://www.who.int/publications/m/item/draft-landscape-of-covid-19-candidate-vaccines> (accessed on 1 February 2021).
7. Krammer, F. SARS-CoV-2 vaccines in development. *Nature* **2020**, *586*, 516–527. [CrossRef]
8. Weisberg, E.; Sattler, M.; Yang, P.L.; Parent, A.; Gray, N.; Griffin, J.D. Current therapies under investigation for COVID-19: Potential COVID-19 treatments. *Can. J. Physiol. Pharmacol.* **2020**, *98*, 483–489. [CrossRef]
9. Asai, A.; Konno, M.; Ozaki, M.; Otsuka, C.; Vecchione, A.; Arai, T.; Kitagawa, T.; Ofusa, K.; Yabumoto, M.; Hirotsu, T.; et al. COVID-19 drug discovery using intensive approaches. *Int. J. Mol. Sci.* **2020**, *21*, 2839. [CrossRef]
10. Beigel, J.H.; Tomashek, K.M.; Dodd, L.E.; Mehta, A.K.; Zingman, B.S.; Kalil, A.C.; Hohmann, E.; Chu, H.Y.; Luetkemeyer, A.; Kline, S.; et al. ACTT-1 study group members. Remdesivir for the treatment of Covid-19—final report. *N. Engl. J. Med.* **2020**, *383*, 1813–1826. [CrossRef]
11. Recovery Collaborative Group; Horby, P.; Lim, W.S.; Emberson, J.R.; Mafham, M.; Bell, J.L.; Linsell, L.; Staplin, N.; Brightling, C.; Ustianowski, A.; Elmahi, E.; et al. Dexamethasone in hospitalized patients with Covid-19—Preliminary report. *N. Engl. J. Med.* **2020**. [CrossRef]
12. McKee, D.L.; Sternberg, A.; Stange, U.; Laufer, S.; Naujokat, C. Candidate drugs against SARS-CoV-2 and COVID-19. *Pharmacol. Res.* **2020**, *157*, 104859. [CrossRef]
13. He, Y.Q.; Zhou, C.C.; Yu, L.Y.; Wang, L.; Deng, J.L.; Tao, Y.L.; Zhang, F.; Chen, W.S. Natural product derived phytochemicals in managing acute lung injury by multiple mechanisms. *Pharmacol. Res.* **2020**, *163*, 105224. [CrossRef]
14. Mamber, S.W.; Krakowka, S.; Osborn, J.; Saberski, L.; Rhodes, R.G.; Dahlberg, A.E.; Pond-Tor, S.; Fitzgerald, K.; Wright, N.; Beseme, S.; et al. Can unconventional immunomodulatory agents help alleviate COVID-19 symptoms and severity? *MSphere* **2020**, *5*, e00288. [CrossRef]
15. Onaivi, E.S.; Sharma, V. Cannabis for COVID-19: Can cannabinoids quell the cytokine storm? *Future Sci.* **2020**, *6*, FSO625. [CrossRef] [PubMed]
16. Ni, W.; Yang, X.; Yang, D.; Bao, J.; Li, R.; Xiao, Y.; Hou, C.; Wang, H.; Liu, J.; Yang, D.; et al. Role of angiotensin-converting enzyme 2 (ACE2) in COVID-19. *Crit. Care* **2020**, *24*, 422. [CrossRef]
17. Kielian, M. Enhancing host cell infection by SARS-CoV-2. *Science* **2020**, *370*, 765–766. [CrossRef]
18. Penna, C.; Mercurio, V.; Tocchetti, C.G.; Pagliaro, P. Sex-related differences in COVID-19 lethality. *Br. J. Pharmacol.* **2020**, *177*, 4375–4385. [CrossRef] [PubMed]
19. Gemmati, D.; Bramanti, B.; Serino, M.L.; Secchiero, P.; Zauli, G.; Tisato, V. COVID-19 and individual genetic susceptibility/receptivity: Role of ACE1/ACE2 genes, immunity, inflammation and coagulation. Might the double X-chromosome in females be protective against SARS-CoV-2 compared to the single X-Chromosome in males? *Int. J. Mol. Sci.* **2020**, *21*, 3474. [CrossRef] [PubMed]
20. Pagliaro, P.; Penna, C. ACE/ACE2 ratio: A key also in 2019 Coronavirus disease (Covid-19)? *Front. Med.* **2020**, *7*, 335. [CrossRef]
21. Gheblawi, M.; Wang, K.; Viveiros, A.; Nguyen, Q.; Zhong, J.C.; Turner, A.J.; Raizada, M.K.; Grant, M.B.; Oudit, G.Y. Angiotensin-Converting Enzyme 2: SARS-CoV-2 receptor and regulator of the renin-angiotensin system: Celebrating the 20th anniversary of the discovery of ACE2. *Circ. Res.* **2020**, *126*, 1456–1474. [CrossRef]
22. Guzik, T.J.; Mohiddin, S.A.; Dimarco, A.; Patel, V.; Savvatis, K.; Marelli-Berg, F.M.; Madhur, M.S.; Tomaszewski, M.; Maffia, P.; D'Acquisto, F.; et al. COVID-19 and the cardiovascular system: Implications for risk assessment, diagnosis, and treatment options. *Cardiovasc. Res.* **2020**, *16*, 1666–1687. [CrossRef]
23. Coperchini, F.; Chiovato, L.; Croce, L.; Magri, F.; Rotondi, M. The cytokine storm in COVID-19: An overview of the involvement of the chemokine/chemokine-receptor system. *Cytokine Growth Factor Rev.* **2020**, *53*, 25–32. [CrossRef]
24. Arendse, L.B.; Danser, A.; Poglitsch, M.; Touyz, R.M.; Burnett, J.C.; Llorens-Cortes, C.; Ehlers, M.R.; Sturrock, E.D. Novel therapeutic approaches targeting the renin-angiotensin system and associated peptides in hypertension and heart failure. *Pharmacol. Rev.* **2019**, *71*, 539–570. [CrossRef]
25. Verdecchia, P.; Cavallini, C.; Spanevello, A.; Angeli, F. COVID-19: ACE2centric infective disease? *Hypertension* **2020**, *76*, 294–299. [CrossRef]
26. Samidurai, A.; Das, A. Cardiovascular Complications associated with COVID-19 and potential therapeutic strategies. *Int. J. Mol. Sci.* **2020**, *21*, 6790. [CrossRef]
27. Iwasaki, M.; Saito, J.; Zhao, H.; Sakamoto, A.; Hirota, K.; Ma, D. Inflammation triggered by SARS-CoV-2 and ACE2 augment drives multiple organ failure of severe COVID-19: Molecular mechanisms and implications. *Inflammation* **2020**, *44*, 13–34. [CrossRef]
28. Cremer, P.C. SARS-CoV-2 and myocardial injury: Few answers, many questions. *Cleve. Clin. J. Med.* **2020**, *87*, 521–525. [CrossRef] [PubMed]

29. Evans, P.C.; Rainger, G.E.; Mason, J.C.; Guzik, T.J.; Osto, E.; Stamataki, Z.; Neil, D.; Hoefler, I.E.; Fragiadaki, M.; Waltenberger, J.; et al. Endothelial dysfunction in COVID-19: A position paper of the ESC Working Group for Atherosclerosis and Vascular Biology, and the ESC Council of Basic Cardiovascular Science. *Cardiovasc. Res.* **2020**, *116*, 2177–2184. [[CrossRef](#)] [[PubMed](#)]
30. Kochi, A.N.; Tagliari, A.P.; Forleo, G.B.; Fassini, G.M.; Tondo, C. Cardiac and arrhythmic complications in patients with COVID-19. *J. Cardiovasc. Electrophysiol.* **2020**, *31*, 1003–1008. [[CrossRef](#)] [[PubMed](#)]
31. Zheng, K.I.; Feng, G.; Liu, W.Y.; Targher, G.; Byrne, C.D.; Zheng, M.H. Extrapulmonary complications of COVID-19: A multisystem disease? *J. Med. Virol.* **2020**, *10*. [[CrossRef](#)]
32. De Vries, A. SARS-CoV-2/COVID-19: A primer for cardiologists. *Neth. Heart J.* **2020**, *28*, 366–383. [[CrossRef](#)]
33. Zaim, S.; Chong, J.H.; Sankaranarayanan, V.; Harky, A. COVID-19 and multiorgan response. *Curr. Prob. Cardiol.* **2020**, *45*, 100618. [[CrossRef](#)]
34. Gąsecka, A.; Borovac, J.A.; Guerreiro, R.A.; Giustozzi, M.; Parker, W.; Caldeira, D.; Chiva-Blanch, G. Thrombotic complications in patients with COVID-19: Pathophysiological mechanisms, diagnosis, and treatment. *Cardiovasc. Drugs Ther.* **2020**, 1–15. [[CrossRef](#)] [[PubMed](#)]
35. Jayarangaiah, A.; Kariyanna, P.T.; Chen, X.; Jayarangaiah, A.; Kumar, A. COVID-19-associated coagulopathy: An exacerbated immunothrombosis response. *Clin. Appl. Thromb. Hemost.* **2020**, *26*, 1076029620943293. [[CrossRef](#)]
36. Vinayagam, S.; Sattu, K. SARS-CoV-2 and coagulation disorders in different organs. *Life Sci.* **2020**, *260*, 118431. [[CrossRef](#)]
37. Chueh, T.I.; Zheng, C.M.; Hou, Y.C.; Lu, K.C. Novel evidence of acute kidney injury in COVID-19. *J. Clin. Med.* **2020**, *9*, 3547. [[CrossRef](#)] [[PubMed](#)]
38. Gan, R.; Rosoman, N.P.; Henshaw, D.; Noble, E.P.; Georgius, P.; Sommerfeld, N. COVID-19 as a viral functional ACE2 deficiency disorder with ACE2 related multi-organ disease. *Med. Hypotheses* **2020**, *144*, 110024. [[CrossRef](#)]
39. Galanopoulos, M.; Gkeros, F.; Doukatas, A.; Karianakis, G.; Pontas, C.; Tsoukalas, N.; Viazis, N.; Liatsos, C.; Mantzaris, G.J. COVID-19 pandemic: Pathophysiology and manifestations from the gastrointestinal tract. *World J. Gastroenterol.* **2020**, *26*, 4579–4588. [[CrossRef](#)]
40. Hunt, R.H.; East, J.E.; Lanas, A.; Malfertheiner, P.; Satsangi, J.; Scarpignato, C.; Webb, G.J. COVID-19 and gastrointestinal disease. Implications for the gastroenterologist. *Dig. Dis.* **2020**, 1–21. [[CrossRef](#)]
41. Asadi-Pooya, A.A. Seizures associated with coronavirus infections. *Seizure* **2020**, *79*, 49–52. [[CrossRef](#)] [[PubMed](#)]
42. Pennisi, M.; Lanza, G.; Falzone, L.; Fiscaro, F.; Ferri, R.; Bella, R. SARS-CoV-2 and the nervous system: From clinical features to molecular mechanisms. *Int. J. Mol. Sci.* **2020**, *21*, 5475. [[CrossRef](#)]
43. Varatharaj, A.; Thomas, N.; Ellul, M.A.; Davies, N.W.S.; Pollak, T.A.; Tenorio, E.L.; Sultan, M.; Easton, A.; Breen, G.; Zandi, M.; et al. CoroNerve Study Group. Neurological and neuropsychiatric complications of COVID-19 in 153 patients: A UK-wide surveillance study. *Lancet Psychiatry* **2020**, *7*, 875–882. [[CrossRef](#)]
44. Luís, M.E.; Hipólito-Fernandes, D.; Mota, C.; Maleita, D.; Xavier, C.; Maio, T.; Cunha, J.P.; Tavares Ferreira, J. A review of neuro-ophthalmological manifestations of human coronavirus infection. *Eye Brain* **2020**, *12*, 129–137. [[CrossRef](#)] [[PubMed](#)]
45. Somasundaram, N.P.; Ranathunga, I.; Ratnasamy, V.; Wijewickrama, P.; Dissanayake, H.A.; Yogendranathan, N.; Gamage, K.; de Silva, N.L.; Sumanatilleke, M.; Katulanda, P.; et al. The impact of SARS-Cov-2 virus infection on the endocrine system. *J. Endocr. Soc.* **2020**, *4*, bvaa082. [[CrossRef](#)]
46. Lundholm, M.D.; Poku, C.; Emanuele, N.; Emanuele, M.A.; Lopez, N. SARS-CoV-2 (COVID-19) and the endocrine system. *J. Endocr. Soc.* **2020**, *4*, bvaa144. [[CrossRef](#)] [[PubMed](#)]
47. Juan, J.; Gil, M.M.; Rong, Z.; Zhang, Y.; Yang, H.; Poon, L.C. Effect of coronavirus disease 2019 (COVID-19) on maternal, perinatal and neonatal outcome: Systematic review. *Ultrasound Obstet. Gynecol.* **2020**, *56*, 15–27. [[CrossRef](#)]
48. Gonzalez, A.; Orozco-Aguilar, J.; Achiardi, O.; Simon, F.; Cabello-Verrugio, C. SARS-CoV-2/renin-angiotensin system: Deciphering the clues for a couple with potentially harmful effects on skeletal muscle. *Int. J. Mol. Sci.* **2020**, *21*, 7904. [[CrossRef](#)]
49. Rahimi, H.; Tehraninia, Z. A comprehensive review of cutaneous manifestations associated with COVID-19. *Biomed. Res. Int.* **2020**, *2020*, 1236520. [[CrossRef](#)]
50. Mawhirt, S.L.; Frankel, D.; Diaz, A.M. Cutaneous manifestations in adult patients with COVID-19 and dermatologic conditions related to the COVID-19 pandemic in health care workers. *Curr. Allergy Asthma Rep.* **2020**, *20*, 75. [[CrossRef](#)]
51. Kicman, A.; Toczek, M. The effects of cannabidiol, a non-intoxicating compound of cannabis, on the cardiovascular system in health and disease. *Int. J. Mol. Sci.* **2020**, *21*, 6740. [[CrossRef](#)] [[PubMed](#)]
52. Atalay, S.; Jarocka-Karpowicz, I.; Skrzydlewska, E. Antioxidative and anti-inflammatory properties of cannabidiol. *Antioxidants* **2019**, *9*, 21. [[CrossRef](#)]
53. Ligresti, A.; De Petrocellis, L.; Di Marzo, V. Phytocannabinoids to cannabinoid receptors and endocannabinoids: Pleiotropic physiological and pathological roles through complex pharmacology. *Physiol. Rev.* **2016**, *96*, 1593–1659. [[CrossRef](#)]
54. Britch, S.C.; Babalonis, S.; Walsh, S.L. Cannabidiol: Pharmacology and therapeutic targets. *Psychopharmacology* **2020**, *238*, 9–28. [[CrossRef](#)]
55. Pisanti, S.; Malfitano, A.M.; Ciaglia, E.; Lamberti, A.; Ranieri, R.; Cuomo, G.; Abate, M.; Faggiana, G.; Proto, M.C.; Fiore, D.; et al. Cannabidiol: State of the art and new challenges for therapeutic applications. *Pharmacol. Ther.* **2017**, *175*, 133–150. [[CrossRef](#)] [[PubMed](#)]

56. Esposito, G.; Pesce, M.; Seguela, L.; Sanseverino, W.; Lu, J.; Corpetti, C.; Sarnelli, G. The potential of cannabidiol in the COVID-19 pandemic. *Br. J. Pharmacol.* **2020**, *177*, 4967–4970. [[CrossRef](#)]
57. Janmohamed, K.; Soale, A.N.; Forastiere, L.; Tang, W.; Sha, Y.; Demant, J.; Airoidi, E.; Kumar, N. Intersection of the web-based vaping narrative with COVID-19: Topic modeling study. *J. Med. Internet Res.* **2020**, *22*, e21743. [[CrossRef](#)] [[PubMed](#)]
58. Sexton, M. Cannabis in the time of coronavirus disease 2019: The Yin and Yang of the endocannabinoid system in immunocompetence. *J. Altern. Complement. Med.* **2020**, *26*, 444–448. [[CrossRef](#)] [[PubMed](#)]
59. Costiniuk, C.T.; Jenabian, M.A. Acute inflammation and pathogenesis of SARS-CoV-2 infection: Cannabidiol as a potential anti-inflammatory treatment? *Cytokine Growth Factor Rev.* **2020**, *53*, 63–65. [[CrossRef](#)] [[PubMed](#)]
60. Rossi, F.; Tortora, C.; Argenziano, M.; Di Paola, A.; Punzo, F. Cannabinoid receptor type 2: A possible target in SARS-CoV-2 (CoV-19) infection? *Int. J. Mol. Sci.* **2020**, *21*, 3809. [[CrossRef](#)]
61. Shover, C.L.; Humphreys, K. Debunking cannabidiol as a treatment for COVID-19: Time for the FDA to adopt a focused deterrence model? *Cureus* **2020**, *12*, e8671. [[CrossRef](#)]
62. Biali, M.; Broers, B.; Besson, M.; Demeules, J. Cannabinoids and COVID-19. *Med. Cannabis Cannabinoids* **2020**, *3*, 111–115. [[CrossRef](#)]
63. Khalsa, J.H.; Bunt, G.; Maggirwar, S.B.; Kottlil, S. COVID-19 and cannabidiol (CBD). *J. Addict. Med.* **2020**. [[CrossRef](#)] [[PubMed](#)]
64. Brown, J.D. Cannabidiol as prophylaxis for SARS-CoV-2 and COVID-19? Unfounded claims versus potential risks of medications during the pandemic. *Res. Soc. Adm. Pharm.* **2021**, *17*, 2053. [[CrossRef](#)]
65. Hill, K.P. Cannabinoids and the coronavirus. *Cannabis Cannabinoid Res.* **2020**, *5*, 118–120. [[CrossRef](#)]
66. Booz, G.W. Cannabidiol as an emergent therapeutic strategy for lessening the impact of inflammation on oxidative stress. *Free Radic. Biol. Med.* **2011**, *51*, 1054–1061. [[CrossRef](#)]
67. Wang, B.; Kovalchuk, A.; Li, D.; Rodriguez-Juarez, R.; Illytskyy, Y.; Kovalchuk, I.; Kovalchuk, O. In search of preventive strategies: Novel high-CBD *Cannabis sativa* extracts modulate ACE2 expression in COVID-19 gateway tissues. *Aging* **2020**, *2*, 22425–22444. [[CrossRef](#)]
68. Raj, V.; Park, J.G.; Cho, K.H.; Choi, P.; Kim, T.; Ham, J.; Lee, J. Assessment of antiviral potencies of cannabinoids against SARS-CoV-2 using computational and in vitro approaches. *Int. J. Biol. Macromol.* **2020**, *168*, 474–485. [[CrossRef](#)] [[PubMed](#)]
69. Lowe, H.I.; Toyang, N.J.; McLaughlin, W. Potential of cannabidiol for the treatment of viral hepatitis. *Pharmacognosy. Res.* **2017**, *9*, 116–118. [[CrossRef](#)] [[PubMed](#)]
70. Maor, Y.; Yu, J.; Kuzontkoski, P.M.; Dezube, B.J.; Zhang, X.; Groopman, J.E. Cannabidiol inhibits growth and induces programmed cell death in kaposi sarcoma-associated herpesvirus-infected endothelium. *Genes. Cancer* **2012**, *3*, 512–520. [[CrossRef](#)]
71. Khodadadi, H.; Salles, É.L.; Jarrahi, A.; Chibane, F.; Costigliola, V.; Yu, J.C.; Vaibhav, K.; Hess, D.C.; Dhandapani, K.M.; Baban, B. Cannabidiol modulates cytokine storm in acute respiratory distress syndrome induced by simulated viral infection using synthetic RNA. *Cannabis Cannabinoid Res.* **2020**, *5*, 197–201. [[CrossRef](#)] [[PubMed](#)]
72. Salles, É.L.; Khodadadi, H.; Jarrahi, A.; Ahluwalia, M.; Paffaro, V.A.; Costigliola, V.; Yu, J.C.; Hess, D.C.; Dhandapani, K.M.; Baban, B. Cannabidiol (CBD) modulation of apelin in acute respiratory distress syndrome. *J. Cell. Mol. Med.* **2020**, *24*, 12869–12872. [[CrossRef](#)]
73. Patel, D.C.; Wallis, G.; Fujinami, R.S.; Wilcox, K.S.; Smith, M.D. Cannabidiol reduces seizures following CNS infection with Theiler’s murine encephalomyelitis virus. *Epilepsia Open* **2019**, *4*, 431–442. [[CrossRef](#)]
74. Mecha, M.; Feliú, A.; Iñigo, P.M.; Mestre, L.; Carrillo-Salinas, F.J.; Guaza, C. Cannabidiol provides long-lasting protection against the deleterious effects of inflammation in a viral model of multiple sclerosis: A role for A2A receptors. *Neurobiol. Dis.* **2013**, *59*, 141–150. [[CrossRef](#)] [[PubMed](#)]
75. Ribeiro, A.; Ferraz-de-Paula, V.; Pinheiro, M.L.; Vitoretti, L.B.; Mariano-Souza, D.P.; Quinteiro-Filho, W.M.; Akamine, A.T.; Almeida, V.I.; Quevedo, J.; Dal-Pizzol, F.; et al. Cannabidiol, a non-psychoactive plant-derived cannabinoid, decreases inflammation in a murine model of acute lung injury: Role for the adenosine A2A receptor. *Eur. J. Pharmacol.* **2012**, *678*, 78–85. [[CrossRef](#)] [[PubMed](#)]
76. Ribeiro, A.; Almeida, V.I.; Costola De Souza, C.; Ferraz De Paula, V.; Pinheiro, M.L.; Vitoretti, L.B.; Gimenes-Junior, J.A.; Akamine, A.T.; Crippa, J.A.; Tavares De Lima, W.; et al. Cannabidiol improves lung function and inflammation in mice submitted to LPS-induced acute lung injury. *Immunopharmacol. Immunotoxicol.* **2015**, *37*, 35–41. [[CrossRef](#)]
77. Sadowska, O.; Baranowska-Kuczko, M.; Gromotowicz-Popławska, A.; Biernacki, M.; Kicman, A.; Malinowska, B.; Kasacka, I.; Krzyżewska, A.; Kozłowska, H. Cannabidiol ameliorates monocrotaline-induced pulmonary hypertension in rats. *Int. J. Mol. Sci.* **2020**, *21*, 7077. [[CrossRef](#)] [[PubMed](#)]
78. Arruza, L.; Pazos, M.R.; Mohammed, N.; Escribano, N.; Lafuente, H.; Santos, M.; Alvarez-Díaz, F.J. Cannabidiol reduces lung injury induced by hypoxic-ischemic brain damage in newborn piglets. *Pediatr. Res.* **2017**, *82*, 79–86. [[CrossRef](#)] [[PubMed](#)]
79. Vuolo, F.; Petronilho, F.; Sonai, B.; Ritter, C.; Hallak, J.E.; Zuardi, A.W.; Crippa, J.A.; Dal-Pizzol, F. Evaluation of serum cytokines levels and the role of cannabidiol treatment in animal model of asthma. *Mediators Inflamm.* **2015**, *2015*, 538670. [[CrossRef](#)] [[PubMed](#)]
80. Vuolo, F.; Abreu, S.C.; Michels, M.; Xisto, D.G.; Blanco, N.G.; Hallak, J.E.; Zuardi, A.W.; Crippa, J.A.; Reis, C.; Bahl, M.; et al. Cannabidiol reduces airway inflammation and fibrosis in experimental allergic asthma. *Eur. J. Pharmacol.* **2019**, *843*, 251–259. [[CrossRef](#)]

81. Dudášová, A.; Keir, S.D.; Parsons, M.E.; Molleman, A.; Page, C.P. The effects of cannabidiol on the antigen-induced contraction of airways smooth muscle in the guinea-pig. *Pulm. Pharmacol. Ther.* **2013**, *26*, 373–379. [[CrossRef](#)] [[PubMed](#)]
82. Baranowska-Kuczko, M.; Kozłowska, H.; Kloza, M.; Sadowska, O.; Kozłowski, M.; Kusaczuk, M.; Kasacka, I.; Malinowska, B. Vasodilatory effects of cannabidiol in human pulmonary and rat small mesenteric arteries: Modification by hypertension and the potential pharmacological opportunities. *J. Hypertens.* **2020**, *38*, 896–911. [[CrossRef](#)]
83. Karmaus, P.W.; Wagner, J.G.; Harkema, J.R.; Kaminski, N.E.; Kaplan, B.L. Cannabidiol (CBD) enhances lipopolysaccharide (LPS)-induced pulmonary inflammation in C57BL/6 mice. *J. Immunotoxicol.* **2013**, *10*, 321–328. [[CrossRef](#)] [[PubMed](#)]
84. Makwana, R.; Venkatasamy, R.; Spina, D.; Page, C. The effect of phytocannabinoids on airway hyper-responsiveness, airway inflammation, and cough. *J. Pharmacol. Expl. Ther.* **2015**, *353*, 169–180. [[CrossRef](#)]
85. Feng, Y.; Chen, F.; Yin, T.; Xia, Q.; Liu, Y.; Huang, G.; Zhang, J.; Oyen, R.; Ni, Y.J. Effects of cannabidiol on acute reperfused myocardial infarction in rabbits: Evaluated with 3.0T Cardiac Magnetic Resonance Imaging and Histopathology. *J. Cardiovasc. Pharmacol.* **2015**, *66*, 354–363. [[CrossRef](#)] [[PubMed](#)]
86. Durst, R.; Danenberg, H.; Gallily, R.; Mechoulam, R.; Meir, K.; Grad, E.; Beerli, R.; Pugatsch, T.; Tarsish, E.; Lotan, C. Cannabidiol, a nonpsychoactive Cannabis constituent, protects against myocardial ischemic reperfusion injury. *Am. J. Physiol. Heart Circ. Physiol.* **2007**, *293*, 3602–3607. [[CrossRef](#)]
87. Walsh, S.K.; Hepburn, C.Y.; Kane, K.A.; Wainwright, C.L. Acute administration of cannabidiol in vivo suppresses ischaemia-induced cardiac arrhythmias and reduces infarct size when given at reperfusion. *Br. J. Pharmacol.* **2010**, *160*, 1234–1242. [[CrossRef](#)]
88. Gonca, E.; Darici, F. The effect of cannabidiol on ischemia/reperfusion-induced ventricular arrhythmias: The role of adenosine A1 receptors. *J. Cardiovasc. Pharmacol. Ther.* **2015**, *20*, 76–83. [[CrossRef](#)]
89. Lee, W.S.; Erdelyi, K.; Matyas, C.; Mukhopadhyay, P.; Varga, Z.V.; Liaudet, L.; Haskú, G.; Čiháková, D.; Mechoulam, R.; Pacher, P. Cannabidiol limits T cell-mediated chronic autoimmune myocarditis: Implications to autoimmune disorders and organ transplantation. *Mol. Med.* **2016**, *22*, 136–146. [[CrossRef](#)] [[PubMed](#)]
90. Rajesh, M.; Mukhopadhyay, P.; Bátkai, S.; Patel, V.; Saito, K.; Matsumoto, S.; Kashiwaya, Y.; Horváth, B.; Mukhopadhyay, B.; Becker, L.; et al. Cannabidiol attenuates cardiac dysfunction, oxidative stress, fibrosis, and inflammatory and cell death signaling pathways in diabetic cardiomyopathy. *J. Am. Coll. Cardiol.* **2010**, *56*, 2115–2125. [[CrossRef](#)]
91. Remiszewski, P.; Jarocka-Karpowicz, I.; Biernacki, M.; Jastrzab, A.; Schlicker, E.; Toczek, M.; Harasim-Symbor, E.; Pędzińska-Betiuk, A.; Malinowska, B. Chronic Cannabidiol administration fails to diminish blood pressure in rats with primary and secondary hypertension despite its effects on cardiac and plasma endocannabinoid system, oxidative stress and lipid metabolism. *Int. J. Mol. Sci.* **2020**, *21*, 1295. [[CrossRef](#)]
92. Pędzińska-Betiuk, A.; Weresa, J.; Schlicker, E.; Harasim-Symbor, E.; Toczek, M.; Kasacka, I.; Gajo, B.; Malinowska, B. Chronic cannabidiol treatment reduces the carbachol-induced coronary constriction and left ventricular cardiomyocyte width of the isolated hypertensive rat heart. *Toxicol. Appl. Pharmacol.* **2020**, *411*, 115368. [[CrossRef](#)]
93. Wheal, A.J.; Jadoon, K.; Randall, M.D.; O'Sullivan, S.E. In Vivo cannabidiol treatment improves endothelium-dependent vasorelaxation in mesenteric arteries of Zucker diabetic fatty rats. *Front. Pharmacol.* **2017**, *8*, 248. [[CrossRef](#)] [[PubMed](#)]
94. Grambow, E.; Strüder, D.; Klar, E.; Hinz, B.; Vollmar, B. Differential effects of endogenous, phyto and synthetic cannabinoids on thrombogenesis and platelet activity. *BioFactors* **2016**, *42*, 581–590. [[CrossRef](#)]
95. Fouad, A.A.; Al-Mulhim, A.S.; Jresat, I. Cannabidiol treatment ameliorates ischemia/reperfusion renal injury in rats. *Life Sci.* **2012**, *91*, 284–292. [[CrossRef](#)]
96. Soares, R.Z.; Vuolo, F.; Dall'Igna, D.M.; Michels, M.; Crippa, J.A.; Hallak, J.E.; Zuardi, A.W.; Dal-Pizzol, F. Evaluation of the role of the cannabidiol system in an animal model of ischemia/reperfusion kidney injury. *Rev. Bras. Ter. Intensiva* **2015**, *27*, 383–389. [[CrossRef](#)] [[PubMed](#)]
97. Baban, B.; Khodadadi, H.; Vaibhav, K.; Marchetti, C.; Riccardi, C.; Mozaffari, M.S. Regulation of innate lymphoid cells in acute kidney injury: Crosstalk between cannabidiol and GILZ. *J. Immunol. Res.* **2020**, *2020*, 6056373. [[CrossRef](#)] [[PubMed](#)]
98. Rock, E.M.; Sullivan, M.T.; Collins, S.A.; Goodman, H.; Limebeer, C.L.; Mechoulam, R.; Parker, L.A. Evaluation of repeated or acute treatment with cannabidiol (CBD), cannabidiolic acid (CBDA) or CBDA methyl ester (HU-580) on nausea and/or vomiting in rats and shrews. *Psychopharmacology* **2020**, *37*, 2621–2631. [[CrossRef](#)]
99. Kossakowski, R.; Schlicker, E.; Toczek, M.; Weresa, J.; Malinowska, B. Cannabidiol affects the Bezold-Jarisch reflex via TRPV1 and 5-HT₃ receptors and has peripheral sympathomimetic effects in spontaneously hypertensive and normotensive rats. *Front. Pharmacol.* **2019**, *10*, 500. [[CrossRef](#)]
100. Mukhopadhyay, P.; Rajesh, M.; Horváth, B.; Bátkai, S.; Park, O.; Tanchian, G.; Gao, R.Y.; Patel, V.; Wink, D.A.; Liaudet, L.; et al. Cannabidiol protects against hepatic ischemia/reperfusion injury by attenuating inflammatory signaling and response, oxidative/nitrative stress, and cell death. *Free Radic. Biol. Med.* **2011**, *50*, 1368–1381. [[CrossRef](#)] [[PubMed](#)]
101. Wang, Y.; Mukhopadhyay, P.; Cao, Z.; Wang, H.; Feng, D.; Haskó, G.; Mechoulam, R.; Gao, B.; Pacher, P. Cannabidiol attenuates alcohol-induced liver steatosis, metabolic dysregulation, inflammation and neutrophil-mediated injury. *Sci. Rep.* **2017**, *7*, 12064. [[CrossRef](#)] [[PubMed](#)]
102. Avraham, Y.; Grigoriadis, N.; Poutahidis, T.; Vorobiev, L.; Magen, I.; Ilan, Y.; Mechoulam, R.; Berry, E. Cannabidiol improves brain and liver function in a fulminant hepatic failure-induced model of hepatic encephalopathy in mice. *Br. J. Pharmacol.* **2010**, *162*, 1650–1658. [[CrossRef](#)]

103. Vilela, L.R.; Gomides, L.F.; David, B.A.; Antunes, M.M.; Diniz, A.B.; Moreira, F.; Menezes, G.B. Cannabidiol rescues acute hepatic toxicity and seizure induced by cocaine. *Mediators Inflamm.* **2015**, *2015*, 523418. [[CrossRef](#)]
104. Li, K.; Feng, J.Y.; Li, Y.Y.; Yuce, B.; Lin, X.H.; Yu, L.Y.; Li, Y.N.; Feng, Y.J.; Storr, M. Anti-inflammatory role of cannabidiol and O-1602 in cerulein-induced acute pancreatitis in mice. *Pancreas* **2013**, *42*, 123–129. [[CrossRef](#)] [[PubMed](#)]
105. Capasso, R.; Borrelli, F.; Aviello, G.; Romano, B.; Scalisi, C.; Capasso, F.; Izzo, A.A. Cannabidiol, extracted from *Cannabis sativa*, selectively inhibits inflammatory hypermotility in mice. *Br. J. Pharmacol.* **2008**, *154*, 1001–1008. [[CrossRef](#)] [[PubMed](#)]
106. Pagano, E.; Capasso, R.; Piscitelli, F.; Romano, B.; Parisi, O.A.; Finizio, S.; Lauritano, A.; Marzo, V.D.; Izzo, A.A.; Borrelli, F. An orally active cannabis extract with high content in cannabidiol attenuates chemically-induced intestinal inflammation and hypermotility in the mouse. *Front. Pharmacol.* **2016**, *7*, 341. [[CrossRef](#)] [[PubMed](#)]
107. Gáll, Z.; Farkas, S.; Albert, Á.; Ferencz, E.; Vancea, S.; Urkon, M.; Kolcsár, M. Effects of chronic cannabidiol treatment in the rat chronic unpredictable mild stress model of depression. *Biomolecules* **2020**, *10*, 801. [[CrossRef](#)] [[PubMed](#)]
108. Fiani, B.; Sarhadi, K.J.; Soula, M.; Zafar, A.; Quadri, S.A. Current application of cannabidiol (CBD) in the management and treatment of neurological disorders. *Neurol. Sci.* **2020**, *41*, 3085–3098. [[CrossRef](#)]
109. Patra, P.H.; Barker-Haliski, M.; White, H.S.; Whalley, B.J.; Glyn, S.; Sandhu, H.; Jones, N.; Bazelot, M.; Williams, C.M.; McNeish, A.J. Cannabidiol reduces seizures and associated behavioral comorbidities in a range of animal seizure and epilepsy models. *Epilepsia* **2019**, *60*, 303–314. [[CrossRef](#)]
110. Vilela, L.R.; Lima, I.V.; Kunsch, É.B.; Pinto, H.; De Miranda, A.S.; Vieira, É.; De Oliveira, A.; Moraes, M.; Teixeira, A.L.; Moreira, F.A. Anticonvulsant effect of cannabidiol in the pentylenetetrazole model: Pharmacological mechanisms, electroencephalographic profile, and brain cytokine levels. *Epilepsy Behav.* **2017**, *75*, 29–35. [[CrossRef](#)]
111. Pazos, M.R.; Mohammed, N.; Lafuente, H.; Santos, M.; Martínez-Pinilla, E.; Moreno, E.; Valdizan, E.; Romero, J.; Pazos, A.; Franco, R.; et al. Mechanisms of cannabidiol neuroprotection in hypoxic-ischemic newborn pigs: Role of 5HT_{1A} and CB2 receptors. *Neuropharmacology* **2013**, *71*, 282–291. [[CrossRef](#)]
112. Lafuente, H.; Pazos, M.R.; Alvarez, A.; Mohammed, N.; Santos, M.; Arizti, M.; Alvarez, F.J.; Martinez-Orgado, J.A. Effects of cannabidiol and hypothermia on short-term brain damage in new-born piglets after acute hypoxia-ischemia. *Front. Neurosci.* **2016**, *10*, 323. [[CrossRef](#)]
113. Ruiz-Valdepeñas, L.; Martínez-Orgado, J.A.; Benito, C.; Millán, A.; Tolón, R.M.; Romero, J. Cannabidiol reduces lipopolysaccharide-induced vascular changes and inflammation in the mouse brain: An intravital microscopy study. *J. Neuroinflamm.* **2011**, *8*, 5. [[CrossRef](#)] [[PubMed](#)]
114. Hayakawa, K.; Mishima, K.; Irie, K.; Hazekawa, M.; Mishima, S.; Fujioka, M.; Orito, K.; Egashira, N.; Katsurabayashi, S.; Takasaki, K.; et al. Cannabidiol prevents a post-ischemic injury progressively induced by cerebral ischemia via a high-mobility group box1-inhibiting mechanism. *Neuropharmacology* **2008**, *55*, 1280–1286. [[CrossRef](#)] [[PubMed](#)]
115. Mishima, K.; Hayakawa, K.; Abe, K.; Ikeda, T.; Egashira, N.; Iwasaki, K.; Fujiwara, M. Cannabidiol prevents cerebral infarction via a serotonergic 5-hydroxytryptamine 1A receptor-dependent mechanism. *Stroke* **2005**, *36*, 1077–1082. [[CrossRef](#)] [[PubMed](#)]
116. Liou, G.; El-Remessy, A.; Ibrahim, A.; Caldwell, R.; Khalifa, Y.; Gunes, A.; Nussbaum, J. Cannabidiol as a putative novel therapy for diabetic retinopathy: A postulated mechanism of action as an entry point for biomarker-guided clinical development. *Curr. Pharmacogen. Person Med.* **2009**, *7*, 215–222. [[CrossRef](#)] [[PubMed](#)]
117. Liou, G.I.; Auchampach, J.A.; Hillard, C.J.; Zhu, G.; Yousufzai, B.; Mian, S.; Khan, S.; Khalifa, Y. Mediation of cannabidiol anti-inflammation in the retina by equilibrative nucleoside transporter and A2A adenosine receptor. *Invest. Ophthalmol. Vis. Sci.* **2008**, *9*, 5526–5531. [[CrossRef](#)]
118. Bielawiec, P.; Harasim-Symbor, E.; Konstantynowicz-Nowicka, K.; Sztolszter, K.; Chabowski, A. Chronic cannabidiol administration attenuates skeletal muscle de novo ceramide synthesis pathway and related metabolic effects in a rat model of high-fat diet-induced obesity. *Biomolecules* **2020**, *10*, 1241. [[CrossRef](#)]
119. Zorzenon, M.; Santiago, A.N.; Mori, M.A.; Piovan, S.; Jansen, C.A.; Perina Padilha, M.E.; Ciotta, S.R.; Cezar De Freitas Mathias, P.; Guimarães, F.S.; Weffort De Oliveira, R.M.; et al. Cannabidiol improves metabolic dysfunction in middle-aged diabetic rats submitted to a chronic cerebral hypoperfusion. *Chem. Biol. Interact.* **2019**, *312*, 108819. [[CrossRef](#)]
120. Paoli, A.; Gorini, S.; Caprio, M. The dark side of the spoon-glucose, ketones and COVID-19: A possible role for ketogenic diet? *J. Transl. Med.* **2020**, *18*, 441. [[CrossRef](#)]
121. Iannotti, F.A.; Pagano, E.; Moriello, A.S.; Alvino, F.G.; Sorrentino, N.C.; D’Orsi, L.; Gazzero, E.; Capasso, R.; De Leonibus, E.; De Petrocellis, L.; et al. Effects of non-euphoric plant cannabinoids on muscle quality and performance of dystrophic mdx mice. *Br. J. Pharmacol.* **2019**, *176*, 1568–1584. [[CrossRef](#)]
122. Atalay, S.; Gegotek, A.; Wroński, A.; Domigues, P.; Skrzydlewska, E. Therapeutic application of cannabidiol on UVA and UVB irradiated rat skin. A proteomic study. *J. Pharm. Biomed. Anal.* **2021**, *192*, 113656. [[CrossRef](#)]
123. Oláh, A.; Tóth, B.I.; Borbíró, I.; Sugawara, K.; Szöllösi, A.G.; Czifra, G.; Pál, B.; Ambrus, L.; Kloepper, J.; Camera, E.; et al. Cannabidiol exerts sebostatic and antiinflammatory effects on human sebocytes. *J. Clin. Invest.* **2014**, *124*, 3713–3724. [[CrossRef](#)]
124. Petrosino, S.; Verde, R.; Vaia, M.; Allarà, M.; Iuvone, T.; Di Marzo, V. Anti-inflammatory properties of cannabidiol, a nonpsychoactive cannabinoid, in experimental allergic contact dermatitis. *J. Pharmacol. Exp. Ther.* **2018**, *365*, 652–663. [[CrossRef](#)]
125. Cassol, O.J.; Comim, C.M.; Silva, B.R.; Hermani, F.V.; Constantino, L.S.; Felisberto, F.; Petronilho, F.; Hallak, J.E.; De Martinis, B.S.; Zuardi, A.W.; et al. Treatment with cannabidiol reverses oxidative stress parameters, cognitive impairment and mortality in rats submitted to sepsis by cecal ligation and puncture. *Brain Res.* **2010**, *1348*, 128–138. [[CrossRef](#)]

126. Abdallah, S.J.; Smith, B.M.; Ware, M.A.; Moore, M.; Li, P.Z.; Bourbeau, J.; Jensen, D. Effect of vaporized cannabis on exertional breathlessness and exercise endurance in advanced chronic obstructive pulmonary disease. A Randomized Controlled Trial. *Ann. Am. Thorac. Soc.* **2018**, *15*, 1146–1158. [[CrossRef](#)]
127. Pickering, E.E.; Semple, S.J.; Nazir, M.S.; Murphy, K.; Snow, T.M.; Cummin, A.R.; Moosavi, S.H.; Guz, A.; Holdcroft, A. Cannabinoid effects on ventilation and breathlessness: A pilot study of efficacy and safety. *Chron. Respir. Dis.* **2011**, *8*, 109–118. [[CrossRef](#)] [[PubMed](#)]
128. Duran, M.; Pérez, E.; Abanades, S.; Vidal, X.; Saura, C.; Majem, M.; Arriola, E.; Rabanal, M.; Pastor, A.; Farré, M.; et al. Preliminary efficacy and safety of an oromucosal standardized cannabis extract in chemotherapy-induced nausea and vomiting. *Br. J. Clin. Pharmacol.* **2010**, *70*, 656–663. [[CrossRef](#)] [[PubMed](#)]
129. Luftner, D.; Possinger, K.; Ernst, G.; Ruhstaller, T.; Meissner, W.; Ko, Y.D.; Schnelle, M.; Reif, M.; Cerny, T. Comparison of orally administered cannabis extract and delta-9-tetrahydrocannabinol in treating patients with cancer-related anorexia-cachexia syndrome: A multicenter, phase III, randomized, double-blind, placebo-controlled clinical trial from the Cannabis-In-Cachexia-Study-Group. *J. Clin. Oncol.* **2006**, *24*, 3394–3400. [[CrossRef](#)]
130. Couch, D.G.; Cook, H.; Ortori, C.; Barrett, D.; Lund, J.N.; O'Sullivan, S.E. Palmitoylethanolamide and cannabidiol prevent inflammation-induced hyperpermeability of the human gut in vitro and in vivo—A randomized, placebo-controlled, double-blind controlled trial. *Inflamm. Bowel Dis.* **2019**, *25*, 1006–1018. [[CrossRef](#)]
131. Naftali, T.; Mechulam, R.; Marii, A.; Gabay, G.; Stein, A.; Bronshtain, M.; Laish, I.; Benjaminov, F.; Konikoff, F.M. Low-dose cannabidiol is safe but not effective in the treatment of Crohn's disease, a randomized controlled trial. *Dig. Dis. Sci.* **2017**, *62*, 1615–1620. [[CrossRef](#)] [[PubMed](#)]
132. Irving, P.M.; Iqbal, T.; Nwokolo, C.; Subramanian, S.; Bloom, S.; Prasad, N.; Hart, A.; Murray, C.; Lindsay, J.O.; Taylor, A.; et al. A randomized, double-blind, placebo-controlled, parallel-group, pilot study of cannabidiol-rich botanical extract in the symptomatic treatment of ulcerative colitis. *Inflamm. Bowel Dis.* **2018**, *24*, 714–724. [[CrossRef](#)]
133. Zuardi, A.W.; Cosme, R.A.; Graeff, F.G.; Guimarães, F.S. Effects of ipsapirone and cannabidiol on human experimental anxiety. *J. Psychopharmacol.* **1993**, *7*, 82–88. [[CrossRef](#)]
134. Crippa, J.A.; Zuardi, A.W.; Garrido, G.E.; Wichert-Ana, L.; Guarnieri, R.; Ferrari, L.; Azevedo-Marques, P.M.; Hallak, J.E.; McGuire, P.K.; Filho Busatto, G. Effects of cannabidiol (CBD) on regional cerebral blood flow. *Neuropsychopharmacology* **2004**, *29*, 417–426. [[CrossRef](#)] [[PubMed](#)]
135. Zuardi, A.W.; Rodrigues, N.P.; Silva, A.L.; Bernardo, S.A.; Hallak, J.; Guimarães, F.S.; Crippa, J. Inverted U-shaped dose-response curve of the anxiolytic effect of cannabidiol during public speaking in real life. *Front. Pharmacol.* **2017**, *8*, 259. [[CrossRef](#)]
136. Bergamaschi, M.M.; Queiroz, R.H.; Chagas, M.H.; De Oliveira, D.C.; De Martinis, B.S.; Kapczinski, F.; Quevedo, J.; Roesler, R.; Schröder, N.; Nardi, A.E.; et al. Cannabidiol reduces the anxiety induced by simulated public speaking in treatment-naïve social phobia patients. *Neuropsychopharmacology* **2011**, *36*, 1219–1226. [[CrossRef](#)]
137. Crippa, J.A.; Derenusson, G.N.; Ferrari, T.B.; Wichert-Ana, L.; Duran, F.L.; Martin-Santos, R.; Simões, M.V.; Bhattacharyya, S.; Fusar-Poli, P.; Atakan, Z.; et al. Neural basis of anxiolytic effects of cannabidiol (CBD) in generalized social anxiety disorder: A preliminary report. *J. Psychopharmacol.* **2011**, *25*, 121–130. [[CrossRef](#)]
138. Elms, L.; Shannon, S.; Hughes, S.; Lewis, N. Cannabidiol in the treatment of post-traumatic stress disorder: A case series. *J. Altern. Complement. Med.* **2019**, *25*, 392–397. [[CrossRef](#)]
139. Shannon, S.; Lewis, N.; Lee, H.; Hughes, S. Cannabidiol in anxiety and sleep: A large case series. *Perm. J.* **2019**, *23*, 18–41. [[CrossRef](#)] [[PubMed](#)]
140. Appiah-Kusi, E.; Petros, N.; Wilson, R.; Colizzi, M.; Bossong, M.G.; Valmaggia, L.; Mondelli, V.; McGuire, P.; Bhattacharyya, S. Effects of short-term cannabidiol treatment on response to social stress in subjects at clinical high risk of developing psychosis. *Psychopharmacology* **2020**, *237*, 1121–1130. [[CrossRef](#)]
141. Wilson, R.; Bossong, M.G.; Appiah-Kusi, E.; Petros, N.; Brammer, M.; Perez, J.; Allen, P.; McGuire, P.; Bhattacharyya, S. Cannabidiol attenuates insular dysfunction during motivational salience processing in subjects at clinical high risk for psychosis. *Transl. Psychiatry* **2019**, *9*, 203. [[CrossRef](#)] [[PubMed](#)]
142. Allsop, D.J.; Copeland, J.; Lintzeris, N.; Dunlop, A.J.; Montebello, M.; Sadler, C.; Rivas, G.R.; Holland, R.M.; Muhleisen, P.; Norberg, M.M.; et al. Nabiximols as an agonist replacement therapy during cannabis withdrawal: A randomized clinical trial. *JAMA Psychiatry* **2014**, *71*, 281–291. [[CrossRef](#)]
143. Solowij, N.; Broyd, S.J.; Beale, C.; Prick, J.A.; Greenwood, L.M.; Van Hell, H.; Suo, C.; Galettis, P.; Pai, N.; Fu, S.; et al. Therapeutic effects of prolonged cannabidiol treatment on psychological symptoms and cognitive function in regular cannabis users: A pragmatic open-label clinical trial. *Cannabis Cannabinoid Res.* **2018**, *3*, 21–34. [[CrossRef](#)] [[PubMed](#)]
144. Hurd, Y.L.; Spriggs, S.; Alishayev, J.; Winkel, G.; Gurgov, K.; Kudrich, C.; Oprescu, A.M.; Salsitz, E. Cannabidiol for the reduction of cue-induced craving and anxiety in drug-abstinent individuals with heroin use disorder: A double-blind randomized placebo-controlled trial. *Am. J. Psychiatry* **2019**, *176*, 911–922. [[CrossRef](#)]
145. Hundal, H.; Lister, R.; Evans, N.; Antley, A.; Englund, A.; Murray, R.M.; Freeman, D.; Morrison, P.D. The effects of cannabidiol on persecutory ideation and anxiety in a high trait paranoid group. *J. Psychopharmacol.* **2018**, *32*, 276–282. [[CrossRef](#)]
146. Leweke, F.M.; Piomelli, D.; Pahlisch, F.; Muhl, D.; Gerth, C.W.; Hoyer, C.; Klosterkötter, J.; Hellmich, M.; Koethe, D. Cannabidiol enhances anandamide signaling and alleviates psychotic symptoms of schizophrenia. *Transl. Psychiatry* **2012**, *2*, e94. [[CrossRef](#)] [[PubMed](#)]

147. McGuire, P.; Robson, P.; Cubala, W.J.; Vasile, D.; Morrison, P.D.; Barron, R.; Taylor, A.; Wright, S. Cannabidiol (CBD) as an adjunctive therapy in schizophrenia: A multicenter randomized controlled trial. *Am. J. Psychiatry* **2018**, *175*, 225–231. [[CrossRef](#)] [[PubMed](#)]
148. Boggs, D.L.; Surti, T.; Gupta, A.; Gupta, S.; Niciu, M.; Pittman, B.; Schnakenberg Martin, A.M.; Thurnauer, H.; Davies, A.; D'Souza, D.C.; et al. The effects of cannabidiol (CBD) on cognition and symptoms in outpatients with chronic schizophrenia a randomized placebo controlled trial. *Psychopharmacology* **2018**, *235*, 1923–1932. [[CrossRef](#)]
149. Capano, A.; Weaver, R.; Burkman, E. Evaluation of the effects of CBD hemp extract on opioid use and quality of life indicators in chronic pain patients: A prospective cohort study. *Postgrad. Med.* **2020**, *132*, 56–61. [[CrossRef](#)]
150. Cuñetti, L.; Manzo, L.; Peyraube, R.; Arnaiz, J.; Curi, L.; Orihuela, S. Chronic pain treatment with cannabidiol in kidney transplant patients in Uruguay. *Transplant. Proc.* **2018**, *50*, 461–464. [[CrossRef](#)]
151. Van De Donk, T.; Niesters, M.; Kowal, M.A.; Olofsen, E.; Dahan, A.; Van Velzen, M. An experimental randomized study on the analgesic effects of pharmaceutical-grade cannabis in chronic pain patients with fibromyalgia. *Pain* **2019**, *160*, 860–869. [[CrossRef](#)]
152. Russo, M.; Naro, A.; Leo, A.; Sessa, E.; D'Aleo, G.; Bramanti, P.; Calabrò, R.S. Evaluating Sativex® in neuropathic pain management: A clinical and neurophysiological assessment in multiple sclerosis. *Pain Med.* **2016**, *17*, 1145–1154. [[CrossRef](#)]
153. Hoggart, B.; Ratcliffe, S.; Ehler, E.; Simpson, K.H.; Hovorka, J.; Lejčko, J.; Taylor, L.; Lauder, H.; Serpell, M. A multicentre, open-label, follow-up study to assess the long-term maintenance of effect, tolerance and safety of THC/CBD oromucosal spray in the management of neuropathic pain. *J. Neurol.* **2015**, *262*, 27–40. [[CrossRef](#)] [[PubMed](#)]
154. Selvarajah, D.; Gandhi, R.; Emery, C.J.; Tesfaye, S. Randomized placebo-controlled double-blind clinical trial of cannabis-based medicinal product (Sativex) in painful diabetic neuropathy: Depression is a major confounding factor. *Diabetes Care* **2010**, *33*, 128–130. [[CrossRef](#)] [[PubMed](#)]
155. Lichtman, A.H.; Lux, E.A.; McQuade, R.; Rossetti, S.; Sanchez, R.; Sun, W.; Wright, S.; Kornyeveva, E.; Fallon, M.T. Results of a double-blind, randomized, placebo-controlled study of nabiximols oromucosal spray as an adjunctive therapy in advanced cancer patients with chronic uncontrolled pain. *J. Pain Symptom Manag.* **2018**, *55*, 179–188. [[CrossRef](#)] [[PubMed](#)]
156. Miller, I.; Scheffer, I.E.; Gunning, B.; Sanchez-Carpintero, R.; Gil-Nagel, A.; Perry, M.S.; Saneto, R.P.; Checketts, D.; Dunayevich, E.; Knappertz, V. GWPCARE2 Study Group. Dose-Ranging effect of adjunctive oral cannabidiol vs placebo on convulsive seizure frequency in Dravet syndrome: A randomized clinical trial. *JAMA Neurol.* **2020**, *77*, 613–621. [[CrossRef](#)] [[PubMed](#)]
157. Devinsky, O.; Cross, J.H.; Laux, L.; Marsh, E.; Miller, I.; Nabbout, R.; Scheffer, I.E.; Thiele, E.A.; Wright, S. Cannabidiol in Dravet Syndrome Study Group. Trial of cannabidiol for drug-resistant seizures in the Dravet syndrome. *N. Engl. J. Med.* **2017**, *376*, 2011–2020. [[CrossRef](#)]
158. Devinsky, O.; Patel, A.D.; Cross, J.H.; Villanueva, V.; Wirrell, E.C.; Privitera, M.; Greenwood, S.M.; Roberts, C.; Checketts, D.; Van Landingham, K.E.; et al. GWPCARE3 Study Group. Effect of Cannabidiol on drop seizures in the Lennox-Gastaut syndrome. *N. Engl. J. Med.* **2018**, *378*, 1888–1897. [[CrossRef](#)]
159. Thiele, E.A.; Marsh, E.D.; French, J.A.; Mazurkiewicz-Beldzinska, M.; Benbadis, S.R.; Joshi, C.; Lyons, P.D.; Taylor, A.; Roberts, C.; GWPCARE4 Study Group; et al. Cannabidiol in patients with seizures associated with Lennox-Gastaut syndrome (GWPCARE4): A randomised, double-blind, placebo-controlled phase 3 trial. *Lancet* **2018**, *391*, 1085–1096. [[CrossRef](#)]
160. Wade, D.T.; Makela, P.; Robson, P.; House, H.; Bateman, C. Do cannabis-based medicinal extracts have general or specific effects on symptoms in multiple sclerosis? A double-blind, randomized, placebo-controlled study on 160 patients. *Mult. Scler.* **2004**, *10*, 434–441. [[CrossRef](#)]
161. Wade, D.T.; Makela, P.M.; House, H.; Bateman, C.; Robson, P. Long-term use of a cannabis-based medicine in the treatment of spasticity and other symptoms in multiple sclerosis. *Mult. Scler.* **2006**, *12*, 639–645. [[CrossRef](#)]
162. Contin, M.; Mancinelli, L.; Perrone, A.; Sabbatini, L.; Mohamed, S.; Scandellari, C.; Foschi, M.; Vacchiano, V.; Lugaresi, A.; Riva, R. Tetrahydrocannabinol/cannabidiol oromucosal spray in patients with multiple sclerosis: A pilot study on the plasma concentration-effect relationship. *Clin. Neuropharmacol.* **2018**, *41*, 171–176. [[CrossRef](#)]
163. Haupts, M.; Vila, C.; Jonas, A.; Witte, K.; Álvarez-Ossorio, L. Influence of previous failed antispasticity therapy on the efficacy and tolerability of THC:CBD oromucosal spray for multiple sclerosis spasticity. *Eur. Neurol.* **2016**, *75*, 236–243. [[CrossRef](#)] [[PubMed](#)]
164. Palmieri, B.; Laurino, C.; Vadalà, M. A therapeutic effect of CBD-enriched ointment in inflammatory skin diseases and cutaneous scars. *Clin. Ter.* **2019**, *170*, e93–e99. [[CrossRef](#)]
165. Singh, J.; Antimisiaris, M.F. Epidiolex-induced skin rash. *Epileptic Disord.* **2020**, *22*, 511–514. [[CrossRef](#)]
166. Černe, K. Toxicological properties of Δ^9 -tetrahydrocannabinol and cannabidiol. *Arh. Ind. Hyg. Toksikol.* **2020**, *71*, 1–11. [[CrossRef](#)]
167. Chesney, E.; McGuire, P.; Freeman, T.P.; Strang, J.; Englund, A. Lack of evidence for the effectiveness or safety of over-the-counter cannabidiol products. *Ther. Adv. Psychopharmacol.* **2020**, *10*, 2045125320954992. [[CrossRef](#)] [[PubMed](#)]
168. Dos Santos, R.G.; Guimarães, F.S.; Crippa, J.; Hallak, J.; Rossi, G.N.; Rocha, J.M.; Zuardi, A.W. Serious adverse effects of cannabidiol (CBD): A review of randomized controlled trials. *Expert Opin. Drug Metab. Toxicol.* **2020**, *16*, 517–526. [[CrossRef](#)]
169. Huestis, M.A.; Solimini, R.; Pichini, S.; Pacifici, R.; Carlier, J.; Busardò, F.P. Cannabidiol adverse effects and toxicity. *Curr. Neuropharmacol.* **2019**, *17*, 974–989. [[CrossRef](#)] [[PubMed](#)]
170. Bass, J.; Linz, D.R. A Case of toxicity from cannabidiol gummy ingestion. *Cureus* **2020**, *12*, e7688. [[CrossRef](#)]
171. Herbst, J.; Musgrave, G. Respiratory depression following an accidental overdose of a CBD-labeled product: A pediatric case report. *J. Am. Pharm. Assoc.* **2020**, *60*, 248–252. [[CrossRef](#)]

172. Devinsky, O.; Cilio, M.R.; Cross, H.; Fernandez-Ruiz, J.; French, J.; Hill, C.; Katz, R.; Di Marzo, V.; Jutras-Aswad, D.; Notcutt, W.G.; et al. Cannabidiol: Pharmacology and potential therapeutic role in epilepsy and other neuropsychiatric disorders. *Epilepsia* **2014**, *55*, 791–802. [CrossRef]
173. Adapa, S.; Gayam, V.; Konala, V.M.; Annangi, S.; Raju, M.P.; Bezwada, V.; McMillan, C.; Dalal, H.; Mandal, A.; Naramala, S. Cannabis vaping-induced acute pulmonary toxicity: Case series and review of literature. *J. Investig. Med. High. Impact Case Rep.* **2020**, *8*, 2324709620947267. [CrossRef]
174. Millar, S.A.; Stone, N.L.; Bellman, Z.D.; Yates, A.S.; England, T.J.; O'Sullivan, S.E. A systematic review of cannabidiol dosing in clinical populations. *Br. J. Clin. Pharmacol.* **2019**, *85*, 1888–1900. [CrossRef]
175. Arkell, T.R.; Lintzeris, N.; Kevin, R.C.; Ramaekers, J.G.; Vandrey, R.; Irwin, C.; Haber, P.S.; McGregor, I.S. Cannabidiol (CBD) content in vaporized cannabis does not prevent tetrahydrocannabinol (THC)-induced impairment of driving and cognition. *Psychopharmacology* **2019**, *236*, 2713–2724. [CrossRef]
176. Greene, N.Z.; Wiley, J.L.; Yu, Z.; Clowers, B.H.; Craft, R.M. Cannabidiol modulation of antinociceptive tolerance to Δ^9 -Tetrahydrocannabinol. *Psychopharmacology* **2018**, *235*, 3289–3302. [CrossRef]
177. Haney, M.; Malcolm, R.J.; Babalonis, S.; Nuzzo, P.A.; Cooper, Z.D.; Bedi, G.; Gray, K.M.; McRae-Clark, A.; Lofwall, M.R.; Sparenborg, S.; et al. Oral cannabidiol does not alter the subjective, reinforcing or cardiovascular effects of smoked Cannabis. *Neuropsychopharmacology* **2016**, *41*, 1974–1982. [CrossRef] [PubMed]
178. Karschner, E.L.; Darwin, W.D.; Goodwin, R.S.; Wright, S.; Huestis, M.A. Plasma cannabinoid pharmacokinetics following controlled oral delta9-tetrahydrocannabinol and oromucosal cannabis extract administration. *Clin. Chem.* **2011**, *57*, 66–75. [CrossRef] [PubMed]
179. Solowij, N.; Broyd, S.; Greenwood, L.M.; Van Hell, H.; Martellozzo, D.; Rueb, K.; Todd, J.; Liu, Z.; Galettis, P.; Martin, J.; et al. A randomised controlled trial of vaporised Δ^9 -tetrahydrocannabinol and cannabidiol alone and in combination in frequent and infrequent cannabis users: Acute intoxication effects. *Eur. Arch. Psychiatry Clin. Neurosci.* **2019**, *269*, 17–35. [CrossRef]
180. World Health Organization. Cannabidiol (CBD). Critical Review Report. Available online: <https://www.who.int/medicines/access/controlled-substances/CannabidiolCriticalReview.pdf> (accessed on 1 February 2021).
181. Iffland, K.; Grotenhermen, F. An update on safety and side effects of cannabidiol: A review of clinical data and relevant animal studies. *Cannabis Cannabinoid Res.* **2017**, *2*, 139–154. [CrossRef]
182. Patsalos, P.N.; Szaflarski, J.P.; Gidal, B.; Van Landingham, K.; Critchley, D.; Morrison, G. Clinical implications of trials investigating drug-drug interactions between cannabidiol and enzyme inducers or inhibitors or common antiseizure drugs. *Epilepsia* **2020**, *61*, 1854–1868. [CrossRef] [PubMed]
183. Cortopassi, J. Warfarin dose adjustment required after cannabidiol initiation and titration. *Am. J. Health Syst. Pharm.* **2020**, *77*, 1846–1851. [CrossRef] [PubMed]
184. McNamara, N.A.; Dang, L.T.; Sturza, J.; Ziobro, J.M.; Fedak Romanowski, E.M.; Smith, G.C.; Joshi, S.M.; Leber, S.M.; Carlson, M.; Robertson, P.; et al. Thrombocytopenia in pediatric patients on concurrent cannabidiol and valproic acid. *Epilepsia* **2020**, *61*, e85–e89. [CrossRef]
185. Wilson-Morkeh, H.; Al-Abdulla, A.; Sien, L.; Mohamed, H.; Youngstein, T. Important drug interactions exist between cannabidiol oil and commonly prescribed drugs in rheumatology practice. *Rheumatology* **2020**, *59*, 249–251. [CrossRef]
186. Hsu, A.; Painter, N.A. Probable interaction between warfarin and inhaled and oral administration of cannabis. *J. Pharm. Pract.* **2020**, *33*, 915–918. [CrossRef]
187. Dey, T.; Chatterjee, S.; Manna, S.; Nandy, A.; Basak, S.C. Identification and computational analysis of mutations in SARS-CoV-2. *Comput. Biol. Med.* **2020**, *129*, 104166. [CrossRef] [PubMed]
188. Santos, R.; Sampaio, W.O.; Alzamora, A.C.; Motta-Santos, D.; Alenina, N.; Bader, M.; Campagnole-Santos, M.J. The ACE2/Angiotensin-(1-7)/MAS axis of the renin-angiotensin system: Focus on Angiotensin-(1-7). *Physiol. Rev.* **2018**, *98*, 505–553. [CrossRef] [PubMed]
189. Jia, H. Pulmonary angiotensin-converting enzyme 2 (ACE2) and inflammatory lung disease. *Shock* **2016**, *46*, 239–248. [CrossRef]
190. Feng, Y.; Wan, H.; Liu, J.; Zhang, R.; Ma, Q.; Han, B.; Xiang, Y.; Che, J.; Cao, H.; Fei, X.; et al. The angiotensin-converting enzyme 2 in tumor growth and tumor-associated angiogenesis in non-small cell lung cancer. *Oncol. Rep.* **2010**, *23*, 941–948. [CrossRef]
191. Imai, Y.; Kuba, K.; Rao, S.; Huan, Y.; Guo, F.; Guan, B.; Yang, P.; Sarao, R.; Wada, T.; Leong-Poi, H.; et al. Angiotensin-converting enzyme 2 protects from severe acute lung failure. *Nature* **2005**, *436*, 112–116. [CrossRef] [PubMed]
192. He, H.; Liu, L.; Chen, Q.; Liu, A.; Cai, S.; Yang, Y.; Lu, X.; Qiu, H. Mesenchymal stem cells overexpressing angiotensin-converting enzyme 2 rescue lipopolysaccharide-induced lung injury. *Cell. Transplant.* **2015**, *24*, 1699–1715. [CrossRef] [PubMed]
193. Shenoy, V.; Kwon, K.C.; Rathinasabapathy, A.; Lin, S.; Jin, G.; Song, C.; Shil, P.; Nair, A.; Qi, Y.; Li, Q.; et al. Oral delivery of Angiotensin-converting enzyme 2 and Angiotensin-(1-7) bioencapsulated in plant cells attenuates pulmonary hypertension. *Hypertension* **2014**, *64*, 1248–1259. [CrossRef]
194. Rey-Parra, G.J.; Vadivel, A.; Coltan, L.; Hall, A.; Eaton, F.; Schuster, M.; Loibner, H.; Penninger, J.M.; Kassiri, Z.; Oudit, G.Y.; et al. Angiotensin converting enzyme 2 abrogates bleomycin-induced lung injury. *J. Mol. Med.* **2012**, *90*, 637–647. [CrossRef]
195. Marquez, A.; Wysocki, J.; Pandit, J.; Batlle, D. An update on ACE2 amplification and its therapeutic potential. *Acta. Physiol.* **2021**, *231*, e13513. [CrossRef] [PubMed]
196. Marshall, R.P.; Gohlke, P.; Chambers, R.C.; Howell, D.C.; Bottoms, S.E.; Unger, T.; McAnulty, R.J.; Laurent, G.J. Angiotensin II and the fibroproliferative response to acute lung injury. *Am. J. Physiol. Lung. Cell. Mol. Physiol.* **2004**, *286*, L156–L164. [CrossRef]

197. Tan, W.; Liao, W.; Zhou, S.; Mei, D.; Wong, W.F. Targeting the renin-angiotensin system as novel therapeutic strategy for pulmonary diseases. *Curr. Opin. Pharmacol.* **2018**, *40*, 9–17. [[CrossRef](#)]
198. Kuba, K.; Imai, Y.; Ohto-Nakanishi, T.; Penninger, J.M. Trilogy of ACE2: A peptidase in the renin-angiotensin system, a SARS receptor, and a partner for amino acid transporters. *Pharmacol. Ther.* **2010**, *128*, 119–128. [[CrossRef](#)] [[PubMed](#)]
199. Ingraham, N.E.; Barakat, A.G.; Reilkoff, R.; Bezdicek, T.; Schacker, T.; Chipman, J.G.; Tignanelli, C.J.; Puskarich, M.A. Understanding the renin-angiotensin-aldosterone-SARS-CoV axis: A comprehensive review. *Eur. Respir. J.* **2020**, *56*, 2000912. [[CrossRef](#)]
200. Marshall, R.P.; Webb, S.; Bellingan, G.J.; Montgomery, H.E.; Chaudhari, B.; McAnulty, R.J.; Humphries, S.E.; Hill, M.R.; Laurent, G.J. Angiotensin converting enzyme insertion/deletion polymorphism is associated with susceptibility and outcome in acute respiratory distress syndrome. *Am. J. Respir. Crit. Care. Med.* **2002**, *166*, 646–650. [[CrossRef](#)]
201. Cruces, P.; Díaz, F.; Puga, A.; Erranz, B.; Donoso, A.; Carvajal, C.; Wilhelm, J.; Repetto, G.M. Angiotensin-converting enzyme insertion/deletion polymorphism is associated with severe hypoxemia in pediatric ARDS. *Intensive Care. Med.* **2012**, *38*, 113–119. [[CrossRef](#)] [[PubMed](#)]
202. Bourgonje, A.R.; Abdulle, A.E.; Timens, W.; Hillebrands, J.L.; Navis, G.J.; Gordijn, S.J.; Bolling, M.C.; Dijkstra, G.; Voors, A.A.; Osterhaus, A.D.; et al. Angiotensin-converting enzyme 2 (ACE2), SARS-CoV-2 and the pathophysiology of coronavirus disease 2019 (COVID-19). *J. Pathol.* **2020**, *251*, 228–248. [[CrossRef](#)]
203. Guan, W.J.; Ni, Z.Y.; Hu, Y.; Liang, W.H.; Ou, C.Q.; He, J.X.; Liu, L.; Shan, H.; Lei, C.L.; Hui, D.; et al. Clinical characteristics of coronavirus disease 2019 in China. *N. Engl. J. Med.* **2020**, *382*, 1708–1720. [[CrossRef](#)]
204. Sharma, R.K.; Stevens, B.R.; Obukhov, A.G.; Grant, M.B.; Oudit, G.Y.; Li, Q.; Richards, E.M.; Pepine, C.J.; Raizada, M.K. ACE2 (angiotensin-converting enzyme 2) in cardiopulmonary diseases: Ramifications for the control of SARS-CoV-2. *Hypertension* **2020**, *76*, 651–661. [[CrossRef](#)]
205. Leung, J.M.; Niikura, M.; Yang, C.; Sin, D.D. COVID-19 and COPD. *Eur. Respir. J.* **2020**, *56*, 2002108. [[CrossRef](#)]
206. Bilinska, K.; Butowt., R. Anosmia in COVID-19: A bumpy road to establishing a cellular mechanism. *ACS Chem. Neurosci.* **2020**, *11*, 2152–2155. [[CrossRef](#)]
207. Kimura, H.; Francisco, D.; Conway, M.; Martinez, F.D.; Vercelli, D.; Polverino, F.; Billheimer, D.; Kraft, M. Type 2 inflammation modulates ACE2 and TMPRSS2 in airway epithelial cells. *J. Allergy. Clin. Immunol.* **2020**, *146*, 80–88. [[CrossRef](#)]
208. Jiang, F.; Yang, J.; Zhang, Y.; Dong, M.; Wang, S.; Zhang, Q.; Liu, F.F.; Zhang, K.; Zhang, C. Angiotensin-converting enzyme 2 and angiotensin 1-7: Novel therapeutic targets. *Nat. Rev. Cardiol.* **2014**, *11*, 413–426. [[CrossRef](#)]
209. Zhong, J.; Basu, R.; Guo, D.; Chow, F.L.; Byrns, S.; Schuster, M.; Loibner, H.; Wang, X.H.; Penninger, J.M.; Kassiri, Z.; et al. Angiotensin-converting enzyme 2 suppresses pathological hypertrophy, myocardial fibrosis, and cardiac dysfunction. *Circulation* **2010**, *122*, 717–728. [[CrossRef](#)]
210. Patel, V.B.; Bodiga, S.; Fan, D.; Das, S.K.; Wang, Z.; Wang, W.; Basu, R.; Zhong, J.; Kassiri, Z.; Oudit, G.Y. Cardioprotective effects mediated by angiotensin II type 1 receptor blockade and enhancing angiotensin 1-7 in experimental heart failure in angiotensin-converting enzyme 2-null mice. *Hypertension* **2012**, *59*, 1195–1203. [[CrossRef](#)] [[PubMed](#)]
211. Patel, V.B.; Zhong, J.C.; Grant, M.B.; Oudit, G.Y. Role of the ACE2/Angiotensin 1-7 axis of the renin-angiotensin system in heart failure. *Circ. Res.* **2016**, *118*, 1313–1326. [[CrossRef](#)]
212. Donoghue, M.; Wakimoto, H.; Maguire, C.T.; Acton, S.; Hales, P.; Stagliano, N.; Fairchild-Huntress, V.; Xu, J.; Lorenz, J.N.; Kadambi, V.; et al. Heart block, ventricular tachycardia, and sudden death in ACE2 transgenic mice with downregulated connexins. *J. Mol. Cell. Cardiol.* **2003**, *35*, 1043–1053. [[CrossRef](#)]
213. Saponaro, F.; Rutigliano, G.; Sestito, S.; Bandini, L.; Storti, B.; Bizzarri, R.; Zucchi, R. ACE2 in the era of SARS-CoV-2: Controversies and novel perspectives. *Front. Mol. Biosci.* **2020**, *7*, 588618. [[CrossRef](#)]
214. Oudit, G.Y.; Kassiri, Z.; Patel, M.P.; Chappell, M.; Butany, J.; Backx, P.H.; Tsushima, R.G.; Scholey, J.W.; Khokha, R.; Penninger, J.M. Angiotensin II-mediated oxidative stress and inflammation mediate the age-dependent cardiomyopathy in ACE2 null mice. *Cardiovasc. Res.* **2007**, *75*, 29–39. [[CrossRef](#)] [[PubMed](#)]
215. Yamamoto, K.; Ohishi, M.; Katsuya, T.; Ito, N.; Ikushima, M.; Kaibe, M.; Tatara, Y.; Shiota, A.; Sugano, S.; Takeda, S.; et al. Deletion of angiotensin-converting enzyme 2 accelerates pressure overload-induced cardiac dysfunction by increasing local angiotensin II. *Hypertension* **2006**, *47*, 718–726. [[CrossRef](#)] [[PubMed](#)]
216. Burrell, L.M.; Risvanis, J.; Kubota, E.; Dean, R.G.; MacDonald, P.S.; Lu, S.; Tikellis, C.; Grant, S.L.; Lew, R.A.; Smith, A.I.; et al. Myocardial infarction increases ACE2 expression in rat and humans. *Eur. Heart. J.* **2005**, *26*, 369–375. [[CrossRef](#)] [[PubMed](#)]
217. Kassiri, Z.; Zhong, J.; Guo, D.; Basu, R.; Wang, X.; Liu, P.P.; Scholey, J.W.; Penninger, J.M.; Oudit, G.Y. Loss of angiotensin-converting enzyme 2 accelerates maladaptive left ventricular remodeling in response to myocardial infarction. *Circ. Heart. Fail.* **2009**, *2*, 446–455. [[CrossRef](#)] [[PubMed](#)]
218. Epelman, S.; Shrestha, K.; Troughton, R.W.; Francis, G.S.; Sen, S.; Klein, A.L.; Tang, W.H. Soluble angiotensin-converting enzyme 2 in human heart failure: Relation with myocardial function and clinical outcomes. *J. Card. Fail.* **2009**, *15*, 565–571. [[CrossRef](#)]
219. Li, Y.; Zhou, W.; Yang, L.; You, R. Physiological and pathological regulation of ACE2, the SARS-CoV-2 receptor. *Pharmacol. Res.* **2020**, *157*, 104833. [[CrossRef](#)]
220. Hamming, I.; Timens, W.; Bulthuis, M.L.; Lely, A.T.; Navis, G.; van Goor, H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J. Pathol.* **2004**, *203*, 631–637. [[CrossRef](#)]

221. Lovren, F.; Pan, Y.; Quan, A.; Teoh, H.; Wang, G.; Shukla, P.C.; Levitt, K.S.; Oudit, G.Y.; Al-Omran, M.; Stewart, D.J.; et al. Angiotensin converting enzyme-2 confers endothelial protection and attenuates atherosclerosis. *Am. J. Physiol. Heart. Circ. Physiol.* **2008**, *295*, H1377–H1384. [[CrossRef](#)]
222. Sahara, M.; Ikutomi, M.; Morita, T.; Minami, Y.; Nakajima, T.; Hirata, Y.; Nagai, R.; Sata, M. Deletion of angiotensin-converting enzyme 2 promotes the development of atherosclerosis and arterial neointima formation. *Cardiovasc. Res.* **2014**, *101*, 236–246. [[CrossRef](#)]
223. Tang, N.; Li, D.; Wang, X.; Sun, Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J. Thromb. Haemost.* **2020**, *18*, 844–847. [[CrossRef](#)]
224. Bonow, R.O.; Fonarow, G.C.; O’Gara, P.T.; Yancy, C.W. Association of coronavirus disease 2019 (COVID-19) with myocardial injury and mortality. *JAMA Cardiol.* **2020**, *5*, 751–753. [[CrossRef](#)]
225. Sawalha, A.H.; Zhao, M.; Coit, P.; Lu, Q. Epigenetic dysregulation of ACE2 and interferon-regulated genes might suggest increased COVID-19 susceptibility and severity in lupus patients. *medRxiv* **2020**. [[CrossRef](#)]
226. Fraga-Silva, R.A.; Sorg, B.S.; Wankhede, M.; Dedeugd, C.; Jun, J.Y.; Baker, M.B.; Li, Y.; Castellano, R.K.; Katovich, M.J.; Raizada, M.K.; et al. ACE2 activation promotes antithrombotic activity. *Mol. Med.* **2010**, *16*, 210–215. [[CrossRef](#)] [[PubMed](#)]
227. Samavati, L.; Uhal, B.D. ACE2, much more than just a receptor for SARS-CoV-2. *Front. Cell. Infect. Microbiol.* **2020**, *10*, 317. [[CrossRef](#)]
228. Konturek, P.C.; Harsch, I.A.; Neurath, M.F.; Zopf, Y. COVID-19 - more than respiratory disease: A gastroenterologist’s perspective. *J. Physiol. Pharmacol.* **2020**, *71*. [[CrossRef](#)]
229. Hashimoto, T.; Perlot, T.; Rehman, A.; Trichereau, J.; Ishiguro, H.; Paolino, M.; Sigl, V.; Hanada, T.; Hanada, R.; Lipinski, S.; et al. ACE2 links amino acid malnutrition to microbial ecology and intestinal inflammation. *Nature* **2012**, *487*, 477–481. [[CrossRef](#)]
230. Garg, M.; Christensen, B.; Lubel, J.S. Gastrointestinal ACE2, COVID-19 and IBD: Opportunity in the face of tragedy? *Gastroenterology* **2020**, *159*, 1623–1624. [[CrossRef](#)] [[PubMed](#)]
231. Garg, M.; Royce, S.G.; Tikellis, C.; Shallue, C.; Batu, D.; Velkoska, E.; Burrell, L.M.; Patel, S.K.; Beswick, L.; Jackson, A.; et al. Imbalance of the renin-angiotensin system may contribute to inflammation and fibrosis in IBD: A novel therapeutic target? *Gut* **2020**, *69*, 841–851. [[CrossRef](#)]
232. Cao, X.; Lu, X.M.; Tuo, X.; Liu, J.Y.; Zhang, Y.C.; Song, L.N.; Cheng, Z.Q.; Yang, J.K.; Xin, Z. Angiotensin-converting enzyme 2 regulates endoplasmic reticulum stress and mitochondrial function to preserve skeletal muscle lipid metabolism. *Lipids. Health. Dis.* **2019**, *18*, 207. [[CrossRef](#)] [[PubMed](#)]
233. Osterreicher, C.H.; Taura, K.; De Minicis, S.; Seki, E.; Penz-Osterreicher, M.; Kodama, Y.; Kluwe, J.; Schuster, M.; Oudit, G.Y.; Penninger, J.M.; et al. Angiotensin-converting-enzyme 2 inhibits liver fibrosis in mice. *Hepatology* **2009**, *50*, 929–938. [[CrossRef](#)] [[PubMed](#)]
234. Chai, X.; Hu, L.; Zhang, Y.; Han, W.; Lu, Z.; Ke, A.; Zhou, J.; Shi, G.; Fang, N.; Fan, J.; et al. Specific ACE2 expression in cholangiocytes may cause liver damage after 2019-nCoV infection. *medRxiv* **2020**.
235. Nadarajah, R.; Milagres, R.; Dilauro, M.; Gutsol, A.; Xiao, F.; Zimpelmann, J.; Kennedy, C.; Wysocki, J.; Battle, D.; Burns, K.D. Podocyte-specific overexpression of human angiotensin-converting enzyme 2 attenuates diabetic nephropathy in mice. *Kidney Int.* **2012**, *82*, 292–303. [[CrossRef](#)] [[PubMed](#)]
236. Ortiz-Melo, D.I.; Gurley, S.B. Angiotensin converting enzyme 2 and the kidney. *Curr. Opin. Nephrol. Hypertens.* **2016**, *25*, 59–66. [[CrossRef](#)] [[PubMed](#)]
237. Cheng, Y.; Luo, R.; Wang, K.; Zhang, M.; Wang, Z.; Dong, L.; Li, J.; Yao, Y.; Ge, S.; Xu, G. Kidney disease is associated with in-hospital death of patients with COVID-19. *Kidney Int.* **2020**, *97*, 829–838. [[CrossRef](#)] [[PubMed](#)]
238. Li, Z.; Wu, M.; Yao, J.; Guo, J.; Liao, X.; Song, S.; Li, J.; Duan, G.; Zhou, Y.; Wu, X.; et al. Caution on kidney dysfunctions of COVID-19 patients. *medRxiv* **2020**.
239. Xia, H.; Lazartigues, E. Angiotensin-converting enzyme 2 in the brain: Properties and future directions. *J. Neurochem.* **2008**, *107*, 1482–1494. [[CrossRef](#)]
240. Xiao, L.; Gao, L.; Lazartigues, E.; Zucker, I.H. Brain-selective overexpression of angiotensin-converting enzyme 2 attenuates sympathetic nerve activity and enhances baroreflex function in chronic heart failure. *Hypertension* **2011**, *58*, 1057–1065. [[CrossRef](#)] [[PubMed](#)]
241. Katsi, V.; Maragkoudakis, S.; Marketou, M.; Tsioufis, C.; Parthenakis, F.; Tousoulis, D. The role of angiotensin-(1-7)/Mas axis and angiotensin type 2 receptors in the central nervous system in cardiovascular disease and therapeutics: A riddle to be solved. *Curr. Vasc. Pharmacol.* **2019**, *17*, 319–325. [[CrossRef](#)]
242. Alenina, N.; Bader, M. ACE2 in brain physiology and pathophysiology: Evidence from transgenic animal models. *Neurochem. Res.* **2019**, *44*, 1323–1329. [[CrossRef](#)]
243. Kangussu, L.M.; Marzano, L.; Souza, C.F.; Dantas, C.C.; Miranda, A.S.; Simões E Silva, A.C. The renin-angiotensin system and the cerebrovascular diseases: Experimental and clinical evidence. *Protein Pept. Lett.* **2020**, *27*, 463–475. [[CrossRef](#)]
244. Gupta, A.; Madhavan, M.V.; Sehgal, K.; Nair, N.; Mahajan, S.; Sehrawat, T.S.; Bikdeli, B.; Ahluwalia, N.; Ausiello, J.C.; Wan, E.Y.; et al. Extrapulmonary manifestations of COVID-19. *Nat. Med.* **2020**, *26*, 1017–1032. [[CrossRef](#)]
245. Machhi, J.; Herskovitz, J.; Senan, A.M.; Dutta, D.; Nath, B.; Oleynikov, M.D.; Blomberg, W.R.; Meigs, D.D.; Hasan, M.; Patel, M.; et al. The natural history, pathobiology, and clinical manifestations of SARS-CoV-2 infections. *J. Neuroimmune Pharmacol.* **2020**, *15*, 359–386. [[CrossRef](#)]

246. Zhou, Z.; Kang, H.; Li, S.; Zhao, X. Understanding the neurotropic characteristics of SARS-CoV-2: From neurological manifestations of COVID-19 to potential neurotropic mechanisms. *J. Neurol.* **2020**, *267*, 2179–2184. [[CrossRef](#)] [[PubMed](#)]
247. Najjar, S.; Najjar, A.; Chong, D.J.; Pramanik, B.K.; Kirsch, C.; Kuzniecky, R.I.; Pacia, S.V.; Azhar, S. Central nervous system complications associated with SARS-CoV-2 infection: Integrative concepts of pathophysiology and case reports. *J. Neuroinflamm.* **2020**, *17*, 231. [[CrossRef](#)]
248. Holappa, M.; Vapaatalo, H.; Vaajanen, A. Local ocular renin-angiotensin-aldosterone system: Any connection with intraocular pressure? A comprehensive review. *Ann. Med.* **2020**, *52*, 191–206. [[CrossRef](#)]
249. Holappa, M.; Vapaatalo, H.; Vaajanen, A. Many faces of renin-angiotensin system - Focus on eye. *Open. Ophthalmol. J.* **2017**, *11*, 122–142. [[CrossRef](#)]
250. Verma, A.; Shan, Z.; Lei, B.; Yuan, L.; Liu, X.; Nakagawa, T.; Grant, M.B.; Lewin, A.S.; Hauswirth, W.W.; Raizada, M.K.; et al. ACE2 and Ang-(1-7) confer protection against development of diabetic retinopathy. *Mol. Ther.* **2012**, *20*, 28–36. [[CrossRef](#)] [[PubMed](#)]
251. Zhu, P.; Verma, A.; Prasad, T.; Li, Q. Expression and function of Mas-related G Protein-Coupled Receptor D and its ligand alamandine in retina. *Mol. Neurobiol.* **2020**, *57*, 513–527. [[CrossRef](#)]
252. Duan, Y.; Beli, E.; Li Calzi, S.; Quigley, J.L.; Miller, R.C.; Moldovan, L.; Feng, D.; Salazar, T.E.; Hazra, S.; Al-Sabah, J.; et al. Loss of angiotensin-converting enzyme 2 exacerbates diabetic retinopathy by promoting bone marrow dysfunction. *Stem. Cells.* **2018**, *36*, 1430–1440. [[CrossRef](#)] [[PubMed](#)]
253. Verma, A.; Xu, K.; Du, T.; Zhu, P.; Liang, Z.; Liao, S.; Zhang, J.; Raizada, M.K.; Grant, M.B.; Li, Q. Expression of human ACE2 in Lactobacillus and beneficial effects in diabetic retinopathy in mice. *Mol. Ther. Methods. Clin. Dev.* **2019**, *14*, 161–170. [[CrossRef](#)]
254. Lazartigues, E.; Qadir, M.; Mauvais-Jarvis, F. Endocrine significance of SARS-CoV-2's reliance on ACE2. *Endocrinology* **2020**, *161*, bqaa108. [[CrossRef](#)] [[PubMed](#)]
255. Niu, M.J.; Yang, J.K.; Lin, S.S.; Ji, X.J.; Guo, L.M. Loss of angiotensin-converting enzyme 2 leads to impaired glucose homeostasis in mice. *Endocrine* **2008**, *34*, 56–61. [[CrossRef](#)]
256. Liu, F.; Long, X.; Zhang, B.; Zhang, W.; Chen, X.; Zhang, Z. ACE2 expression in pancreas may cause pancreatic damage after SARS-CoV-2 infection. *Clin. Gastroenterol. Hepatol.* **2020**, *18*, 2128–2130.e2. [[CrossRef](#)] [[PubMed](#)]
257. Gupte, M.; Thatcher, S.E.; Boustany-Kari, C.M.; Shoemaker, R.; Yiannikouris, F.; Zhang, X.; Karounos, M.; Cassis, L.A. Angiotensin converting enzyme 2 contributes to sex differences in the development of obesity hypertension in C57BL/6 mice. *Arterioscler. Thromb. Vasc. Biol.* **2012**, *32*, 1392–1399. [[CrossRef](#)]
258. Shoemaker, R.; Tannock, L.R.; Su, W.; Gong, M.; Gurley, S.B.; Thatcher, S.E.; Yiannikouris, F.; Ensor, C.M.; Cassis, L.A. Adipocyte deficiency of ACE2 increases systolic blood pressures of obese female C57BL/6 mice. *Biol. Sex. Differ.* **2019**, *10*, 45. [[CrossRef](#)] [[PubMed](#)]
259. Pan, P.P.; Zhan, Q.T.; Le, F.; Zheng, Y.M.; Jin, F. Angiotensin-converting enzymes play a dominant role in fertility. *Int. J. Mol. Sci.* **2013**, *14*, 21071–21086. [[CrossRef](#)]
260. Shan, T.; Shang, W.; Zhang, L.; Zhao, C.; Chen, W.; Zhang, Y.; Li, G. Effect of angiotensin-(1-7) and angiotensin II on the proliferation and activation of human endometrial stromal cells in vitro. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 8948–8957.
261. Domińska, K. Involvement of ACE2/Ang-(1-7)/MAS1 axis in the regulation of ovarian function in mammals. *Int. J. Mol. Sci.* **2020**, *21*, 4572. [[CrossRef](#)]
262. Reis, F.M.; Reis, A.M. Angiotensin-converting enzyme 2 (ACE2), angiotensin-(1-7) and Mas receptor in gonadal and reproductive functions. *Clin. Sci (Lond)*. **2020**, *134*, 2929–2941. [[CrossRef](#)] [[PubMed](#)]
263. Honorato-Sampaio, K.; Andrade, R.F.; Bader, M.; Martins, A.S.; Santos, R.; Reis, A.M. Genetic deletion of the Angiotensin-(1-7) receptor Mas leads to a reduced ovulatory rate. *Peptides* **2018**, *107*, 83–88. [[CrossRef](#)]
264. Jing, Y.; Run-Qian, L.; Hao-Ran, W.; Hao-Ran, C.; Ya-Bin, L.; Yang, G.; Fei, C. Potential influence of COVID-19/ACE2 on the female reproductive system. *Mol. Hum. Reprod.* **2020**, *26*, 367–373. [[CrossRef](#)]
265. Palumbo, A.; Ávila, J.; Naftolin, F. The Ovarian Renin-Angiotensin System (OVRAS): A Major Factor in Ovarian Function and Disease. *Reprod. Sci.* **2016**, *23*, 1644–1655. [[CrossRef](#)]
266. Dhaundiyal, A.; Kumari, P.; Jawalekar, S.S.; Chauhan, G.; Kalra, S.; Navik, U. Is highly expressed ACE 2 in pregnant women “a curse” in times of COVID-19 pandemic? *Life Sci.* **2021**, *264*, 118676. [[CrossRef](#)]
267. Gilbert, J.S.; LaMarca, B.B.; Granger, J.P. ACE2 and ANG-(1-7) in the gravid uterus: The new players on the block. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **2008**, *294*, R915–R916. [[CrossRef](#)]
268. Bharadwaj, M.S.; Strawn, W.B.; Groban, L.; Yamaleyeva, L.M.; Chappell, M.C.; Horta, C.; Atkins, K.; Firmes, L.; Gurley, S.B.; Brosnihan, K.B.; et al. Angiotensin-converting enzyme 2 deficiency is associated with impaired gestational weight gain and fetal growth restriction. *Hypertension* **2011**, *58*, 852–858. [[CrossRef](#)]
269. Yamaleyeva, L.M.; Pulgar, V.M.; Lindsey, S.H.; Yamane, L.; Varagic, J.; McGee, C.; daSilva, M.; Lopes Bonfa, P.; Gurley, S.B.; Brosnihan, K.B. Uterine artery dysfunction in pregnant ACE2 knockout mice is associated with placental hypoxia and reduced umbilical blood flow velocity. *Am. J. Physiol. Endocrinol. Metab.* **2015**, *309*, E84–E94. [[CrossRef](#)]
270. Lumbers, E.R.; Delforce, S.J.; Pringle, K.G.; Smith, G.R. The lung, the heart, the novel coronavirus, and the renin-angiotensin system; The need for clinical trials. *Front. Med.* **2020**, *7*, 248. [[CrossRef](#)]
271. Pascolo, L.; Zito, G.; Zupin, L.; Luppi, S.; Giolo, E.; Martinelli, M.; De Rocco, D.; Crovella, S.; Ricci, G. Renin angiotensin system, COVID-19 and male fertility: Any risk for conceiving? *Microorganisms* **2020**, *8*, 1492. [[CrossRef](#)]

272. Nozato, S.; Yamamoto, K.; Takeshita, H.; Nozato, Y.; Imaizumi, Y.; Fujimoto, T.; Yokoyama, S.; Nagasawa, M.; Takeda, M.; Hongyo, K.; et al. Angiotensin 1-7 alleviates aging-associated muscle weakness and bone loss, but is not associated with accelerated aging in ACE2-knockout mice. *Clin. Sci.* **2019**, *133*, 2005–2018. [[CrossRef](#)] [[PubMed](#)]
273. Ferrandi, P.J.; Alway, S.E.; Mohamed, J.S. The interaction between SARS-CoV-2 and ACE2 may have consequences for skeletal muscle viral susceptibility and myopathies. *J. Appl. Physiol.* **2020**, *129*, 864–867. [[CrossRef](#)] [[PubMed](#)]
274. Jin, M.; Tong, Q. Rhabdomyolysis as potential late complication associated with COVID-19. *Emerg. Infect. Dis.* **2020**, *26*, 1618–1620. [[CrossRef](#)] [[PubMed](#)]
275. Sampaio, W.O.; Nascimento, A.A.; Santos, R.A. Systemic and regional hemodynamic effects of angiotensin-(1-7) in rats. *Am. J. Physiol. Heart. Circ. Physiol.* **2003**, *284*, H1985–H1994. [[CrossRef](#)] [[PubMed](#)]
276. Krueger, J.G.; Murrell, D.F.; Garcet, S.; Navrazhina, K.; Lee, P.C.; Muscianisi, E.; Blauvelt, A. Secukinumab lowers expression of ACE2 in affected skin of patients with psoriasis. *J. Allergy Clin. Immunol.* **2020**. [[CrossRef](#)] [[PubMed](#)]
277. Xue, X.; Mi, Z.; Wang, Z.; Pang, Z.; Liu, H.; Zhang, F. High expression of ACE2 on keratinocytes reveals skin as a potential target for SARS-CoV-2. *J. Invest. Dermatol.* **2021**, *41*, 206–209.e1. [[CrossRef](#)] [[PubMed](#)]
278. Garduño-Soto, M.; Choreño-Parra, J.A.; Cazarín-Barrientos, J. Dermatological aspects of SARS-CoV-2 infection: Mechanisms and manifestations. *Arch. Dermatol. Res.* **2020**, 1–12. [[CrossRef](#)]
279. Tikellis, C.; Thomas, M.C. Angiotensin-converting enzyme 2 (ACE2) is a key modulator of the renin angiotensin system in health and disease. *Int. J. Pept.* **2012**, *2012*, 256294. [[CrossRef](#)] [[PubMed](#)]