



On Having No Head: Cognition throughout Biological Systems

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The central nervous system (CNS) underlies memory, perception, decision-making, and behavior in numerous organisms. However, neural networks have no monopoly on the signaling functions that implement these remarkable algorithms. It is often forgotten that neurons optimized cellular signaling modes that existed long before the CNS appeared during evolution, and were used by somatic cellular networks to orchestrate physiology, embryonic development, and behavior. Many of the key dynamics that enable information processing can, in fact, be implemented by different biological hardware. This is widely exploited by organisms throughout the tree of life. Here, we review data on memory, learning, and other aspects of cognition in a range of models, including single celled organisms, plants, and tissues in animal bodies. We discuss current knowledge of the molecular mechanisms at work in these systems, and suggest several hypotheses for future investigation. The study of cognitive processes implemented in aneural contexts is a fascinating, highly interdisciplinary topic that has many implications for evolution, cell biology, regenerative medicine, computer science, and synthetic bioengineering.

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INTRODUCTION

Survival in a complex, dynamic, and highly competitive environment requires biological systems to make numerous decisions with respect to possible activities (Conrad, 1996; Holcombe and Paton, 1998). Evolutionary pressure to optimize decision-making has led to the inevitable exploitation of past history (memory) and information processing (computation). Importantly however, decisions are made at every level of biological organization. For example, multicellular organisms, such as animals and higher plants, exhibit multilayer complex goal-directed behaviors also at their cellular and subcellular levels. Underlying physiological systems must maintain homeostasis and predict future conditions (Freddolino and Tavazoie, 2012) in the face of unpredictable changes in environmental conditions, while cells must coordinate their activity in an exquisite 3dimensional ballet of embryogenesis and complex organ regeneration. At the extremes of the scale of organization, dynamic self-organizing subcellular components like cytoskeleton and molecular networks (Albrecht-Buehler, 1985; Craddock et al., 2012; for plant cells see Volkmann and Baluška, 1999; Barlow and Baluška, 2000) and colonies of organisms (Shapiro, 1998; Couzin, 2009) perform similar functions in their own contexts. Here, "cognition" refers to the total set of mechanisms and processes that underlie information acquisition, storage, processing, and use, at any level of organization (Lyon, 2015).

Memory is an essential component of these processes, at all levels. For our purposes, memory can be defined as experiencedependent modification of internal structure, in a stimulusspecific manner that alters the way the system will respond to stimuli in the future as a function of its past. This requires a labile yet stable medium, to provide the necessary latency. The process may or may not involve a degree of intelligence, in the sense of the ability to compress prior stimuli into informationallycompact representations (inference). In essence, sensory memory is a message to one's future self - a view reminds us that memory is thus another instance of biological communication (which, as exchange of signals, is ubiquitous among all levels of biological organization). Put this way, we can see that in principle many biological mechanisms can be exploited for this purpose. The updates in the configuration (or state) of a system, as occurs also during sensory memory formation in all organisms, is formally known as "computation" in computer science.

These concepts are quite general. However, outside of the unconventional cognition community (Calvo and Baluška, 2015) or biological computation community (Adamatzky et al., 2008), it is widely assumed that memory is the exclusive province of brains, or even complex animals. Older work exploring these issues in plants (reviewed by Gremiaux et al., 2014), non-neural somatic tissues (Mackie, 1970), and even inorganic media (Bose, 1926), have been largely forgotten in favor of the remarkable advances in recent cognitive neurosciences with their focus on the brain. Nevertheless, plant cells are known to be able to use action potentials to control their movements and behavior since times when Charles Darwin and Jagadis Chandra Bose turned their interest toward plants (Darwin, 1880; Shepherd, 2005; Baluška et al., 2009a). Currently, surprisingly, higher plants are emerging as behaviorally active organisms, enjoying bio-communication and showing plant-specific cognition and intelligence (Trewavas, 2005, 2014; Karban et al., 2014a,b; Calvo and Baluška, 2015; Calvo, 2016).

Here, we survey a wide-ranging literature on memory and sensory systems-based cognition in organisms (biological systems) lacking animal/human-type brains. Our goal is to acquaint readers interested in cognition with numerous aneural model systems in which this subject can be pursued, and to draw the attention of bench biologists working on those systems to cognitive, information-focused perspectives on the mechanisms they are studying. Importantly, in discussing cognitive performance in the various systems, we do not mean the full-blown human-like cognitive performance, or humantype of self-awareness and consciousness. We are avoiding issues of the 'Hard' problem of cognitive science, and do not claim anything like higher-order symbolic representations. Our definition is purely functional and minimalist (Calvo and Baluška, 2015), drawing attention to the similarities in computational tasks performed by diverse biological systems, at all levels of complexity, other than animal and human brains. Figure 1 illustrates the full spectrum of cognitive levels and capabilities upon which the various systems we discuss can be placed (Rosenblueth et al., 1943). Our review begins with a consideration of the familiar substrate of cognition: neural dynamics, and of mechanisms that blur the boundaries

between neural and non-neural cell functions. We then proceed through progressively more divergent cognitive systems, considering molecular networks, single cell behaviors, networks of cells in various tissues, and organism-wide information processing during regenerative repair. We conclude with some common threads of cognition across levels of organization, which suggest a unified perspective on these highly diverse systems.

NEURONS: THEIR POWERS, EVOLUTIONARY HISTORY, AND BEYOND

Recent work has begun to encompass cognition in *ex vivo* systems, with studies that have shown training and learning in cultured minimal neural networks (DeMarse and Dockendorf, 2005; Dranias et al., 2013; Pimashkin et al., 2013). Even *in vivo*, it is increasingly recognized how much processing happens before signals get to the brain of the central nervous system (CNS); a recent example is the discovery that neurons in the skin perform edge detection (Pruszynski and Johansson, 2014).

Importantly, CNS neurons do not embody cognition due to any magical, unique property. Their computational powers derive from the dynamics of networks of linked elements that propagate and integrate signals, and the ability to alter connectivity among those elements (network topology) based on prior activity. In fact, these basic properties are present in biological systems at many complexity scales (from subcellular protein networks to coupled tissues). Might they too underlie some aspects of cognitive-like information processing? Indeed, neurons did not invent their special tricks - they merely optimized them for speed to drive adaptive behavior. These functions, and the molecular mechanisms that implement them - ion channels, electrical synapses (gap junctions), and neurotransmitter molecules are all ancient (Goldsworthy, 1983; Baluška, 2010; Brunet and Arendt, 2016; Moroz and Kohn, 2016). Neural networks evolved from far older signaling pathways that orchestrated development, physiology, and other cellular functions long before the CNS arrived on the evolutionary scene (Buznikov et al., 1996; Levin et al., 2006; Keijzer et al., 2013). Already simple cells of bacteria enjoy sensory systems feeding into cognitive-behavioral circuits and showing many other neural features (Miller and Koshland, 1977; Koshland, 1980; Lyon, 2015). Electrical long-distance signaling and information exchange via spatially propagating waves of potassium is synchronizing bacterial biofilms (Beagle and Lockless, 2015; Nunes-Alves, 2015; Prindle et al., 2015). Integrated bacteria within the biofilm community appear to act as some kind of 'microbial brain'. Obviously, the neuronal communication has bacterial origins (Baluška and Mancuso, 2009).

The main principles by which neural networks store and process information – plasticity, excitability, and experiencedependent change (Daoudal and Debanne, 2003) are readily applicable to numerous cell types. Indeed, the computational powers of glia and other non-spiking cells in the brain are increasingly appreciated in their contributions to mammalian



cognition and intelligence (Oberheim et al., 2009; Goldman et al., 2015). Astrocyte networks perform computations (Schummers et al., 2008), and models of memory have long been proposed that rely on non-spiking neurons (Aur, 2012), revealing that neural-specific, discrete action potentials are not a pre-requisite for memory dynamics.

At the same time, tissues other than neurons are able to conduct the kind of signaling impulses that are considered the sine qua non of cognition. For example, excitation and impulse propagation have been shown in skin (Roberts and Stirling, 1971; James and Soffe, 2011). The evolution of neurons from excitable precursors has been reviewed elsewhere (Mackie, 1970; Baluška and Mancuso, 2009; Baluška, 2010; Moroz and Kohn, 2016), as have the many similarities between neurons and other cell types (Bharti and Arnheiter, 2005; Yaar and Park, 2012). In this overview, we cast our net even broader, examining examples of cognition outside of the CNS domain of life (Calvo and Baluška, 2015; Lyon, 2015; Calvo, 2016), with or without spiking, in cellular networks of complex metazoans, or within singlecells. We also review some of the mechanisms that underlie this cognition which is inherent to cellular life at all levels of biological complexity, and suggest a few novel experimental directions that may exploit the deep lessons suggested by the ubiquitous nature of aneural cognition.

CROSSOVER BETWEEN NON-NEURAL AND NEURAL MECHANISMS

The interplay of neural and non-neural signaling has been shown in several regenerative systems. Neural inputs are required for amphibian limb regeneration (Singer, 1952; Kumar and Brockes, 2012), although curiously, this is not a hardwired requirement but must be learned: limbs that grew without the presence of a nerve later do not require nerve to regenerate, unlike normal limbs. This phenomenon has been termed "nerve addiction" (Yntema, 1959a,b; Filoni et al., 1995), extending the principle of experience-dependent long term change to limb regeneration. Neural inputs are also needed for maintenance of tissue structure in the rodent tongue (Takeda et al., 1996; Sollars et al., 2002), suppression of tumors in rabbits (Pawlowski and Weddell, 1967; Pawlowski, 1970), and regulation of specific pattern in distal tail regeneration in amphibia (Mondia et al., 2011).

Neural and non-neural information systems must cooperate especially when pattern formation and memory intersect. A unique model system for these studies is planaria, the free-living flatworm (Gentile et al., 2011); this is a unique model system that regenerates every part of its body (Reddien and Sánchez Alvarado, 2004) and also possesses a true centralized brain (Sarnat and Netsky, 1985; Pagán, 2014) and learning capabilities (Wells, 1967; Sheiman and Tiras, 1996; Nicolas et al., 2008). In this model species, the dynamics of behavioral memory can be studied during complete brain regeneration [in the axolotl, this can also be done, albeit with only partial brain regeneration (Pietsch and Schneider, 1969)]. Classical studies (McConnell et al., 1959; Corning, 1966), as well as more recent work performed using automated analysis methods (Shomrat and Levin, 2013), showed that memories in planaria survive decapitation - tail fragments trained on a task regenerate brains and then show evidence of recall of the original information. This requires the body to store learned information and imprint it on the nascent brain after it is rebuilt. The mechanisms of this interaction are completely unknown, but offer an unprecedented opportunity to study transfer between neural and somatic memory systems.

MOLECULAR MECHANISMS OF NON-NEURAL COGNITION

Memory, and often the intermediate processes of computation, requires that "stimuli produce a permanent record written on the irritable substance" (Semon and Simon, 1921). What underlying mechanisms have been implicated in non-neural memory and related processes?

One of the best candidates for mechanisms underlying information processing at the single cell level is the cytoskeleton (Albrecht-Buehler, 1985; Craddock et al., 2010; Sahu et al., 2013), which has all of the necessary properties: it is a large, complex structure that is readily modified by a variety of molecular pathways (writing data), is interpreted by numerous motor proteins and other machinery (reading data), and implements a rich set of discrete transition states that could implement computational operations (Hameroff and Watt, 1982; Lahoz-Beltra et al., 1993; Volkmann and Baluška, 1999; Craddock et al., 2012). The cytoskeleton has long been a favorite locus of information storage and control in single-celled organisms, where it regulates behavior (Eisenstein, 1967; Hamilton, 1975) and serves as a non-genomic repository for permanent somatic changes such as cell surface chirality changes (Nelsen et al., 1989). The dynamic actin cytoskeleton behaves as excitable medium (Khan et al., 2012).

Another medium for information processing is within chemical networks, such as reaction-diffusion (RD) dynamics that underlie pattern formation in embryogenesis (Kondo, 2002; Kondo and Miura, 2010; Raspopovic et al., 2014). Recent work has revealed that RD systems and similar excitable chemical media can be designed so as to execute specific computations, and are being used for the design of minimal cognition controllers (Dale and Husbands, 2010) and other kinds of computation including planning (Adamatzky et al., 2003; Adamatzky et al., 2008; Costello et al., 2009). Remarkably, it was shown long ago (Rosen, 1968) that Rashevsky's 2-factor systems (a model for neuronal excitation) is formally equivalent to Turing's RD scheme for self-organizing morphogenesis (Turing, 1952). Grossberg then described extensive parallelism between signal processing in chemical gradients during development and neural memory and visual processing (Grossberg, 1978). RD systems are Turingcomplete (Scarle, 2009) and support semantical interpretations (Schumann and Adamatzky, 2009), making them an excellent candidate for complex computations. Recent work used *in silico* evolution of chemical networks to show that simple, plausible reactions can be found which perform associative learning and Bayesian behavior which includes memory traces (McGregor et al., 2012). These data are especially exciting in that they imply that associative learning can readily evolve in metabolic, gene regulatory, or intracellular signaling networks.

The transcriptional control machinery that guides embryogenesis has also been modeled as cognitive processes. Gene regulatory networks can be modeled as neural networks (Watson et al., 2010), with genes representing nodes and functional links representing inductive or repressive relationships among those genes. That landmark study showed that changes to the connections in the regulatory net represent a kind of Hebbian plasticity (as genes whose expression is up-regulated in specific environments tend to become co-regulated and thus expressed together). In part due to this fire-together-wire-together process, a GRN will develop an associative memory of phenotypes selected in the past. This view sheds important light on the relationship between homeostasis and evolvability and shows that a transcriptional network can develop memory and recall capabilities often thought to be reserved for classical cognitive systems. As a consequence of memory, genetic networks can exhibit predictive ability, enabling anticipatory behavior with respect to physiological stimuli (Tagkopoulos et al., 2008). A similar result was obtained for protein networks, showing that signaling via the tumor suppressor P53 could be modeled as a neural net (Ling et al., 2013), while MAP kinase pathways implement specific decision-making processes (McClean et al., 2007). Embryos make use of genetically encoded cellular memory, for example in the case of HOX gene expression patterns, which constitute a form of positional memory - "an internal representation by a cell of where it is located within a multicellular organism" (Chang et al., 2002; Rinn et al., 2006; Wang et al., 2009), and hysteresis in Hedgehog protein signaling (Balaskas et al., 2012), all of which are used to guide the subsequent activity of cells as a function of prior "experience".

Additional memory media include the extracellular matrix (Becchetti et al., 2010; for plant cell walls see Humphrey et al., 2007; Seifert and Blaukopf, 2010; Hamann, 2015) and chromatin complex markings (Francis and Kingston, 2001; Maurange and Paro, 2002; Ringrose and Paro, 2004), both of which are ideal media for recording traces representing specific environmental and/or physiological events. These are examples of internal stigmergy – activity that leaves traces in a labile intracellular or extracellular medium which can be read as memories in the future by cells making decisions for migration, differentiation, apoptosis, or signaling (Theraulaz and Bonabeau, 1999; Ricci et al., 2007).

Importantly, many cell types communicate electrically, not just excitable nerve and muscle (McCaig et al., 2005; Levin, 2007a,b, 2012a; Bates, 2015). Recent molecular data show that developmental bioelectricity is an important modality by which cell networks process information that instructs patterning during regeneration, development, and cancer suppression (Levin, 2014a,b,c). Thus, one obvious candidate for cognition outside the brain is via the same mechanism used in the brain bioelectrical networks (Levin and Stevenson, 2012; Mustard and Levin, 2014). Indeed it is likely that the processing in the brain is a direct extension (and speed optimization) of far older mechanisms used originally for morphogenesis (Buznikov and Shmukler, 1981; Levin et al., 2006). Developmental bioelectricity in animal systems features slowly-changing, continuous voltage changes as opposed to millisecond discrete (binary) spiking usually studied in the brain. However, the brain also includes non-spiking neurons (Victor, 1999) that have computational compartments similar to the membrane voltage domains observed in embryonic and other non-neural cells (Levin, 2007b; Adams and Levin, 2012). It has recently been proposed (Levin, 2012b, 2013; Mustard and Levin, 2014) that nonneural tissues support the same two types of plasticity as seen in the brain: changes of connectivity via electrical synapses (gap junctions) which corresponds to synaptic plasticity, and changes of ion channel function which corresponds to intrinsic plasticity (Marder et al., 1996; Turrigiano et al., 1996; Daoudal and Debanne, 2003). In addition to computation via changes in resting potential, which is a primary regulator of pattern memory in embryogenesis and regeneration (Adams, 2008; Funk, 2013; Levin, 2014b), as well as of processing in the brain (Sachidhanandam et al., 2013; Yamashita et al., 2013), ion pumps such as the ubiquitous sodium-potassium ATPase, have been suggested as computational elements (Forrest, 2014).

Most of these bioelectrically active systems are based on ion dynamics at membranes which modify bioelectric fields via activities of ion channels and transporters (Taylor, 1974; Wayne, 1993, 1994; Hille, 2001). These membrane-associated electric fields feed-back on membranes and associated proteins (Jaffe, 1977; Tsong and Astumian, 1986; Westerhoff et al., 1986; Bezanilla, 2002, 2006, 2008). They also control endocytosis and vesicle trafficking (Antov et al., 2005; Baluška and Wan, 2012). Relevantly, even biochemical reactions are under electric control (Aragoněs et al., 2016; Xiang and Tao, 2016), as is transcription (Pai et al., 2015b) and chromatin modification (Carneiro et al., 2011; Chernet and Levin, 2014).

COGNITIVE CAPABILITIES OF SINGLE CELLS

While the dominant model of neural-based cognition relies on the signaling dynamics among networks of neurons, it's becoming increasingly appreciated that single neurons can execute subtraction, addition, low- and band-pass filtering, normalization, gain control, saturation, amplification, multiplication, and thresholding with respect to the inputoutput relations they implement (Koch and Segev, 2000). Memory and computation is thus not exclusively a multi-cellular phenomenon, and is not restricted to somatic neural cells. Recent computational studies have revealed conditions under which cells expressing ion channels can keep a stable memory with respect to resting potential, and these conditions do not specifically require neuronal cell identity – they can be fulfilled by numerous cell types, somatic as well as free-living (Ramanathan and Broach, 2007; Cervera et al., 2014; Law and Levin, 2015).

The amoeba of *Dictyostelium discoideum* migrate by extending pseudopods in an alternating pattern. The specific pattern of the pseudopods' zig-zag behavior was recently shown to be predictable by viewing the cell surface as an excitable medium. In this model, the appearance of a pseudopod makes the local cortex temporarily more excitable (a kind of potentiation), while globally new pseudopods are inhibited. This model thus includes a memory of previous pseudopod locations, and quantitatively fits data from cell tracking experiments and the known chemotactic sensitivity of these cells (Cooper et al., 2012).

Budding yeast also keep a history which influences their future behavior – a memory of past events. They avoid pheromoneinduced cessation of cell cycle after a deceptive mating attempt (failure to reach a putative partner cell within a specific time period). The mechanisms of this are beginning to be unraveled (driven by the dynamics of the maternally segregating G1/S inhibitor Whi3), and the authors term the macromolecular assemblies that mediate this memory "mnemons", cellular structures that encode previous environmental conditions (Caudron and Barral, 2013). With respect to the search for the molecular substrate of specific memories, this yeast work may be ahead of similar efforts in the brain (Ungar, 1972, 1974a,b).

The flexible and versatile responses of bacteria to their environment has drawn significant attention of synthetic, molecular, and evolutionary biologists, as well as those interested in unconventional computational media (Miller and Koshland, 1977; Koshland, 1980; Ben-Jacob, 2009; Norris et al., 2011). Single bacteria are able to migrate toward beneficial targets, and away from noxious stimuli. The control algorithm for this behavior has long been the subject of investigation, with respect to the shortterm memory needed for following gradients (Vladimirov and Sourjik, 2009) as well as "infotaxis" policies that do not require gradient sensing (Vergassola et al., 2007). Especially exciting are the recent findings that bacterial communities (biofilms) process information and make decisions about nutrient distribution and metabolism as an integrated whole, using ion channels (Prindle et al., 2015) and a kind of volume transmission as occurs in the brain (Agnati et al., 2006; Fuxe et al., 2013; Zhang et al., 2013). Ciliates (protozoa) exhibit learning and a form of memory, which even survives loss of nuclei and some cytoplasm (Gelber, 1962; Applewhite et al., 1969; Hamilton, 1975; Clark, 2013). The mechanism is unknown, but may involve electrical signaling (Applewhite, 1972; Kunita et al., 2014).

In addition to cells that make their living independently, single somatic cells from metazoan organisms also exhibit memory and decision-making (Albrecht-Buehler, 1985) during directed steering (Albrecht-Buehler, 1982) – a capability that also extends to cell fragments (Albrecht-Buehler, 1980) and even human sperm that adjust their flagellar beat to reach the egg via calciumdependent tracking of chemical attractants (Alvarez et al., 2013).

The immune system has long been a paradigm of pattern recognition and classification (Carter, 2000). While the mainstream view of immune function is that of an evolutionary

system driven by selection, a cognitive perspective has been proposed (Cohen, 1992a,b) as an alternative theoretical framework for understanding the body's remarkable ability to distinguish self from non-self and adapt via immunological memory. It is interesting that the converse proposal has been made as well, to understand brain dynamics via a selectionist model (Fernando et al., 2012). Thus, in some sense, evolutionary and cognitive dynamics could be parallel (isomorphic?) ways to explain complex systems. If true in general, it may have significant implications for evolutionary theory.

An interesting kind of cognitive process is revealed by drug addiction. The increased tolerance with exposure is desensitization (one kind of basic memory element). Drug addiction reactions have been shown in somatic mammalian cells in culture (Corssen and Skora, 1964; Manner et al., 1974; Higgins et al., 1978), suggesting that this form of memory is not always a body-level phenomenon that necessarily involves the brain.

SLIME MOLDS: BETWEEN UNICELLULAR LIFE AND METAZOAN BODIES

Physarum polycephalum is a slime mold that has been extensively used in studies of biological information processing (Nakagaki et al., 2004; Saigusa et al., 2008). By computing optimal paths for nutrients throughout its syncytial body, the organism can implement behavior that solves challenging spatial optimization problems, such as solving mazes and finding efficient highway layouts (Nakagaki et al., 2004; Saigusa et al., 2008; Adamatzky and Alonso-Sanz, 2011). This organism shows how the internal dynamics of morphogenesis, even at this primitive step toward a multicellular bodyplan, can implement decision-making and computation. It is particularly interesting that in this system, a kind of variational (least-action, or minimization) principle is explicitly implemented by a biological medium (Friston, 2010; Friston et al., 2015), providing a much-needed "base case" for starting to understand the common features of goal-directed activity across levels of organization from cells to body structures to organism behavior.

Physarum also shows evidence of memory. In their study of the Traveling Salesman Problem (requiring an optimal strategy for connecting regions in space), Zhu et al. (2013) found that when two individuals were created by dividing one individual, they remained correlated in their exploration even though they were spatially separated, suggesting the presence of a long-term memory in the intrinsic dynamics.

COGNITION IN PLANTS

Although plants are still considered generally to be outside of neuronal and cognitive organisms, due to their lacking of animaltype of neurons and brains, plant cells have many features which are considered neuronal, including plasma membrane excitability supporting action potentials, acentriolar microtubules, motile Trans Golgi Networks, and synaptic-like actin-enriched cell-cell adhesion domains (Wayne, 1993, 1994; Barlow and Baluška, 2000; Baluška et al., 2003, 2005, 2008, 2009b; Baluška, 2010). Especially cells in root apices are very active in these neuronallike activities and act as *brain-like* command centers (Baluška et al., 2004, 2009a,b, 2010; Baluška and Mancuso, 2009, 2013), navigating growing roots in their search for water and mineral nutrients in soil, and active root avoidance or escape from toxic, stressful and dangerous situations (Burbach et al., 2012; Yokawa et al., 2014; Yokawa and Baluška, 2015, 2016).

The classic studies on plants showing animal-like features and activities were accomplished more that 150 years ago by Charles Darwin, assisted with his son Francis Darwin, and Claude Bernard (Darwin, 1880; Bancroft and Richter, 1930; Perouansky, 2012). Later, Jagadis Bose accomplished his sophisticated experiments on plants, confirming and extending the previous results obtained by Charles Darwin and Claude Bernard (Shepherd, 2005). Despite the fact that plant action potentials are known for more than 150 years now, and these are known to control many plant processes (Wayne, 1993, 1994; Masi et al., 2009; Volkov et al., 2010; Sukhov et al., 2011; Böhm et al., 2016; Hedrich et al., 2016), plant action potentials are still ignored by the mainstream. For example, there is no single mention of plant action potentials in the book Plant Physiology by Lincoln Taiz, which represent the most accepted view of plants in biology (Taiz, 2010).

Claude Bernard performed many anesthetic experiments. He expanded experimental materials from animals to plants. He showed that the Mimosa plant (Mimosa pudica), closing leaves upon touch, was unresponsive when exposed to a diethyl ether atmosphere which immobilized mice. Claude Bernard arrived at the conclusion that plants and animals share a common biological principle that is disrupted by anesthetics. He hypothesized that similarly as animals, also plants are able to actively sense their environment. He called this ability plant "sensitivity". In order to test his ideas, he performed anesthesia on plants and the results of these experiments were presented in 1878 in "Leçons sur les phénomènes de la vie communs aux animaux et aux végétaux" (Bernard, 1878; Bancroft and Richter, 1930). Later, sensitivity of plants to anesthetics was confirmed not only for Mimosa and Dionea, but also for many other plants (Milne and Beamish, 1999; De Luccia, 2012; Gremiaux et al., 2014).

Similarly as neurons, plant cells are excitable and plant-specific action potentials serve for long-distance communication and integration of plant bodies. Action potential also control rapid plant organ movements such as closing the Dionea traps or touch-induced movements of Mimosa leafs (Volkov et al., 2010; Böhm et al., 2016; Hedrich et al., 2016). Our preliminary data with Dionea traps suggest that anesthetics block action potentials (Yokawa et al., in preparation). Moreover, action potentials control also nutrient transporters in Dionea prey-stimulated traps (Böhm et al., 2016; Hedrich et al., 2016). In the root apex, the transition zone is very active not only in electric activities (Masi et al., 2009), and synaptic-like cell-cell communication (Baluška et al., 2003, 2004, 2005, 2009a,b, 2010; Baluška and Mancuso, 2013), but also in sensory-based control of root growth navigation associated with high electric activity. Root apex navigation is based on complex computations as roots sample continuously huge amounts of abiotic and biotic information from their environment in order to find water and nutrient rich zones in soil; and to avoid dry, toxic and dangerous zones. Our data suggest that root navigation is controlled via computations accomplished at the root apex synapses and associated with electric activities (Masi et al., 2009).

Plants are emerging as excellent biological computational systems. For example, leaves maintain stable temperature near their surfaces despite large fluctuations of temperature in the atmosphere (Helliker and Richter, 2008; Pincebourde and Woods, 2012). They relay in leaf stomata which acts as plant thermostats tissue, with individual stomata acting as autonomous units showing collective behavior (Hetherington and Woodward, 2003; Peak et al., 2004). In the case of plant leaves, stomata are simultaneously the sensors of external information, the processing units that calculate gas exchange rates and sensitively regulate their controls. Plants solved the dilemma of optimal gas exchanges via elegant parsimonious computational techniques in which input, output, and processing are all accomplished by using the same hardware.

Additional nice examples of plant computation include the ability of plants to compute starch synthesis and degradation rates (Scialdone et al., 2013; Webb and Satake, 2015), root apex computation of numerous abiotic and biotic parameters to navigate optimally root growth in complex environment of patchy soil (Baluška et al., 2009a,b, 2010; Masi et al., 2009; Baluška and Mancuso, 2013), as well as computations accomplished via Dionea leaf traps (Volkov et al., 2010; Böhm et al., 2016). Action potentials are relevant for most (perhaps all) of plant-specific computations (Masi et al., 2009; Volkov et al., 2010; Böhm et al., 2016; Hedrich et al., 2016).

In the root apex transition zone, cells and their membranes oscillate in almost all their activities (Baluška and Mancuso, 2013). These root apex transition zones resemble presomitic mesoderm segmentation clocks underlying vertebrate embryo segmentation (Moreno-Risueno et al., 2010; Traas and Vernoux, 2010; Moreno-Risueno and Benfey, 2011).

ANIMAL CELL PHYSIOLOGY AS INFORMATION PROCESSING

A number of non-neural cells have been shown to exhibit memory, with respect to somatic position (Carlson, 1983; Chang et al., 2002; McCusker and Gardiner, 2014) or differentiation (Xiong and Ferrell, 2003), implemented via long-term stable changes in bioelectric state (Marder et al., 1996; Turrigiano et al., 1996; Rosen and Cohen, 2006) and transcriptional profile (Kragl et al., 2009; Wang et al., 2009). These are now beginning to be understood via physiological modeling and dynamical systems theory that views memories as attractors in transcriptional, bioelectric, or epigenetic state space (Huang et al., 2005; Cervera et al., 2015; Law and Levin, 2015).

Moving up in organization, several tissues have been suggested to exhibit memory. One is bone, which has many similarities to a neural network, both molecularly and functionally (Turner et al., 2002). For example, the neurotransmitter glutamate plays a role in cell-to-cell communication among bone cells. Glutamate of course is a key neurotransmitter for learning and memory in the hippocampus. Bone cells exhibit habituation (to repeated mechanical stimuli) and sensitization (to mechanical loading) two of the most basic components of memory. Skull bones react quite differently to mechanical loading and hormones than do long bones, and it has been speculated that the past history of weight bearing imparts long-term cellular memory to the bone cell network, manifesting as differential responses to a variety of stimuli. A model involving long-term potentiation via the NMDA receptor has been proposed to explain memory of past stresses, and its subsequent influence over growth control, has been proposed (Spencer and Genever, 2003; Ho et al., 2005). Muscle comprises of some of the largest cells of animals, and also process, store and retrieve information via muscle-specific memory which can last from 15 years up to the entire lifetime in humans (Bruusgaard et al., 2010; Gundersen, 2016).

A most interesting set of studies have examined the phenomenon of cardiac memory. This is a clinically important pathway, in which specific changes of heartbeat pattern can persist stably (Otani and Gilmour, 1997; Goldberger and Kadish, 1999; Rosen and Plotnikov, 2002). This phenomenon has been modeled as a simple memory-like quantity that determines the relationship among the durations and amplitudes of action potentials (Otani and Gilmour, 1997). Most importantly, a specific mathematical model has been proposed for cardiac memory, taken after Hebbian plasticity in the brain (Chakravarthy and Ghosh, 1997; Zoghi, 2004).

The most recent addition to this body of work is the study of pancreas physiology (Goel and Mehta, 2013), which studied gap junctions (electrical synapses used for ionic communication in the brain, heart, and other organs), and their role in secretion of insulin from the pancreatic islets of Langerhans in response to glucose stimulation. Gap junctions synchronize oscillations of resting potentials among beta cells, and thus control insulin secretion. Past measurements of gap junctional conductance was unable to explain systemic properties, such as diminished junctional coupling in type-2 diabetes. In contrast to the prevailing tendency to focus on bottom-up views of the molecules involved and their interactions, Goel and Mehta viewed the process top-down, as a learning-like adaptation. Modeling gap junctions as links in a network of beta cells, subjected to homeostatic plasticity, they elucidated the systemlevel properties of this tissue, explaining why reductions in gap junction-mediated coupling in diabetes is necessary for an increase in blood insulin levels following hyperglycemia. It is not yet known if these mechanisms also underlie classical studies by Pavlov and others (Gantt, 1974, 1981; Gantt et al., 1987) on the classical conditioning of body organs to sugar, adrenaline, histamine, and other physiological stimuli.

We next consider larger-scale multicellular systems, at the level of organs or whole bodyplans (Levin, 2012b). Many species' bodies exhibit pattern memory during regeneration (Baddour et al., 2012; Lobo et al., 2014), and even transplanted organs maintain spatial information, such as transplanted eyes which send out optical axons to penetrate the brain on the side corresponding to its former location in a donor animal (Koo and Graziadei, 1995). It should be noted that one challenge to multicellularity is the ever-present danger of cancer - defection of somatic cells from the anatomical goals of the organism toward more primitive "every man for himself" behavior of individual cells and tumors (Johnston et al., 1992; Vincent, 2012; Chen and He, 2016). The interplay between the tumor and host has been analyzed using game theory (Dingli et al., 2009; McEvoy, 2009), consistent with each being an autonomous system with internal and external information channels, goals, and functional capabilities. Control networks regulating cancer have been analyzed from the perspective of learning (Gyurko et al., 2013), which represents an interesting new area for further research. Interestingly, recent data implicate in carcinogenesis the same bioelectric mechanisms that orchestrate pattern regulation and keep cells away from tumorigenesis (Chernet B. and Levin, 2013; Chernet B.T. and Levin, 2013; Yang and Brackenbury, 2013; Bates, 2015).

SOMATIC PATTERN MEMORIES: NON-NEURAL BIOELECTRICITY

The first task of any animal body is to assemble the progeny of a fertilized egg cell into a specific 3-dimensional pattern during embryogenesis. Then comes the need to maintain anatomical integrity over the lifespan, despite individual cell senescence, injury, and neoplastic conversion. Thus, long before animals developed brains to execute adaptive behaviors, cells had to have ways to coordinate their activity in an exquisite ballet that self-assembles, and then continuously remodels and repairs, a complex anatomical form. Some animals (e.g., salamanders) can regenerate their limbs, eyes, jaws, hearts, and portions of the brain (Sanchez Alvarado and Tsonis, 2006). Mammals have reduced powers of regeneration, but deer regenerate antler bone (adding up to 1 cm per day) every year, while humans regenerate their livers, and children regenerate their fingertips. Tails grafted onto the sides of salamanders slowly remodel into limbs (a structure more appropriate to their new location), and mammalian embryos can be split in half or combined together, resulting in normal embryos (reviewed in (Mustard and Levin, 2014)). All of these capabilities require significant information storage and processing, and many take place prior to (or without) the presence of the CNS.

Embryogenesis, regeneration, and metamorphosis stop precisely when the correct anatomical shape has been produced; this is a process akin to goal-directed behaviors, in the sense that the system can pursue multiple paths toward the same (anatomical) goal state, can accommodate unpredictable external perturbations (is not hardwired but flexible), and rests when it is satisfied (can recognize when its goal is achieved). All of these examples show the remarkable information processing that cells carry out, in order to create and maintain specific shapes (Levin, 2012b). Analogously to how brains implement goal-seeking behavior via information processing, non-neural cell networks process information about current and future anatomical shape. While the brain operates muscles and glands in service of activity in ecological space, the computational processes of non-neural somatic networks control cell behaviors (differentiation, migration, proliferation) to optimize the body's movement through morphospace (Stone, 1997; Rasskin-Gutman and Izpisua-Belmonte, 2004; Newman and Bhat, 2009).

A primary goal of developmental biology, synthetic bioengineering, and regenerative medicine is to learn to understand and control patterning networks, for applications in birth defects, organ regeneration, and cancer reprogramming (Ingber and Levin, 2007; Doursat et al., 2013). In particular it is crucial to tame the endogenous closed-loop pattern regulatory systems (flexible remodeling and regeneration pathways), as these offer the opportunity to exploit modularity to achieve needed changes in growth and form without micromanaging the details. What mechanisms underlie the ability of tissues to measure large-scale shape, detect deviations from a "remembered" correct target morphology, implement remodeling toward repairing that shape, and know when to stop (Levin, 2011)? Recent work has shown that as in the brain, these control networks make use of ion channels, gap junctions (electrical synapses), and neurotransmitters (Levin, 2012a; Tseng and Levin, 2013). A parsimonious hypothesis is that this is no coincidence, and that the brain learned its prodigious computational tricks from far more ancient pathways, co-opting developmental bioelectricity and optimizing it for the speed needed for behavior. While the brain operates on millisecond-scale bioelectric spiking, developmental bioelectricity involves steady, slow changes in ion fluxes, resting potentials, and electric fields.

A long history of work implicated bioelectric events in patterning (Jaffe, 1981; Nuccitelli, 2003; McCaig et al., 2005). Recent advances in molecular physiology have revealed that gap junctions, ion channels, and neurotransmitter pathway molecules – workhorses of cognitive processes in the CNS – are broadly expressed throughout the body, beginning prior to fertilization. Analogously to the brain, non-neural tissues continuously regulate resting potential (V_{mem}) and local field potentials (extracellular electric fields), as well as regulate the movement of neurotransmitters among cells (Pullar, 2011; Bates, 2015).

Signaling mediated by bioelectric events plays a crucial, instructive role in pattern formation (Funk, 2013; Levin, 2014b). Ion channel-mediated changes in V_{mem} not only affects individual cell behaviors such as proliferation, differentiation, apoptosis, and migration (Sundelacruz et al., 2009), but also determines large-scale parameters such as organ size, shape, and axial patterning of the entire body (Beane et al., 2011; Perathoner et al., 2014). In a range of model systems, V_{mem} regulates the formation of the brain, eye, wing, and face, and controls patterning along the anterior-posterior and left-right axes during embryonic development (Levin et al., 2002; Dahal et al., 2012; Pai et al., 2015a). Moreover, experimental control of bioelectric gradients has enabled induction of regenerative ability in non-regenerative contexts (Tseng et al., 2010; Leppik et al., 2015), induced reprogramming of gut tissue into complete eyes (Pai et al., 2012), and normalized tumors (Chernet B. and Levin, 2013; Chernet B.T. and Levin, 2013). Electrical synapses (gap junctions, GJs) and neurotransmitters like serotonin are a key component of several patterning systems, having been

implicated in embryonic left–right asymmetry, bone patterning, tumor suppression, and brain size control (Levin and Mercola, 1998; Iovine et al., 2005; Chernet et al., 2015; Pai et al., 2015a). As in the brain, these elements often work together, such as the bioelectrically controlled movement of serotonin through GJs during left-right patterning and control of nerve growth (Levin et al., 2006; Blackiston et al., 2015). The molecular pieces are now being identified, but the idea of neurotransmitters being ancient "pre-nervous" developmental signaling molecules is an old one (Buznikov and Shmukler, 1981).

The analogy between the brain and somatic pattern control (Figure 2) makes several specific predictions. One is that ion channels, GJs, and neurotransmitters should play a role in development; this has been amply demonstrated by the identification of patterning channelopathies (Levin, 2013), functional experiments in regenerative and developmental biology (Stewart et al., 2007), and the teratogenic effects of numerous psychoactive drugs (Hernandez-Diaz and Levin, 2014). Another key prediction concerns the encoding of instructive information. In the brain, genetics establish the hardware - genes encode the available components and thus define the limits of cellular activity. However, the information content of the brain is not directly encoded by the genome, but rather arises dynamically through environmental stimuli (learning) and self-organizing dynamics of the electrochemical circuitry (plasticity). Is this the case in pattern formation as well?

Can "long term somatic memory" be edited, in the context of a wild-type genome, leading to a permanent change? A first example of this was shown in a different species of planaria (Nogi and Levin, 2005), where targeting GJs for just 48 hours in a chunk of tissue caused it to regenerate 2 heads - one at the former anterior end (normal), and one at the posterior-facing end (which would normally grow a tail). Strikingly, these 2headed worms continue to regenerate as 2-headed when cut in subsequent rounds of regeneration, in plain water, months after the GJ blocking reagent is long gone from the tissue (Oviedo et al., 2010). The target morphology - the shape to which this animal regenerates upon damage - has been permanently re-written by temporarily editing the physiological network. This finding has clear similarity to plasticity [well-known to be exhibited by electrical synapses (Pereda et al., 2013)]: a brief induced change of GJ connectivity becomes stabilized to a long-term change (Levin, 2014a). This interaction between bioelectric activity and voltagegated GJs makes developmental bioelectrical networks especially suitable as a labile yet stable memory medium (Palacios-Prado and Bukauskas, 2009). Another brain-like property exhibited in this effect is its holographic nature: in each round of cutting, the ectopic head (perhaps "epigenetically reprogrammed") is removed, and a middle fragment of the gut still knows it must make 2 heads if cut. The patterning information is distributed non-locally throughout the network.

This field is advancing rapidly in its mechanistic details at the cellular level: the genetics of endogenous ion channels causing the gradients, the transduction mechanisms that control transcription after V_{mem} change, and the gene expression changes downstream of bioelectrical signaling are all becoming clear (Yang and Brackenbury, 2013; Pai et al., 2015b). Techniques,

such as optogenetics (Adams et al., 2013, 2014), are allowing imposition of specific voltage patterns onto tissue *in vivo*. As in the brain, where optogenetics is used to insert memories directly into brains (Ramirez et al., 2013; Liu et al., 2014), these techniques will be crucial to learn to rewrite pattern memories during regeneration or embryogenesis. However, as in neuroscience, there is more than one level at which progress needs to be made. A mature understanding of the brain requires synthesis of data from people working on the genetics and biochemistry of specific neurotransmitter receptors and their downstream molecular signaling, with the insights of workers at the level of circuits, behavior, cognitive science, and psychology.

Classic work explored the extensive parallels between chemical gradients during development and signal processing in the visual system (Grossberg, 1978), and indeed early quantitative models of patterning (explaining self-regulatory features like proportion regulation) were based on visual system function (Hartline et al., 1956; Gierer and Meinhardt, 1972). More recent efforts include the notion of memory for position during regeneration (Chang et al., 2002; Kragl et al., 2009; Wang et al., 2009) and development (Beloussov, 1997) and for signaling hysteresis during development (Balaskas et al., 2012), excitable cortex memory models of pseudopod dynamics (Cooper et al., 2012), and neural network models of chemical signaling (Ling et al., 2013) (which showed formal isomorphisms between gene regulation networks and Hebbian learning in neural nets) (Watson et al., 2010; Ling et al., 2013). In addition to classical neuroscience concepts, more exotic group cognition models have been applied to patterning (Gunji and Ono, 2012), while a few recent studies investigated the decision-making and formal computational capabilities of RD systems - a chemical signaling modality often used to model morphogenesis (Adamatzky et al., 2003, 2008; Costello et al., 2009; Dale and Husbands, 2010, which is now known to be Turing-complete (Scarle, 2009) and support semantic interpretations (Schumann and Adamatzky, 2009). Despite these fascinating efforts to identify elements of cognitivelike processing in well-known elements of pattern formation, developmental biology is still firmly centered in a mechanistic perspective, seeking explanations in terms of pathways and not information (systems that know things and make decisions based on that understanding). However, it is crucial to note that attributing true knowledge and memory to biological systems is not mystical thinking - computational neuroscience shows us a clear proof of concept that information-level, cognitive approaches to cellular networks are viable, and in fact necessary, strategy for understanding a system at all of its salient levels.

Thus, neuroscience offers developmental biology more than just tools and molecular mechanisms: it offers a unique paradigm, otherwise unavailable to molecular and cell biologists, of the emergence of higher levels of organization that have both causal potency and experimental tractability. The field is in need of new formalisms and conceptual tools for linking the dynamics of physiological circuits with downstream patterning outcomes. Developmental biology is currently focused entirely in a bottom-up mode, with molecules being the preferred level of explanation. Neuroscience teaches us that we must look upward as well as downward, for emergent levels with their own rules



FIGURE 2 | Parallelism between neural and somatic computational systems. Complex, flexible, goal-seeking behavior (A) is implemented by information processing in the brain (B), which consists of networks of electrically communicating neural cells networks executing physiological circuits (C), which operate because of electrically gated ion channel and electrical synapse proteins (D). Similarly, large-scale goal-directed pattern remodeling and regeneration (E) occurs via bioelectric gradients that coordinate cell activity (F), implemented by physiological circuits in non-neural cells (G) which operate because of the same set of ion channels and electrical synapses (H). The behavior of these systems at the lowest level is achieved by regulating gap junction state and ion channel activity in specific cells (I). Circuit activity is beginning to be tractable in both contexts using optogenetics (J). In behavioral settings, the most effective path toward desired outcomes is to interact with the system at the highest level, rewarding for desired behavior (K). This strategy remains to be tried in patterning contexts, where the current paradigm has been focused on bottom-up approaches and has not yet investigated the top-down strategies that have paid off so well for cognitive science. (A-H) drawn by Alexis Pietak. (I,K) drawn by Jeremy Guay of Peregrine Creative. (J) used with permission from Liu et al. (2014).





and advantages (Friston et al., 2015). For example, training an animal to a particular complex behavior is far more efficient than attempting to elicit the same behavior by manipulating individual neurons in their brains. We now know that beneficial changes at the genetic and chemical levels can be induced by *cognitive* therapies – top–down control of tissue structure and function induced by specific thoughts and experiences. If patterning tissues are "primitive cognitive agents", in the sense that they can be profitably understood as memory-bearing, information

processing, goal-directed cybernetic systems (Pezzulo and Levin, 2015), then a whole new set of approaches becomes available for regenerative medicine. If we understood the bioelectric code, we could interact with it at these higher levels of organization, taking advantage of endogenous modularity and perhaps rationally controlling anatomical outcomes without having to micromanage molecular networks. In this field, cognitive science, unconventional computation, and developmental biology intersect. A fundamental open direction is the search for

a computational pipeline to extract goal patterns from bioelectric state data, parallel to efforts to extract image data from brain measurements (Nishimoto et al., 2011). The flow of knowledge will likely not all be unidirectional: cracking the bioelectric code in patterning tissues is likely to in turn benefit fundamental neuroscience by showing, in perhaps a simpler context how to extract semantic content from bioelectrical cell states in the brain.

CONCLUSION

How does biological matter give rise to decision-making, representation, and goal-directed memory, activity? Implementation-independence is a core principle of computer science: an algorithm does what it does regardless of what kind of medium is implementing the steps. However, in the biological sciences, the study of memory and other cognitive functions has largely been the province of neurobiology, which studies the information processing and computational functions of one type of system: collections of neurons. Instead, we have surveyed a broad range of systems at various scales, from molecular to organismal, which have their own distinct ability to process information, make decisions, and achieve specific goal states (Figure 3). Neural-like computation, decision-making, and memory have been reported in sperm (Alvarez et al., 2013), amoebae (Zhu et al., 2013), yeast (Caudron and Barral, 2013), and plants (Gagliano et al., 2014), using ubiquitous mechanisms like cytoskeletal elements which appear to be also involved in neural information processing (Sahu et al., 2013). It is clear that neural networks have no monopoly on such functions. Remarkably, it is not only the positive (adaptive) cognitive functions that are widely conserved: some of the same illusions to which advanced brains' perception and rational reasoning fall prey are being found in systems from slime molds to multi-animal colonies (Beekman and Latty, 2015; Sakiyama and Gunji, 2016).

McCulloch said "Why the mind is in the head? Because there, and only there, are hosts of possible connections to be performed as time and circumstance demand it" (McCulloch, 1951). Given the facts of protein, cytoskeletal, transcriptional, and bioelectric networks, it appears that many different media at various scales have the ability to form and rewire experiencedependent connections. The "dynamical hypothesis" (van Gelder, 1998) asks, what if the brain is better understood as a dynamical system, than a computational one? We invert this hypothesis, and ask what if some dynamical systems are better understood as cognitive agents? The appearance of memory and computation at many levels of biological organization suggests a fractal organization of cognitive subsystems within systems molecular, cellular, tissue, and body-wide (Figure 2). This has been suggested in the brain [Smythies' nested doll hypothesis, (Smythies, 2015)] but may indeed exist throughout the biological world. Whether each successive level of organization is in some sense smarter than the ones below it, or whether structures derive their cognitive powers from those of lower levels, remains to be discovered. It should be noted, however, that even in advanced brains, the relationship between cognitive capacity and biological

structure is not trivial to pin down, as shown by the occasional example of potent function in the presence of severe structural deficits (Lorber, 1978, 1981; Nahm et al., 2012).

The hypothesis of nested, widely prevalent cognitive layers suggests a rich research program, including: (1) the development of improved methods for reading/writing bioelectrical state information into somatic tissues and sculpting non-neural bioelectric circuits (optogenetics beyond excitable cells and in the synthetic biology of gap junction and neurotransmitter signaling; Adams et al., 2013), (2) continued work on cracking the bioelectric code (bioelectric state information maps onto the topology of various patterning outcomes in tractable model systems such as planaria; Tseng and Levin, 2013), (3) formulation and testing of quantitative, molecular models of LTP, habituation, sensitization, plasticity, and higherorder learning applied to protein interaction networks, gene regulatory circuits, cytoskeletal dynamics, and cell behavior during morphogenesis, (4) use of reagents that impact cognition (hallucinogens, anesthetics; Kawamoto et al., 2005), stimulants, nootropics/cognitive enhancers, etc.) in cellular, developmental, and regenerative patterning assays to probe conservation of pathways between neuroscience and morphogenesis, (5) creation of larger-scale computational models of regeneration and functional experiments in morphogenesis based on goal-seeking and error minimization algorithms with molecularly specified metrics (Slack, 1980; Chao et al., 2008), (6) exploration of molecular models of cognitive concepts (attention, autism spectrum, sleep, visual illusions/hallucinations, addiction) in specific patterning and mispatterning contexts, (7) bioengineering platforms that reward and punish in vitro patterning systems for specific changes in growth and morphogenesis (instrumental learning and top-down control of shape in developmental or regenerative contexts), and (8) a mechanistic investigation of the mechanism of persistence of memories through massive brain regeneration, which is likely to reveal the interface between somatic and neural memories (Blackiston et al., 2008; Shomrat and Levin, 2013).

We have avoided here the thorny issues of philosophy of mind that arise from trying to define exactly under what conditions words like "knowledge" are appropriate, in favor of an intentional stance-like pragmatic, engineering approach grounded in cybernetics. The coverage of cognitive terms across biology can expand to the extent that information-centered approaches are shown to be effective in predicting and controlling the behavior of biological systems. The practical implications for biotechnology, unconventional computation, and regenerative medicine are enormous. Equally likely, the lessons we learn from unconventional cognitive systems will deeply impact our most basic understanding of how mind emerges from the brain.

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ML and FB both contributed sections to the review according to their specialties. Both provided novel ideas, edited each other's text, and prepared the finished product.

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REFERENCES

- Adamatzky, A., and Alonso-Sanz, R. (2011). Rebuilding Iberian motorways with slime mould. *Biosystems* 105, 89–100. doi: 10.1016/j.biosystems.2011.03.007
- Adamatzky, A., Costello, B. D., Melhuish, C., and Ratcliffe, N. (2003). Experimental reaction-diffusion chemical processors for robot path planning. J. Intell. Robot. Syst. 37, 233–249. doi: 10.1023/A:1025414424756
- Adamatzky, A., Costello, B. D. L., and Shirakawa, T. (2008). Universal computation with limited resources: belousov-zhabotinsky and physarum computers. *Int. J. Bifurcat. Chaos* 18, 2373–2389. doi: 10.1142/S0218127408021750
- Adams, D. S. (2008). A new tool for tissue engineers: ions as regulators of morphogenesis during development and regeneration. *Tiss. Eng. Part A* 14, 1461–1468. doi: 10.1089/ten.tea.2008.0080
- Adams, D. S., Lemire, J. M., Kramer, R. H., and Levin, M. (2014). Optogenetics in developmental biology: using light to control ion flux-dependent signals in *Xenopus embryos. Int. J. Dev. Biol.* 58, 851–861. doi: 10.1387/ijdb.140207ml
- Adams, D. S., and Levin, M. (2012). General principles for measuring resting membrane potential and ion concentration using fluorescent bioelectricity reporters. *Cold Spring Harb. Prot.* 2012, 385–397. doi: 10.1101/pdb.top067710
- Adams, D. S., Tseng, A. S., and Levin, M. (2013). Light-activation of the Archaerhodopsin H+-pump reverses age-dependent loss of vertebrate regeneration: sparking system-level controls in vivo. *Biol. Open* 2, 306–313. doi: 10.1242/bio.20133665
- Agnati, L. F., Leo, G., Zanardi, A., Genedani, S., Rivera, A., Fuxe, K., et al. (2006). Volume transmission and wiring transmission from cellular to molecular networks: history and perspectives. *Acta Physiol.* 187, 329–344. doi: 10.1111/j.1748-1716.2006.01579.x
- Albrecht-Buehler, G. (1980). Autonomous movements of cytoplasmic fragments. Proc. Natl. Acad. Sci. U.S.A. 77, 6639–6643. doi: 10.1073/pnas.77.11.6639
- Albrecht-Buehler, G. (1982). Control of tissue cell movement. Natl. Cancer Inst. Monogr. 60, 117–122.
- Albrecht-Buehler, G. (1985). Is cytoplasm intelligent too? *Cell Muscl Motil.* 6, 1–21. doi: 10.1007/978-1-4757-4723-2_1
- Alvarez, L., Friedrich, B. M., Gompper, G., and Kaupp, U. B. (2013). The computational sperm cell. *Trends Cell Biol.* 24, 198–207. doi: 10.1016/j.tcb.2013.10.004
- Antov, Y., Barbul, A., Mantsur, H., and Korenstein, R. (2005). Electroendocytosis: exposure of cells to pulsed low electric fields enhances adsorption and uptake of macromolecules. *Biophys. J.* 88, 2206–2223. doi: 10.1529/biophysj.104.051268
- Applewhite, P. B. (1972). The flow of ions in learning and memory. J. Theor. Biol. 36, 419–423. doi: 10.1016/0022-5193(72)90109-9
- Applewhite, P. B., Lapan, E. A., and Gardner, F. T. (1969). Protozoan habituation learning after loss of macronuclei and cytoplasm. *Nature* 222, 491–492. doi: 10.1038/222491a0
- Aragonès, A. C., Haworth, N. L., Darwish, N., Ciampi, S., Bloomfield, N. J., Wallace, G. G., et al. (2016). Electrostatic catalysis of a Diels–Alder reaction. *Nature* 531, 88–91. doi: 10.1038/nature16989
- Aur, D. (2012). From neuroelectrodynamics to thinking machines. *Cogn. Comput.* 4, 4–12. doi: 10.1007/s12559-011-9106-3
- Baddour, J. A., Sousounis, K., and Tsonis, P. A. (2012). Organ repair and regeneration: an overview. Birth Def. Res. C Embr. Today 96, 1–29. doi: 10.1002/bdrc.21006
- Balaskas, N., Ribeiro, A., Panovska, J., Dessaud, E., Sasai, N., Page, K. M., et al. (2012). Gene regulatory logic for reading the Sonic Hedgehog signaling gradient in the vertebrate neural tube. *Cell* 148, 273–284. doi: 10.1016/j.cell.2011.10.047
- Baluška, F. (2010). Recent surprising similarities between plant cells and neurons. *Plant Signal. Behav.* 5, 87–89. doi: 10.4161/psb.5.2.11237
- Baluška, F., and Mancuso, S. (2009). Deep evolutionary origins of neurobiology: turning the essence of 'neural' upside-down. *Commun. Integr. Biol.* 2, 60–65. doi: 10.4161/cib.2.1.7620

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- Baluška, F., and Mancuso, S. (2013). Root apex transition zone as oscillatory zone. *Front. Plant Sci.* 4:354. doi: 10.3389/fpls.2013.00354
- Baluška, F., Mancuso, S., Volkmann, D., and Barlow, P. W. (2004). Root apices as plant command centres: the unique 'brain-like' status of the root apex transition zone. *Biologia* 59, 9–17.
- Baluška, F., Mancuso, S., Volkmann, D., and Barlow, P. W. (2009a). The 'root-brain' hypothesis of Charles and Francis Darwin: revival after more than 125 years. *Plant Signal. Behav.* 4, 1121–1127. doi: 10.4161/psb.4.12. 10574
- Baluška, F., Mancuso, S., Volkmann, D., and Barlow, P. W. (2010). Root apex transition zone: a signalling – response nexus in the root. *Trends Plant Sci.* 15, 402–408. doi: 10.1016/j.tplants.2010.04.007
- Baluška, F., Šamaj, J., and Menzel, D. (2003). Polar transport of auxin: carriermediated flux across the plasma membrane or neurotransmitter-like secretion? *Trends Cell Biol.* 13, 282–285. doi: 10.1016/S0962-8924(03)00084-9
- Baluška, F., Schlicht, M., Volkmann, D., and Mancuso, S. (2008). Vesicular secretion of auxin: evidences and implications. *Plant Signal. Behav.* 3, 254–256. doi: 10.4161/psb.3.4.5183
- Baluška, F., Schlicht, M., Wan, Y.-L., Burbach, C., and Volkmann, D. (2009b). Intracellular domains and polarity in root apices: from synaptic domains to plant neurobiology. *Nova Acta Leopold* 96, 103–122.
- Baluška, F., Volkmann, D., and Menzel, D. (2005). Plant synapses: actin-based adhesion domains for cell-to-cell communication. *Trends Plant Sci.* 10, 106– 111. doi: 10.1016/j.tplants.2005.01.002
- Baluška, F., and Wan, Y. L. (2012). "Physical control overendocytosis," in Endocytosis in Plants, ed. J. Šamaj (Berlin: Springer-Verlag), 123–149.
- Bancroft, W. D., and Richter, G. H. (1930). Claude Bernard's theory of narcosis. Proc. Natl. Acad. Sci. U.S.A. 16, 573–577. doi: 10.1073/pnas.16.9.573
- Barlow, P. W., and Baluška, F. (2000). Cytoskeletal perspectives on root growth and morphogenesis. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* 51, 289–322. doi: 10.1146/annurev.arplant.51.1.289
- Bates, E. (2015). Ion channels in development and cancer. Annu. Rev. Cell. Dev. Biol. 31, 231–247. doi: 10.1146/annurev-cellbio-100814-125338
- Beagle, S. D., and Lockless, S. W. (2015). Microbiology: electrical signalling goes bacterial. *Nature* 527, 44–45. doi: 10.1038/nature15641
- Beane, W. S., Morokuma, J., Adams, D. S., and Levin, M. (2011). A chemical genetics approach reveals H,K-ATPase-mediated membrane voltage is required for planarian head regeneration. *Chem. Biol.* 18, 77–89. doi: 10.1016/j.chembiol.2010.11.012
- Becchetti, A., Pillozzi, S., Morini, R., Nesti, E., and Arcangeli, A. (2010). New insights into the regulation of ion channels by integrins. *Int. Rev. Cell. Mol. Biol.* 279, 135–190. doi: 10.1016/S1937-6448(10)79005-5
- Beekman, M., and Latty, T. (2015). Brainless but multi-headed: decision making by the acellular slime mould *Physarum polycephalum*. J. Mol. Biol. 427, 3734–3743. doi: 10.1016/j.jmb.2015.07.007
- Beloussov, L. V. (1997). On the active memory in developing systems. *Riv. Biol.* 90, 31–46.
- Ben-Jacob, E. (2009). Learning from bacteria about natural information processing. Ann. N. Y. Acad. Sci. 1178, 78–90. doi: 10.1111/j.1749-6632.2009.05022.x
- Bernard, C. (1878). Lectures on Phenomena of Life Common to Animals and Plants. Paris: Ballliere, and Son.
- Bezanilla, F. (2002). Voltage sensor movements. J. Gen. Physiol. 120, 465–473. doi: 10.1085/jgp.20028660
- Bezanilla, F. (2006). The action potential: from voltage-gated conductances to molecular structures. *Biol. Res.* 39, 425–435. doi: 10.4067/S0716-97602006000300005
- Bezanilla, F. (2008). How membrane proteins sense voltage. Nat. Rev. Mol. Cell. Biol. 9, 323–332. doi: 10.1038/nrm2376
- Bharti, K., and Arnheiter, H. (2005). When pigment cells turn into neurons. *J. Invest. Dermatol.* 125, 10–11. doi: 10.1111/j.0022-202X.2005.23876.x

- Blackiston, D. J., Anderson, G. M., Rahman, N., Bieck, C., and Levin, M. (2015). A novel method for inducing nerve growth via modulation of host resting potential: gap junction-mediated and serotonergic signaling mechanisms. *Neurotherapeutics* 12, 170–184. doi: 10.1007/s13311-014-0317-7
- Blackiston, D. J., Silva Casey, E., and Weiss, M. R. (2008). Retention of memory through metamorphosis: can a moth remember what it learned as a caterpillar? *PLoS ONE* 3:e1736. doi: 10.1371/journal.pone.0001736
- Böhm, J., Scherzer, S., Krol, E., Kreuzer, I., von Meyer, K., Lorey, C., et al. (2016). The Venus flytrap *Dionaea muscipula* counts prey-induced action potentials to induce sodium uptake. *Curr. Biol.* 26, 286–295. doi: 10.1016/j.cub.2015.11.057
- Bose, J. (1926). Response in the Living and the Non-Living. London: Longmans, Green & Co.
- Brunet, T., and Arendt, D. (2016). From damage response to action potentials: early evolution of neural and contractile modules in stem eukaryotes. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 371, 1685. doi: 10.1098/rstb.2015.0043
- Bruusgaard, J. C., Johansen, I. B., Egner, I. M., Rana, Z. A., and Gundersen, K. (2010). Myonuclei acquired by overload exercise precede hypertrophy and are not lost on detraining. *Proc. Natl. Acad. Sci. U.S.A.* 107, 15111–15116. doi: 10.1073/pnas.0913935107
- Burbach, C., Markus, K., Yin, Z., Schlicht, M., and Baluška, F. (2012). Photophobic behaviour of maize roots. *Plant Signal. Behav.* 7, 876–880. doi: 10.4161/psb.21012
- Buznikov, G., Shmukler, Y., and Lauder, J. (1996). From oocyte to neuron: do neurotransmitters function in the same way throughout development? *Cell Mol. Neurobiol.* 16, 537–559. doi: 10.1007/BF02152056
- Buznikov, G. A., and Shmukler, Y. B. (1981). Possible role of "prenervous" neurotransmitters in cellular interactions of early embryogenesis: a hypothesis. *Neurochem. Res.* 6, 55–68. doi: 10.1007/BF00963906
- Calvo, P. (2016). The philosophy of plant neurobiology: a manifesto. *Synthese* 193, 1323–1343. doi: 10.1007/s11229-016-1040-1
- Calvo, P., and Baluška, F. (2015). Conditions for minimal intelligence across eukaryota: a cognitive science perspective. *Front. Psychol.* 6:1329. doi: 10.3389/fpsyg.2015.01329
- Carlson, B. M. (1983). Positional memory in vertebrate limb development and regeneration. *Prog. Clin. Biol. Res.* 110, 433-443.
- Carneiro, K., Donnet, C., Rejtar, T., Karger, B. L., Barisone, G. A., Diaz, E., et al. (2011). Histone deacetylase activity is necessary for left-right patterning during vertebrate development. *BMC Dev. Biol.* 11:29. doi: 10.1186/1471-213X-11-29
- Carter, J. H. (2000). The immune system as a model for pattern recognition and classification. J. Am. Med. Inform. Assoc. 7, 28–41. doi: 10.1136/jamia.2000.0070028
- Caudron, F., and Barral, Y. (2013). A super-assembly of Whi3 encodes memory of deceptive encounters by single cells during yeast courtship. *Cell* 155, 1244–1257. doi: 10.1016/j.cell.2013.10.046
- Cervera, J., Alcaraz, A., and Mafe, S. (2014). Membrane potential bistability in nonexcitable cells as described by inward and outward voltage-gated ion channels. *J. Phys. Chem. B* 118, 12444–12450. doi: 10.1021/jp508304h
- Cervera, J., Manzanares, J. A., and Mafe, S. (2015). Electrical coupling in ensembles of nonexcitable cells: modeling the spatial map of single cell potentials. *J. Phys. Chem. B* 119, 2968–2978. doi: 10.1021/jp512900x
- Chakravarthy, S. V., and Ghosh, J. (1997). On Hebbian-like adaptation in heart muscle: a proposal for 'cardiac memory'. *Biol. Cybern.* 76, 207–215. doi: 10.1007/s004220050333
- Chang, H. Y., Chi, J. T., Dudoit, S., Bondre, C., van de Rijn, M., Botstein, D., et al. (2002). Diversity, topographic differentiation, and positional memory in human fibroblasts. *Proc. Natl. Acad. Sci. U.S.A.* 99, 12877–12882. doi: 10.1073/pnas.162488599
- Chao, Z. C., Bakkum, D. J., and Potter, S. M. (2008). Shaping embodied neural networks for adaptive goal-directed behavior. *PLoS Comp. Biol.* 4:e1000042. doi: 10.1371/journal.pcbi.1000042
- Chen, H., and He, X. (2016). The convergent cancer evolution toward a single cellular destination. *Mol. Biol. Evol.* 33, 4–12. doi: 10.1093/molbev/msv212
- Chernet, B., and Levin, M. (2013). Endogenous voltage potentials and the microenvironment: bioelectric signals that reveal, induce and normalize cancer. *J. Clin. Exp. Oncol. Suppl.* 1, S1–S2.
- Chernet, B. T., Fields, C., and Levin, M. (2015). Long-range gap junctional signaling controls oncogene-mediated tumorigenesis in *Xenopus laevis* embryos. *Front. Physiol.* 5:519. doi: 10.3389/fphys.2014.00519

- Chernet, B. T., and Levin, M. (2013). Transmembrane voltage potential is an essential cellular parameter for the detection and control of tumor development in a *Xenopus* model. *Dis. Mod. Mech.* 6, 595–607. doi: 10.1242/dmm. 010835
- Chernet, B. T., and Levin, M. (2014). Transmembrane voltage potential of somatic cells controls oncogene-mediated tumorigenesis at long-range. *Oncotarget* 5, 3287–3306. doi: 10.18632/oncotarget.1935
- Clark, K. B. (2013). Ciliates learn to diagnose and correct classical error syndromes in mating strategies. *Front. Microbiol.* 4:229. doi: 10.3389/fmicb.2013.00229
- Cohen, I. R. (1992a). The cognitive paradigm and the immunological homunculus. Immunol. Today 13, 490–494. doi: 10.1016/0167-5699(92)90024-2
- Cohen, I. R. (1992b). The cognitive principle challenges clonal selection. *Immunol. Today* 13, 441–444. doi: 10.1016/0167-5699(92)90071-E
- Conrad, M. (1996). Cross-scale information processing in evolution, development and intelligence. *BioSystems* 38, 97–109. doi: 10.1016/0303-2647(95)01579-5
- Cooper, R. M., Wingreen, N. S., and Cox, E. C. (2012). An excitable cortex and memory model successfully predicts new pseudopod dynamics. *PLoS ONE* 7:e33528. doi: 10.1371/journal.pone.0033528
- Corning, W. C. (1966). Retention of a position discrimination after regeneration in planarians. *Psychol. Sci.* 5, 17–18. doi: 10.3758/BF03328256
- Corssen, G., and Skora, I. A. (1964). "Addiction" reactions in cultured Hhuman cells. *JAMA* 187, 328–332. doi: 10.1001/jama.1964.03060180014003
- Costello, B. D., Toth, R., Stone, C., Adamatzky, A., and Bull, L. (2009). Implementation of glider guns in the light-sensitive Belousov-Zhabotinsky medium. *Phys. Rev. Stat. Nonlin Soft. Matter Phys.* 79(Pt 2), 026114. doi: 10.1103/PhysRevE.79.026114
- Couzin, I. D. (2009). Collective cognition in animal groups. *Trends Cogn. Sci.* 13, 36–43. doi: 10.1016/j.tics.2008.10.002
- Craddock, T. J., Tuszynski, J. A., and Hameroff, S. (2012). Cytoskeletal signaling: is memory encoded in microtubule lattices by CaMKII phosphorylation? *PLoS Comp. Biol.* 8:e1002421. doi: 10.1371/journal.pcbi.1002421
- Craddock, T. J., Tuszynski, J. A., Priel, A., and Freedman, H. (2010). Microtubule ionic conduction and its implications for higher cognitive functions. J. Integr. Neurosci. 9, 103–122. doi: 10.1142/S0219635210002421
- Dahal, G. R., Rawson, J., Gassaway, B., Kwok, B., Tong, Y., Ptacek, L. J., et al. (2012). An inwardly rectifying K+ channel is required for patterning. *Development* 139, 3653–3664. doi: 10.1242/dev.078592
- Dale, K., and Husbands, P. (2010). The evolution of reaction-diffusion controllers for minimally cognitive agents. *Artif. Life* 16, 1–19. doi: 10.1162/artl.2009.16.1.16100
- Daoudal, G., and Debanne, D. (2003). Long-term plasticity of intrinsic excitability: learning rules and mechanisms. *Learn. Memory* 10, 456–465. doi: 10.1101/lm.64103
- Darwin, C. (1880). The Power of Movement in Plants. London: John Murray.
- De Luccia, T. P. (2012). Mimosa pudica, *Dionaea muscipula* and anesthetics. *Plant Signal. Behav.* 7, 1163–1167. doi: 10.4161/psb.21000
- DeMarse, T. B., and Dockendorf, K. P. (2005). Adaptive flight control with living neuronal networks on microelectrode arrays. *Proc. IEEE Explor. IJCNN* 3, 1548–1551.
- Dingli, D., Chalub, F. A., Santos, F. C., Van Segbroeck, S., and Pacheco, J. M. (2009). Cancer phenotype as the outcome of an evolutionary game between normal and malignant cells. *Br. J. Cancer.* 101, 1130–1136. doi: 10.1038/sj.bjc. 6605288
- Doursat, R., Sayama, H., and Michel, O. (2013). A review of morphogenetic engineering. Nat. Comput. 12, 517–535. doi: 10.1007/s11047-013-9398-1
- Dranias, M. R., Ju, H., Rajaram, E., and Van Dongen, A. M. (2013). Short-term memory in networks of dissociated cortical neurons. J. Neurosci. 33, 1940–1953. doi: 10.1523/JNEUROSCI.2718-12.2013
- Eisenstein, E. M. (1967). "The use of invertebrate systems for studies on the bases of learning and memory," in *The Neurosciences—A Study Program*, eds G. C. Quarton, T. Melnechuk, and F. U. Schmitt (New York, NY: Rockefeller University Press), 653–665.
- Fernando, C., Szathmary, E., and Husbands, P. (2012). Selectionist and evolutionary approaches to brain function: a critical appraisal. *Front. Comput. Neurosci.* 6:24. doi: 10.3389/fncom.2012.00024
- Filoni, S., Velloso, C. P., Bernardini, S., and Cannata, S. M. (1995). Acquisition of nerve dependence for the formation of a regeneration blastema in amputated

hindlimbs of larval *Xenopus laevis*: the role of limb innervation and that of limb differentiation. *J. Exp. Zool.* 273, 327–341. doi: 10.1002/jez.1402730407

- Forrest, M. D. (2014). The sodium-potassium pump is an information processing element in brain computation. *Front. Physiol.* 5:472. doi: 10.3389/fphys.2014.00472
- Francis, N. J., and Kingston, R. E. (2001). Mechanisms of transcriptional memory. Nat. Rev. Mol. Cell. Biol. 2, 409–421. doi: 10.1038/35073039
- Freddolino, P. L., and Tavazoie, S. (2012). Beyond homeostasis: a predictivedynamic framework for understanding cellular behavior. Annu. Rev. Cell. Dev. Biol. 28, 363–384. doi: 10.1146/annurev-cellbio-092910-154129
- Friston, K. (2010). The free-energy principle: a unified brain theory? Nat. Rev. Neurosci. 11, 127–138. doi: 10.1038/nrn2787
- Friston, K., Levin, M., Sengupta, B., and Pezzulo, G. (2015). Knowing one's place: a free-energy approach to pattern regulation. J. R. Soc. Interface 12. doi: 10.1098/rsif.2014.1383

Funk, R. (2013). Ion gradients in tissue and organ biology. Biol. Syst. 2:1000105.

- Fuxe, K., Borroto-Escuela, D. O., Romero-Fernandez, W., Zhang, W. B., and Agnati, L. F. (2013). Volume transmission and its different forms in the central nervous system. *Chin. J. Integr. Med.* 19, 323–329. doi: 10.1007/s11655-013-1455-1
- Gagliano, M., Renton, M., Depczynski, M., and Mancuso, S. (2014). Experience teaches plants to learn faster and forget slower in environments where it matters. *Oecologia* 175, 63–72. doi: 10.1007/s00442-013-2873-7
- Gantt, W. H. (1974). Autokinesis, schizokinesis, centrokinesis and organ-system responsibility: concepts and definition. *Pavlov J. Biol. Sci.* 9, 187–191.
- Gantt, W. H. (1981). Organ-system responsibility, schizokinesis, and autokinesis in behavior. *Pavlov J. Biol. Sci.* 16, 64–66.
- Gantt, W. H., McGuigan, F. J., and Ban, T. A. (1987). Critical issues in Psychology, Psychiatry, and Physiology. Memorial to W. Horsley Gantt. New York, NY: Gordon and Breach Science Publishers.
- Gelber, B. (1962). Acquisition in Paramecium aurelia during spaced training. Psychol. Rec. 12, 165–177.
- Gentile, L., Cebria, F., and Bartscherer, K. (2011). The planarian flatworm: an in vivo model for stem cell biology and nervous system regeneration. *Dis. Model. Mech.* 4, 12–19. doi: 10.1242/dmm.006692
- Gierer, A., and Meinhardt, H. (1972). A theory of biological pattern formation. *Kybernetik* 12, 30–39. doi: 10.1007/BF00289234
- Goel, P., and Mehta, A. (2013). Learning theories reveal loss of pancreatic electrical connectivity in diabetes as an adaptive response. *PLoS ONE* 8:e70366. doi: 10.1371/journal.pone.0070366
- Goldberger, J. J., and Kadish, A. H. (1999). Cardiac memory. *Pacing Clin. Electrophysiol.* 22, 1672–1679. doi: 10.1111/j.1540-8159.1999.tb00388.x
- Goldman, S. A., Nedergaard, M., and Windrem, M. S. (2015). Modeling cognition and disease using human glial chimeric mice. *Glia* 63, 1483–1493. doi: 10.1002/glia.22862
- Goldsworthy, A. (1983). The evolution of plant action potentials. J. Theor. Biol. 103, 645–648. doi: 10.1016/0022-5193(83)90287-4
- Gremiaux, A., Yokawa, K., Mancuso, S., and Baluška, F. (2014). Plant anesthesia supports similarities between animals and plants: claude Bernard's forgotten studies. *Plant Signal. Behav.* 9:e27886. doi: 10.4161/psb.27886
- Grossberg, S. (1978). "Communication, memory, and development," in *Progress in Theoretical Biology*, eds R. Rosen and F. Snell (Amsterdam: Elsevier).
- Gundersen, K. (2016). Muscle memory and a new cellular model for muscle atrophy and hypertrophy. J. Exp. Biol. 219, 235–242. doi: 10.1242/jeb. 124495
- Gunji, Y. P., and Ono, R. (2012). Sociality of an agent during morphogenetic canalization: asynchronous updating with potential resonance. *BioSystems* 109, 420–429. doi: 10.1016/j.biosystems.2012.05.005
- Gyurko, D. M., Veres, D. V., Modos, D., Lenti, K., Korcsmaros, T., and Csermely, P. (2013). Adaptation and learning of molecular networks as a description of cancer development at the systems-level: potential use in anti-cancer therapies. *Semin. Cancer Biol.* 23, 262–269. doi: 10.1016/j.semcancer.2013. 06.005
- Hamann, T. (2015). The plant cell wall integrity maintenance mechanism a case study of a cell wall plasma membrane signaling network. *Phytochemistry* 112, 100–109. doi: 10.1016/j.phytochem.2014.09.019
- Hameroff, S. R., and Watt, R. C. (1982). Information processing in microtubules. J. Theor. Biol. 98, 549–561. doi: 10.1016/0022-5193(82)90137-0

- Hamilton, T. C. (1975). "Behavioral plasticity in protozoans," in Aneural Organisms in Neurobiology, ed. E. M. Eisenstein (New York: Plenum Press), 111–130.
- Hartline, H. K., Wagner, H. G., and Ratliff, F. (1956). Inhibition in the eye of Limulus. J. Gen. Physiol. 39, 651–673. doi: 10.1085/jgp.39.5.651
- Hedrich, R., Salvador-Recatalà, V., and Dreyer, I. (2016). Electrical wiring and long-distance plant communication. *Trends Plant Sci* 21, 376–387. doi: 10.1016/j.tplants.2016.01.016
- Helliker, B. R., and Richter, S. L. (2008). Subtropical to boreal convergence of tree-leaf temperatures. *Nature* 454, 511–515. doi: 10.1038/nature07031
- Hernandez-Diaz, S., and Levin, M. (2014). Alteration of bioelectrically-controlled processes in the embryo: a teratogenic mechanism for anticonvulsants. *Reprod. Toxicol.* 47, 111–114. doi: 10.1016/j.reprotox.2014.04.008
- Hetherington, A. M., and Woodward, F. I. (2003). The role of stomata in sensing and driving environmental change. *Nature* 424, 901–908. doi: 10.1038/nature01843
- Higgins, M. L., Stadnicki, S. S., Shaw, T. J., and Leach, F. R. (1978). Effects of addicting and nonaddicting drugs on HeLa cells. *Tex. Rep. Biol. Med.* 36, 95–110.
- Hille, B. (2001). *Ion Channels of Excitable Membranes*. Sunderland, MA: Sinauer Associates Inc.
- Ho, M.-L., Tsai, T. N., Chang, J. K., Shao, T. S., Jeng, Y. R., and Hsu, C. (2005). Down-regulation of N-methyl D-aspartate receptor in rat-modeled disuse osteopenia. *Osteop. Int.* 16, 1780–1788. doi: 10.1007/s00198-005-1928-y
- Holcombe, M., and Paton, R. (1998). *Information Processing in Cells and Tissues*. New York, NY: Plenum Press.
- Huang, S., Eichler, G., Bar-Yam, Y., and Ingber, D. E. (2005). Cell fates as highdimensional attractor states of a complex gene regulatory network. *Phys. Rev. Lett.* 94:128701. doi: 10.1103/PhysRevLett.94.128701
- Humphrey, T. V., Bonetta, D. T., and Goring, D. R. (2007). Sentinels at the wall: cell wall receptors and sensors. *New Phytol.* 176, 7–21. doi: 10.1111/j.1469-8137.2007.02192.x
- Ingber, D. E., and Levin, M. (2007). What lies at the interface of regenerative medicine and developmental biology? *Development* 134, 2541–2547. doi: 10.1242/dev.003707
- Iovine, M. K., Higgins, E. P., Hindes, A., Coblitz, B., and Johnson, S. L. (2005). Mutations in connexin43 (GJA1) perturb bone growth in zebrafish fins. *Dev. Biol.* 278, 208–219. doi: 10.1016/j.ydbio.2004.11.005
- Jaffe, L. F. (1977). Electrophoresis along cell membranes. *Nature* 265, 600–602. doi: 10.1038/265600a0
- Jaffe, L. F. (1981). The role of ionic currents in establishing developmental pattern. *Philos. Trans. R. Soc. Lond. B* 295, 553–566. doi: 10.1098/rstb.1981.0160
- James, L. J., and Soffe, S. R. (2011). Skin impulse excitation of spinal sensory neurons in developing *Xenopus laevis* (Daudin) tadpoles. *J. Exp. Biol.* 214, 3341–3350. doi: 10.1242/jeb.058446
- Johnston, R. N., Pai, S. B., and Pai, R. B. (1992). The origin of the cancer cell: oncogeny reverses phylogeny. *Biochem. Cell Biol.* 70, 831–834. doi: 10.1139/o92-130
- Karban, R., Wetzel, W. C., Shiojiri, K., Ishizaki, S., Ramirez, S. R., and Blande, J. D. (2014a). Deciphering the language of plant communication: volatile chemotypes of sagebrush. *New Phytol.* 204, 380–385. doi: 10.1111/nph.12887
- Karban, R., Yang, L. H., and Edwards, K. F. (2014b). Volatile communication between plants that affects herbivory: a meta-analysis. *Ecol. Lett.* 17, 44–52. doi: 10.1111/ele.12205
- Kawamoto, S., Yoshida-Noro, C., and Tochinai, S. (2005). Bipolar head regeneration induced by artificial amputation in *Enchytraeus japonensis* (Annelida, Oligochaeta). *J. Exp. Zoolog. A Comp. Exp. Biol.* 303, 615–627. doi: 10.1002/jez.a.205
- Keijzer, F., van Duijn, M., and Lyon, P. (2013). What nervous systems do: early evolution, input-output, and the skin brain thesis. *Adapt. Behav.* 21, 67–85. doi: 10.1177/1059712312465330
- Khan, S. M., Ali, R., Asi, N., and Molloy, J. E. (2012). Active actin gels. *Commun. Integr. Biol.* 5, 39–42. doi: 10.4161/cib.18286
- Koch, C., and Segev, I. (2000). The role of single neurons in information processing. Nat. Neurosci. 3, 1171–1177. doi: 10.1038/81444
- Kondo, S. (2002). The reaction-diffusion system: a mechanism for autonomous pattern formation in the animal skin. *Genes Cells* 7, 535–541. doi: 10.1046/j.1365-2443.2002.00543.x

- Kondo, S., and Miura, T. (2010). Reaction-diffusion model as a framework for understanding biological pattern formation. *Science* 329, 1616–1620. doi: 10.1126/science.1179047
- Koo, H., and Graziadei, P. P. (1995). Eye primordium transplantation in Xenopus embryo. Anat. Embryol. 191, 155–170. doi: 10.1007/BF00186787
- Koshland, D. E. Jr. (1980). Bacterial chemotaxis in relation to neurobiology. Annu. Rev. Neurosci. 3, 43–75. doi: 10.1146/annurev.ne.03.030180. 000355
- Kragl, M., Knapp, D., Nacu, E., Khattak, S., Maden, M., Epperlein, H. H., et al. (2009). Cells keep a memory of their tissue origin during axolotl limb regeneration. *Nature* 460, 60–65. doi: 10.1038/nature08152
- Kumar, A., and Brockes, J. P. (2012). Nerve dependence in tissue, organ, and appendage regeneration. *Trends Neurosci.* 35, 691–699. doi: 10.1016/j.tins.2012.08.003
- Kunita, I., Kuroda, S., Ohki, K., and Nakagaki, T. (2014). Attempts to retreat from a dead-ended long capillary by backward swimming in *Paramecium. Front. Microbiol.* 5:270. doi: 10.3389/fmicb.2014.00270
- Lahoz-Beltra, R., Hameroff, S. R., and Dayhoff, J. E. (1993). Cytoskeletal logic: a model for molecular computation via Boolean operations in microtubules and microtubule-associated proteins. *Biosystems* 29, 1–23. doi: 10.1016/0303-2647(93)90078-Q
- Law, R., and Levin, M. (2015). Bioelectric memory: modeling resting potential bistability in amphibian embryos and mammalian cells. *Theor. Biol. Med. Model*, 12:22. doi: 10.1186/s12976-015-0019-9
- Leppik, L. P., Froemel, D., Slavici, A., Ovadia, Z. N., Hudak, L., Henrich, D., et al. (2015). Effects of electrical stimulation on rat limb regeneration, a new look at an old model. *Sci. Rep.* 5:18353. doi: 10.1038/srep18353
- Levin, M. (2007a). Gap junctional communication in morphogenesis. Prog. Biophys. Mol. Biol. 94, 186–206. doi: 10.1016/j.pbiomolbio.2007. 03.005
- Levin, M. (2007b). Large-scale biophysics: ion flows and regeneration. Trends Cell Biol. 17, 262–271. doi: 10.1016/j.tcb.2007.04.007
- Levin, M. (2011). The wisdom of the body: future techniques and approaches to morphogenetic fields in regenerative medicine, developmental biology and cancer. *Reg. Med.* 6, 667–673. doi: 10.2217/rme.11.69
- Levin, M. (2012a). Molecular bioelectricity in developmental biology: new tools and recent discoveries: control of cell behavior and pattern formation by transmembrane potential gradients. *Bioessays* 34, 205–217. doi: 10.1002/bies.201100136
- Levin, M. (2012b). Morphogenetic fields in embryogenesis, regeneration, and cancer: non-local control of complex patterning. *Biosystems* 109, 243–261. doi: 10.1016/j.biosystems.2012.04.005
- Levin, M. (2013). Reprogramming cells and tissue patterning via bioelectrical pathways: molecular mechanisms and biomedical opportunities. *Syst. Biol. Med.* 5, 657–676. doi: 10.1002/wsbm.1236
- Levin, M. (2014a). Endogenous bioelectrical networks store non-genetic patterning information during development and regeneration. J. Physiol. 592, 2295–2305. doi: 10.1113/jphysiol.2014.271940
- Levin, M. (2014b). Molecular bioelectricity: how endogenous voltage potentials control cell behavior and instruct pattern regulation in vivo. *Mol. Biol. Cell* 25, 3835–3850. doi: 10.1091/mbc.E13-12-0708
- Levin, M. (2014c). Transmembrane voltage potential of somatic cells controls oncogene-mediated tumorigenesis at long-range. Oncotarget 5, 3287–3306. doi: 10.18632/oncotarget.1935
- Levin, M., Buznikov, G. A., and Lauder, J. M. (2006). Of minds and embryos: leftright asymmetry and the serotonergic controls of pre-neural morphogenesis. *Dev. Neurosci.* 28, 171–185. doi: 10.1159/000091915
- Levin, M., and Mercola, M. (1998). Gap junctions are involved in the early generation of left-right asymmetry. *Dev. Biol.* 203, 90–105. doi: 10.1006/dbio.1998.9024
- Levin, M., and Stevenson, C. G. (2012). Regulation of cell behavior and tissue patterning by bioelectrical signals: challenges and opportunities for biomedical engineering. *Annu. Rev. Biomed. Eng.* 14, 295–323. doi: 10.1146/annurevbioeng-071811-150114
- Levin, M., Thorlin, T., Robinson, K. R., Nogi, T., and Mercola, M. (2002). Asymmetries in H+/K+-ATPase and cell membrane potentials comprise a very early step in left-right patterning. *Cell* 111, 77–89. doi: 10.1016/S0092-8674(02)00939-X

- Ling, H., Samarasinghe, S., and Kulasiri, D. (2013). Novel recurrent neural network for modelling biological networks: oscillatory p53 interaction dynamics. *Biosystems* 114, 191–205. doi: 10.1016/j.biosystems.2013.08.004
- Liu, X., Ramirez, S., and Tonegawa, S. (2014). Inception of a false memory by optogenetic manipulation of a hippocampal memory engram. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 369, 20130142. doi: 10.1098/rstb.2013.0142
- Lobo, D., Solano, M., Bubenik, G. A., and Levin, M. (2014). A linear-encoding model explains the variability of the target morphology in regeneration. J. R. Soc. Interf. 11:20130918. doi: 10.1098/rsif.2013.0918
- Lorber, J. (1978). Is your brain really necessary. Arch. Dis. Child. 53, 834-834.
- Lorber, J. (1981). Is your brain really necessary? Nurs. Mirror 152, 29-30.
- Lyon, P. (2015). The cognitive cell: bacterial behavior reconsidered. Front. Microbiol. 6:264. doi: 10.3389/fmicb.2015.00264
- Mackie, G. O. (1970). Neuroid conduction and the evolution of conducting tissues. *Quart. Rev. Biol.* 45, 319–332. doi: 10.1086/406645
- Manner, G., Foldes, F. F., Kuleba, M., and Deery, A. M. (1974). Morphine tolerance in a human neuroblastoma line: changes in choline acetylase and cholinesterase activities. *Experientia* 30, 137–138. doi: 10.1007/BF01927692
- Marder, E., Abbott, L. F., Turrigiano, G. G., Liu, Z., and Golowasch, J. (1996). Memory from the dynamics of intrinsic membrane currents. *Proc. Natl. Acad. Sci. U.S.A.* 93, 13481–13486. doi: 10.1073/pnas.93.24.13481
- Masi, E., Ciszak, M., Stefano, G., Renna, L., Azzarello, E., Pandolfi, C., et al. (2009). Spatiotemporal dynamics of the electrical network activity in the root apex. *Proc. Natl. Acad. Sci. U.S.A.* 106, 4048–4053. doi: 10.1073/pnas.0804640106
- Maurange, C., and Paro, R. (2002). A cellular memory module conveys epigenetic inheritance of hedgehog expression during *Drosophila* wing imaginal disc development. *Genes Dev.* 16, 2672–2683. doi: 10.1101/gad.242702
- McCaig, C. D., Rajnicek, A. M., Song, B., and Zhao, M. (2005). Controlling cell behavior electrically: current views and future potential. *Physiol. Rev.* 85, 943–978. doi: 10.1152/physrev.00020.2004
- McClean, M. N., Mody, A., Broach, J. R., and Ramanathan, S. (2007). Cross-talk and decision making in MAP kinase pathways. *Nat. Genet.* 39, 409–414. doi: 10.1038/ng0407-567b
- McConnell, J. V., Jacobson, A. L., and Kimble, D. P. (1959). The effects of regeneration upon retention of a conditioned response in the planarian. *J. Comp. Physiol. Psychol.* 52, 1–5. doi: 10.1037/h0048028
- McCulloch, W. S. (1951). "Why the mind is in the head?," in *Cerebral Mechanisms in Behavior: The Hixon Symposium*, ed. L. A. Jeffress (New York: John Wiley & Sons), 42–81.
- McCusker, C. D., and Gardiner, D. M. (2014). Understanding positional cues in salamander limb regeneration: implications for optimizing cell-based regenerative therapies. *Dis. Mod. Mech.* 7, 593–599. doi: 10.1242/dmm.013359
- McEvoy, J. W. (2009). Evolutionary game theory: lessons and limitations, a cancer perspective. *Br. J. Cancer* 101, 2060–2061. doi: 10.1038/sj.bjc.6605444
- McGregor, S., Vasas, V., Husbands, P., and Fernando, C. (2012). Evolution of associative learning in chemical networks. *PLoS Comp. Biol.* 8:e1002739. doi: 10.1371/journal.pcbi.1002739
- Miller, J. B., and Koshland, D. E. Jr. (1977). Sensory electrophysiology of bacteria: relationship of the membrane potential to motility and chemotaxis in Bacillus subtilis. *Proc. Natl. Acad. Sci. U.S.A.* 74, 4752–4756. doi: 10.1073/pnas.74.11.4752
- Milne, A., and Beamish, T. (1999). Inhalational and local anesthetics reduce tactile and thermal responses in *Mimosa pudica*. Can. J. Anaesth. 46, 287–289. doi: 10.1007/BF03012612
- Mondia, J. P., Levin, M., Omenetto, F. G., Orendorff, R. D., Branch, M. R., and Adams, D. S. (2011). Long-distance signals are required for morphogenesis of the regenerating *Xenopus* tadpole tail, as shown by femtosecond-laser ablation. *PLoS ONE* 6:e24953. doi: 10.1371/journal.pone.0024953
- Moreno-Risueno, M. A., and Benfey, P. N. (2011). Time-based patterning in development: the role of oscillating gene expression. *Transcription* 2, 124–129. doi: 10.4161/trns.2.3.15637
- Moreno-Risueno, M. A., Van Norman, J. M., Moreno, A., Zhang, J., Ahnert, S. E., and Benfey, P. N. (2010). Oscillating gene expression determines competence for periodic *Arabidopsis* root branching. *Science* 329, 1306–1311. doi: 10.1126/science.1191937
- Moroz, L. L., and Kohn, A. B. (2016). Independent origins of neurons and synapses: insights from ctenophores. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 371:20150041. doi: 10.1098/rstb.2015.0041

- Mustard, J., and Levin, M. (2014). Bioelectrical mechanisms for programming growth and form: taming physiological networks for soft body robotics. *Soft Robotics* 1, 169–191. doi: 10.1089/soro.2014.0011
- Nahm, M., Greyson, B., Kelly, E. W., and Haraldsson, E. (2012). Terminal lucidity: a review and a case collection. Arch. Gerontol. Geriatr. 55, 138–142. doi: 10.1016/j.archger.2011.06.031
- Nakagaki, T., Kobayashi, R., Nishiura, Y., and Ueda, T. (2004). Obtaining multiple separate food sources: behavioural intelligence in the *Physarum plasmodium*. *Proc. Biol. Sci. R. Soc.* 271, 2305–2310. doi: 10.1098/rspb.2004.2856
- Nelsen, E. M., Frankel, J., and Jenkins, L. M. (1989). Non-genic inheritance of cellular handedness. *Development* 105, 447–456.
- Newman, S. A., and Bhat, R. (2009). Dynamical patterning modules: a "pattern language" for development and evolution of multicellular form. *Int. J. Dev. Biol.* 53, 693–705. doi: 10.1387/ijdb.072481sn
- Nicolas, C., Abramson, C., and Levin, M. (2008). "Analysis of behavior in the planarian model," in *Planaria: A Model for Drug Action and Abuse*, eds R. Raffa and S. Rawls (Austin: RG Landes Co), 83–94.
- Nishimoto, S., Vu, A. T., Naselaris, T., Benjamini, Y., Yu, B., and Gallant, J. L. (2011). Reconstructing visual experiences from brain activity evoked by natural movies. *Curr. Biol.* 21, 1641–1646. doi: 10.1016/j.cub.2011.08.031
- Nogi, T., and Levin, M. (2005). Characterization of innexin gene expression and functional roles of gap-junctional communication in planarian regeneration. *Dev. Biol.* 287, 314–335. doi: 10.1016/j.ydbio.2005.09.002
- Norris, V., Zemirline, A., Amar, P., Audinot, J. N., Ballet, P., Ben-Jacob, E., et al. (2011). Computing with bacterial constituents, cells and populations: from bioputing to bactoputing. *Theor. Biosci.* 130, 211–228. doi: 10.1007/s12064-010-0118-4
- Nuccitelli, R. (2003). A role for endogenous electric fields in wound healing. *Curr. Top. Dev. Biol.* 58, 1–26. doi: 10.1016/S0070-2153(03)58001-2
- Nunes-Alves, C. (2015). Biofilms: electrifying long-range signalling. Nat. Rev. Microbiol. 13:737. doi: 10.1038/nrmicro3588
- Oberheim, N. A., Takano, T., Han, X., He, W., Lin, J. H., Wang, F., et al. (2009). Uniquely hominid features of adult human astrocytes. J. Neurosci. 29, 3276–3287. doi: 10.1523/JNEUROSCI.4707-08.2009
- Otani, N. F., and Gilmour, R. F. Jr. (1997). Memory models for the electrical properties of local cardiac systems. J. Theor. Biol. 187, 409–436. doi: 10.1006/jtbi.1997.0447
- Oviedo, N. J., Morokuma, J., Walentek, P., Kema, I. P., Gu, M. B., Ahn, J. M., et al. (2010). Long-range neural and gap junction protein-mediated cues control polarity during planarian regeneration. *Dev. Biol.* 339, 188–199. doi: 10.1016/j.ydbio.2009.12.012
- Pagán, O. R. (2014). The First Brain: The Neuroscience of Planarians. New York: Oxford University Press.
- Pai, V. P., Aw, S., Shomrat, T., Lemire, J. M., and Levin, M. (2012). Transmembrane voltage potential controls embryonic eye patterning in *Xenopus laevis*. *Development* 139, 313–323. doi: 10.1242/dev.073759
- Pai, V. P., Lemire, J. M., Pare, J. F., Lin, G., Chen, Y., and Levin, M. (2015a). Endogenous gradients of resting potential instructively pattern embryonic neural tissue via Notch signaling and regulation of proliferation. *J. Neurosci.* 35, 4366–4385. doi: 10.1523/JNEUROSCI.1877-14.2015
- Pai, V. P., Martyniuk, C. J., Echeverri, K., Sundelacruz, S., Kaplan, D. L., and Levin, M. (2015b). Genome-wide analysis reveals conserved transcriptional responses downstream of resting potential change in *Xenopus embryos*, axolotl regeneration, and human mesenchymal cell differentiation. *Regeneration* 3, 3–25. doi: 10.1002/reg2.48
- Palacios-Prado, N., and Bukauskas, F. F. (2009). Heterotypic gap junction channels as voltage-sensitive valves for intercellular signaling. *Proc. Natl. Acad. Sci. U.S.A.* 106, 14855–14860. doi: 10.1073/pnas.0901923106
- Pawlowski, A. (1970). [The role of nerve elements of the skin in the course of experimental carcinogenesis and of basal cell epithelioma and squamous cell carcinoma in man]. *Neuropatol. Pol.* 8, 205–239.
- Pawlowski, A., and Weddell, G. (1967). Induction of tumours in denervated skin. *Nature* 213, 1234–1236. doi: 10.1038/2131234a0
- Peak, D., West, J. D., Messinger, S. M., and Mott, K. A. (2004). Evidence for complex, collective dynamics and emergent, distributed computation in plants. *Proc. Natl. Acad. Sci. U.S.A.* 101, 918–922. doi: 10.1073/pnas.0307 811100

- Perathoner, S., Daane, J. M., Henrion, U., Seebohm, G., Higdon, C. W., Johnson, S. L., et al. (2014). Bioelectric signaling regulates size in zebrafish fins. *PLoS Genet.* 10:e1004080. doi: 10.1371/journal.pgen.1004080
- Pereda, A. E., Curti, S., Hoge, G., Cachope, R., Flores, C. E., and Rash, J. E. (2013). Gap junction-mediated electrical transmission: regulatory mechanisms and plasticity. *Biochim. Biophys. Acta* 1828, 134–146. doi: 10.1016/j.bbamem.2012.05.026
- Perouansky, M. (2012). The quest for a unified model of anesthetic action. A century in Claude Bernard's shadow. Anesthesiology 117, 465–474. doi: 10.1097/ALN.0b013e318264492e
- Pezzulo, G., and Levin, M. (2015). Re-membering the body: applications of computational neuroscience to the top-down control of regeneration of limbs and other complex organs. *Integr. Biol. (Camb.)* 7, 1487–1517. doi: 10.1039/c5ib00221d
- Pietsch, P., and Schneider, C. W. (1969). Brain transplantation in Salamanders an approach to memory transfer. *Brain Res.* 14, 707–715. doi: 10.1016/0006-8993(69)90210-8
- Pimashkin, A., Gladkov, A., Mukhina, I., and Kazantsev, V. (2013). Adaptive enhancement of learning protocol in hippocampal cultured networks grown on multielectrode arrays. *Front. Neural. Circ.* 7:87. doi: 10.3389/fncir.2013. 00087
- Pincebourde, S., and Woods, H. A. (2012). Climate uncertainty on leaf surfaces: the biophysics of leaf microclimates and their consequences for leaf-dwelling organisms. *Funct. Ecol.* 26, 844–853. doi: 10.1111/j.1365-2435.2012.02013.x
- Prindle, A., Liu, J., Asally, M., Ly, S., Garcia-Ojalvo, J., and Suel, G. M. (2015). Ion channels enable electrical communication in bacterial communities. *Nature* 527, 59–63. doi: 10.1038/nature15709
- Pruszynski, J. A., and Johansson, R. S. (2014). Edge-orientation processing in first-order tactile neurons. *Nat. Neurosci.* 17, 1404–1409. doi: 10.1038/nn.3804
- Pullar, C. E. (2011). The Physiology of Bioelectricity in Development, Tissue Regeneration, and Cancer. Boca Raton, FL: CRC Press.
- Ramanathan, S., and Broach, J. R. (2007). Do cells think? Cell. Mol. Life Sci. 64, 1801–1804. doi: 10.1007/s00018-007-7001-6
- Ramirez, S., Liu, X., Lin, P. A., Suh, J., Pignatelli, M., Redondo, R. L., et al. (2013). Creating a false memory in the hippocampus. *Science* 341, 387–391. doi: 10.1126/science.1239073
- Raspopovic, J., Marcon, L., Russo, L., and Sharpe, J. (2014). Modeling digits. Digit patterning is controlled by a Bmp-Sox9-Wnt Turing network modulated by morphogen gradients. *Science* 345, 566–570. doi: 10.1126/science.1252960

Rasskin-Gutman, D., and Izpisua-Belmonte, J. C. (2004). Theoretical morphology of developmental asymmetries. *Bioessays* 26, 405–412. doi: 10.1002/bies.10410

Reddien, P. W., and Sánchez Alvarado, A. (2004). Fundamentals of planarian regeneration. Annu. Rev. Cell Dev. Biol. 20, 725–757. doi: 10.1146/annurev.cellbio.20.010403.095114

Ricci, A., Micini, A., Viroli, M., Gardelli, L., and Oliva, E. (2007). "Cognitive stigmergy: towards a framework based on agents and artifacts," in *E4MAS*, eds D. Weyns, H. V. D. Parunak, and F. Michel (Berlin: Springer-Verlag), 124–140.

- Ringrose, L., and Paro, R. (2004). Epigenetic regulation of cellular memory by the *Polycomb* and Trithorax group proteins. *Annu. Rev. Genet.* 38, 413–443. doi: 10.1146/annurev.genet.38.072902.091907
- Rinn, J. L., Bondre, C., Gladstone, H. B., Brown, P. O., and Chang, H. Y. (2006). Anatomic demarcation by positional variation in fibroblast gene expression programs. *PLoS Genet.* 2:e119. doi: 10.1371/journal.pgen.0020119
- Roberts, A., and Stirling, C. A. (1971). Properties and propagation of a cardiaclike impulse in skin of young tadpoles. *Zeitschr. Vergl. Physiol.* 71, 295–310. doi: 10.1007/BF00298141
- Rosen, M., and Plotnikov, A. (2002). The pharmacology of cardiac memory. *Pharmacol. Ther.* 94, 63–75. doi: 10.1016/S0163-7258(02)00172-9
- Rosen, M. R., and Cohen, I. S. (2006). Cardiac memory. new insights into molecular mechanisms. J. Physiol. 570, 209–218. doi: 10.1113/jphysiol.2005.097873
- Rosen, R. (1968). Turing's morphogens, two-factor systems and active transport. Bull. Math. Biophys. 30, 493–499. doi: 10.1007/BF02476609
- Rosenblueth, A., Wiener, N., and Bigelow, J. (1943). Behavior, purpose, and teleology. *Philos. Sci.* 10, 18–24. doi: 10.1086/286788
- Sachidhanandam, S., Sreenivasan, V., Kyriakatos, A., Kremer, Y., and Petersen, C. C. (2013). Membrane potential correlates of sensory perception in mouse barrel cortex. *Nat. Neurosci.* 16, 1671–1677. doi: 10.1038/nn.3532

- Sahu, S., Ghosh, S., Hirata, K., Fujita, D., and Bandyopadhyay, A. (2013). Multilevel memory-switching properties of a single brain microtubule. *Appl. Phys. Lett.* 102:123701. doi: 10.1063/1.4793995
- Saigusa, T., Tero, A., Nakagaki, T., and Kuramoto, Y. (2008). Amoebae anticipate periodic events. *Phys. Rev. Lett.* 100:018101. doi: 10.1103/PhysRevLett.100.018101
- Sakiyama, T., and Gunji, Y. P. (2016). The Kanizsa triangle illusion in foraging ants. *Biosystems* 14, 9–14. doi: 10.1016/j.biosystems.2016.02.003
- Sanchez Alvarado, A., and Tsonis, P. A. (2006). Bridging the regeneration gap: genetic insights from diverse animal models. *Nat. Rev. Genet.* 7, 873–884. doi: 10.1038/nrg1923
- Sarnat, H. B., and Netsky, M. G. (1985). The brain of the planarian as the ancestor of the human brain. *Can. J. Neurol. Sci.* 12, 296–302.
- Scarle, S. (2009). Implications of the Turing completeness of reaction-diffusion models, informed by GPGPU simulations on an XBox 360: Cardiac arrhythmias, re-entry and the Halting problem. *Comput. Biol. Chem.* 33, 253– 260.
- Schumann, A., and Adamatzky, A. (2009). Toward semantical model of reaction-diffusion computing. *Kybernetes* 38, 1518–1531. doi: 10.1108/03684920910991504
- Schummers, J., Yu, H., and Sur, M. (2008). Tuned responses of astrocytes and their influence on hemodynamic signals in the visual cortex. *Science* 320, 1638–1643. doi: 10.1126/science.1156120
- Scialdone, A., Mugford, S. T., Feike, D., Skeffington, A., Borrill, P., Graf, A., et al. (2013). Arabidopsis plants perform arithmetic division to prevent starvation at night. eLife 2:e00669. doi: 10.7554/eLife.00669
- Seifert, G. J., and Blaukopf, C. (2010). Irritable walls: the plant extracellular matrix and signaling. *Plant Physiol*. 153, 467–478. doi: 10.1104/pp.110.153940
- Semon, R. W., and Simon, L. (1921). *The MNEME*. London: The Macmillan company.
- Shapiro, J. A. (1998). Thinking about bacterial populations as multicellular organisms. *Annu. Rev. Microbiol.* 52, 81–104. doi: 10.1146/annurev.micro.52.1.81
- Sheiman, I. M., and Tiras, K. L. (1996). "Memory and morphogenesis in planaria and beetle," in *Russian Contributions to Invertebrate Behavior*, eds C. I. Abramson, Z. P. Shuranova, and Y. M. Burmistrov (Westport, CT: Praeger), 43–76.
- Shepherd, V. A. (2005). From semi-conductors to the rhythms of sensitive plants: the research of J.C. Bose. Cell. Mol. Biol. 51, 607–619.
- Shomrat, T., and Levin, M. (2013). An automated training paradigm reveals longterm memory in planarians and its persistence through head regeneration. *J. Exp. Biol.* 216, 3799–3810. doi: 10.1242/jeb.087809
- Singer, M. (1952). The influence of the nerve in regeneration of the amphibian extremity. *Q. Rev. Biol.* 27, 169–200. doi: 10.1086/398873
- Slack, J. M. (1980). A serial threshold theory of regeneration. J. Theor. Biol. 82, 105–140. doi: 10.1016/0022-5193(80)90092-2
- Smythies, J. (2015). On the possible role of protein vibrations in information processing in the brain: three Russian dolls. *Front. Mol. Neurosci.* 8:38. doi: 10.3389/fnmol.2015.00038
- Sollars, S. I., Smith, P. C., and Hill, D. L. (2002). Time course of morphological alterations of fungiform papillae and taste buds following chorda tympani transection in neonatal rats. J. Neurobiol. 51, 223–236. doi: 10.1002/neu.10055
- Spencer, G. J., and Genever, P. G. (2003). Long-term potentiation in bone-a role for glutamate in strain-induced cellular memory? *BMC Cell Biol.* 4:9. doi: 10.1186/1471-2121-4-9
- Stewart, S., Rojas-Munoz, A., and Izpisua Belmonte, J. C. (2007). Bioelectricity and epimorphic regeneration. *Bioessays* 29, 1133–1137. doi: 10.1002/bies.20656
- Stone, J. R. (1997). The spirit of D'arcy Thompson dwells in empirical morphospace. Math. Biosci. 142, 13–30. doi: 10.1016/S0025-5564(96)00186-1
- Sukhov, V., Nerush, V., Orlova, L., and Vodeneev, V. (2011). Simulation of action potential propagation in plants. J. Theor. Biol. 291C, 47–55. doi: 10.1016/j.jtbi.2011.09.019
- Sundelacruz, S., Levin, M., and Kaplan, D. L. (2009). Role of membrane potential in the regulation of cell proliferation and differentiation. *Stem Cell Rev. Rep.* 5, 231–246. doi: 10.1007/s12015-009-9080-2
- Tagkopoulos, I., Liu, Y. C., and Tavazoie, S. (2008). Predictive behavior within microbial genetic networks. *Science* 320, 1313–1317. doi: 10.1126/science.1154456

Taiz, L. (2010). Plant Physiology, 5th Edn. Sunderland, MA: Sinauer Associates Inc.

- Takeda, M., Suzuki, Y., Obara, N., and Nagai, Y. (1996). Apoptosis in mouse taste buds after denervation. *Cell Tissue Res.* 286, 55–62. doi: 10.1007/s0044100 50674
- Taylor, R. E. (1974). Excitable membranes. Annu. Rev. Phys. Chem. 25, 387–405. doi: 10.1146/annurev.pc.25.100174.002131
- Theraulaz, G., and Bonabeau, E. (1999). A brief history of stigmergy. *Artif. Life* 5, 97–116. doi: 10.1162/106454699568700
- Traas, J., and Vernoux, T. (2010). Oscillating roots. Science 329, 1290–1291. doi: 10.1126/science.1195572
- Trewavas, A. (2005). Plant intelligence. Naturwissenschaften 92, 401–413. doi: 10.1007/s00114-005-0014-9
- Trewavas, A. (2014). *Plant Behaviour and Intelligence*. Oxford: Oxford University Press.
- Tseng, A., and Levin, M. (2013). Cracking the bioelectric code: probing endogenous ionic controls of pattern formation. *Commun. Integr. Biol.* 6, 1–8. doi: 10.4161/cib.22595
- Tseng, A. S., Beane, W. S., Lemire, J. M., Masi, A., and Levin, M. (2010). Induction of vertebrate regeneration by a transient sodium current. J. Neurosci. 30, 13192–13200. doi: 10.1523/JNEUROSCI.3315-10.2010
- Tsong, T. Y., and Astumian, R. D. (1986). Absorption and conversion of electric field energy by membrane bound ATPases. *Bioelectrochem. Bioenerg.* 15, 457– 476. doi: 10.1016/0302-4598(86)85034-6
- Turing, A. M. (1952). The chemical basis of morphogenesis. *Philos. Trans. Roy Soc.* B. 237, 37–72. doi: 10.1098/rstb.1952.0012
- Turner, C. H., Robling, A. G., Duncan, R. L., and Burr, D. B. (2002). Do bone cells behave like a neuronal network? *Calcif. Tissue Int.* 70, 435–442. doi: 10.1007/s00223-001-1024-z
- Turrigiano, G. G., Marder, E., and Abbott, L. F. (1996). Cellular short-term memory from a slow potassium conductance. J. Neurophysiol. 75, 963–966.
- Ungar, G. (1972). Molecular coding of information in nervous system. Naturwissenschaften 59, 85–91. doi: 10.1007/BF00591779
- Ungar, G. (1974a). Molecular coding of memory. Life Sci. 14, 595–604. doi: 10.1016/0024-3205(74)90394-4
- Ungar, G. (1974b). Peptides and memory. *Biochem. Pharmacol.* 23, 1553–1558. doi: 10.1016/0006-2952(74)90366-9
- van Gelder, T. (1998). The dynamical hypothesis in cognitive science. *Behav. Brain Sci.* 21, 615–628; discussion 629–665. doi: 10.1017/S0140525X98001733
- Vergassola, M., Villermaux, E., and Shraiman, B. I. (2007). 'Infotaxis' as a strategy for searching without gradients. *Nature* 445, 406–409. doi: 10.1038/nature 05464
- Victor, J. D. (1999). Temporal aspects of neural coding in the retina and lateral geniculate. *Network* 10, R1–R66. doi: 10.1088/0954-898X_10_4_201
- Vincent, M. (2012). Cancer: a de-repression of a default survival program common to all cells? A life-history perspective on the nature of cancer. *Bioessays* 34, 72–82. doi: 10.1002/bies.201100049
- Vladimirov, N., and Sourjik, V. (2009). Chemotaxis: how bacteria use memory. Biol. Chem. 390, 1097–1104. doi: 10.1515/BC.2009.130
- Volkmann, D., and Baluška, F. (1999). The actin cytoskeleton in plants: from transport networks to signaling networks. *Microsc. Res. Technol.* 47, 135–154. doi: 10.1002/(SICI)1097-0029(19991015)47:2<135::AID-JEMT6>3.0.CO;2-1
- Volkov, A. G., Foster, J. C., Ashby, T. A., Walker, R. K., Johnson, J. A., and Markin, V. S. (2010). Mimosa pudica: electrical and mechanical stimulation of plant movements. *Plant Cell Environ.* 33, 163–173. doi: 10.1111/j.1365-3040.2009.02066.x
- Wang, K. C., Helms, J. A., and Chang, H. Y. (2009). Regeneration, repair and remembering identity: the three Rs of Hox gene expression. *Trends Cell Biol.* 19, 268–275. doi: 10.1016/j.tcb.2009.03.007
- Watson, R. A., Buckley, C. L., Mills, R., and Davies, A. (2010). "Associative memory in gene regulation networks," in *Proceedings of the Artificial Life Conference XII*, Odense, 194–201.
- Wayne, R. (1993). Excitability in plant cells. Am. Sci. 81, 140-143.
- Wayne, R. (1994). The excitability of plant cells: with a special emphasis on Characean internodal cells. *Bot. Rev.* 60, 265–367. doi: 10.1007/BF029 60261
- Webb, A. A., and Satake, A. (2015). Understanding circadian regulation of carbohydrate metabolism in Arabidopsis using mathematical models. *Plant Cell Physiol.* 56, 586–593. doi: 10.1093/pcp/pcv033

- Wells, P. H. (1967). "Training flatworms in a van oye maze," in *Chemistry of Learning*, eds W. C. Corning and S. C. Ratner (New York, NY: Plenum), 251–254.
- Westerhoff, H. V., Tsong, T. Y., Chock, P. B., Chen, Y. D., and Astumian, R. D. (1986). How enzymes can capture and transmit free energy from an oscillating electric field. *Proc. Natl. Acad. Sci. U.S.A.* 83, 4734–4738. doi: 10.1073/pnas.83.13.4734
- Xiang, L., and Tao, N. J. (2016). Reactions triggered electrically. *Nature* 531, 38–39. doi: 10.1038/531038a
- Xiong, W., and Ferrell, J. E. Jr., (2003). A positive-feedback-based bistable 'memory module' that governs a cell fate decision. *Nature* 426, 460–465. doi: 10.1038/nature02089
- Yaar, M., and Park, H. Y. (2012). Melanocytes: a window into the nervous system. J. Invest. Dermatol. 132, 835-845. doi: 10.1038/jid.2011.386
- Yamashita, T., Pala, A., Pedrido, L., Kremer, Y., Welker, E., and Petersen, C. C. (2013). Membrane potential dynamics of neocortical projection neurons driving target-specific signals. *Neuron* 80, 1477–1490. doi: 10.1016/j.neuron.2013.10.059
- Yang, M., and Brackenbury, W. J. (2013). Membrane potential and cancer progression. Front. Physiol. 4:185. doi: 10.3389/fphys.2013.00185
- Yntema, C. L. (1959a). Blastema formation in sparsely innervated and aneurogenic forelimbs of amblystoma larvae. J. Exp. Zool. 142, 423–439. doi: 10.1002/jez.1401420119
- Yntema, C. L. (1959b). Regeneration in sparsely innervated and aneurogenic forelimbs of Amblystoma larvae. J. Exp. Zool. 140, 101–123. doi: 10.1002/jez.1401400106
- Yokawa, K., and Baluška, F. (2015). C. elegans and Arabidopsis thaliana show similar behavior: ROS induce escape tropisms both in

illuminated nematodes and roots. Plant Signal. Behav. 10:e1073870. doi: 10.1080/15592324.2015.1073870

- Yokawa, K., and Baluška, F. (2016). The TOR complex: an emergency switch for root behavior. *Plant Cell Physiol.* 57, 14–18. doi: 10.1093/pcp/pcv191
- Yokawa, K., Fassano, R., Kagenishi, T., and Baluška, F. (2014). Light as stress factor to plant roots – case of root halotropism. *Front. Plant Sci.* 5:718. doi: 10.3389/fpls.2014.00718
- Zhang, W. B., Zhao, Y., and Kjell, F. (2013). Understanding propagated sensation along meridians by volume transmission in peripheral tissue. *Chin. J. Integr. Med.* 19, 330–339. doi: 10.1007/s11655-013-1456-0
- Zhu, L., Aono, M., Kim, S. J., and Hara, M. (2013). Amoeba-based computing for traveling salesman problem: long-term correlations between spatially separated individual cells of *Physarum polycephalum*. *Biosystems* 112, 1–10. doi: 10.1016/j.biosystems.2013.01.008
- Zoghi, M. (2004). Cardiac memory: do the heart and the brain remember the same? *J. Interv. Card. Electrophysiol.* 11, 177–182. doi: 10.1023/B:JICE.0000048567.18088.a2

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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