# **Ecosystem Ecology**

# Carbon Neutral: The Failure of Dung Beetles (Coleoptera: Scarabaeidae) to Affect Dung-Generated Greenhouse Gases in the Pasture

Fallon Fowler, 1,3 Steve Denning, 1 Shuijin Hu, 2 and Wes Watson 1

<sup>1</sup>Department of Entomology and Plant Pathology, North Carolina State University, Grinnells Animal Health Laboratories, 3200 Faucette Boulevard, Raleigh, NC 27607, <sup>2</sup>Department of Entomology and Plant Pathology, North Carolina State University, Varsity Research Building, 1575 Varsity Drive, Raleigh, NC 27606, and <sup>3</sup>Corresponding author, e-mail: fefowler@ncsu.edu

Subject Editor: Jason Schmidt

Received 11 May 2020; Editorial decision 14 July 2020

#### **Abstract**

Research suggests dung beetles can churn, aerate, and desiccate dung in ways that influence the dung and soil microbes producing greenhouse gases (GHGs). We examined the impacts of the tunneling beetle, *Onthophagus taurus* (Schreber), and the dwelling beetle, *Labarrus pseudolividus* (Balthasar), on the carbon dioxide ( $CO_2$ ), methane ( $CO_4$ ), and nitrous oxide ( $O_2$ ) emitted from pasture-laid bovine dung as well as their sum-total ( $CO_2$  +  $CO_4$  +  $O_2$ ) effect on global warming, or their carbon dioxide equivalent ( $O_2$ ). Despite dung beetles potential effects on  $O_4$  and  $O_2$ 0, the existing literature shows no ultimate  $O_2$ 0 ereductions. We hypothesized that more dung beetles would degrade pats faster and reduce  $O_2$ 0, and so we increased the average dung beetle biomass per dung volume 6.22x above previously published records, and visually documented any dung damage. However, the time effects were 2–5x greater for any GHG and  $O_2$ 0 ( $O_2$ 1) than dung beetle effects alone ( $O_2$ 1). This suggests that dung beetle communities cannot adequately reduce GHGs unless they can accelerate dung decomposition faster than time alone.

Key words: dung beetle, tunneling, dwelling, greenhouse gas, dung decomposition

Global CO<sub>2</sub>e estimates for animal agriculture range from 18% (Steinfeld et al. 2006) to 51% (Goodland and Anhang 2009) depending on how direct and indirect GHG sources are calculated. Direct emissions include cattle respiration, manure emissions, and enteric fermentation from ruminants. Meanwhile, indirect emissions include the land-sparing effect (emissions from deforested, degraded, or polluted habitats; reduced environmental sequestration capacity), supply-chain emissions (storage, refrigeration, transportation, processing, food waste), and third-world impacts (food imports and their associated environmental impact outside the country-of-interest). The IPCC (Intergovernmental Panel on Climate Change) reports that animal agriculture directly produces 23% of anthropogenic CO<sub>2</sub>e by producing 33% CH<sub>4</sub>, 13% CO<sub>2</sub>, and >33% N<sub>2</sub>O globally—primarily through a combination of enteric fermentation, deforestation, and manure deposition (Jia et al. 2019). However, when also considering indirect emissions, animal agriculture produces up to 37% of global CO<sub>2</sub>e emissions (Mbow et al. 2019). This is because animal agriculture is part of an interconnected web of industries (oil, coal, deforestation, plastic-use, etc.) and trade economies; only focusing on by-sector categories (transportation, electricity, industrial, etc.) or by-country categories (neglects importation considerations) unintentionally masks strong climate-affecting connections.

A key aspect in GHG production is habitat replacement, degradation, and/or destruction as this hinders the ecosystem's ability to provide benefits, support diverse wildlife, and sequester GHGs (Steinfeld et al. 2006, Shukla et al. 2019). For example, land-use change from 1750 to 2011 produces nearly half of the CO<sub>2</sub> generated by fossil fuels alone (Jia et al. 2019), hence why habitat-affecting industries are targeted for climate change reform; they hinder the global effort to reduce and reverse global warming. Animal agriculture occupies ~70% of all agricultural land (Steinfeld et al. 2006) and 30-47% of the global ice-free surface (Shukla et al. 2019), with cattle alone accounting for 65-77% of global livestock emissions (Smith et al. 2019). Cattle systems are highly interconnected with deforestation, fossil fuels, and intensive resource use as their environmental impacts includes the impact of everything they consume as well (Poore and Nemecek 2018). Thus, reducing animal agriculture's land-use footprint-often from localized pollution (i.e., waste, pesticides, antibiotics, fertilizer for feed crops), habitat replacement/destruction (deforestation, overgrazing), and resource overuse and inefficiency (land, water, oil)—is key. Converting these lands to wildlife ecosystems would both reduce and sequester existing GHGs (Poore and Nemecek 2018), and limit any consequences. Climate change affects extreme weather phenomena, such as acidic rains

and oceans, glacial melting, droughts and wildfires, rising sea levels, and hurricanes/cyclones, which can easily decimate animals and habitats *en masse* (Veron 2008, Ceballos et al. 2015). This explains why five mass extinction periods, wherein ≥75% of all species went extinct, were documented throughout earth's history due to climate-induced habitat loss and ecological cascading (Veron 2008, Ceballos et al. 2015).

The two most popular solutions to reduce GHG production are: 1) reduce animal consumption and production (Mbow et al. 2019) by moving towards predominately whole foods, plant-based diets that are more sustainable (Poore and Nemecek 2018) and healthy (Willett et al. 2019); and 2) reform remaining animal operations into more resource-efficient systems. Dung beetles improve pasture health by actively dismantling and burying dung, which: 1) increases soil organic matter ('humification') and refills water tables through increased soil percolation; 2) reduces pollution, pests, and pestilence through competitive exclusion and removal; and 3) encourages cooperation among earthworms, soil arthropods, and plant species to bury seeds, enrich the soil, and/or pollinate plants (Lavelle et al. 2006, Nichols et al. 2008, Ridsdill-Smith and Edwards 2011, Doube 2018). Only recently have researchers studied the relationship between soil, dung, dung beetles, and GHGs as part of a larger resource recycling strategy (Penttilä et al. 2013, Iwasa et al. 2015, Hammer et al. 2016a, Slade et al. 2016a, Piccini et al. 2017, Evans et al. 2019). Enteric fermentation and deposited, applied, and flooded manure and dung (Mbow et al. 2019) produce 66% CH<sub>4</sub> of all agricultural emissions, with deposited manure and dung also producing >50% N<sub>2</sub>O net agricultural emissions (Jia et al. 2019)—this does not include the emissions generated by applied synthetic fertilizer to animal food crops. Thus, dung management by dung beetles present an opportunity for reducing GHGs in addition to reducing disease, pollution, compaction, improving C and N storage/use, and overall sustainability.

Dung beetles generally decrease CH<sub>4</sub>, increase N<sub>2</sub>O, and yield no CO<sub>2</sub>e effect—though in some cases they increased CO<sub>2</sub>e (Fowler et al. 2020c). We propose that increasing dung beetle biomass may sufficiently alter the dung structure and reduce CO<sub>2</sub>e. Therefore,

we increased dung beetle biomass per unit dung above previously reported estimates (Table 1) and compared if major dung altering behaviors (mixing vs. burial) accelerated dung decomposition and reduced more GHGs.

#### **Materials and Methods**

#### **Experimental Design**

At the beef unit (latitude 35°43′47.40′′N, longitude 78°41′15.50′′W) of NCSU Lake Wheeler Road Field Lab (Raleigh, NC), we set up a randomized complete block design (n = 2 blocks per experiment) of our treatments (Table 2) on fresh, untouched cattle dung (grass and hay fed) to test whether different dung beetle behaviors and abundances reduce dung-produced CO<sub>2</sub>e. We did not standardize dung so as not to disrupt/alter its natural physical and microbial properties nor confuse dung beetle activity with human activity, thus the dung's volume (via dung width, length, and depth) was calculated instead (Supp Fig. G5 [online only]), and additional replicates were processed. We measured GHGs (CH4, CO2, N2O) over a week (0, 1, 3, 5,and 7 d) using mobile GHG chambers (n = 1) treatment per block) between May and October 2018 (n = 12 experiments) totaling 24 replicates per treatment and day. On non-sampling days, cages (Fig. 1) protected treatments from dung beetle entry and exit while allowing for airflow and avoiding chamber burial (Fowler et al. 2020b). Sampling sites were enclosed with electric fences to prevent animal tampering. We photographed dung damage on 7 and 14 d for dung-based treatments and field pats, respectively (Fig. 2). We informally included naturally colonized dung pats ('field pats') in the experiment to ensure and show whether the treatment dung pats were dissembled by dung arthropods faster and more intensely than that found naturally in the surrounding area—thereby showing if natural dung beetle colonization rates are adequate for GHG reduction. See Fowler et al. (2020b) for the measurements, costs, limitations, building procedures, and verification experiments of the physical chamber designs, gas sampling strategies, and caged designs.

**Table 1.** A summary of reported GHG differences (+: increase, -: decrease, 0: no effect) focused on aggregate treatment effects of *t*-tests and ANOVAs (*P* < 0.05) that exclude strong and effect-masking predictors such as time or vegetation

Article	$n_{\rm dung}/{\rm d}$	Biomass/dung (g/L)	$CH_4$	$CO_2$	$N_2O$	CO <sub>2</sub> e	
Yokoyama et al. (1991a,b)	koyama et al. (1991a,b) 2–3		_	_	+1.93×		
Penttilä et al. (2013)	10	1.075	-1.65×	0	+27.2×	0	
Iwasa et al. $(2015)^a$	3	$0.60^{e}$	-2.61×	+7.87×	+10.81×	+1.91×	
Slade et al. (2016a,b) <sup>a</sup>	$30,20^{c}$	_	0	$0^g$	0	0	
Hammer et al. (2016a,b)	$10^d$	$0.31^{e}$	-1.59×	0	+3.02×	$O^a$	
Piccini et al. $(2017)^b$	8	1.48	$0^f$	$0^{h}$	$O^i$	$O^j$	
Evans et al. (2019)	32	<u>_e</u>	+3×	0	+3×	0	
Fowler et al. (2020c)	24	3.09	0	0	0	0	
Our data	20	3.75-7.5	-1.40×	0	+1.56×	0	

These values look at how dung beetles affect the dung GHGs relative to the dung-only for each GHG. The experimental information from the current literature (2020) includes: the dung-containing sample size total per day ( $n_{\text{dung}}/d$ ) and the dung beetle biomass per reported dung volumes (maximums only). An updated version from Fowler et al. (2020c).

<sup>&</sup>quot;Traditional *t*-test not reported in the published paper. Based on reported and overlapping SE values or a *t*-test was performed on the available raw data (Fowler et al. 2020c - Supp Table D7 [online only]).

<sup>&</sup>lt;sup>b</sup>Based only on the reported *P*-adjusted value (familywise error corrected).

<sup>&</sup>lt;sup>c</sup>Numbers represent dung beetle treatments, dung-only was n = 3 for both 2016a and b.

<sup>&</sup>lt;sup>d</sup>Considering only the non-antibiotic dung to avoid confounding effects.

<sup>&</sup>lt;sup>e</sup>Biomass unreported, so: study-reported abundance\*biomass of tunneler (Piccini, 0.20 g) or dweller (Penttilä, 0.0261 g).

<sup>&</sup>lt;sup>f</sup>Five of six dung beetle treatment reported no difference, 1 treatment increased CH<sub>4</sub> emissions (T4).

gBased on fig. A2 from Slade et al. (2016b), all other GHGs come from Slade et al.'s (2016a) supplementary materials.

<sup>&</sup>lt;sup>h</sup>Five of six dung beetle treatments reported no difference, 1 treatment decreased CO<sub>2</sub> emissions (T4).

<sup>&#</sup>x27;Six of six dung beetle treatments reported no difference for N<sub>2</sub>O.

Five of six dung beetle treatments reported no difference, 1 treatment decreased CO<sub>2</sub>e emissions (T6).

#### **Treatment Descriptions**

Dung beetle nesting behaviors include (Bertone et al. 2004): 1) endocoprids or 'dwellers', which shred dung by dwelling within cavities in and around dung; 2) paracoprids or 'tunnelers', which bury brood balls (balls of dung created by dung beetles to house and feed progeny over the course of their development) beneath pats; and 3) telecoprids or 'rollers', which form, roll, and bury brood balls elsewhere. We particularly focused on the tunneler, Onthophagus taurus, and dweller, Labarrus (Aphodius) pseudolividus, since they are the most abundant dung beetles on North Carolina cattle pastures (Bertone et al. 2005) and so could be reliably gathered throughout the season. These species also represented the major dung handling behaviors (mixing vs. burial). We determined representative dung beetle biomasses by mimicking the dung damage commonly found in local pastures to simulate reasonable to intense dung beetle activities (Fig. 2). The dung-only treatments were either completely untouched ('Unmixed') or the dung (no soil or vegetation) was mixed for ~1 min on 0 d ('Mixed'). Meanwhile the pasture-only controls ('Grass') represented nearby pasture conditions. We included a mixed dung-only to further confirm/refute if mixing itself changes GHG production pathways, though recent studies thus far show that dung homogenization, translocation, and placement do not alter them (Fowler et al. 2020c).

**Table 2.** A summary of the treatment descriptions (Tunn = Tunneler, Dwell = Dweller) including the estimated dung beetle abundances per treatment for a single replicate (n = 24)

Treatment	Beetles	Dung	Grass	Biomass (g)	Tunn (#)	Dwell (#)
Unmixed		/	<b>✓</b>	_	_	
Mixed		1	1	_	_	_
High Tunn	/	1	/	15	205	_
High Dwell	✓	1	1	15	_	2,489
Low Tunn	/	1	/	7.5	102	_
Low Dwell	/	1	/	7.5	_	1,244
Grass			✓	-	-	-

#### **Dung Beetle Collection**

Dung beetle treatments used biomass rather than abundance to avoid confounding dung beetle size and behavior. For example, O. taurus ('Tunn') is ~12× larger than L. pseudolividus ('Dwell') and so abundance varies greatly (Fowler et al. 2020a). We weighed 7.5 ('Low') and 15 g ('High') of each species (Table 2) and used the biomass-abundance conversion rate (described by Fowler et al. 2020a) to estimate beetle abundance. We wet-sieved the dung and collected dung beetles 1–2 d before experimentation (for survival stats and photographed methodology, see Fowler et al. 2020a). We replaced any dung beetles (<1%) that could not walk on 0 d with healthy dung beetles to maintain high performance activity. Dung beetles were added after GHG sampling on 0 d.

#### **Statistics**

First, we conducted power tests (packages: 'pwr', 'pwr2') in the free R statistical program (R Development Team, Geneva, Switzerland; http://www.r-project.org) to estimate the required sample sizes for 70% power given our number of contrasts (Supp Mat. P [online only]). Second, we acknowledged any heterogeneous variation, extreme outliers, and positive skewness (Erceg-Hurn and Mirosevich 2008) using Wilcox's Robust Statistics (package: WRS) (Wilcox 2013) by:

- Winsorizing extreme outliers to allow focus on gas majority representation,
- 2. identifying skewed outliers using modified M-estimators,
- 3. bootstrapping the data ( $n_{\text{boot}} = 500-600$ ) to calculate the proportion showing  $P \le 0.05$ , and
- 4. using the Benjamini–Hochberg procedure to account for familywise error, which creates Type II errors ('false positives') by chance when evaluating multiple comparisons.

Lastly, because the analyses are identical and yielded the same conclusions (Supp Table D1 [online only] vs. Supp Table D2 [online only]), we have provided a simplified treatment layout combining all dung beetle treatments (High Tunn/Dwell and Low Tunn/Dwell, where '/' means 'or') into a single treatment ('Beetle').



Fig. 1. An example of the cages (3.7 L) within a fenced-in cattle pasture. Both the window screens and dirt-covered skirt prevented dung beetle entry or exit but allowed natural airflow. Cage designs, assembly, costs, and limitations are further discussed in Fowler et al. (2020b).

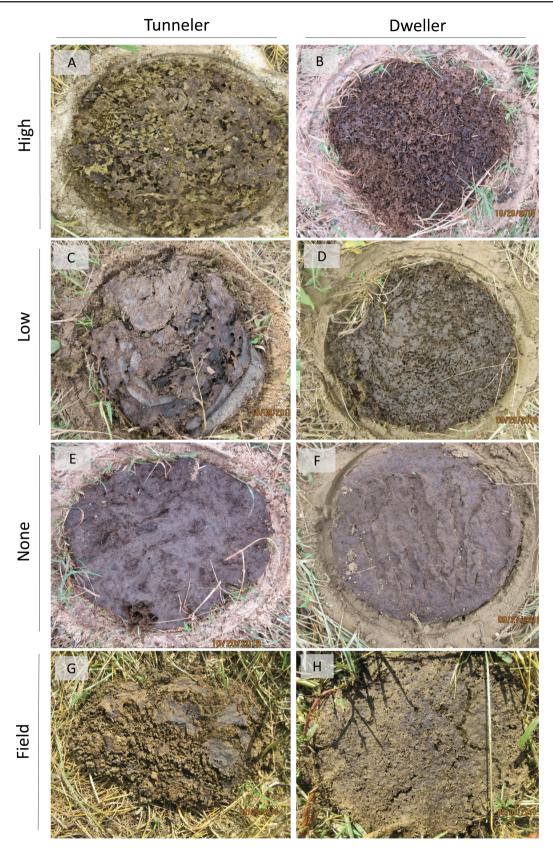


Fig. 2. Typical examples of the various treatment intensities ('Low' or 'High') of Tunneler and Dweller damage, as described in Table 2, on Unmixed dung at 7 d ('None'), with naturally colonized (no-cage) dung pats recorded at 14 d ('Field'). The lettered labels (A–H) describe both the treatment (y-axis) and dung beetle activity (x-axis) assigned to each dung pat.

#### Conversion, Tables, and Graphs

We converted our original GHG measurements (ppm) into fluxes (mg gas/m<sup>2</sup>/d) using sampling time, headspace volume (Chamber Volume - Dung Volume), and temperature expansion ratios, and by applying the modified Hutchinson and Mosier model (package: HMR) to calculate linear/nonlinear HMR fluxes (Venterea and Parkin 2012). Total CO<sub>2</sub>e were calculated as the sum of the global warming potential impact factor over 100 yr (IPCC 2007), as follows: CH<sub>4</sub> (=25 CO<sub>2</sub>e), CO<sub>2</sub> (=1 CO<sub>2</sub>e), and N<sub>2</sub>O (=298 CO<sub>2</sub>e). We included treatment, time, and treatment:time interactions (marginal means), graphs, and tables for both the original treatment designation (Supp Mat. G [online only] and Supp Table D1 [online only]) and the simplified beetle designation (Figs. 3-6; Supp Table D2 [online only]). The relevant statistics—mean-based analyses of variance (ANOVA) and median-based effect sizes—are presented on all figures, but we have also provided supplementary tables (Supp Mat. D [online only], including pseudoreplication, effect size, and comparison tables provided in Supp. Table D3-D5), and graphs (Supp Mat. G [online only]).

#### Results

Both the beetle design (Figs. 3–6) and original design (Supp Figs. G1–G3 [online only]) supplement one another by testing if dung beetles affected GHGs overall, and, if so, which groups. We examined two types of statistics here to present a more nuanced view of our data: the *P*-value and effect sizes. The *P*-value is the likelihood that the between-group mean difference ( $\neq$ 0) seen is potentially a false positive a certain percentage ( $\sim$ 5% when  $\alpha$  = 0.05) of the time given accurate ANOVA assumptions and sufficient power (Colquhoun 2014). Meanwhile the effect *size* reflects the *magnitude* of reported differences to help aid interpretative conclusions (a small/weak, medium, and large/strong explanatory measure of effect is considered at least E = 0.15, 0.30, and 0.50, respectively; Wilcox et al. 2013).

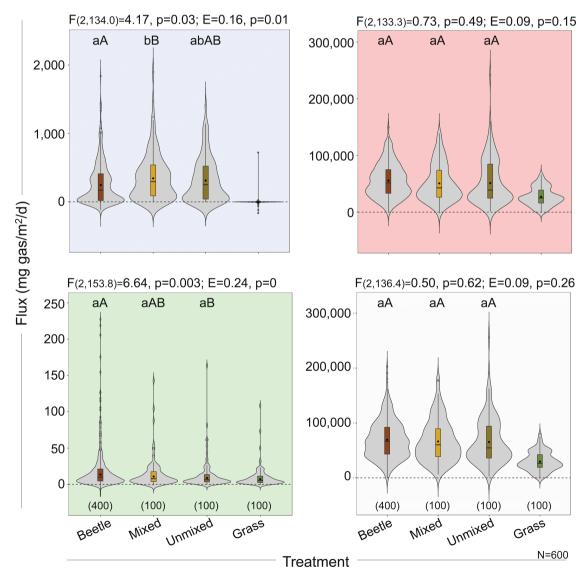


Fig. 3. Violin box plots of HMR Flux by Treatment. Each quadrant represents a GHG including  $CH_4$  (top left),  $CO_2$  (top right),  $N_2O$  (bottom left), and  $CO_2$ e (bottom right) with their respective omnibus mean-based ANOVAs ( $F_{dt num, dt den}$ ) and median-based explanatory effect (E) sizes shown above. Pairwise comparisons of ANOVAs (lowercase letters) and effect sizes (uppercase letters) are shown within the graph. Sample sizes for each treatment (n) are shown underneath each box plot, with the total sample size (N) shown along the x-axis. Differing letters between groups show differences ( $P \le 0.05$ ). Exact means, medians, and measures of variations are given in SuppTable D2 (online only).

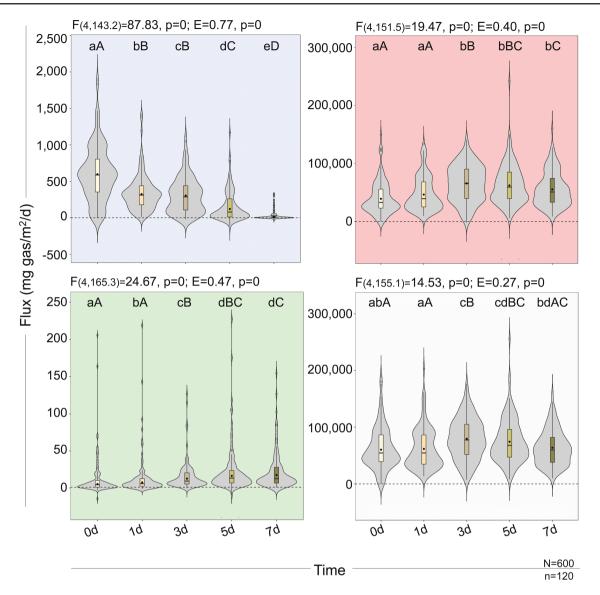


Fig. 4. Violin box plots of HMR Flux by Time. Each quadrant represents a GHG including  $CH_4$  (top left),  $CO_2$  (top right),  $N_2O$  (bottom left), and  $CO_2e$  (bottom right) with their respective omnibus mean-based ANOVAs ( $F_{df_{num,df_den}}$ ) and median-based explanatory effect (E) sizes shown above. Pairwise comparisons of ANOVAs (lowercase letters) and effect sizes (uppercase letters) are shown within the graph. Sample size (n) and total samples (N) are shown along the x-axis. Differing letters between groups show differences ( $P \le 0.05$ ). Exact means, medians, and measures of variations are found in SuppTable D2 (online only).

#### Methane

By 7 d, all dung-based groups reduced CH<sub>4</sub> by >94% compared with its 0 d flux; however, other dung-based treatments did not differ from the Unmixed dung-only (Fig. 3), thus suggesting this reduction was strongly time-related (Fig. 4). For example, time reduced CH<sub>4</sub> by 1.85-1.96× for 1 and 3 d, by 4.98× on 5 d, and by 49.83× on 7 d; meanwhile, dung beetles only reduced CH<sub>4</sub> by 1.40× across a week-not even surpassing the effect of a single day. The largest beetle reductions were seen on 3 d and 5 d when the Unmixed and Mixed dung showed 1.90-2× and 1.28-3.97× greater mean CH<sub>4</sub> fluxes than dung beetle treatments, respectively (Fig. 5). Thus, dung beetle groups inhibited CH<sub>4</sub> spikes (trajectories without peaks, Supp Fig. G3 [online only]) while dung-only encouraged it (trajectories with peaks-Fig. 6). Considering that vegetation was a weak sink  $(-1 \text{ to } -3 \text{ mg CH}_a/\text{m}^2/\text{d})$  and dung increased CH<sub>a</sub> flux by ~300× we can surmise that dung was the predominant CH4 source until 7 d, when it nears vegetative fluxes (Fig. 6). Ultimately, time was

the main effect—it explained 60% of the variation, yielded nearly 50-fold  $CH_4$  reductions (E = 0.77), predicted negative  $CH_4$  trajectory ( $R^2 = 0.68$ ;  $y = -90.30 \times + 426.60$ ), and closely approximated interaction effects (E = 0.75), while treatment did not (E = 0.16).

#### Carbon Dioxide

Dung presence enhanced  $\mathrm{CO}_2$  generation, producing  $1.94\times\mathrm{CO}_2$  overall or  $1.53-2.51\times$  on any given day than grass-only. Yet, theory holds that dung beetles significantly aerate the dung, but beetle treatments did not significantly increase  $\mathrm{CO}_2$  overall (Fig. 3), though time did (Fig. 4). However, dung beetles did produce  $1.55\times$  more  $\mathrm{CO}_2$  than the Unmixed dung-only on 1 d (Fig. 5) before declining, but all treatments showed this same negative-skewed bell curve (Fig. 6). While dung beetles had larger  $\mathrm{CO}_2$  fluxes more frequently (P=0.01) compared with Unmixed dung-only (Supp Fig. G3 [online only]), it did not compare with the time effect. For example,  $1.67\times$ ,  $1.59\times$ , and  $1.41\times$  more  $\mathrm{CO}_2$  was produced on 3, 5, and 7 d compared with 0 d. Thus, time (E=0.40), rather than treatment

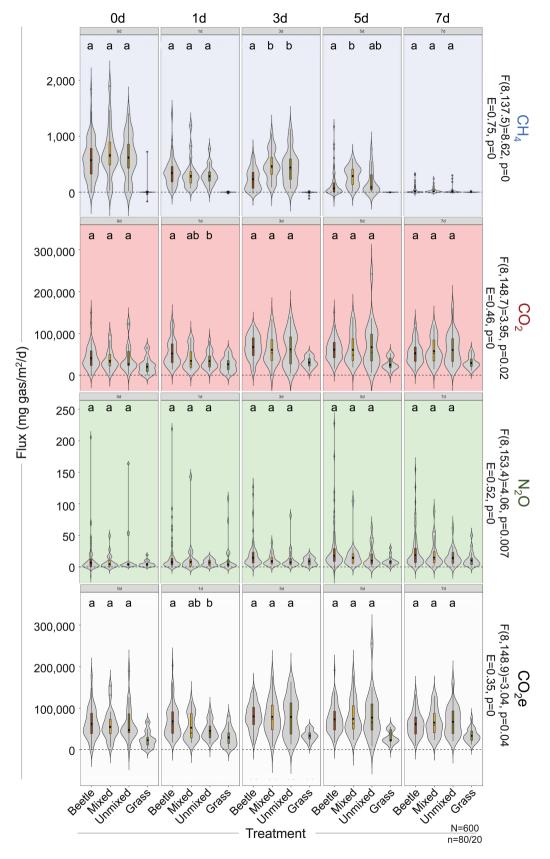


Fig. 5. Violin box plots of HMR Flux by Treatment within Time. Each quadrant represents a GHG with their respective omnibus mean-based ANOVAs ( $F_{\text{df num, df den}}$ ) and median-based explanatory effect sizes (E) shown on the graph's right. Pairwise comparisons of ANOVAs (lowercase letters) are shown within the graph. Sample size (n = beetle/other groups) and total samples (N) are shown along the x-axis. Differing letters between groups show differences ( $P \le 0.05$ ). Exact means, medians, and measures of variations are found in SuppTable D2 (online only).

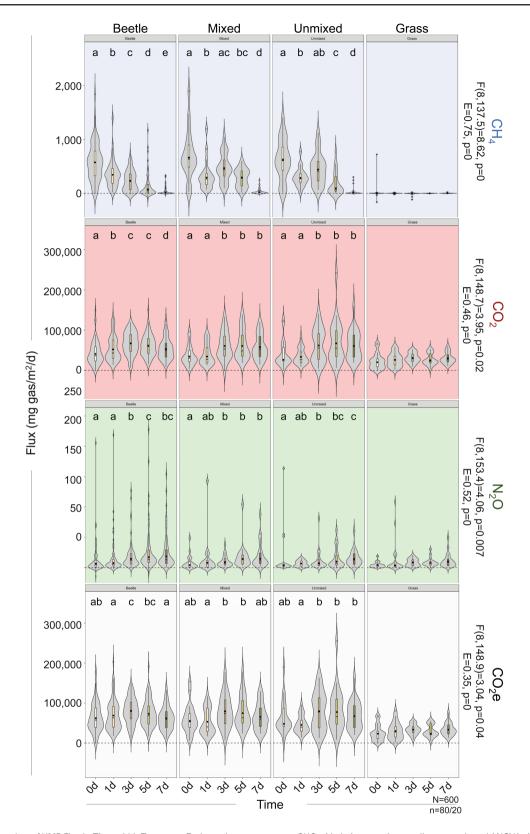


Fig. 6. Violin box plots of HMR Flux by Time within Treatment. Each quadrant represents a GHG with their respective omnibus mean-based ANOVAs ( $F_{dfnum, dfden}$ ) and median-based explanatory effect sizes (E) shown on the graph's right. Pairwise comparisons of ANOVAs (lowercase letters) are shown within the graph. Sample size (n = beetle/other groups) and total samples (N) are shown along the x-axis. Differing letters between groups show differences ( $P \le 0.05$ ). Exact means, medians, and measures of variations are found in SuppTable D2 (online only).

(E = 0.09), predicted a positive CO<sub>2</sub> trajectory ( $R^2 = 0.11$ ; y = 4,354x + 35,070) and more closely approximated the interactive effect (E = 0.46).

#### Nitrous Oxide

Cumulatively, dung beetles produced 1.56× and 1.67× greater N<sub>2</sub>O means and medians than the Unmixed dung-only, but only the medians were statistically larger (Fig. 3). This is because dung beetles produced larger N<sub>2</sub>O maximums, smaller N<sub>2</sub>O minimums, and larger N<sub>2</sub>O fluxes more frequently (P < 0.005; Supp Fig. G6 [online only]) than the Unmixed dung-only (Fig. 6), despite means showing different results (Fig. 5). Tunneler treatments were responsible for the largest fluxes, especially on 1 d (Supp Table D1 [online only] and Supp Fig. G1 [online only]). Mean-median inconsistencies indicate that reliable mean differences are infrequent and/or weak when comparing beetle and dung-only treatments (E = 0.24). This mean difference arises as dung beetle groups increased N<sub>2</sub>O emissions from 3-5 d (Supp Table D1 [online only]) and dung-only treatments only did so from 5-7 d (Fig. 6; Supp Table D2 [online only]). These earlier arising N<sub>2</sub>O emissions, when compared with Grass-only, suggests a beetle-time interaction-albeit, a minor influence. Comparatively, time surpassed treatment effects by increasing both mean and median N<sub>2</sub>O fluxes (Fig. 4) across a week (Fig. 6). From 0 d onward, both means and medians increased by ~1.5x, ~2.4x, and ~3.3x on 1, 3, and 5—7 d, respectively (E = 0.47). However, four of the seven treatments are dung beetle-based and all dung beetle groups raised N<sub>2</sub>O fluxes preemptively (Fig. 6), thus indicating that time's effect may be part of a dung beetle-time interaction. As with the other GHGs, dung presence increased total N<sub>2</sub>O generation by 1.62× compared to Grass-only, with dung-based treatments generating 1.6-2× more N<sub>2</sub>O on any given day except on 0 and 3 d (<1.25×). Interestingly, all treatments, including the Grass-only, had both N2O and CO2 follow the same increasing pattern ( $R^2 = 0.14$ ; y = 1.76x + 2.23) over time (Fig. 6), particularly for the dung beetle groups.

#### Total Greenhouse Gas Effect

CO<sub>2</sub>e is the sum of each GHG's relative effect rather than their total volume. For example, CH4 and N2O accounted for less than 0.5% of the total gases collected, but contributed 9.26 and 11.18% of the total CO<sub>2</sub>e, respectively (Supp Fig. G4 [online only]). Comparatively, CO, accounted for nearly 80% of the total CO,e, and so CO,e and CO, are identical (Figs. 3 and 5) or nearly so (Figs. 4 and 6). Once again, treatment had no reliable effect on CO<sub>2</sub>e (Fig. 3), except on 3 d in which dung beetles produced 1.06x more CO, than the Unmixed dung-only (Fig. 5). Time, meanwhile, showed a negative-skewed bell curve (Fig. 4) in which CO<sub>2</sub>e weakly increased over time for all treatments ( $R^2 = 0.008$ ; y = 1,794x + 56,960), except for 1 d, wherein dung-only treatments were at their lowest (Fig. 6). So while the dung-only treatments tended toward a higher frequency of larger CO<sub>2</sub>e fluxes (Supp Fig. G3 [online only]), this effect was inconsistent (Fig. 3) and weak (E = 0.09), especially when compared with time's consistent (Fig. 4), strong effect (E = 0.27), and its close relationship with the interaction effect (E = 0.35).

In summary, both the beetle design and original design analyses revealed the same conclusions: dung beetles weakly reduced  $\mathrm{CH}_4$ , inconsistently increased  $\mathrm{N}_2\mathrm{O}$ , and had no effect on  $\mathrm{CO}_2$  and  $\mathrm{CO}_2\mathrm{e}$  when compared with either the Mixed or Unmixed treatments. Tunneler treatments (Supp Fig. G1 [online only]) were likely responsible for the greatest  $\mathrm{CH}_4$  reductions and  $\mathrm{N}_2\mathrm{O}$  productions compared with the Mixed or Unmixed treatments. However, Beetle-Unmixed and Mixed-Unmixed contrasts prominently explain the dung beetle effect and homogenization effect, respectively. By these standards, dung beetles did not reduce  $\mathrm{CH}_4$  or  $\mathrm{CO}_2\mathrm{e}$  nor did mixing affect any GHG.

### **Discussion**

Dung beetle activity disassembles, dries, and/or buries dung (Fowler et al. 2020c) over time (Fig. 2), which changes the microbial community through the mixing of dung and soil (Yokoyama et al. 1991a, Slade et al. 2016b). Furthermore, this releases and increases dung nutrients (Yokoyama et al. 1991a, Nichols et al. 2008, Evans et al. 2019) and moisture (Penttilä et al. 2013, Evans et al. 2019, Fowler et al. 2020c) in surrounding soils; this sparks soil microbial consumption (Fowler et al. 2020c), particularly for N<sub>2</sub>O-producing microbes within the soil (Yokoyama et al. 1991a), because of the increased dung surface area. Therefore, we hypothesized that more constant activity (manual mixing on 1 d vs. dung beetle activity for 7 d), greater abundance (7 g vs. 15 g dung beetles), and soil-disrupting activities (dwelling vs. tunneling) would result in treatment-based GHG reductions—this was not the case.

#### **GHGTrends Across the Literature**

The production of CH<sub>4</sub> and N<sub>2</sub>O occur under similar anaerobic conditions (Sylvia et al. 2005), so it is surprising that the dung-only treatments saw brief CH, spikes during the week whereas dung beetle-based treatments solely decreased CH4 over time-with the situation reversed for N<sub>2</sub>O (Figs. 5 and 6). Periodic CH<sub>4</sub> (Penttilä et al. 2013, Slade et al. 2016a, Evans et al. 2019) and N2O spikes (Penttilä et al. 2013, Iwasa et al. 2015, Slade et al. 2016a, Fowler et al. 2020c) are common and likely related to dung/soil moisture (Penttilä et al. 2013, Evans et al. 2019) and/or soil disturbance (Fowler et al. 2020c)—abiotic factors heavily influenced by dung's natural decay process. Understandably, since dung beetles affect both the dung and soil matrix simultaneously, studies often focus on these two GHGs, despite beetle related CH4 reductions and N2O productions occurring only 50-63% of the time, respectively (Table 1). While our Tunneler treatments hint at this similar trend when compared to the Unmixed dung-only (Supp Fig. G1 [online only]), the effect is too weak. Meanwhile, CO, is produced under aerobic conditions and saw different trends.

Generally, dung-only showed highly variable CO, trajectories, with some studies showing increases (Slade et al. 2016b, Evans et al. 2019, Fowler et al. our data) or decreases (Penttilä et al. 2013, Iwasa et al. 2015, Piccini et al. 2017, Fowler et al. 2020c) despite using identical methodologies in some cases. Thus, CO, does not follow a familiar process as it does for CH<sub>4</sub> and N<sub>5</sub>O. More importantly, if dung beetles aerated the dung, we would expect more dung-based CO<sub>2</sub> from increased microbial respiration; however, only 12.5% of studies showed that dung beetles increased CO<sub>2</sub> production (Table 2) and our results showed only a weak effect on 1 of 5 d (Fig. 5). Explanations for this increase included: 1) dung beetle respiration (Iwasa et al. 2015), 2) enhanced gas transport (Evans et al. 2019), or 3) increased microbial populations and subsequent respiration from fresh resources (Fowler et al. 2020c). While gas transport is unknown, our recent trace gas analysis indicates CO, changes were not based on dung beetle respiration (Fowler et al. 2020c, Piccini et al. 2017). In fact, CH<sub>4</sub>, N<sub>2</sub>O, and CO<sub>2</sub> fluxes dwarfed vegetative controls by >300x, 1.94x, and 1.62x, respectively; this suggests that microbial consumption and respiration of dung-specific resources is likely the primary source of increased GHGs generally. So while dung beetle microtunnels partially explain CH<sub>4</sub>, N<sub>2</sub>O, and CO<sub>3</sub> increases in wet dung (Evans et al. 2019)—it does not explain why dried dung (Fig. 2) showed both linear increases (Fowler et al. our data) and decreases (Fowler et al. 2020c) in CO, over time using identical methodologies. Like many biological processes, CO2 production is

unclear and is guided by a myriad of processes for which there may be multiple reasons for an experiments' reported trajectory.

Overall, dung beetles weakly decreased  $\mathrm{CH_4}$  and increased  $\mathrm{N_2O}$ , while time strongly decreased  $\mathrm{CH_4}$ , increased  $\mathrm{N_2O}$ , and increased  $\mathrm{CO_2}$  over time (E = 0.27–0.77). Time effects were ~2–5× greater than any dung beetle effect alone (E = 0.09–0.24), nor were interaction effects greatly enhanced by dung beetle presence (E = 0.35–0.75), except for  $\mathrm{N_2O}$ . Dung beetles interacting with dung increased  $\mathrm{N_2O}$  compared with dung-only, which suggests an unexplored potential link between the soil and dung decay processes (Fowler et al. 2020c). Theoretically, dung beetles can drastically affect dung/soil-generated GHGs, since both time and dung beetles similarly affect dung (Penttilä et al. 2013) and soil moisture content (Evans et al. 2019, Fowler et al. 2020c), but it is likely that this effect will only show when dung beetles accelerate dung decay as strongly as time does—which ours and previous experiments do not reflect (Table 2).

#### Potential Reasons for Negligible Dung Beetle Effects

Perhaps there is no dung beetle effect on CO<sub>2</sub>e because of: 1) low dung beetle biomass per liter dung; 2) low productivity (dung damage) and cage efficiency; or 3) unrepresentative dung beetle populations.

- 1. Dung Beetle Biomass. While abundance is a useful metric, it weakly correlates with dung beetle ecosystem services and functionality when compared with biomass (Nichols et al. 2008) as individual numbers cannot predict large weight differences between and within species. For example, Piccini et al. (2017) measured the mass of the small tunneler, Onthophagus coenobita, and the larger tunneler, Copris lunaris, which showed a 10× difference in individual biomass despite being from the same functional group. Therefore, we created the largest dung beetle abundances our treatments could naturally sustain throughout the late spring, summer, and fall. Our limiting dung beetle species was L. pseudolividus and collecting >50 g dung beetle per experiment throughout the season was unreasonable. Thus, we chose biomasses similar to and 2x as large as the heaviest treatments to date (Fowler et al. 2020c) to reveal any differences, but no practical GHG differences were found despite our 15 g treatment being 6.22× larger (min: 2.42×, max: 23.94×) than the mean reported beetle biomass per dung (Table 1).
- 2. Productivity. Since no other visual dung pat damage data exist for other studies (Penttilä et al. 2013; Iwasa et al. 2015; Hammer et al. 2016a,b; Slade et al. 2016a, b; Piccini et al. 2017; Evans et al. 2019), we hypothesized greater dung beetle biomass per unit dung would also increase dung damage—it did (Fig. 2). Both the Unmixed and Mixed dung-only pats were completely solid throughout (Fig. 2E and F) and, when broken apart, gooey on the inside, which is the ideal control. Meanwhile, Tunneler treatments showed shredded damage (Fig. 2A) and/or dung tunnels or soil mound combos that increased when tunneler biomass increased. Dweller treatments were shredded at higher abundances as well (Fig. 2B), though lacked the rigor of tunneling damage despite identical biomasses. Low Dwell treatments showed pockmarked dung pats (Fig. 2D). Because of the protective measures taken with the cages (Fig. 1), the difference in visual damage between all treatments (Fig. 2), and the progressively worsening dung damage over a week-it is unlikely that dung beetle escape was an issue nor a reason for seeing minimal GHG changes.
- Population Representation. Most studies, including this one, were assumed to adequately represent rigorous dung beetle populations when this may not be the case. In our experiment, our treatment pats

were ideally over-colonized (Fig. 2A and B) compared with the surrounding field pats (Fig. 2G and H), thus giving our treatments the best chance at finding any GHG differences from dung beetle activity, or so we thought. On one rare occasion, we happened upon an unusual dung pat (Supp Fig. A [online only]) completely dissembled within 2 d—an activity not reported by any other authors (Penttilä et al. 2013; Iwasa et al. 2015; Hammer et al. 2016a, b; Slade et al. 2016a, b; Piccini et al. 2017; Evans et al. 2019; Fowler et al. 2020c), but frequently reported in wild, tropical habitats (chapter 17: Hanski and Cambefort 2014; Jones 2017) and in studies showing decimation of local fly larvae populations (Nichols et al. 2008). We found this pat (Supp Fig. A [online only]) after setting up the electric fences surrounding our experimental site on 0 d where the curious cattle lingered and defecated just outside of our sampling site. We marked the dung pats to informally compare our treatment pats to field pats (Fig. 2A and B vs. Fig. 2G and H). Even the damage sustained by 15 g dung beetles per liter dung did not compare to this intense, wild dung beetle activity. Caging dung beetles may potentially force them to withstand unfavorable conditions over time that they may not have chosen for themselves (=reduced activity)—especially when considering that dung beetles show soil, dung age, dung quality, diel, and seasonal preferences (chapter 18: Hanski and Cambefort 2014). Additionally, field pats may show rapid dung decomposition (Supp Fig. A [online only]) because of a constant daily influx of attracted dung beetles, thereby any single dung pat may experience greater beetle biomasses over time compared with restricted, caged dung pats. As evidence, when determining our experimental biomasses for this study (7.5 and 15 g), we tested various tunneler biomasses (5, 10, 15, 20, and 25 g) and noticed that >15 g dung beetle biomass showed similar dung damages to our highest biomass treatment (Fig. 2, High Tunn). Initially, we thought this meant a certain dung beetle biomass threshold guaranteed adequate dung damage, but alternatively it could mean that limited space (extreme competition) or forced cohabitation with non-preferred, aging dung result in somewhat dissembled dung pats (Fig. 2 vs. Supp Fig. A [online only]). We therefore wonder if studying the dung beetle effect from extremely active populations (disassembling dung pats within days) at their leisure may reveal their true GHG reductive potential.

#### Conclusion

Our experiment, despite reporting the highest reported dung beetle biomass per dung pat (Table 1) with observed dung damage (Fig. 2), saw no practical dung beetle effect on any GHG nor on CO<sub>2</sub>e. It is possible that dung beetles do not play a significant role in reducing dung-generated GHGs, but after finding that dung beetles can dissemble dung pats within 2 d (Supp Fig. A [online only]), it is also possible that current reported dung beetle abundances and biomasses may be insufficient in impacting GHG emissions. Thus, studying extremely active dung beetles in and around wildlife habitats may finally explain why current research reports negligible dung beetle effects on dung-generated GHGs (Fowler et al. 2020c).

## **Supplementary Data**

Supplementary data are available at *Environmental Entomology* online.

# Acknowledgments

We thank Annabel Parker for assisting with field experiments and beetle capture. We appreciate Cong Tu for conveying the basic concepts of GHG collection and measurement, as well as the NCSU Lake Wheeler Road Field Lab for

collaborating with us on cattle pasture use. This material is based upon work supported by the National Science Foundation Graduate Research Fellowship Program under grant no. DGE-1746939. Any opinions, findings, and conclusions or recommendations expressed in this material are those of the author(s) and do not necessarily reflect the views of the National Science Foundation. This research was also supported by the Center for Environmental Farming Systems Graduate Student Fellowship at North Carolina State University. FF (70%) acquired the data based on her experimental design, scripted the R code, analyzed and interpreted the data, wrote the manuscript, designed all figures, and helped with the tasks written for each author listed below. SD (20%) contributed by partially designing the GHG chambers and cages, renovating the GC-FID-ECD system, ordering materials for the chambers and GC-FID-ECD, and helping with the engineering and physics of the entire project. SH (5%) provided the GC-FID-ECD. WW (5%) contributed by suggesting the topic of how dung beetles affect GHG, providing partial funding, and editing.

#### **References Cited**

- Bertone, M., W. Watson, M. Stringham, J. Green, S. Washburn, M. Poore, and M. Hucks. 2004. Dung beetles of central and eastern North Carolina cattle pastures. North Carolina Cooperative Extension, North Carolina State University, Raleigh, NC.
- Bertone, M., J. Green, S. Washburn, M. Poore, C. Sorenson, and D. W. Watson. 2005. Seasonal activity and species composition of dung beetles (Coleoptera: Scarabaeidae and Geotrupidae) inhabiting cattle pastures in North Carolina. Ann. Entomol. Soc. Am. 98: 309–321.
- Colquhoun, D. 2014. An investigation of the false discovery rate and the misinterpretation of p-values. R. Soc. Open Sci. 1: 140216.
- Ceballos, G., P. R. Ehrlich, A. D. Barnosky, A. García, R. M. Pringle, and T. M. Palmer. 2015. Accelerated modern human-induced species losses: entering the sixth mass extinction. Sci. Adv. 1: e1400253.
- Doube, B. M. 2018. Ecosystem services provided by dung beetles in Australia. Basic Appl. Ecol. 26: 35–49.
- Erceg-Hurn, D. M., and V. M. Mirosevich. 2008. Modern robust statistical methods: an easy way to maximize the accuracy and power of your research. Am. Psychol. 63: 591–601.
- Evans, K. S., M. Mamo, A. Wingeyer, W. H. Schacht, K. M. Eskridge, J. Bradshaw, and D. Ginting. 2019. Dung beetles increase greenhouse gas fluxes from dung pats in a north temperate grassland. J. Environ. Qual. 48: 537–548
- Fowler, F., S. Wilcox, S. Nuss, and W. Watson. 2020a. Chapter 1: sampling efficacy and mortality rates of Labarrus pseudolividus (Coleoptera: Scarabaeidae) and Onthophagus taurus (Coleoptera: Scarabaeidae) using flotation and sieve-separation methodology. *In* How dung beetles affect dung-generated greenhouse gases in cattle pastures: experimental studies and literature review. PhD dissertation, North Carolina State University, Raleigh, NC.
- Fowler, F., S. Denning, and W. Watson. 2020b. Chapter 2: gases on the go: mobile chamber designs and gas techniques for field research. *In* How dung beetles affect dung-generated greenhouse gases in cattle pastures: experimental studies and literature review. PhD dissertation, North Carolina State University, Raleigh, NC.
- Fowler, F., S. Denning, S. Hu, and W. Watson. 2020c. Chapter 3: mixing the message: do dung beetles (Coleoptera: Scarabaeidae) affect dung-generated greenhouse gas emissions? In How dung beetles affect dung-generated greenhouse gases in cattle pastures: experimental studies and literature review. PhD dissertation, North Carolina State University, Raleigh, NC.
- Goodland, R., and J. Anhang. 2009. Livestock and climate change: what if the key actors in climate change are cows, pigs, and chickens? Environmental Science. Worldwatch Institute, Washington, USA.
- Hammer, T. J., N. Fierer, B. Hardwick, A. Simojoki, E. Slade, J. Taponen, H. Viljanen, and T. Roslin. 2016a. Treating cattle with antibiotics affects greenhouse gas emissions, and microbiota in dung and dung beetles. Proc. R. Soc. B 283: 20160150.
- Hammer, T. J., N. Fierer, B. Hardwick, A. Simojoki, E. Slade, J. Taponen, H. Viljanen, and T. Roslin. 2016b. Data from: Treating cattle with antibiotics affects greenhouse gas emissions, and microbiota in dung and dung beetles. Proc. R. Soc. B 283:20160150. doi:10.5061/dryad.6bs01

- Hanski, I., and Y. Cambefort. 2014. Dung beetle ecology, vol. 1195. Princeton University Press, Princeton, NJ.
- IPCC. 2007. Climate change 2007: synthesis report. Contribution of Working Groups I, II and III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Geneva, Switzerland. Intergovernmental Panel on Climate Change (IPCC), Geneva, Switzerland.
- Iwasa, M., Y. Moki, and J. Takahashi. 2015. Effects of the activity of coprophagous insects on greenhouse gas emissions from cattle dung pats and changes in amounts of nitrogen, carbon, and energy. Environ. Entomol. 44: 106–113.
- Jia, G., E. Shevliakova, P. E. Artaxo Netto, N. De Noblet-Ducoudre, R. Houghton, J. House, K. Kitajima, C. Lennard, A. Popp, A. Sirin, et al. 2019. Chapter 2: land-climate interactions. In J. Skea, E. C. Buendía, V. Masson-Delmotte, H. Pörtner, D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. van Diemen, M. Ferrat, et al. (eds.), Climate change and land: an IPCC special report on climate change, desertification, land degradation, sustainable land management, food security, and greenhouse gas fluxes in terrestrial ecosystems. In press. https://www.ipcc.ch/srccl/chapter/ chapter-2/. Accessed March 2020.
- Jones, R. 2017. Call of nature: the secret life of dung. Pelagic Publishing Ltd., Exeter, UK.
- Lavelle, P., T. Decaëns, M. Aubert, S. Barot, M. Blouin, F. Bureau, P. Margerie, P. Mora, and J. Rossi. 2006. Soil invertebrates and ecosystem services. Eur. J. Soil Biol. 42: S3–S15.
- Mbow, C., C. Rosenzweig, L. G. Barioni, T. Benton, M. Herrero, M. V. Krishnapillai, E. Liwenga, P. Pradhan, M. G. Rivera-Ferre, T. Sapkota, et al. 2019. Chapter 5: food security. In J. Skea, E. C. Buendía, V. Masson-Delmotte, H. Pörtner, D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. van Diemen, M. Ferrat, et al. (eds.), Climate change and land: an IPCC special report on climate change, desertification, land degradation, sustainable land management, food security, and greenhouse gas fluxes in terrestrial ecosystems. In press. https://www.ipcc.ch/srccl/chapter/chapter-5/. Accessed March 2020.
- Nichols, E., S. Spector, J. Louzada, T. Larsen, S. Amezquita, and M. E. Favila. 2008. Ecological functions and ecosystem services provided by Scarabaeinae dung beetles. Biol. Conserv. 141: 1461–1474.
- Penttilä, A., E. M. Slade, A. Simojoki, T. Riutta, K. Minkkinen, and T. Roslin. 2013. Quantifying beetle-mediated effects on gas fluxes from dung pats. PLoS ONE 8: e71454.
- Piccini, I., F. Arnieri, E. Caprio, B. Nervo, S. Pelissetti, C. Palestrini, T. Roslin, and A. Rolando. 2017. Greenhouse gas emissions from dung pats vary with dung beetle species and with assemblage composition. PLoS ONE 12: e0178077.
- Poore, J., and T. Nemecek. 2018. Reducing food's environmental impacts through producers and consumers. Science 360: 987–992.
- Ridsdill-Smith, T. J., and P. B. Edwards. 2011. Chapter 12: Biological control: ecosystem functions provided by dung beetles, pp. 245–266. In L. W. Simmons and T. J. Ridsdill-Smith (eds.), Ecology and evolution of dung beetles. John Wiley & Sons Ltd., West Sussex, UK.
- Shukla, P. R., J. Skea, R. Slade, R. van Diemen, E. Haughey, J. Malley, M. Pathak, and J. Portugal Pereira. 2019. Technical summary. In J. Skea, E. C. Buendía, V. Masson-Delmotte, H. Pörtner, D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. van Diemen, M. Ferrat, et al. (eds.), Climate change and land: an IPCC special report on climate change, desertification, land degradation, sustainable land management, food security, and greenhouse gas fluxes in terrestrial ecosystems. In press. https://www.ipcc.ch/srccl/chapter/technical-summary/. Accessed March 2020.
- Slade, E. M., T. Riutta, T. Roslin, and H. L. Tuomisto. 2016a. The role of dung beetles in reducing greenhouse gas emissions from cattle farming. Sci. Rep. 6: 18140.
- Slade, E. M., T. Roslin, M. Santalahti, and T. Bell. 2016b. Disentangling the 'brown world' faecal-detritus interaction web: dung beetle effects on soil microbial properties. Oikos 125: 629–635.
- Smith, P., J. Nkem, K. Calvin, D. Campbell, F. Cherubini, G. Grassi, K. Vladimir, A. Hoang Le, L. Shuaib, P. McElwee, et al. 2019. Chapter 6: interlinkages between desertification, land degradation, food security and greenhouse gas fluxes: synergies, trade-offs and integrated response options. In J. Skea, E. C. Buendía, V. Masson-Delmotte, H. Pörtner,

- D. C. Roberts, P. Zhai, R. Slade, S. Connors, R. van Diemen, M. Ferrat, et al. (eds.), Climate change and land: an IPCC special report on climate change, desertification, land degradation, sustainable land management, food security, and greenhouse gas fluxes in terrestrial ecosystems. In press. https://www.ipcc.ch/srccl/chapter/chapter-6/. Accessed March 2020.
- Steinfeld, H., P. Gerber, T. D. Wassenaar, V. Castel, M. Rosales, M. Rosales, and C. de Haan. 2006. Livestock's long shadow: environmental issues and options. Food & Agriculture Org, Rome, Italy.
- Sylvia, D. M., J. J. Fuhrmann, P. G. Hartel, and D. A. Zuberer. 2005. Principles and applications of soil microbiology. Pearson Prentice Hall, Upper Saddle River, NJ.
- Venterea, R. T., and T. B. Parkin. 2012. Chapter 19: Quantifying biases in non-steady state chamber measurements of soil-atmosphere gas exchange, pp. 327–343. *In M. A. Liebig, A. J. Franzluebbers, and R. F. Follett (eds.)*, Managing agricultural greenhouse gases. Elsevier, Boston, MA.

- Veron, J. E. 2008. Mass extinctions and ocean acidification: biological constraints on geological dilemmas. Coral Reefs 27: 459–472.
- Wilcox, R. 2013. Introduction to robust estimation and hypothesis testing, 3rd ed. Elsevier Science, Waltham, MA.
- Willett, W., J. Rockström, B. Loken, M. Springmann, T. Lang, S. Vermeulen, T. Garnett, D. Tilman, F. DeClerck, A. Wood, et al. 2019. Food in the Anthropocene: the EAT-Lancet Commission on healthy diets from sustainable food systems. Lancet 393: 447–492.
- Yokoyama, K., H. Kai, T. Koga, and T. Aibe. 1991a. Nitrogen mineralization and microbial populations in cow dung, dung balls and underlying soil affected by paracoprid dung beetles. Soil Biol. Biochem. 23: 649–653.
- Yokoyama, K., H. Kai, and H. Tsuchiyama. 1991b. Paracoprid dung beetles and gaseous loss of nitrogen from cow dung. Soil Biol. Biochem. 23: 643–647