



CENPA is one of the potential key genes associated with the proliferation and prognosis of ovarian cancer based on integrated bioinformatics analysis and regulated by MYBL2

Jing Han¹, Rongkai Xie¹, Ying Yang¹, Diangang Chen², Li Liu³, Jiayang Wu¹, Sufen Li¹

¹Department of Obstetrics and Gynecology, Xinqiao Hospital, Army Medical University, Chongqing, China; ²Cancer Institute of PLA, Xinqiao Hospital, Army Medical University, Chongqing, China; ³Department of Orthopedics, Chengdu Seventh People's Hospital, Chengdu, China

Contributions: (I) Conception and design: J Han, S Li; (II) Administrative support: R Xie, S Li; (III) Provision of study materials or patients: Y Yang, D Chen, L Liu; (IV) Collection and assembly of data: J Han, J Wu, Y Yang, D Chen, L Liu; (V) Data analysis and interpretation: J Han, R Xie, S Li; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Sufen Li. Department of Obstetrics and Gynecology, Xinqiao Hospital, Army Medical University, Xinqiao Street No. 2r, Chongqing 400037, China. Email: lifei858828@163.com.

Background: Ovarian cancer (OV) is a highly lethal disease, and the fifth leading cause of all cancer-related deaths in women. The study aimed to identify potential key genes associated with the proliferation and prognosis of OV.

Methods: Differentially expressed genes (DEGs) between ovarian cancer and normal tissues were screened by the robust rank aggregation (RRA) method. The expression of CENPA and MYBL2 were examined in SKOV3 and A2780 ovarian cancer cell lines and tumor tissues by qRT-PCR and western blot. Small RNA interference assays, plasmid overexpression assays and EdU assays were used to validate the proliferative effect of the MYBL2-CENPA axis in ovarian cancer cell lines. The ChIP assay was used to verify the direct regulation of MYBL2 on CENPA.

Results: 133 up-regulated genes and 158 down-regulated genes were identified, and the up-regulated genes mainly enrichment in cell cycle. The three up-regulated gene with DNA separation (CENPA, CENPF and CEP55) might be tightly correlated with proliferation and prognosis of OV. Knockdown CENPA expression inhibited the proliferation of A2780 and SKOV3 cells. After the knockout of MYBL2, the expression of CENPA significantly decreased. MYBL2 directly binds to the promoter region of CENPA.

Conclusions: The MYBL2-CENPA pathway plays an important role in the proliferation of ovarian cancer cells, suggesting that this pathway may be a potential target for the treatment of ovarian cancer.

Keywords: MYBL2; CENPA; ovarian cancer; proliferation; robust rank aggregation (RRA)

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Introduction

Ovarian cancer is a highly lethal disease, and the fifth leading cause of all cancer-related deaths in women. Furthermore, there have been approximately 295,414 (1.6%) new cases and 184,799 (1.9%) deaths in 185 countries (1), and 22,530 new cases and 13,980 deaths in the United States per year (2). Ovarian cancer can be classified as high-grade serous carcinoma (HGSC) and low-grade serous (LGSC), and that's

only a small fraction of all cases. HGSCs are characterized by rapid growth and aggressive behavior (3). The combination of cytoreductive surgery and chemotherapy has been the most important treatment strategy for HGSC, in terms of improving patient survival. However, the treatment efficacy remains unsatisfactory (4). Therefore, in order to better treat ovarian cancer, it is very important to understand the pathogenesis in greater detail.

MYB-related protein B (B-MYB/MYBL2) is a transcription factor and plays important roles in the regulation of cell cycle, survival and differentiation. In addition, it is often abnormally expressed in cancer, suggesting that it functions in driving the onset and/or progression of cancer (5,6). In glioma, demethylzeylasteral can inhibit glioma cell growth *via* the miR-30e-5p/MYBL2 axis (7). In addition, studies have shown that MYBL2 is regulated by the Akt/FoxM1 signaling pathway in gliomas. This may become a biomarker for glioma radiation therapy (8). In non-small-cell lung cancer (NSCLC), MYBL2 is aberrantly upregulated. Its overexpression can significantly promote NSCLC cell growth and motility by suppressing IGFBP3 (9). Furthermore, miR-30a may suppress the proliferation of NSCLC by targeting MYBL2 (10). In breast cancer, MYBL2 regulates cell proliferation and apoptosis, and is targeted by miR-143-3p (11). Mybl2 upregulation leads to an imbalance in cell cycle regulation that promotes liver cancer progression (12). In leukemic blasts, MYBL2 is regulated by CCNA1, and promotes cell proliferation (13).

The present study evaluates the prognostic value of MYBL2 in ovarian cancer, and explores the molecular mechanisms underlying its role in the disease. The results indicate that the abnormally high expression of MYBL2 promotes the proliferation of ovarian cancer cells mainly through the direct transcriptional regulation of centromere protein A (CENPA) expression. Therefore, targeting the MYBL2-CENPA axis may provide a new approach to the treatment of ovarian cancer.

We present the following article in accordance with the MDAR checklist (available at <https://dx.doi.org/10.21037/tcr-21-175>).

Methods

Gene expression profile data

Microarray data [GSE4122, GSE18520 (14), GSE12470 (15), GSE38666 (16), GSE27651 (17) and GSE26712 (18)] were downloaded from Gene Expression Omnibus (GEO). All of them meet the following criteria: (I) they all including human ovarian cancer samples. (II) Containing normal ovary samples. (III) Containing at least twenty samples. The R analysis process is borrowed from Liu *et al.* (19). The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Integrated analysis of microarray datasets

The Limma software package (20) in the R software was used to normalize the matrix data of each GEO data set and the base 2 logarithm conversion was used, and the DEGs between tumor and normal tissues was screened by Limma package. The R package “RobustRankAggreg” (21) is based on the Robust Rank Aggregation (RRA) method to perform genetic integration on DEGs determined from six data sets. Based on the null hypothesis of irrelevant inputs, the genes screened by this RRA method are always better than expected (21). $|\log_2FC| \geq 0.5$, and adjust P value < 0.05 were used.

Functional enrichment analysis of DEGs

To elucidate possible differences in biological processes, molecular functions, and cellular components with DEGs, GO enrichment analysis (22) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis (23) were performed.

Cell lines and culture

The SKOV3 and A2780 ovarian cell lines were purchased from ATCC (Manassas, VA, USA). Cells were cultured at 37 °C in DMEM (Hyclone, Logan, UT, USA) with 10% FBS (Hyclone) in a humidified incubator with 5% CO₂.

Small interfering RNA transfection

Lipofectamine 3000 (Thermo Fisher Scientific, Waltham, MA, USA) was used to transfect the MYBL2 or CENPA siRNA (GenePharma, Shanghai, China) into cells, according to manufacturer's instructions. The siRNA sequences were as follows: siMYBL2: 5'-CCCU GUCAGGUAUCAAGA-3' (24). siCENPA-1: 5'-UACACAUUUUCUCUUGCCA-3'; siCENPA-2: 5'-AAUUUAACACAUUUUCUCU-3'.

The qRT-PCR analysis

TRIzol reagent (Thermo Fisher Scientific) was used to extract the total RNA. Each experiment was performed for three times. The primer sequences were, as follows: *GAPDH* forward, 5'-CTGGGCTACACTGAGCACC-3', *GAPDH* reverse, 5'-AAGTGGTCGTTGAGGGCAATG-3'; *MYBL2* forward, 5'-CTTGAGCGAGTCCAAAGACTG-3', *MYBL2*

Table 1 The details of the samples in the 6 data sets from the GEO datasets

Dataset	Reference	Platform	Number of samples (control/tumor)
GSE4122	Tate DL, Mostafavi ZB, Mostafavi MT	GPL201 [HG-Focus] Affymetrix Human HG-Focus Target Array	67 (14/34)
GSE12470	Yoshihara K, Tajima A, Komata D, Yamamoto T, <i>et al.</i>	GPL887 Agilent-012097 Human 1A Microarray (V2) G4110B (Feature Number version)	53 (10/42)
GSE18520	Mok SC, Bonome T, Vathipadiekal V, Bell A, <i>et al.</i>	GPL570 [HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array	63 (10/53)
GSE26712	Bonome T, Levine DA, Shih J, Randonovich M, <i>et al.</i>	GPL96[HG-U133A] Affymetrix Human Genome U133A Array	195 (10/185)
GSE38666	Lili LN, Matyunina LV, Walker LD, Benigno BB, <i>et al.</i>	GPL570 [HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array	45 (20/25)
GSE27651	King ER, Tung CS, Tsang YT, Zu Z, <i>et al.</i>	GPL570 [HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array	36 (6/30)

reverse, 5'-AGTTGGTCAGAAGACTTCCCT-3'. CENPA forward, 5'-TTCCTCCCATCAACACAGTCG-3'; CENPA reverse, 5'-CACACCAGAGTGAATTTAACAC-3'.

Western blot

The total protein was extracted by RIPA buffer containing PMSF (Beyotime, Jiangsu, China). Then, 40 µg of total protein was separated on 10% polyacrylamide gels (Beyotime). Afterwards, After the 10% polyacrylamide gels (Beyotime) separation, the proteins were transferred onto PVDF membranes and blocked by 5% BSA. Subsequently, these PVDF membranes were incubated overnight. The antibodies are: mouse monoclonal anti-GAPDH (60004-1-Ig; Proteintech, Wuhan, China), Rabbit anti-MYBL2 (18896-1-AP; Proteintech), and Rabbit anti-CENPA (#2186; CST, Danvers, MA, USA). The protein bands were visualized using the ECL Western Blotting Substrate (Pierce, Rockford, IL, USA).

ChIP assay

The CHIP kit (Cell Signaling; Cat. #9005) was used. The sequences were, as follows: forward, 5'-TTATCTCAAAGCCCCGCCG-3', reverse, 5'-TGACAACCCCGTGTCTGATC-3';

Cell proliferation assay

Cells with siRNA-mediated knockdown were seeded in 96-well plates (6×10³ per well). and was detected using a CCK-

8Assay Kit (Dojindo, Kumamoto, Japan).

The EDU Assay Kit (GeneCopoeia, Rockville, MD, USA) was used to evaluate the proliferation of ovarian cells, according to manufacturer's instructions. Briefly, these cells were treated with EdU for five hours.

Statistical analysis

SPSS Statistics was used for the analyses. All data are expressed as mean ± standard deviation (SD). Unpaired Student's *t*-tests were used to evaluate the differences. *P*<0.05 was considered statistically significant.

Results

Identification of DEGs and functional enrichment analysis of DEGs

The details of the samples in the 6 data sets from the GEO datasets were listed in *Table 1*. After the integrated analysis, 291 DEGs were obtained, including 158 down-regulated genes (available online: <https://cdn.amegroups.com/static/public/tcr-21-175-1.xls>) and 133 up-regulated genes (available online: <https://cdn.amegroups.com/static/public/tcr-21-175-2.xls>). Top 20 down and up-regulated genes was showed in *Figure 1A*.

Enrichment analysis of GO and KEGG pathways was used to elucidate the potential biological functions of the 291 DEGs. In the down-regulated group, they were significantly enriched in various biological binding (*Figure 1B*). In the up-regulated group, they showed most

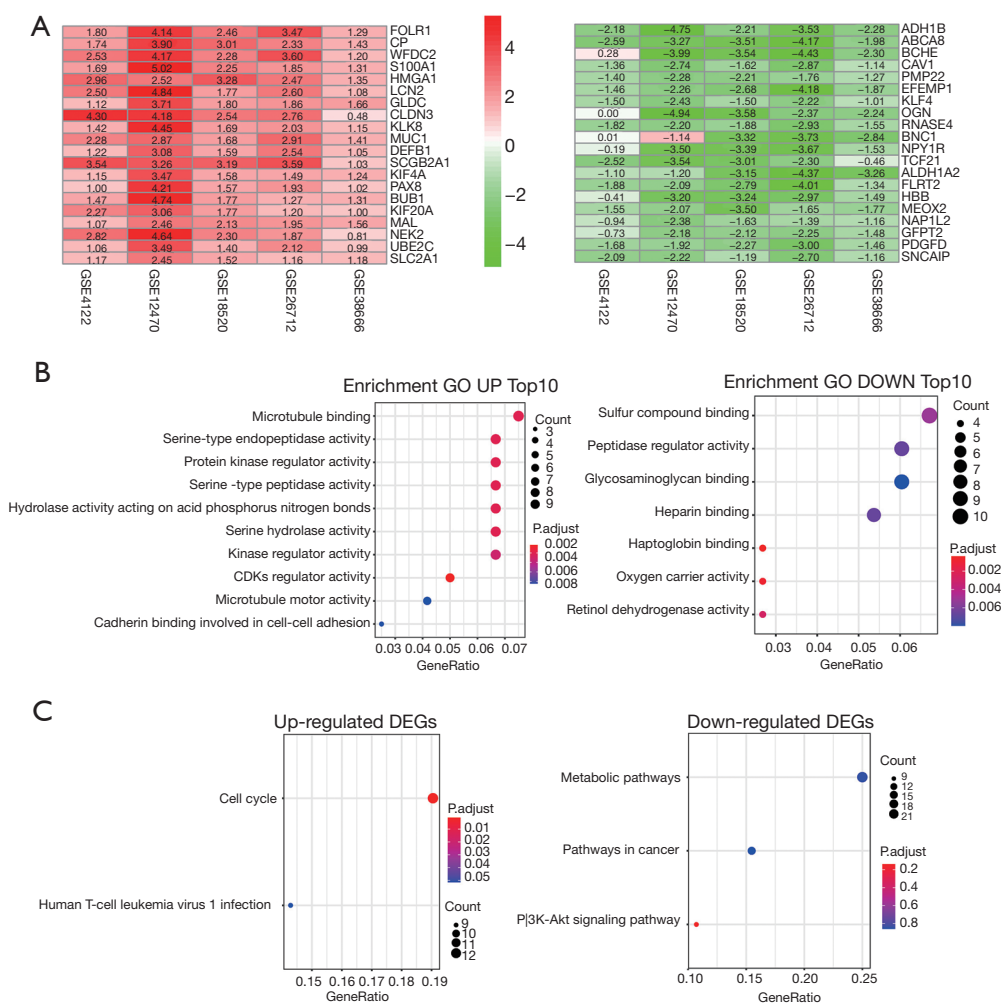


Figure 1 Identification of DEGs and Functional enrichment analysis of the DEGs. (A) Heat maps of the top 20 up-regulated and down-regulated DEGs in the integrated microarray analysis. (B) Top 10 up-regulated and down-regulated GO enrichment analysis of the DEGs. (C) KEGG pathway enrichment analysis of the DEGs.

significantly enriched in microtubule binding. In addition, they showed a close correlation with biological activity, such as serine-type endopeptidase activity, protein kinase regulator activity, serine-type peptidase activity and so on (Figure 1B). The results of KEGG pathway enrichment analysis showed that the up-regulated genes were enriched in the cell cycle (Figure 1C). In addition, down-regulated genes are mainly involved in metabolic pathways, cancer pathways, and PI3K-Akt signaling pathways (Figure 1C).

Expression characteristics of CENPF, CENPA and CEP55 in human ovarian tissues

According to the results of GO enrichment and KEGG

analysis, we know that the up-regulated DEGs are mainly related to the cell cycle, and the centromere protein had a very significant influence on cell cycle. Therefore, CENPF, CENPA and CEP55 proteins were chose for further research. We further studied the expression of these genes in the TCGA database. The GEPIA dataset (<http://gepia.cancer-pku.cn/>) was used to verified the expression patterns of the proteins in normal tissues and ovarian tissues. The mRNA expression levels of CENPF, CEP55 and CENPA in cancer tissues were higher than those in normal tissues (Figure 2A,B,C). The survival analysis was performed in the web (<http://kmplot.com/analysis/index.php>). Ovarian cancer patients with high expression of CENPA, CENPF or CEP55 had a lower overall survival rate than those with

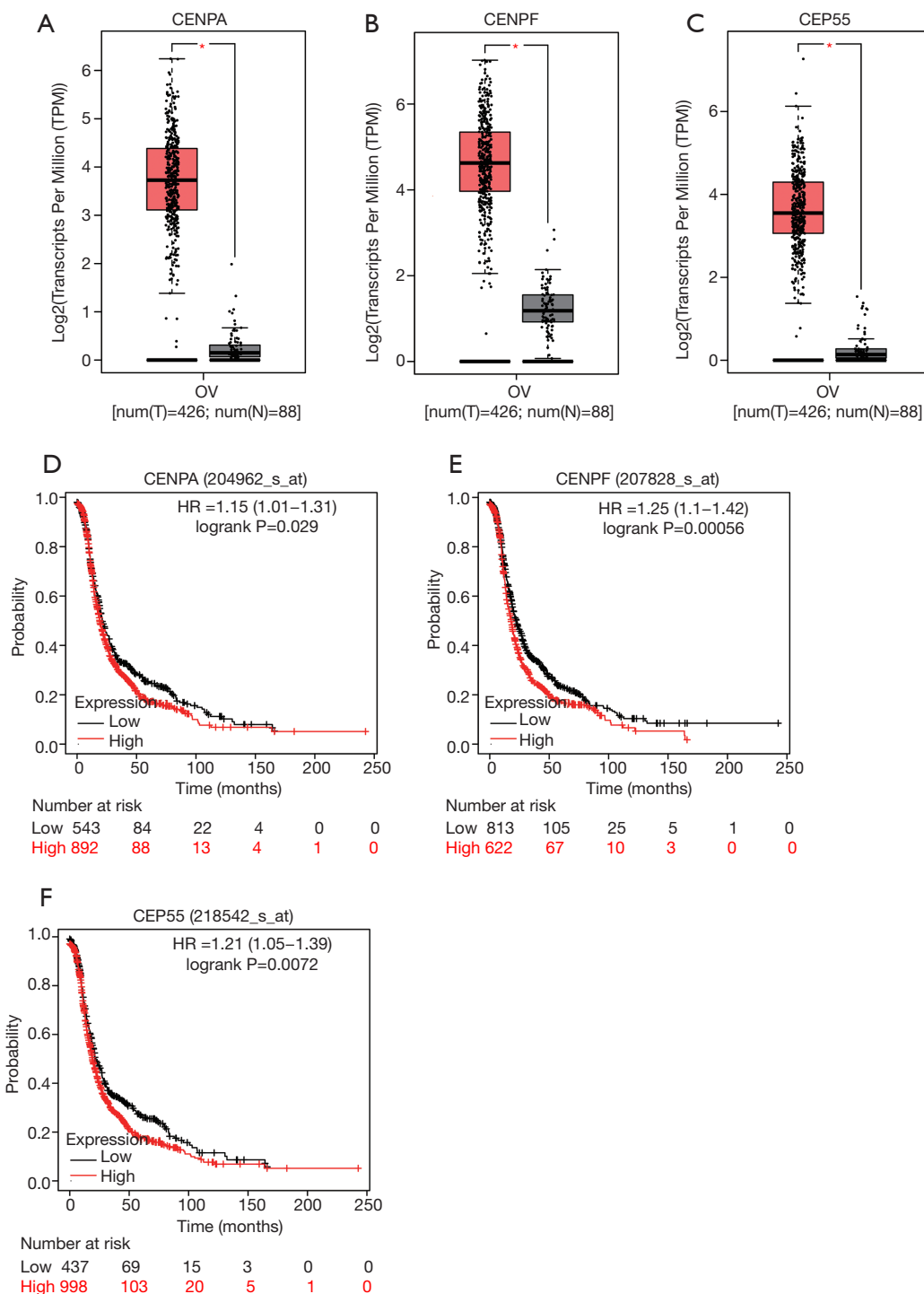


Figure 2 The expression pattern and prognostic value of CENPF, CENPA and CEP55 in ovarian cancer. (A) CENPA is more highly expressed in ovarian cancer tissues than in normal tissues (*P<0.05). (B) CENPF is more highly expressed in ovarian cancer tissues than in normal tissues (*P<0.05). (C) CEP55 is more highly expressed in ovarian cancer tissues than in normal tissues (*P<0.05). (D) The Kaplan-Meier analysis shows that ovarian cancer patients with a high expression of CENPA have the worse overall survival. (E) The Kaplan-Meier analysis shows that ovarian cancer patients with a high expression of CENPF have the worse overall survival. (F) The Kaplan-Meier analysis shows that ovarian cancer patients with a high expression of CEP55 have the worse overall survival.

low expression (Figure 2D,E,F).

The inhibition of CENPA inhibits the proliferation of ovarian cells

Previous experiments have demonstrated that ovarian cancer patients with a high expression of CENPA have poor prognosis. In order to verify the role of CENPA in ovarian cancer, its expression was knocked down in SKOV3 and A2780 cell lines using two different small RNA sequences (siCENPA-1 and siCENPA-2). Compared with levels in the control group, the CENPA mRNA and protein levels were significantly lower in these two cell lines (Figure 3A,B). The CCK-8 results revealed that the decreased expression of CENPA significantly inhibits the proliferation of ovarian cell lines (Figure 3C). Similar results were obtained in the EDU assay (Figure 3D). These results indicate that CENPA greatly promotes the proliferation of ovarian cells.

The expression of CENPA was regulated by MYBL2 in ovarian cancer cells

In order to study the regulatory mechanism of CENPA, we further studied CENPA in the TCGA database. We found that CENPA showed high consistency with MYBL2 (Figure 4A). At the same time, mRNA levels of MYBL2 were higher in cancer tissues compared to normal tissues (Figure 4B). Ovarian cancer patients with high MYBL2 expression showed worse overall survival (Figure 4C). The level of CENPA decreases with the knock down of MYBL2 in SKOV3 cell line (Figure 4D).

CENPA was directly regulated by MYBL2

In order to determine whether MYBL2 directly regulates the CENPA expression in ovarian cancer cells, ChIP assay was performed. The Cistrome database (<http://cistrome.org/db/#/>) was used to predict the binding ability. The results of the ChIP-seq from the Gertz *et al.* (25) revealed that MYBL2 can bind to the CENPA promoter in liver cells (Figure 5A). In order to verify the direct binding in ovarian cancer cells, the SKOV3 cell line was used for the ChIP assays. These results show that MYBL2 binds to the promoter region of CENPA (Figure 5B). In order to further confirm this result, the binding in the NC-SKOV3 and siMYBL2-SKOV3 cell lines was examined, and it was found that the binding of MYBL2 to the CENPA promoter significantly decreased (Figure 5B). Furthermore, the

qRT-PCR results also revealed the same trend ($P < 0.01$, Figure 5C). These data indicate that MYBL2 binds to the CENPA promoter region and promotes the CENPA expression.

Discussion

Unlimited cell proliferation, metastatic, and drug resistance are all the characteristics of cancer cells that can lead to a malignant phenotype and affect prognosis. The inhibition of cell proliferation has become an important strategy for cancer treatment. MYBL2 is an important regulator of cell cycle. When it is dysregulated in cancer cells, abnormal proliferation occurs. In fact, MYBL2 is overexpressed in a variety of cancers and is associated with poor prognosis. Thus, Some investigators have used MYBL2 as a biomarker for prognosis and as a potential therapeutic target (6). For example, the high expression levels of CCNE1, KRT16 and MYBL2 are associated with worse relapse-free survival and overall survival (OS) in ER-negative/HER2-negative breast cancer (26). In patients with primary hepatocellular carcinoma, high expression of MYBL2 is associated with low OS (27). In human hepatocellular carcinoma, high expression of MYBL2 is associated with faster progression (28). MYBL2 can be used as a target for cancer therapy, according to some studies. Ginkgetin regulates b-Myb by modulating miR-34a, and may be used to induce G2 arrest for the treatment of colorectal cancer (29).

An important sign of cancer is the unlimited proliferation of cancer cells (30), and this is mainly caused by the disruption in cell cycle regulation (31). The overexpression of MYBL2 is associated with the increase in cell proliferation, and plays an important role in cancer progression (32). However, the molecular mechanism by which mybl2 promotes cell proliferation in lung cancer remains unclear. A previous study revealed that MYBL2 binds to the A3B promoter, causing the transactivation in breast cancer cells (33). In addition, in the present study, it was found that MYBL2 promotes ovarian cell proliferation by directly regulating the expression of CENPA. This pathway further contributes to the pathogenesis and progression of ovarian cancer, providing a basis for treatment.

CENPA is a fundamental component of the human centromere and plays an important role in cell cycle, survival and genetic stability. CENPA expression is altered in various types of human cancers. In epithelial ovarian cancer, high levels of CENPA are significantly correlated with poor survival (34). In osteosarcoma, the expression

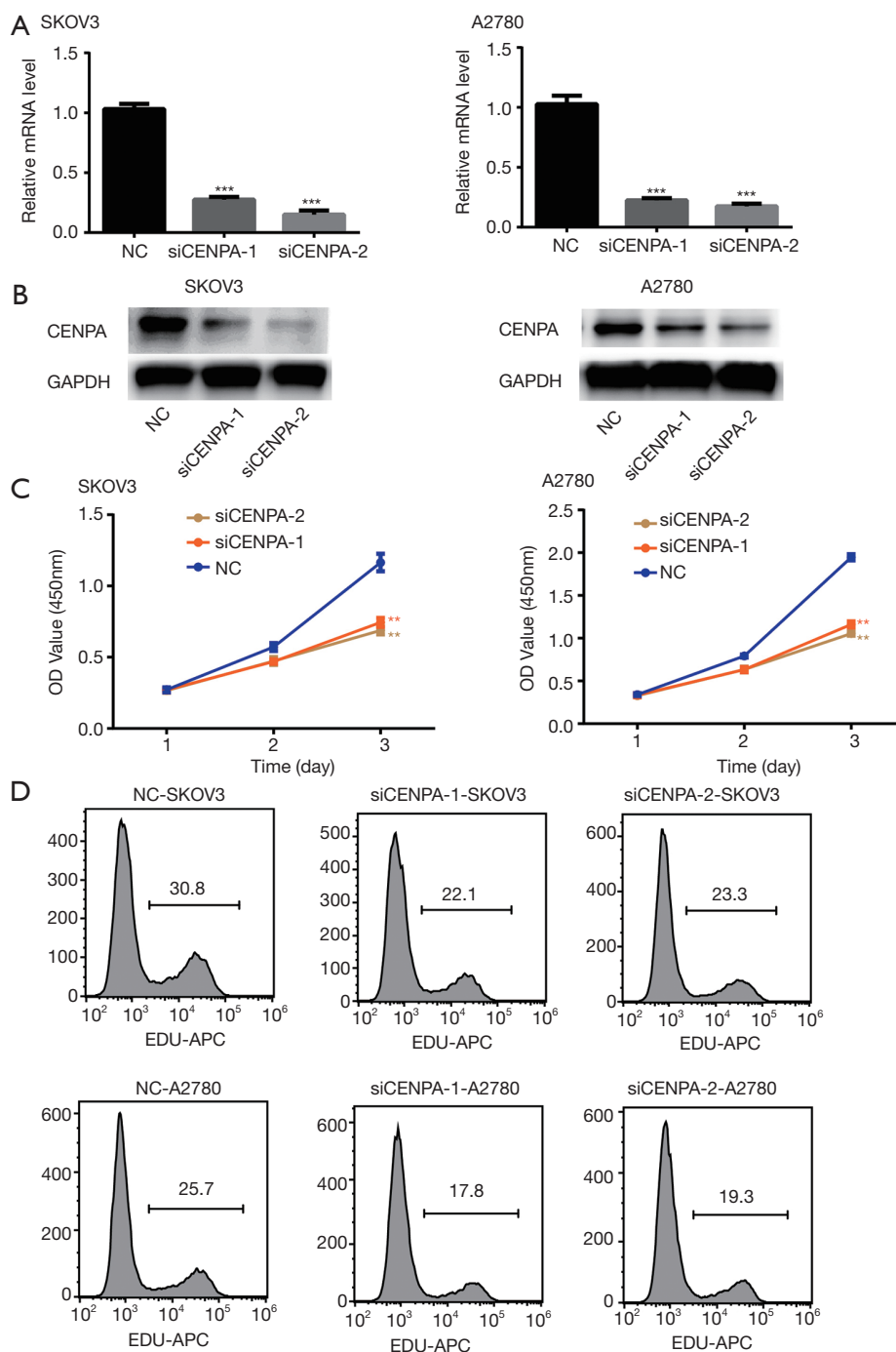


Figure 3 Knockdown of CENPA inhibits the proliferation of ovarian cells. (A) The qRT-PCR analysis of CENPA expression levels in the CENPA knockdown of SKOV3 and A2780 ovarian cell lines ($***P < 0.001$, independent t -test). (B) The western blot analysis of CENPA expression levels in the CENPA knockdown of SKOV3 and A2780 ovarian cell lines. (C) The CCK-8 assays in the CENPA knockdown of SKOV3 and A2780 ovarian cell lines ($**P < 0.01$, independent t -test). (D) The EDU assays in the CENPA knockdown SKOV3 and A2780 ovarian cell lines.

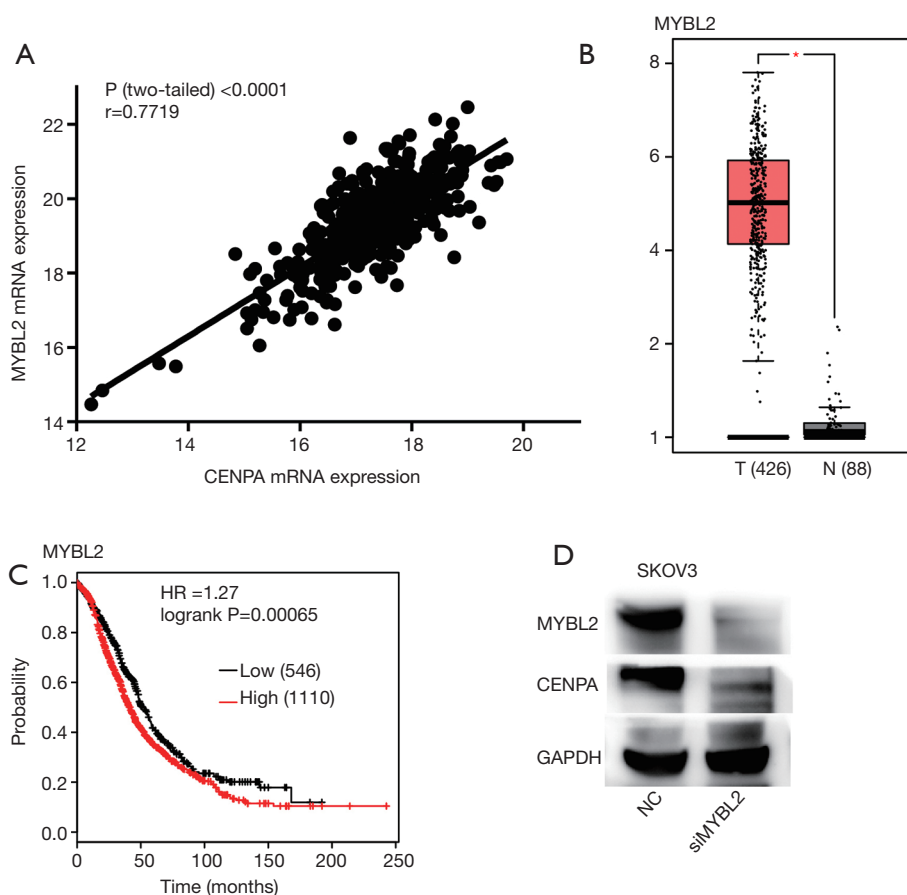


Figure 4 The expression of CENPA was regulated by MYBL2 in ovarian cancer. (A) CENPA expression showed a very high correlation with MYBL2 expression in ovarian cancer tissues (data from TCGA). (B) MYBL2 is more highly expressed in ovarian cancer tissues than in normal tissues (* $P < 0.05$). (C) The Kaplan-Meier analysis shows that ovarian cancer patients with a high expression of MYBL2 have the worse overall survival. (D) The level of CENPA decreases with the knock down of MYBL2 in SKOV3 cell line.

of CENPA is significantly associated with progression, and patients with a high expression of CENPA have poor overall and recurrence-free survival (35). In triple-negative breast cancer, BIRC5, CENPA and FAM64A are specifically upregulated, and the high levels of these three genes are associated with poor prognosis, suggesting the clinical application as therapeutic targets (36). In lung adenocarcinoma, CENPA upregulation predicts a poor OS in patients, and CENPA downregulation may attenuate the aggressive phenotype of lung adenocarcinoma cells (37,38). In glioblastoma, CENPA is consistently expressed in GSC cultures, but is not expressed in NSC cultures, suggesting that this should be further explored as therapeutic targets (39). In hepatocellular carcinoma, CENPA is strongly associated with hepatocellular carcinoma cell growth and division (40).

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Footnote

Reporting Checklist: The authors have completed the MDAR checklist. Available at <https://dx.doi.org/10.21037/tcr-21-175>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://dx.doi.org/10.21037/tcr-21-175>). The authors have no conflicts of interest to declare.

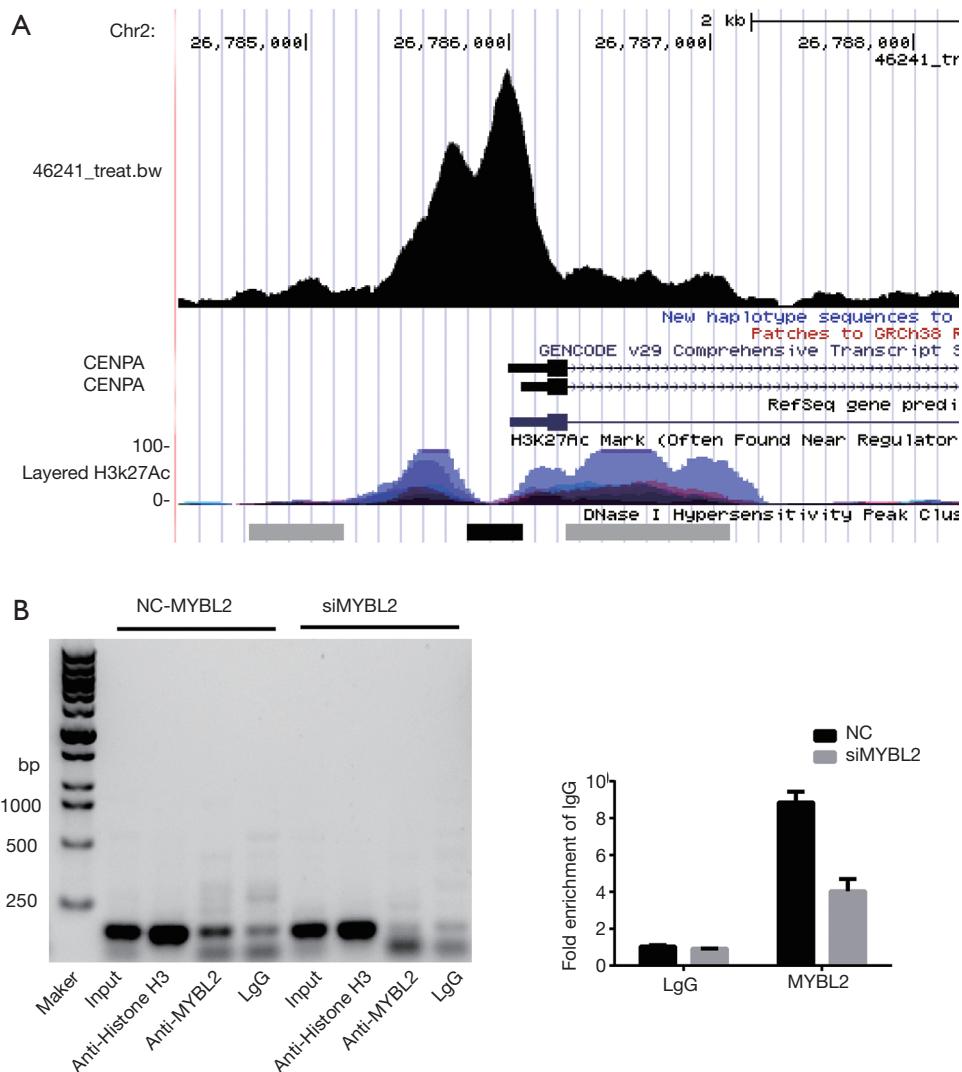


Figure 5 MYBL2 directly regulates the expression of CENPA in SKOV3 cell line. (A) Prediction of the binding ability of MYBL2 to the CENPA promoter region using Cistrome DB. (B) The ChIP assay results for the direct binding of MYBL2 to the promoter of CENPA. (C) The qRT-PCR analysis of the ChIP assay results.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). Institutional ethical approval and informed consent were waived.

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