

REVIEW

Current status of poly(ADP-ribose) polymerase inhibitors and future directions

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Laboratory of Clinical Genomics, National Cancer Center Research Institute, Tokyo, ²Department of Cancer Genome Informatics, Graduate School of Medicine, Faculty of Medicine, Osaka University, Osaka, Japan **Abstract:** Inhibitors of poly(ADP-ribose) polymerases (PARPs), which play a key role in DNA damage/repair pathways, have been developed as antitumor agents based on the concept of synthetic lethality. Synthetic lethality is the idea that cell death would be efficiently induced by simultaneous loss of function of plural key molecules, for example, by exposing tumor cells with inactivating gene mutation of BRCA-mediated DNA repair to chemically induced inhibition of PARPs. Indeed, three PARP inhibitors, olaparib, rucaparib and niraparib have already been approved in the US or Europe, mainly for the treatment of BRCA-mutant ovarian cancer. Clinical trials of various combinations of PARP inhibitors with cytotoxic or moleculartargeted agents are also underway. In particular, expanded applications of PARP inhibitors are anticipated following recent reports that defects in homologous recombination repair (HRR) are associated with mutations in repair genes other than BRCA1/BRCA2, such as ATM, ATR, PALB2, RAD51, CHEK1 and CHEK2, as well as with epigenetic loss of BRCA1 function through promoter methylation or overexpression of the BRCA2-interacting transcriptional repressor EMSY. Current topics of interest include selection of the best agent in each clinical context, identification of new treatment targets for HRR-proficient cases, and development of PARP inhibitor-based regimens that are less toxic and that prolong overall survival as well as progression-free survival. In addition, potential long-term side effects and suitable biomarkers for predicting efficacy and mechanisms of clinical resistance are in discussion. This review summarizes representative preclinical and clinical data for PARP inhibitors and discusses their potential for future applications to treat various malignancies.

Keywords: PARP inhibitors, synthetic lethality, *BRCA* mutation, homologous recombination, drug resistance, biomarkers

Introduction

Poly(ADP-ribose) polymerases (PARPs) are important components of DNA damage/repair pathways, playing key roles in genomic stability and tumor cell survival. PARP inhibitors have attracted great interest as potential antitumor agents since the concept of synthetic lethality was introduced by Bryant et al¹ and Farmer et al² in 2005. This concept is based on the idea that simultaneous loss of function of two or more key gene products can cause cell death, even if a deficiency in only one of them is not lethal. Thus, for example, tumor cells with inactivating mutation of the *BRCA* DNA-repair genes, which is not lethal, might be killed if they are also exposed to chemical inhibition of PARPs. Indeed, preclinical and clinical studies have verified antitumor effects of PARP inhibition in *BRCA*-mutant tumors.¹⁻⁸ So far, three agents (olaparib, rucaparib and niraparib) have been approved by western regulatory agencies, and >70 clinical trials evaluating PARP inhibitors for various kinds of malignancies are underway.⁹ Hereditary breast and ovarian cancer (HBOC) is a syndrome

Correspondence: Akihiro Ohmoto Laboratory of Clinical Genomics, National Cancer Center Research Institute, 5-1-1 Tsukiji, Chuo-ku, Tokyo 1040045, Japan Tel +81 3 3542 2511 Fax +81 3 3545 3567 Email aohmoto@ncc.go.jp associated with germline mutations of BRCA1/BRCA2 and is considered as the most suitable target for PARP inhibitors. A pooled analysis of 26 observational studies with a total of 3,879 ovarian cancer patients showed that germline BRCA mutations were associated with better survival, supporting the position of HBOC as a distinct clinical subtype. 10 Triplenegative breast cancer (TNBC), characterized by lack of expression of estrogen receptor, progesterone receptor and HER2 gene amplification, is a pathologically and clinically aggressive entity and is considered as the most problematic for breast cancer patients and clinicians, in which hormone treatments or anti-HER2 agents are ineffective. Defects of DNA-repair pathways including BRCA1 are reported in many TNBC cases, providing a rationale for PARP inhibitor treatment. 11 Defects in homologous recombination repair (HRR) genes other than BRCA1/BRCA2 and acquired loss of BRCA1 function through promoter methylation have also opened up the possibility of new indications for PARP inhibitor treatment. 12,13 However, only limited information is yet available about mechanisms of clinical resistance to these inhibitors or biomarkers to identify suitable cases for treatment. In this review, we summarize the current status of basic research and clinical trials on PARP inhibitors and discuss prospects for extending their applicability and improving clinical outcomes.

Mechanisms of action

PARP is a nuclear protein first reported in 1963 as a DNAdependent polyadenylic acid-synthesizing nuclear enzyme, and to date, 17 members of the PARP protein family have been identified. 14,15 PARP members are involved in various cellular processes, such as DNA repair, cellular differentiation, gene transcription, inflammation, mitosis, cell death and metabolism. Among them, PARP-1 accounts for >90% of the total PARP activity and is considered to be a key player in DNA base excision repair and repair of DNA single-strand breaks (SSBs). 16-18 PARP-1 binds to SSBs through a series of N-terminal zinc finger DNA-binding domains and catalyzes the polymerization of ADP-ribose (PARylation) with NAD+ as a substrate, resulting in the production of variable-sized polymers of ADP-ribose (PAR). 15,19,20 The amount of PAR production is regulated by the balance between PARP-1 and poly(ADP-ribose) glucohydrolase, which hydrolyzes PAR.²¹ PAR recruits SSBs repair scaffolding proteins, such as X-ray repair cross-complementing protein 1 (XRCC1), DNA ligase III and DNA polymerase beta, and concurrently modifies chromatin structure around lesions. 15,19 PARP-2, which accounts for 5%-10% of the total PARP activity, is also essential for DNA base excision repair, in collaboration with PARP-1.²² Accumulation of SSBs due to inhibition of PARylation or base excision repair produces DNA double-strand breaks (DSBs) through replication fork collapse.²³ HRR and nonhomologous end joining (NHEJ) are the main types of repair mechanisms for DSBs. 3,15,21,24 HRR is an accurate process conducted using a homologous DNA sequence and is composed of gene conversion and single-strand annealing pathways. On the other hand, NHEJ is an inaccurate process in which broken DNA ends are directly connected without using a homologous sequence. The former process occurs exclusively during the S and G2 phases of the cell cycle, while the latter is cell cycle independent. 15 While HRR normally takes a leading role in the repair of DSBs, deficiency of this process often leads to alternative NHEJ activation, and some of the genetic alterations introduced by NHEJ promote carcinogenesis by cancer-driver genes. 12,24,25 PARP-1 affects repair of DSBs and SSBs through activating ATM in the HRR pathway and inactivating DNA-dependent protein kinase in the NHEJ pathway, and thus PARP occupies a central position in the whole picture of DNA damage control and repair. 26,27 Interactions between PARP, HRR and NHEJ are concisely illustrated in a review by O'Sullivan et al.²⁶ PARP-1 expression and activity are increased in actively proliferating cells, and high PARP-1 expression is a feature of various tumors.²⁸ A preclinical study using 318 cell lines from 30 cancer types showed that PARP-1 mRNA expression was extremely high in small cell lung cancer (SCLC), in addition to hematologic malignancies. These results might provide clues to new treatment approaches for intractable tumors such as SCLC or neuroendocrine carcinomas derived from other organs, which are unresponsive to conventional molecular-targeting agents.29

In contrast to the limited efficacy of PARP inhibition alone, the combination of PARP inhibition with DNA-repair defects involving other signaling pathways is often lethal to tumor cells.³⁰ This synthetic lethality is powerful, because BRCA-deficient tumor cells have shown >1,000fold greater sensitivity to PARP inhibitors in preclinical models.^{1,2} Inactivation of BRCA1 and BRCA2, which are essential for HRR, increases the importance of PARP1 for maintaining genomic stability, and addition of PARP inhibition thus leads to cell cycle arrest and apoptosis through stimulating inaccurate NHEJ.30,31 In addition to germline BRCA-mutant tumors, BRCA deficiency is observed in somatic BRCA-mutant tumors or tumors with epigenetic loss of function through BRCA1 promoter methylation or overexpression of the BRCA2 transcription suppressor

EMSY. 12,32 In addition, large-scale genomic data showed that some BRCA-proficient tumors have defects in HRR genes such as ATM, ATR, CHK1, CHK2, PALB2, RAD51 and PTEN.13 These gene products are linked in the HRR signaling pathway as follows: ATM recruited in response to DSBs produces γH2AX and concurrently phosphorylates and activates CHK2, while ATR activated by replication protein A phosphorylates and activates CHK1, and PALB2 facilitates invasion of the BRCA2-RAD51 complex into damaged DNA by localizing BRCA2 to DSBs. 12,16 As mentioned earlier, tumors with acquired loss of BRCA1 function or defects in HRR genes share clinical and biological features with germline BRCA-mutant tumors and are known as BRCAness phenotype tumors. 12,13,24,33 Recent genomic data indicate that the frequency of BRCAness phenotype tumors is not negligible, and thus, the concept of synthetic lethality is adaptable to more cases than might have been expected. Pennington et al34 analyzed germline or somatic mutations of 30 genes, including BRCA1/BRCA2 and other HRR genes, in 390 ovarian cancer patients and showed that 31% of patients possess some deleterious alterations in HRR genes. Mateo et al³⁵ also conducted targeted sequencing of DNA-repair genes, including BRCA1/BRCA2, ATM, and CHEK2 and Fanconi anemia genes in 49 patients with castration-resistant prostate cancer and identified homozygous deletions or deleterious alterations in 33% of the cases. Large-scale integrated genomic analyses of ovarian cancer by The Cancer Genomic Atlas (TCGA) project found gene alterations in HRR genes in 51% of 316 cases.³⁶ These data highlight BRCAness phenotype tumors as a clinically promising target of PARP inhibitors. In addition to inhibiting PARylation, the inhibitors trap PARP1/PARP2 enzymes at sites of DNA damage and induce cytotoxic PARP-DNA complexes.³⁰ The effects of these agents depend mainly on the degree of PARP trapping rather than the ability to inhibit PARylation.^{22,24,37} However, it remains unclear whether an agent with stronger PARP-trapping activity would have greater clinical benefits, and prospective clinical trials are warranted to address this question.

It should be noted that DNA-repair function is intrinsically decreased in tumor cells compared with normal cells, and this disparity accounts for the high selectivity of PARP inhibitors.^{3,38} Nevertheless, it remains important to develop inhibitors with high potency and low toxicity. The clinical efficacy of PARP inhibitors as single agents is actually only moderate, except for *BRCA*-mutant or BRCAness tumors, and therefore, combination therapy is a reasonable approach to improve the utility of these inhibitors, which should

sensitize cells to DNA damage caused by various cytotoxic agents, exhibiting synthetic lethality in the clinical context.³ Some combined regimens have been tested, but it appears that the combination effects are dependent on the partner cytotoxic agent used. 24,39 In in vitro preclinical studies, olaparib showed better synergy than veliparib with temozolomide (TMZ), whereas rucaparib showed the strongest combination effect with topotecan but the weakest effect with paclitaxel (PTX) or gemcitabine (GEM).³⁹⁻⁴¹ In terms of mechanisms, accumulated DNA damage and increased PARP dependency for repair are suggested for the combinations with platinum agents or topoisomerase-1 inhibitors, elevated PARP1 trapping for the combinations with TMZ, and induced BRCAness phenotype for the combinations with taxanes. 42 The next step in evaluating such combinations would be in vivo studies or clinical trials.

Some preclinical studies have recently demonstrated a unique and promising combination effect between PARP inhibitors and alkylating agent TMZ in *isocitrate dehydrogenase 1 (IDH1)*-mutant tumors. ^{43,44} The synthesis of NAD⁺ as a substrate PARylation is reduced in *IDH1*-mutant cases, and this depletion of NAD⁺ is speculated to result in enhancement of PARP inhibition. Here, TMZ promotes PARP recruitment through inducing DNA damage, reasonably explaining interaction of above two agents, which indicates a broader application of PARP inhibitors in various malignancies with NAD metabolic defects, including glioma.

Clinical applications of PARP inhibitors

Clinical applications of PARP inhibitors are the most advanced in ovarian cancer, and so far, olaparib, rucaparib and niraparib are commercially available in the US or Europe. However, many clinical trials are underway, including malignancies such as breast cancer and prostate cancer, as well as ovarian cancer, and tumors without *BRCA* mutations, such as BRCAness phenotype tumors. In this section, we review clinical evidence of the efficacy of five PARP inhibitors (olaparib, talazoparib, rucaparib, veliparib and niraparib; Table 1).

Olaparib

Fong et al⁴⁵ conducted the first Phase I trial of olaparib (AZD-2281; AstraZeneca plc, Cambridge, UK) in 60 patients including 22 *BRCA*-mutant cases, and the maximum tolerated dose (MTD) was established as 400 mg twice daily. An additional cohort study of 50 patients with *BRCA*-mutant ovarian cancer showed that the overall clinical

Table I Summary of Phase II/III trials of PARP inhibitors

,	Phase	Number	Tumor type	ORR	Median PFS	Median OS
ı		of patients				
Olaparib ³¹	2	50	Metastatic castration-	33% (entire cohort), 88% (genomic	9.8 months (genomic defects positive	13.8 months (genomic defects
			resistant prostate cancer	defects positive cases in DNA-repair	cases in DNA-repair genes), 2.7 months	positive cases in DNA-repair
				genes), 6% (genomic defects negative	(genomic defects negative cases in	genes), 7.5 months (genomic defects
				cases in DNA-repair genes)	DNA-repair genes)	negative cases in DNA-repair genes)
Olaparib ⁴¹	2	57	Recurrent BRCA-mutant	33% (400 mg twice daily),	5.8 months (400 mg twice daily),	Not reported
			ovarian cancer	13% (100 mg twice daily)	I.9 months (100 mg twice daily)	
Olaparib ⁴²	2	298	Recurrent BRCA-mutant	26% (entire cohort), 31% (ovarian),	7.0 months (ovarian), 3.7 months	16.6 months (ovarian), 11 months
			cancer	13% (breast), 22% (pancreatic),	(breast), 4.6 months (pancreatic),	(breast), 9.8 months (pancreatic),
				50% (prostate)	7.2 months (prostate)	18.4 months (prostate)
Olaparib ⁴³	2	64	Advanced HGSOC or TNBC	41% (BRCA mutant), 24% (BRCA wild)	7.2 months (ovarian), 1.8 months (breast)	Not reported
Olaparib ⁴⁴	2	265	Recurrent platinum-sensitive	12% (olaparib), 4% (placebo)	8.4 months (olaparib), 4.8 months	29.7 months (olaparib), 29.9 months
			HGSOC		(placebo)	(placebo)
Olaparib ⁴⁶	2	54	Recurrent BRCA-mutant	41% (400 mg twice daily),	5.7 months (400 mg twice daily),	Not reported
			breast cancer	22% (100 mg twice daily)	3.8 months (100 mg twice daily)	
Rucaparib ⁴⁹	7	35	Recurrent BRCA-mutant	%59	Not reached	Not reported
			ovarian cancer			
$Rucaparib^{50}$	2	206	Recurrent platinum-sensitive	80% (BRCA mutant), 29% (BRCA wild/	12.8 months (BRCA mutant), 5.7 months	Not reported
			high-grade ovarian cancer	LOH high),	(BRCA wild/LOH high), 5.2 months	
				10% (BRCA wild/LOH low)	(BRCA wild/LOH low)	
Veliparib ⁵²	2	50	Persistent or recurrent BRCA-	26%	8.2 months	Not reached
			mutant epithelial ovarian,			
			fallopian tube or primary			
			peritoneal cancer			
Niraparib ⁵⁴	~	553	Recurrent platinum-sensitive	Not reported	21.0 months vs 5.5 months (germline	Not reported
			ovarian cancer		BRCA mutant vs placebo), 12.9 months	
					vs 3.8 months (BRCA wild/HRD positive	
					vs placebo), 9.3 months vs 3.9 months	
					(overall BRCA wild vs placebo)	

Abbreviations: PARP, poly(ADP-ribose) polymerase; ORR, overall response rate; PFS, progression-free survival; OS, overall survival; HGSOC, high-grade serous ovarian cancer; TNBC, triple-negative breast cancer; LOH, loss of heterozygosity; HRD, homologous recombination deficiency.

benefit rate (CBR) as a composite radiological and tumor marker factor and the overall response rates (ORRs) were 46% and 28%, respectively, and there was a significant association between CBR and platinum-free interval, defined as the period from last platinum administration to disease progression (platinum sensitive, 69%; platinum resistant, 45%; platinum refractory, 23%).46 In a Phase II trial in 57 patients with recurrent BRCA-mutant ovarian cancer, ORR was 33% in the 400 mg twice daily cohort and 13% in the 100 mg twice daily cohort; ie, higher dose administration of olaparib gave a better clinical outcome.⁴⁷ Kaufman et al also conducted a Phase II trial in 298 patients with recurrent BRCA-mutant cancers, including 193 ovarian cancers, 62 breast cancers, 23 pancreatic cancers and 8 prostate cancers. They reported that ORR was 26% in the entire cohort, while the values were 31%, 13%, 22% and 50% in patients with ovarian, breast, pancreatic and prostate cancers, respectively.4 Another Phase II trial in 64 patients with recurrent high-grade serous ovarian cancer (HGSOC) or TNBC showed a higher ORR in patients with BRCA-mutant ovarian cancer than in ovarian cancer patients without mutation (41% vs 24%).5 These clinical data led to accelerated approval of olaparib by the US Food and Drug Administration (FDA) in 2014 for fourth-line or later treatment of BRCA-mutant ovarian cancer. Some impressive data also support the value of olaparib maintenance treatment. Ledermann et al⁶ conducted a randomized placebo-controlled Phase II trial in 265 patients with recurrent platinum-sensitive HGSOC and reported that progression-free survival (PFS) as a primary end point was significantly longer in olaparib-receiving patients (median PFS: 8.4 months vs 4.8 months; hazard ratio [HR], 0.35; 95% confidence interval [CI], 0.25–0.49; P<0.001). Moreover, the preplanned retrospective analysis for this cohort revealed that PFS benefit was greater in BRCA-mutant ovarian cancer (median PFS: 11.2 months vs 4.3 months; HR, 0.18; 95% CI, 0.10–0.31; P < 0.0001). Based on the above results, the European Medicines Agency (EMA) approved olaparib as a maintenance therapy in 2014 for platinumsensitive relapsed BRCA-mutant HGSOC. Currently, three Phase III trials, termed SOLO1 (NCT01844986), SOLO2 (NCT01874353) and SOLO3 (NCT02282020), are in progress for BRCA-mutant ovarian cancer using the primary end point of PFS. Briefly, SOLO1 is a placebo-controlled Phase III trial in which newly diagnosed patients are randomized to olaparib maintenance (300 mg twice per day) or placebo after a platinum-based regimen. In SOLO2, the usefulness of olaparib maintenance for recurrent platinum sensitivepatients is being evaluated. SOLO3 also targets recurrent platinum-sensitive cases, comparing olaparib maintenance (300 mg twice per day) with physician's choice single-agent chemotherapy (PTX, topotecan, pegylated liposomal doxorubicin or GEM). These Phase III trials are expected to yield definite conclusions as to the clinical efficacy of olaparib.

Clinical investigations of olaparib are in progress for patients with breast and prostate cancers, as well as ovarian cancer. Tutt et al⁴⁹ conducted a Phase II trial in 54 patients with recurrent BRCA-mutant breast cancer and reported that the ORR was 41% in the 400 mg twice daily cohort and 22% in the 100 mg twice daily cohort. While ORR in BRCAmutant cases was generally similar for patients with ovarian cancer and those with breast cancer, the Phase II trial by Tutt et al⁴⁹ found no responders among BRCA-wild breast cancer patients, in contrast to an ORR of 24% in BRCA-wild ovarian cancer patients. 5 The poor sensitivity of BRCA wild-type breast cancer patients might be explained by the biological heterogeneity of breast cancer. Mateo et al35 performed a Phase II trial in 50 cases with metastatic castration-resistant prostate cancer, resulting in an ORR of 33% in the entire cohort. Among them, 16 cases with homozygous deletions, deleterious alterations or both in DNA-repair genes had an ORR of 88%, including 100% in seven patients with BRCA2 loss and 80% in five cases with ATM aberrations. Based on these clinical data, olaparib received FDA breakthrough designation in 2016 for patients with BRCA or ATM-mutant castration-resistant prostate cancer, who have progressed on prior taxane-based chemotherapy and abiraterone or enzalutamide as a new-generation hormonal agent.

Talazoparib

Talazoparib (BMN-673; Medivation, San Francisco, CA, USA) has stronger PARP-trapping activity than other PARP inhibitors. ⁵⁰ In the Phase I trial for patients with advanced solid tumor, MTD was established at 1.0 mg/day and gave ORR values of 50%, 42%, 9%, 20% and 0% in patients with *BRCA*-mutant breast cancer, *BRCA*-mutant ovarian cancer, SCLC, pancreatic cancer and Ewing sarcoma, respectively. ⁵¹ The Phase III EMBRACA trial comparing talazoparib with physician's choice is underway for patients with locally advanced and/or metastatic germline *BRCA*-mutant breast cancer (NCT01945775).

Rucaparib

In a Phase I/II study in patients with advanced solid tumors performed by Kristeleit et al,⁵² the MTD was established as 600 mg twice per day and the disease control rate (complete response [CR] + partial response [PR] + stable disease

 $[SD] \ge 24$ weeks) was 70% in germline BRCA-mutant breast cancer patients receiving rucaparib (AG014699; Clovis Oncology, Boulder, CO, USA) ≥300 mg once per day. A Phase II trial in 35 patients with recurrent BRCA-mutant ovarian cancer gave an ORR of 65% in the entire cohort.53 Swisher et al⁷ conducted the Phase II ARIEL2 trial in 206 patients with recurrent, platinum-sensitive, high-grade ovarian cancer, where patients were classified into BRCA mutant, BRCA wild type plus genomic loss of heterogenity (LOH) high and BRCA wild-type plus LOH low groups using next-generation sequencing. In this trial, the cutoff value of LOH was defined as 14%. PFS as a primary end point was significantly longer in the BRCA-mutant group and the LOH high/BRCA wild group compared with the LOH low/BRCA wild group (HR, 0.27; 95% CI, 0.16–0.44; *P*<0.0001, 0.62; 95% CI, 0.42–0.90; *P*=0.011, respectively). Their study is clinically useful in demonstrating that genomic LOH, as well as BRCA-mutant status, is helpful for identifying rucaparib responders. Based on these trial data, rucaparib obtained FDA breakthrough designation in 2015 for BRCA-mutant ovarian cancer in third-line or later treatment. The Phase III ARIELE3 trial (NCT01968213) is currently in progress for patients with platinum-sensitive ovarian cancer, in which rucaparib maintenance is being compared with placebo with PFS as the primary end point.

Veliparib

Veliparib (ABT-888; AbbVie, North Chicago, IL, USA) is an oral inhibitor possessing relatively weak PARP-trapping activity. ⁵⁰ In a Phase I trial in patients with *BRCA*-mutant cancer, *BRCA*-wild type platinum-refractory ovarian cancer or basal-like breast cancer, MTD was established at 400 mg twice per day and the ORR values were 23% in *BRCA*-mutant and 4% in *BRCA* wild-type patients. ⁵⁴ In a Phase II trial by Coleman et al, ⁵⁵ 50 patients with persistent or recurrent *BRCA*-mutant epithelial ovarian, fallopian tube or primary peritoneal cancer were enrolled to receive veliparib 400 mg twice per day; ORR was 26% in the entire cohort, 20% in platinum-resistant patients and 35% in platinum-sensitive cases.

Niraparib

In the Phase I trial in patients with advanced solid tumor, 300 mg once per day was established as the MTD.⁵⁶ Mirza et al conducted the randomized Phase III NOVA trial for a maintenance situation in 553 recurrent platinum-sensitive ovarian cancer patients, who were divided into two groups based on germline *BRCA* mutation and HRR deficiency (HRD) status using BRACAnalysis and myChoice HRD test

(Myriad Genetics, Salt Lake City, UT, USA), respectively. Niraparib (MK4827; Tesaro, Waltham, MA, USA) maintenance (300 mg once per day) was compared with placebo with PFS as the primary end point.⁸ Remarkably, patients receiving niraparib had a longer PFS, irrespective of germline BRCA mutation and HRD status (BRCA-mutant cohort: median PFS 21.0 months vs 5.5 months; HR, 0.27; 95% CI, 0.17–0.41; P<0.001; non-BRCA-mutant and HRD-positive cohort: 12.9 months vs 3.8 months; HR, 0.38; 95% CI, 0.24–0.59; P<0.001 and overall non-BRCA-mutant cohort: 9.3 months vs 3.9 months; HR, 0.45; 95% CI, 0.34–0.61; P<0.001). Based on the promising results of the NOVA trial, the FDA approved niraparib for recurrent platinum-sensitive ovarian cancer regardless of BRCA mutation and HRD status in March 2017.

In summary, olaparib is currently the best-established agent for *BRCA*-mutant cases, and this agent is available for prostate cancer and ovarian cancer cases. Rucaparib is promising due to its higher ORR for *BRCA*-mutant cases, and niraparib is the first agent demonstrating PFS prolongation for *BRCA*-wild cases in a Phase III trial. Veliparib has modest activity as a single agent, and so combination therapies are expected to become the mainstream of its clinical development. Talazoparib showed strong cytotoxicity in preclinical models, and further evidence is needed in this respect. In addition to these representative agents, clinical evidence has been gradually accumulated for other PARP inhibitors, including iniparib.⁵⁷

Combination therapy with PARP inhibitor and cytotoxic or molecular-targeted agents

Combination trials of PARP inhibitors are actively underway, usually with cytotoxic agents. Veliparib is considered the most suitable for combination trials because of its modest hematopoietic toxicity.⁵⁸ Currently, the Phase III GOG PARTNERS 3005 trial (NCT02470585) is in progress for newly diagnosed ovarian cancer cases. Patients are randomized to the combination of carboplatin (CBDCA)/PTX and veliparib (150 mg twice per day) or CBDCA/PTX alone, followed by maintenance monotherapy with veliparib (400 mg twice per day) or placebo, with PFS as the primary end point.

There are also some impressive data for olaparib combinations. Oza et al⁵⁹ conducted a Phase II trial for patients with recurrent platinum-sensitive ovarian cancer, in which patients were randomized to the combination of CBDCA/PTX and olaparib (200 mg twice per day), followed by maintenance

monotherapy with olaparib (400 mg twice per day) or CBDCA/PTX alone, with PFS as the primary end point. PFS was significantly longer in the combination group (median PFS 12.2 months vs 9.6 months; HR, 0.51; 95% CI, 0.34–0.77; P=0.0012), and it was prominent in BRCAmutant cases (HR, 0.21; 95% CI, 0.08-0.55; P=0.0015). In the randomized Phase II trial by Bang et al, 60 123 patients with recurrent or metastatic gastric cancer were randomized to the combination of olaparib (200 mg twice per day) and PTX or PTX alone, followed by maintenance monotherapy with olaparib (200 mg twice per day) or placebo. While PFS prolongation (primary end point) was not clearly detected, overall survival (OS) was significantly better in the combination group (median OS: 13.1 months vs 8.3 months; HR, 0.56; 80% CI, 0.41-0.75; P=0.005), and the benefit was especially marked in cases with low ATM protein level (median OS: not reached vs 8.2 months; HR, 0.35; 80% CI, 0.22-0.56; P=0.002). Although further consideration of the discrepancy between PFS and OS is needed, this trial is noteworthy for demonstrating the impact of BRCAness phenotype in the clinical situation.

In spite of the low PARP-inhibitory activity of iniparib (BSI201, Sanofi S.A., Paris, France) as a single agent, the open-label Phase II trial by O'Shaughnessy et al⁶¹ found that combination therapy with iniparib and GEM/CBDCA improved not only CBR as a primary end point but also OS compared with GEM/CBDCA alone (56% vs 34%; P<0.01, median OS: 12.3 months vs 7.7 months; HR, 0.57; 95% CI, 0.36–0.90; P=0.01).^{62,63} However, the subsequent Phase III trial for patients with metastatic TNBC failed to demonstrate prolongation of OS (median OS: 11.8 months vs 11.1 months; HR, 0.88; 95% CI, 0.69–1.12; P=0.28), and further evaluation of the survival benefits of this agent is needed.^{63,64}

Combinations with molecular-targeted agents and cytotoxic agents are under investigation and in some cases have moved to the clinical trial phase. Angiogenesis inhibitors seem to be attractive, in that this class of agents downregulates HRR genes through constructing a hypoxic tumor environment. 65 Liu et al 66 reported a randomized Phase II trial of cediranib, which is an oral tyrosine kinase inhibitor of vascular endothelial growth factor (VEGF) receptor (VEGFR)1, VEGFR2 and VEGFR3. In their trial, 90 patients with recurrent platinumsensitive ovarian cancer were randomized to the combination group of olaparib (200 mg twice per day) and cediranib (30 mg per day) or the olaparib (400 mg twice per day) alone group. PFS was significantly longer in the combination group (median PFS: 17.7 months vs 9.0 months; HR, 0.42; 95% CI, 0.23–0.76; P=0.005), and in response to this promising result, Phase III

trials of a combination regimen of olaparib and cediranib are underway (NCT2502266, NCT02446600, ICON9). Clinical trials of combination regimens of PARP inhibitors with cytotoxic agents, molecular-targeted agents or immunotherapies that are currently in progress are summarized in Table 2.

Toxicity profiles of PARP inhibitors

The grade 3-4 toxicity profiles in patients receiving five PARP inhibitors at their MTDs are shown in Table 3. According to previous reports on the aforementioned five inhibitors, nausea and fatigue were detected in >50% of cases in common, but the percentage of grade 3-4 events was <10%. The frequency of hematological toxicity varied depending on the agent and was conspicuous in clinical trials of talazoparib and niraparib. 8,51 In a Phase III trial of niraparib, grade 3-4 thrombocytopenia, anemia and neutropenia were detected in >20% of the patients. On the other hand, in a Phase II trial of veliparib, these events were detected in <2% of the cases. Thus, the moderate hematopoietic toxicity of veliparib could be advantageous for developing combinations with cytotoxic agents.⁵⁵ Moreover, rucaparib increased aspartate transaminase (AST) and alanine aminotransferase (ALT), but these changes were asymptomatic and normalized over time during continuous administration. 7 PARP inhibitors are newcomers in the field of oncology, and detailed toxicity profiles for long-term administration of PARP inhibitors remain to be established. Clinical trial data indicate that the onset of treatment-related myelodysplastic syndrome (MDS) and acute myeloid leukemia (AML) are serious issues.⁶⁷ According to the US Package Insert for olaparib, MDS/AML was reported in 22 of 2,618 patients (0.8%), of which 17 cases were fatal, and the duration from olaparib initiation to the onset of MDS/AML ranged from <6 months to >2 years.⁶⁸ Because all patients had received previous chemotherapy with platinum agents and/or other DNA-damaging agents, it is not clear whether olaparib directly caused MDS/AML. It is possible that the combined effects of PARP inhibition and DNA damage due to cytotoxic agents or the innate defect of BRCA might promote the onset of MDS/AML. DNA-repair inhibitors such as PARP inhibitors might increase intratumor genomic mutation rates and induce evolution of metastatic or resistant clones.⁶⁹ As for other agents, MDS/AML was reported in two of 377 patients (0.5%) treated with rucaparib and seven of 751 patients (0.9%) treated with niraparib. 70,71 Therefore, clinicians should monitor each patient's complete blood count carefully and stop the inhibitor administration immediately if MDS/AML is clinically suspected. Moreover, preclinical models suggested an association of PARP1 with

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Table 2 Ongoing trials of PARP inhibitor combinations

PARP inhibitor	Combination agents Phase Types of malignancies		NCT identifier	
Olaparib	AZD2014 (mTORC 1/2 inhibitor) or	I	Breast cancer, malignant female reproductive	NCT02208375
-	AZD5363 (AKT inhibitor)		system tumors	
Olaparib	Prexasertib	1	Solid tumors	NCT03057145
Olaparib	AZD2281, AZD5363, AZD1775 or AZD2014	II	Tumors	NCT02576444
Olaparib	AT13387 (heat shock protein 90 inhibitor)	1	Ovarian cancer, fallopian tube cancer, peritoneal cancer and TNBC	NCT02898207
Olaparib	Cediranib, MEDI4736 (anti-PD-L1 antibody)	1/11	Lung cancer, breast cancer, ovarian cancer, colorectal cancer, prostate cancer and TNBC	NCT02484404
Olaparib	Temozolomide	1	Ewing sarcoma	NCT01858168
Olaparib	AZD1775 (tyrosine kinase WEE1 inhibitor)	1	Ovarian cancer, breast cancer and SCLC	NCT02511795
Olaparib	Temozolomide	1	Brain and central nervous system tumors	NCT01390571
Olaparib	CRLX101 (nanoparticle camptothecin)	1/11	SCLC	NCT02769962
Olaparib	Neoadjuvant carboplatin and paclitaxel	11/111	TNBC and germline BRCA-positive breast cancer	NCT03150576
Talazoparib	Decitabine	1/11	Acute myeloid leukemia	NCT02878785
Talazoparib	Temozolomide	1/11	Solid tumors	NCT02116777
Veliparib	VX-970 (ATR) and cisplatin	1	Tumors	NCT02723864
Veliparib	Intraperitoneal floxuridine	1	Fallopian tube cancer, ovarian cancer and primary peritoneal cancer	NCT01749397
Veliparib	Carboplatin and paclitaxel	III	BRCA-mutant breast cancer	NCT02163694
Veliparib	FOLFIRI or modified FOLFIRI	II	Pancreatic cancer	NCT02890355
Veliparib	Carboplatin and paclitaxel	III	Ovarian cancer, fallopian tube cancer and peritoneal cancer	NCT02470585
Veliparib	Topotecan hydrochloride	1/11	Solid tumors	NCT01012817
Niraparib	Temozolomide or irinotecan	1	Ewing sarcoma	NCT02044120
Niraparib	Pembrolizumab	1/11	Ovarian cancer, fallopian tube cancer, NCT026579 peritoneal cancer and TNBC	
Fluzoparib	Apatinib and paclitaxel	1	Gastric cancer	NCT03026881
Fluzoparib	Apatinib	1	Ovarian cancer and TNBC	NCT03075462
BGB-290	BGB-A317 (anti-PD-1 monoclonal antibody)	1	Solid tumors	NCT02660034

Abbreviations: FOLFIRI, folinic acid/fluorouracil/ irinotecan; NCT, National Clinical Trial; PD-1, programmed death-1; PD-L1, programmed death-ligand1; TNBC, triplenegative breast cancer; SCLC, small cell lung cancer.

cardiovascular diseases and long-term memory formation, and long-term observation is required to examine the effects on the cardiovascular system and on mental health.31

Identification of biomarkers predicting suitable cases

Identification of responders to PARP inhibitors is important for providing maximum clinical benefit to suitable cases and for preventing development of drug resistance and is also desirable from a cost-benefit point of view. However, there are no established strategies for identifying relevant candidates, although combinations of various kinds of biomarkers with clinicopathological features have been investigated with a focus on detecting HRR-deficient cases.³³ According to the Phase I trial conducted by Fong et al,46 platinum-free interval was significantly associated with CBR for olaparib

Table 3 Grade 3-4 toxicity profiles of PARP inhibitors

Grade 3-4	Olaparib	Talazoparib	Rucaparib	Veliparib	Niraparib
adverse events	(n=298) ⁴²	(n=71) ⁴⁷	(n=204) ⁵⁰	(n=50) ⁵²	(n=367) ⁵⁴
Nausea	0%	0%	4%	4%	3%
Vomiting	2%	Not reported	2%	0%	2%
Fatigue	6%	Not reported	9%	Not reported	8%
Abdominal pain	6%	Not reported	2%	Not reported	1%
Thrombocytopenia	Not reported	18%	2%	2%	34%
Anemia	17%	23%	22%	0%	25%
Neutropenia	Not reported	10%	7%	2%	20%
Hypertension	Not reported	Not reported	Not reported	Not reported	8%
Increased AST/ALT	Not reported	Not reported	12%	Not reported	Not reporte

Abbreviations: AST, aspartate transaminase; ALT, alanine aminotransferase; PARP, poly(ADP-ribose) polymerase.

administration, and Cochrane's review of 599 patients with epithelial ovarian cancer noted that PARP inhibitors appeared to improve PFS in recurrent platinum-sensitive disease, whereas clinical significance for platinum-resistant disease was undetermined.⁷² These data highlight platinum sensitivity as a clinical surrogate marker predicting PARP inhibitor's responsiveness. Assays for the HRR pathway include targeted sequencing, whole-genome sequencing, copy number analyses, gene-expression profiling, protein expression assays and functional assays.⁶⁹

Genomic analysis is a direct and comprehensive approach for predicting suitable cases and has already been embedded in clinical trials, but it is relatively expensive for routine use in many institutions and requires frozen or formalin-fixed paraffin-embedded (FFPE) tumor specimens for analysis, as well as bioinformatic support from experts.³³ In addition, this approach is often affected by different analytical approaches and genomic heterogeneity among tumor specimens; also, genomic alterations do not necessarily cause functional changes in proteins. 73,74 Genomic scars (accumulated patterns of DNA damage and repair) are unique in HRR-deficient cases, and these characteristic signatures are caused by large genomic deletions and LOH owing to the activation of inaccurate NHEJ. Telli et al⁷⁵ defined an HRD score composed of LOH, telomeric allelic imbalance and large-scale state transitions and reported that this index predicted response to neoadjuvant platinum-based regimen in cases with TNBC, regardless of BRCA-mutant status. In a prospective trial setting, two indexes (LOH and HRD score) were adopted for the ARIEL2 trial of rucaparib and the NOVA trial of niraparib, confirming the position of HRD score as an independent prognostic factor.^{7,8} Previous studies indicate that the total number of exome alterations, including nonsynonymous ones, was also associated with clinical outcomes in BRCA-mutant ovarian cancers.⁶⁹

In terms of functional biomarkers, the detection of RAD51 foci with immunofluorescence is useful because of its rapidity. While administration of PARP inhibitors increases expression of RAD51 foci in HRR-proficient tumors, RAD51 foci remain absent in HRR-deficient tumors, indicating a role of RAD51 as a key player in the HRR signaling pathway. 12,76 One weakness of this method is that we cannot predict the change in RAD51 foci prior to actual drug exposure. 12 PARP1 is an essential substrate for PARylation, and immunohistochemical analysis by von Minckwitz et al. 177 showed that high cytoplasmic PARP expression predicted high sensitivity to chemotherapy in breast cancer samples. Moreover, elevated PAR levels detected by Western blot or immunohistochemistry or E26 transformation-specific

sequence (ETS) gene fusions observed in Ewing's sarcoma and prostate cancer are promising biomarkers. ^{78,79} However, the usefulness of the aforementioned factors for the identification of PARP inhibitor responders has mainly been demonstrated in preclinical models, and their clinical relevance needs to be properly assessed in clinical trials.

Mechanisms of clinical resistance

Development of drug resistance is a major problem in patients receiving the same treatment regimen for a long period. Resistance to PARP inhibitors can appear at many steps of the DNA-repair pathways. A conceptual diagram of resistance mechanisms, classified into increased HRR capacity, decreased activity of PARP-1 and decreased intracellular availability of PARP inhibitor, is shown in Figure 1.80

Recovery of HR is an important mechanism. Edwards et al⁸¹ examined BRCA2 c.6174delT mutation of a BRCA2deficient human pancreatic cancer cell line (CAPAN1) and reported that this frameshift mutation resulted in normalization of the functions of BRCA2 protein. Sakai et al⁷³ established seven subclones with restored BRCA2 protein and seven subclones without BRCA2 protein from CAPAN1 and found that the sensitivity to PARP inhibitors was lower in clones with secondary BRCA2 mutation than in those without BRCA2 mutation. Barber et al82 compared genomic information for primary and metastatic samples from a male breast tumor patient developing resistance to olaparib and detected a secondary BRCA2 c.9106C > G mutation leading to the functional restoration of BRCA2 protein. In the context of cisplatin resistance, Swisher et al⁸³ compared recurrent platinum-resistant and platinum-sensitive tumors and reported that four of six in the former group developed secondary genetic changes in BRCA1 leading to functional BRCA1 protein restoration, whereas none of three in the latter group developed BRCA1 sequence alterations. Norquist et al⁸⁴ reviewed 46 recurrent ovarian cancer patients and reported that the percentage of secondary mutations restoring BRCA1/ BRCA2 was significantly higher in platinum-resistant cases than in platinum-sensitive ones (46% vs 5%, P=0.003). Dhillon et al⁸⁵ proposed an interesting model of secondary BRCA restoration from a primary chemotherapy-sensitive tumor. First, BRCA-restored subclones coincidentally arise from initially BRCA-deficient or chemotherapy-sensitive tumor as a result of increased mutation rates associated with DNA-damaging agents. Next, these clones spread through selective pressure from repeated drug administrations. This theory might reasonably explain the induction of PARP inhibitor resistance during repeated cisplatin administrations.

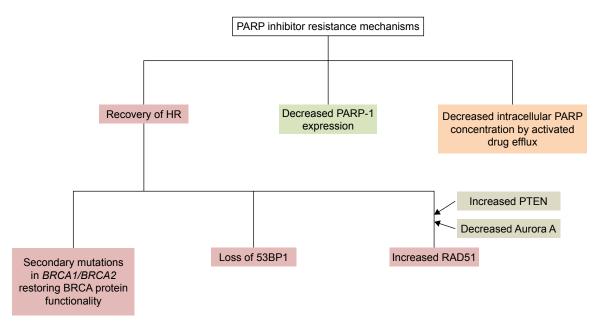


Figure I Conceptual diagram of resistance mechanisms to PARP inhibitors.

Notes: These processes are categorized into three groups (recovery of HRR, decreased PARP-I expression, and decreased intracellular PARP concentration due to activation of drug efflux). Secondary mutations in BRCA1/BRCA2, loss of 53BPI and increased RAD51 finally result in recovery of HRR, where PTEN or Aurora A indirectly regulate RAD51 activity.

Abbreviations: PARP, poly(ADP-ribose) polymerase; HRR, homologous recombination repair.

As for other HRR-related factors, P53-binding protein 1 (53BP1) plays a key role in DNA-repair responses and checkpoint control, and loss of 53BP1 promotes ATM-dependent resection of broken DNA ends in BRCA1-deficient cells and finally restores HRR. 86,87 An association between increased RAD51 activity and resistance to PARP inhibitors is also indicated, and Aurora A and PTEN modulate RAD51 activity indirectly. 80,88,89 Aurora A inhibits the recruitment of RAD51 to DNA damage sites, while PTEN deficiency causes a reduction in RAD51 expression. RAD51 forms a complex with BRCA2, and its combination with damaged DSB is indispensable to HRR, so that increased RAD51 activity leads to resistance to PARP inhibitors. 90 Kondrashova et al 91 recently sequenced HRR pathway genes in PARP inhibitor pretreatment and post-progression tumor samples from patients enrolled in the ARIEL2 trial and detected secondary mutations in RAD51C and RAD51D associated with PARP inhibitor resistance.

P-Glycoprotein (P-gp; multi-drug resistance protein) acts as a drug efflux pump to decrease intracellular drug concentration and is associated with resistance to various kinds of agents. An in vivo model by Rottenberg et al92 showed upregulation of the P-gp-encoding ABCB1A/B genes after long-term PARP administration. In addition, preclinical studies suggest other possibilities, including regulation by micro RNA, epigenetic re-expression of BRCA1, phosphorylation of PARP1 by c-Met and overexpression of HOX family members. 93-97

Large-scale genomic data also revealed the genomic backgrounds of refractory or resistant cases. Patch et al⁹⁸ performed whole-genome sequencing of 92 ovarian cancer patients with primary refractory, resistant, sensitive and matched acquired resistant disease and identified several molecular events associated with acquired resistance, including reverse germline BRCA1/BRCA2 mutation, loss of BRCA1 promoter methylation and ABCB1 rearrangement. The practical relevance of these mechanisms should be considered in future clinical trials.

Conclusion and prospects for the

The concept of synthetic lethality led to great interest in PARP inhibitors as candidate antitumor agents, and clinical applications of this class of agents have so far focused mainly on BRCA-mutant ovarian cancer. Olaparib, rucaparib and niraparib are already available under US health insurance. Detailed assays for HRR repair pathway components, including genomic analysis, are broadening the applicability of PARP inhibitors in HRR-deficient cases, and future investigations are expected to focus on HRR-proficient cases and various kinds of malignancies other than ovarian, prostate and breast cancers (Figure 2). Considering the variety of patients who may receive clinical benefit from PARP inhibitors, we consider that intergroup studies will be essential to improve and extend current clinical outcomes.

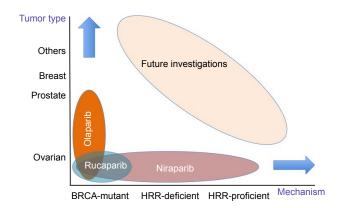


Figure 2 Current indications for olaparib, rucaparib, and niraparib. **Notes:** The differences among these agents, which are available under US health insurance, probably reflect the characteristics of patients who participated in the

relevant clinical trials. Future investigations should be focused on HRR-proficient cases and malignancies other than ovarian, prostate and breast cancers.

Abbreviation: HRR, homologous recombination repair.

In addition to the need for well-established biomarkers to predict suitable cases and resistance mechanisms, there are several areas where further work is needed to realize the full potential of PARP inhibitors (Figure 3). First, although clinical evidence for each agent has been accumulating gradually, direct comparative data of the available agents are lacking, and we cannot draw a clear conclusion as to which agent is the best option in each clinical context.

In this sense, further accumulation of preclinical data and further randomized clinical trials are essential to establish the status of each agent. Second, the FDA approved the first PARP inhibitor, olaparib, in 2014, so that only about 3 years have passed since marketing. Therefore, potential long-term toxicities, including secondary malignancies, are not clear, and further follow-up data in clinical trials and postmarketing surveillance are indispensable to uncover the toxicity profile of this agent. Third, clinical effects on HRR-proficient tumors are not evident, except for niraparib. In addition, it would be desirable to discover new cascades other than the HRR pathway to provide additional indications for PARP inhibitors. Fourth, clinical trials so far performed have not demonstrated OS prolongation, in spite of PFS benefit. The current gold standard for assessment of efficacy of new agents is a statistically significant and clinically meaningful improvement in OS and quality of life (QOL). The surrogacy of PFS for OS is established only in advanced colorectal and ovarian cancers, and the clinical meaning of PFS in other types of tumors is not yet clear.99 A longer follow-up period is required for assessment of OS, and at the same time, development of powerful combination regimens is expected to bring better clinical benefits. Interestingly, the olaparib trial data showed that high-dose olaparib gave a higher ORR, and

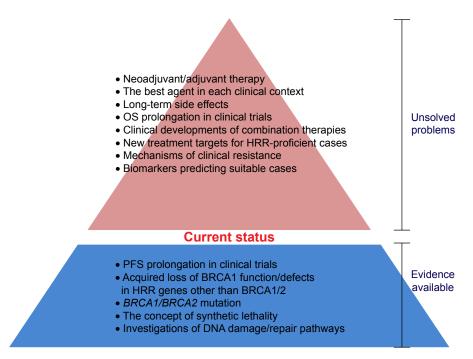


Figure 3 Current status of PARP inhibitors.

Notes: Recent advances in preclinical models have provided fundamental data on synthetic lethality and HRR pathways, and clinical trials have shown PFS prolongation with a focus on *BRCA*-mutant tumors. On the other hand, several issues remain, such as discovering new treatment targets for HRR-proficient cases or selecting the best agent in each clinical context. In the future, it will be important to demonstrate OS prolongation as a real clinical end point through development of PARP inhibitor combination regimens or neoadjuvant/adjuvant therapies.

Abbreviations: PARP, poly(ADP-ribose) polymerase; HRR, homologous recombination repair; PFS, progression-free survival; OS, overall survival.

the combination of high-dose olaparib with relatively lowdose cytotoxic agents seems to be an appropriate approach to concurrently achieve efficacy and tolerance. 42,47,49 However, clinical benefits need to be comprehensively evaluated from the viewpoints of QOL and cost-effectiveness, as well as OS. Finally, further accumulation of evidence about PARP inhibitors may lead to expanded adoption for neoadjuvant or adjuvant therapy in operable patients or preventive administration in BRCA mutation carriers in the future. Based on favorable trial data for pancreatic cancer, clinical development for intractable cancers, including this disease, might be promising. 4,51 PARP inhibitors are considered advantageous for these adaptations in that the target population is small and well-defined and the risk is unlikely to outweigh the clinical benefit, since the complications resulting from the use of this class of agents are mild.100

Disclosure

The authors report no conflicts of interest in this work.

References

- Bryant HE, Schultz N, Thomas HD, et al. Specific killing of BRCA2deficient tumours with inhibitors of poly (ADP-ribose) polymerase. *Nature*. 2005;434(7035):913–917.
- Farmer H, McCabe N, Lord CJ, et al. Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. *Nature*. 2005; 434(7035):917–921.
- 3. del Rivero J, Kohn EC. PARP Inhibitors: the cornerstone of DNA repair-targeted therapies. *Oncology (Williston Park)*. 2017;31(4):265–273.
- Kaufman B, Shapira-Frommer R, Schmutzler RK, et al. Olaparib monotherapy in patients with advanced cancer and a germline BRCA1/2 mutation. *J Clin Oncol*. 2015;33(3):244–250.
- Gelmon KA, Tischkowitz M, Mackay H, et al. Olaparib in patients with recurrent high-grade serous or poorly differentiated ovarian carcinoma or triple-negative breast cancer: a phase 2, multicentre, open-label, non-randomised study. *Lancet Oncol.* 2011;12(9):852–861.
- Ledermann J, Harter P, Gourley C, et al. Olaparib maintenance therapy in platinum-sensitive relapsed ovarian cancer. N Engl J Med. 2012;366(15):1382–1392.
- Swisher EM, Lin KK, Oza AM, et al. Rucaparib in relapsed, platinumsensitive high-grade ovarian carcinoma (ARIEL2 Part 1): an international, multicentre, open-label, phase 2 trial. *Lancet Oncol.* 2017; 18(1):75–87.
- Mirza MR, Monk BJ, Herrstedt J, et al. Niraparib maintenance therapy in platinum-sensitive, recurrent ovarian cancer. N Engl J Med. 2016; 375(22):2154–2164.
- Lin KY, Kraus WL. PARP inhibitors for cancer therapy. Cell. 2017; 169(2):183.
- Bolton KL, Chenevix-Trench G, Goh C, et al. Association between BRCA1 and BRCA2 mutations and survival in women with invasive epithelial ovarian cancer. *JAMA*. 2012;307(4):382–390.
- Gonzalez-Angulo AM, Timms KM, Liu S, et al. Incidence and outcome of BRCA mutations in unselected patients with triple receptor-negative breast cancer. Clin Cancer Res. 2011;17(5):1082–1089.
- Lord CJ, Ashworth A. BRCAness revisited. Nat Rev Cancer. 2016; 16(2):110–120.
- McCabe N, Turner NC, Lord CJ, et al. Deficiency in the repair of DNA damage by homologous recombination and sensitivity to poly (ADPribose) polymerase inhibition. *Cancer Res.* 2006;66(16):8109–8115.

- Chambon P, Weill JD, Mandel P. Nicotinamide mononucleotide activation of new DNA-dependent polyadenylic acid synthesizing nuclear enzyme. *Biochem Biophys Res Commun.* 1963;11:39–43.
- Ashworth A. A synthetic lethal therapeutic approach: poly (ADP) ribose polymerase inhibitors for the treatment of cancers deficient in DNA double-strand break repair. *J Clin Oncol.* 2008;26(22):3785–3790.
- Brown JS, O'Carrigan B, Jackson SP, Yap TA. Targeting DNA repair in cancer: beyond PARP inhibitors. *Cancer Discov*. 2017;7:20–37.
- Amé JC, Rolli V, Schreiber V, et al. PARP-2, a novel mammalian DNA damage-dependent poly(ADP-ribose) polymerase. *J Biol Chem*. 1999;274(25):17860–17868.
- 18. McLornan DP, List A, Mufti GJ. Applying synthetic lethality for the selective targeting of cancer. *N Engl J Med*. 2014;371:1725–1735.
- Schreiber V, Dantzer F, Ame JC, de Murcia G. Poly (ADP-ribose): novel functions for an old molecule. *Nat Rev Mol Cell Biol*. 2006;7(7): 517–528.
- Lord CJ, Ashworth A. The DNA damage response and cancer therapy. Nature. 2012;481:287–294.
- Papa A, Caruso D, Strudel M, Tomao S, Tomao F. Update on Poly-ADPribose polymerase inhibition for ovarian cancer treatment. *J Transl Med*. 2016;14:267.
- Murai J, Huang SY, Das BB, et al. Trapping of PARP1 and PARP2 by clinical PARP inhibitors. *Cancer Res.* 2012;72(21):5588–5599.
- Plummer ER, Calvert H. Targeting poly (ADP-ribose) polymerase: a two-armed strategy for cancer therapy. *Clin Cancer Res*. 2007;13(21): 6252–6256.
- Lord CJ, Ashworth A. PARP inhibitors: synthetic lethality in the clinic. Science. 2017;355(6330):1152–1158.
- Annunziata CM, O'Shaughnessy J. Poly (ADP-ribose) polymerase as a novel therapeutic target in cancer. *Clin Cancer Res.* 2010;16(18): 4517–4526.
- O'Sullivan CC, Moon DH, Kohn EC, Lee JM. Beyond breast and ovarian cancers: PARP inhibitors for BRCA mutation-associated and BRCA-like solid tumors. Front Oncol. 2014;4:42.
- Shrivastav M, De Haro LP, Nickoloff JA. Regulation of DNA doublestrand break repair pathway choice. *Cell Res.* 2008;18(1):134–147.
- Benafif S, Hall M. An update on PARP inhibitors for the treatment of cancer. Onco Targets Ther. 2015;8:519–528.
- Byers LA, Wang J, Nilsson MB, et al. Proteomic profiling identifies dysregulated pathways in small cell lung cancer and novel therapeutic targets including PARP1. *Cancer Discov.* 2012;2(9):798–811.
- Ratnam K, Low JA. Current development of clinical inhibitors of poly (ADP-ribose) polymerase in oncology. *Clin Cancer Res.* 2007; 13(5):1383–1388.
- Rouleau M, Patel A, Hendzel MJ, Kaufmann SH, Poirier GG. PARP inhibition: PARP1 and beyond. *Nat Rev Cancer*. 2010;10(4): 293–301.
- 32. Hughes-Davies L, Huntsman D, Ruas M, et al. EMSY links the BRCA2 pathway to sporadic breast and ovarian cancer. *Cell.* 2003;115(5): 523–535.
- Lim D, Ngeow J. Evaluation of the methods to identify patients who may benefit from PARP inhibitor use. *Endocr Relat Cancer*. 2016; 23(6):R267–R285.
- 34. Pennington KP, Walsh T, Harrell MI, et al. Germline and somatic mutations in homologous recombination genes predict platinum response and survival in ovarian, fallopian tube, and peritoneal carcinomas. *Clin Cancer Res.* 2014;20(3):764–775.
- Mateo J, Carreira S, Sandhu S, et al. DNA-repair defects and olaparib in metastatic prostate cancer. N Engl J Med. 2015;373:1697–1708.
- The Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. *Nature*. 2011;474(7353):609–615.
- Pommier Y, O'Connor MJ, de Bono J. Laying a trap to kill cancer cells: PARP inhibitors and their mechanisms of action. *Sci Transl Med*. 2016;8:362s17.
- Postel-Vinay S, Vanhecke E, Olaussen KA, Lord CJ, Ashworth A, Soria JC. The potential of exploiting DNA-repair defects for optimizing lung cancer treatment. *Nat Rev Clin Oncol*. 2012;9(3):144–155.

 Matulonis UA, Monk BJ. PARP inhibitor and chemotherapy combination trials for the treatment of advanced malignancies: does a development pathway forward exist? *Ann Oncol*. 2017;28:443–447.

- Murai J, Zhang Y, Morris J, et al. Rationale for poly (ADP-ribose) polymerase (PARP) inhibitors in combination therapy with camptothecins or temozolomide based on PARP trapping versus catalytic inhibition. *J Pharmacol Exp Ther*. 2014;349(3):408–416.
- Ihnen M, zu Eulenburg C, Kolarova T, et al. Therapeutic potential of the poly (ADP-ribose) polymerase inhibitor rucaparib for the treatment of sporadic human ovarian cancer. *Mol Cancer Ther*. 2013;12(6): 1002–1015.
- Dréan A, Lord CJ, Ashworth A. PARP inhibitor combination therapy. Crit Rev Oncol Hematol. 2016;108:73–85.
- Lu Y, Kwintkiewicz J, Liu Y, et al. Chemosensitivity of IDH1-mutated gliomas due to an impairment in PARP1-mediated DNA repair. *Cancer Res.* 2017;77(7):1709–1718.
- 44. Tateishi K, Higuchi F, Miller JJ, et al. The alkylating chemotherapeutic temozolomide induces metabolic stress in IDH1-mutant cancers and potentiates NAD+ depletion-mediated cytotoxicity. *Cancer Res.* 2017;77(15):4102–4115.
- Fong PC, Boss DS, Yap TA, et al. Inhibition of poly (ADP-ribose) polymerase in tumors from BRCA mutation carriers. N Engl J Med. 2009;361(2):123–134.
- Fong PC, Yap TA, Boss DS, et al. Poly (ADP)-ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinum-free interval. *J Clin Oncol*. 2010;28(15): 2512–2519
- Audeh MW, Carmichael J, Penson RT, et al. Oral poly (ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet*. 2010;376(9737):245–251.
- 48. Ledermann J, Harter P, Gourley C, et al. Olaparib maintenance therapy in patients with platinum-sensitive relapsed serous ovarian cancer: a preplanned retrospective analysis of outcomes by BRCA status in a randomised phase 2 trial. *Lancet Oncol.* 2014;15:852–861.
- Tutt A, Robson M, Garber JE, et al. Oral poly (ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and advanced breast cancer: a proof-of-concept trial. *Lancet*. 2010; 376(9737):235–244.
- Hopkins TA, Shi Y, Rodriguez LE, et al. Mechanistic dissection of PARP1 trapping and the impact on in vivo tolerability and efficacy of PARP inhibitors. *Mol Cancer Res.* 2015;13(11):1465–1477.
- de Bono J, Ramanathan RK, Mina L, et al. Phase I, dose-escalation, twopart trial of the PARP inhibitor talazoparib in patients with advanced germline BRCA1/2 mutations and selected sporadic cancers. *Cancer Discov.* 2017;7(6):620–629.
- Kristeleit RS, Burris HA, LoRusso P, et al. Phase 1/2 study of oral rucaparib: final phase 1 results. J Clin Oncol. 2014;32(suppl):abstr2573.
- 53. Shapira-Frommer R, Oza AM, Domchek SM, et al. A phase II openlabel, multicenter study of single-agent rucaparib in the treatment of patients with relapsed ovarian cancer and a deleterious BRCA mutation. *J Clin Oncol*. 2015;33(suppl):abstr5513.
- 54. Puhalla S, Beumer JH, Pahuja S, et al. Final results of a phase 1 study of single-agent veliparib (V) in patients (pts) with either BRCA1/2-mutated cancer (BRCA+), platinum-refractory ovarian, or basal-like breast cancer (BRCA-wt). *J Clin Oncol*. 2014;32(suppl): abstr2570.
- 55. Coleman RL, Sill MW, Bell-McGuinn K, et al. A phase II evaluation of the potent, highly selective PARP inhibitor veliparib in the treatment of persistent or recurrent epithelial ovarian, fallopian tube, or primary peritoneal cancer in patients who carry a germline BRCA1 or BRCA2 mutation an NRG Oncology/Gynecologic Oncology Group study. *Gynecol Oncol.* 2015;137(3):386–391.
- Sandhu SK, Schelman WR, Wilding G, et al. The poly (ADP-ribose) polymerase inhibitor niraparib (MK4827) in BRCA mutation carriers and patients with sporadic cancer: a phase 1 dose-escalation trial. *Lancet Oncol*. 2013;14(9):882–892.

- Bell-McGuinn KM, Konner JA, Tew WP, et al. A phase 2, single arm study of iniparib in patients with BRCA1 or BRCA2 associated advanced epithelial ovarian, fallopian tube, or primary peritoneal cancer. *Int J Gynecol Cancer*. 2016;26(2):255–260.
- Wagner LM. Profile of veliparib and its potential in the treatment of solid tumors. Onco Targets Ther. 2015;8:1931–1939.
- Oza AM, Cibula D, Benzaquen AO, et al. Olaparib combined with chemotherapy for recurrent platinum-sensitive ovarian cancer: a randomised phase 2 trial. *Lancet Oncol.* 2015;16:87–97.
- 60. Bang YJ, Im SA, Lee KW, et al. Randomized, double-blind phase II trial with prospective classification by ATM protein level to evaluate the efficacy and tolerability of olaparib plus paclitaxel in patients with recurrent or metastatic gastric cancer. *J Clin Oncol.* 2015;33:3858–3865.
- O'Shaughnessy J, Osborne C, Pippen JE, et al. Iniparib plus chemotherapy in metastatic triple-negative breast cancer. N Engl J Med. 2011;364(3):205–214.
- Licht S, Cao H, Li Z, et al. Mechanism of action of iniparib: stimulation of reactive oxygen species (ROS) production in an iniparib-sensitive breast cancer cell line. *Mol Cancer Ther*. 2011;10(supl 11):abstrA226.
- Guha M. PARP inhibitors stumble in breast cancer. *Nat Biotechnol*. 2011;29:373–374.
- 64. O'Shaughnessy J, Schwartzberg L, Danso MA, et al. Phase III study of iniparib plus gemcitabine and carboplatin versus gemcitabine and carboplatin in patients with metastatic triple-negative breast cancer. *J Clin Oncol*. 2014;32(34):3840–3847.
- Chan N, Bristow RG. "Contextual" synthetic lethality and/or loss of heterozygosity: tumor hypoxia and modification of DNA repair. *Clin Cancer Res*. 2010;16:4553

 –4560.
- Liu JF, Barry WT, Birrer M, et al. Combination cediranib and olaparib versus olaparib alone for women with recurrent platinum-sensitive ovarian cancer: a randomised phase 2 study. *Lancet Oncol*. 2014;15(11): 1207–1214.
- O'Sullivan Coyne G, Chen AP, Meehan R, Doroshow JH. PARP inhibitors in reproductive system cancers: current use and developments. *Drugs*. 2017;77(2):113–130.
- LYNPARZATM (olaparib) capsules, for oral use [prescribing information approved by the US FDA]. Available from: https://www.accessdata.fda. gov/drugsatfda_docs/label/2014/206162lbl.pdf. Accessed August 22, 2017
- Stover EH, Konstantinopoulos PA, Matulonis UA, Swisher EM. Biomarkers of response and resistance to DNA repair targeted therapies. *Clin Cancer Res.* 2016;22(23):5651–5660.
- RUBRACA™ (rucaparib) tablets, for oral use [prescribing information approved by the US FDA]. Available from: https://www.accessdata. fda.gov/drugsatfda_docs/label/2016/209115s000lbl.pdf. Accessed August 22, 2017.
- ZEJULATM (niraparib) capsules, for oral use [prescribing information approved by the US FDA]. Available from: https://www.accessdata.fda. gov/drugsatfda_docs/label/2017/208447lbl.pdf. Accessed August 22, 2017.
- Wiggans AJ, Cass GK, Bryant A, et al. Poly(ADP-ribose) polymerase (PARP) inhibitors for the treatment of ovarian cancer. *Cochrane Database Syst Rev.* 2015;5:CD007929.
- Sakai W, Swisher EM, Karlan BY, et al. Secondary mutations as a mechanism of cisplatin resistance in BRCA2-mutated cancers. *Nature*. 2008;451(7182):1116–1120.
- Yachida S, Jones S, Bozic I, et al. Distant metastasis occurs late during the genetic evolution of pancreatic cancer. *Nature*. 2010;467(7319): 1114–1117.
- Telli ML, Timms KM, Reid J, et al. Homologous recombination deficiency (HRD) score predicts response to platinum-containing neoadjuvant chemotherapy in patients with triple-negative breast cancer. Clin Cancer Res. 2016;22(15):3764–3773.
- Graeser M, McCarthy A, Lord CJ, et al. A marker of homologous recombination predicts pathologic complete response to neoadjuvant chemotherapy in primary breast cancer. *Clin Cancer Res.* 2010;16(24): 6159–6168.

- 77. von Minckwitz G, Müller BM, Loibl S, et al. Cytoplasmic poly(adenosine diphosphate-ribose) polymerase expression is predictive and prognostic in patients with breast cancer treated with neoadjuvant chemotherapy. J Clin Oncol. 2011;29(16):2150-2157.
- 78. Brenner JC, Feng FY, Han S, et al. PARP-1 inhibition as a targeted strategy to treat Ewing's sarcoma. Cancer Res. 2012;72(7):1608–1613.
- 79. Michels J, Vitale I, Galluzzi L, et al. Cisplatin resistance associated with PARP hyperactivation. Cancer Res. 2013;73(7):2271-2280.
- 80. Montoni A, Robu M, Pouliot E, Shah GM. Resistance to PARPinhibitors in cancer therapy. Front Pharmacol. 2013;4:18.
- 81. Edwards SL, Brough R, Lord CJ, et al. Resistance to therapy caused by intragenic deletion in BRCA2. Nature. 2008;451(7182):1111-1115.
- 82. Barber LJ, Sandhu S, Chen L, et al. Secondary mutations in BRCA2 associated with clinical resistance to a PARP inhibitor. J Pathol. 2013; 229(3):422-429.
- 83. Swisher EM, Sakai W, Karlan BY, Wurz K, Urban N, Taniguchi T. Secondary BRCA1 mutations in BRCA1-mutated ovarian carcinomas with platinum resistance. Cancer Res. 2008;68(8):2581-2586.
- 84. Norquist B, Wurz KA, Pennil CC, et al. Secondary somatic mutations restoring BRCA1/2 predict chemotherapy resistance in hereditary ovarian carcinomas. J Clin Oncol. 2011;29(22):3008-3015.
- 85. Dhillon KK, Swisher EM, Taniguchi T. Secondary mutations of BRCA1/2 and drug resistance. Cancer Sci. 2011;102:663-669.
- 86. Bunting SF, Callén E, Wong N, et al. 53BP1 inhibits homologous recombination in Brca1-deficient cells by blocking resection of DNA breaks. Cell. 2010;141(2):243-254.
- 87. Bouwman P, Aly A, Escandell JM, et al. 53BP1 loss rescues BRCA1 deficiency and is associated with triple-negative and BRCA-mutated breast cancers. Nat Struct Mol Biol. 2010;17(6):688-695.
- 88. Sourisseau T, Maniotis D, McCarthy A, et al. Aurora-A expressing tumour cells are deficient for homology-directed DNA double strandbreak repair and sensitive to PARP inhibition. EMBO Mol Med. 2010; 2(4):130-142.
- 89. Mendes-Pereira AM, Martin SA, Brough R, et al. Synthetic lethal targeting of PTEN mutant cells with PARP inhibitors. EMBO Mol Med. 2009;1(6-7):315-322.

- 90. Liu X, Han EK, Anderson M, et al. Acquired resistance to combination treatment with temozolomide and ABT-888 is mediated by both base excision repair and homologous recombination DNA repair pathways. Mol Cancer Res. 2009;7(10):1686-1692.
- 91. Kondrashova O, Nguyen M, Shield-Artin K, et al. Secondary somatic mutations restoring RAD51C and RAD51D associated with acquired resistance to the PARP inhibitor rucaparib in high-grade ovarian carcinoma. Cancer Discov. 2017;7(9):984-998.
- 92. Rottenberg S, Jaspers JE, Kersbergen A, et al. High sensitivity of BRCA1-deficient mammary tumors to the PARP inhibitor AZD2281 alone and in combination with platinum drugs. Proc Natl Acad Sci USA. 2008;105(44):17079-17084.
- 93. Kim Y, Kim A, Sharip A, et al. Reverse the resistance to PARP inhibitors. Int J Biol Sci. 2017;13:198-208.
- 94. Ter Brugge P, Kristel P, van der Burg E, et al. Mechanisms of therapy resistance in patient-derived xenograft models of BRCA1-deficient breast cancer. J Natl Cancer Inst. 2016;108(11):djw148.
- 95. Choi YE, Meghani K, Brault ME, et al. Platinum and PARP inhibitor resistance due to overexpression of microRNA-622 in BRCA1-mutant ovarian cancer. Cell Rep. 2016;14(3):429-439.
- 96. Du Y, Yamaguchi H, Wei Y, et al. Blocking c-Met-mediated PARP1 phosphorylation enhances anti-tumor effects of PARP inhibitors. Nat Med. 2016;22:194-201.
- 97. Esposito MT, Zhao L, Fung TK, et al. Synthetic lethal targeting of oncogenic transcription factors in acute leukemia by PARP inhibitors. Nat Med. 2015;21(12):1481-1490.
- 98. Patch AM, Christie EL, Etemadmoghadam D, et al. Whole-genome characterization of chemoresistant ovarian cancer. Nature. 2015; 521(7553):489-494.
- 99. Booth CM, Eisenhauer EA. Progression-free survival: meaningful or simply measurable? J Clin Oncol. 2012;30(10):1030-1033.
- 100. Sonnenblick A, de Azambuja E, Azim HA Jr, Piccart M. An update on PARP inhibitors - moving to the adjuvant setting. Nat Rev Clin Oncol. 2015;12(1):27-41.

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