#### **RESEARCH**



# Lipid Metabolism-Related Enzyme Inhibition and Antioxidant Potential of the Extracts and Phytochemical Compounds from *Trachystemon orientalis* (L.) D.Don

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#### **Abstract**

Trachystemon orientalis (L.) D.Don is a perennial herb extensively utilized for culinary purposes and the treatment of numerous ailments. Hyperlipidemia is a significant risk factor for atherosclerosis, diabetes, obesity, and coronary heart disease. The inhibition of pancreatic lipase (PL) and cholesterol esterase (CE) aims to mitigate problems associated with lipid metabolism disorders by decreasing hyperlipidemia. In this context, enhancing antioxidant capability is an additional objective. This study assessed the PL and CE inhibitory actions of extracts and compounds from the aerial parts of T. orientalis, utilizing ferric reducing antioxidant power (FRAP) and cupric-reducing antioxidant capacity (CUPRAC) assays to measure antioxidant capabilities. Rosmarinic acid, a combination of rosmarinic acid and danshensu (phenolic acids), and  $\beta$ -sitosterol (phytosterol) were isolated.  $\beta$ -sitosterol (IC<sub>50</sub>: 41.698 ± 1.982  $\mu$ g/mL, 14.249 ± 1.209  $\mu$ g/mL, respectively) followed by rosmarinic acid (IC<sub>50</sub>: 48.213 ± 2.490 μg/mL, 21.941 ± 3.785 μg/mL, respectively) demonstrated significant inhibitory effects on PL and CE. Additionally, rosmarinic acid (FRAP: 1274.400±1.633 μM TEAC, CUPRAC: 1801.389±5.500 μM TEAC) and its combination with danshensu (FRAP: 1405.067±4.491, CUPRAC: 2174.167±9.647 μΜ TEAC) exhibited high antioxidant activity. The ethyl acetate subextract (TOE) from which rosmarinic acid was isolated showed considerable enzyme inhibitory (IC<sub>50</sub>: 47.577±0.931 μg/mL, 12.443±1.233 μg/mL, for PL and CE inhibition, respectively) and antioxidant activity (FRAP: 1770.021±4.583, CUPRAC: 2189.167±5.401 μM TEAC). T. orientalis has been identified as a promising natural resource for the management of lipid metabolism-related disorders and for enhancing antioxidant activity.

### **Clinical Trial**

Not applicable.

**Keywords** Antioxidant · Cholesterol esterase · Danshensu · Pancreatic lipase · Rosmarinic acid ·  $\beta$ -sitosterol

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### Introduction

Hyperlipidemia is a metabolic condition characterized by elevated triglyceride and cholesterol levels in the blood-stream, resulting from high-fat diets and poor lifestyles. Hyperlipidemia is a significant risk factor for illnesses associated with lipid metabolism, including atherosclerosis, coronary heart disease, diabetes, and obesity [1]. The prevalence of metabolic syndrome, encompassing conditions such as hypertension, obesity, insulin resistance, and dyslipidemia, ranges from 20 to 45%. By 2030, it is projected that 89% of men and 85% of women globally would be overweight or obese [2].

Pancreatic lipase (PL) is the principal lipase enzyme that facilitates the transformation of triglycerides into free fatty acids and glycerol, accounting for roughly 70% of digestion and absorption [3]. PL is essential in converting dietary triglycerides into absorbable lipids and is an attractive target for its ability to diminish fat absorption and decelerate lipid metabolism, therefore averting the buildup of body fat. Cholesterol esterase (CE) is found in bile, hydrolyzing cholesterol esters and aiding the transfer of free cholesterol from micelles to enterocytes. Besides cholesterol esters, it also hydrolyzes substrates including fat-soluble vitamins, phospholipids, and triglycerides. The inhibition of phospholipase (PL) and cholesterol esterase (CE) to prevent the absorption of triglycerides and cholesterol esters is a therapy strategy for hyperlipidemia and other lipid metabolism disorders [4]. The PL inhibitor orlistat is utilized for obesity management, whereas simvastatin is employed for hypercholesterolemia; nonetheless, both possess significant adverse effects [5]. Consequently, natural products serve as significant sources in the quest for safe and efficacious antihyperlipidemic medicines.

Trachystemon orientalis (L.) D.Don, referred to in Turkey as "kaldırayak, kaldırık, balıkotu, ıspıt, hodan, tamara, acı hodan," is the sole species under the genus Trachystemon G. Don (Boraginaceae) [6]. It proliferates in the Black Sea Region, Eastern Bulgaria, and the Western Caucasus globally. It is a perennial herb that attains a height of 30-40 cm, with blue-purple blooms, large leaves, and a rhizome [7]. It is extensively utilized, particularly in the Black Sea area of Turkey, where its leaves, petioles, and flowers are ingested as vegetables. The leaves serve as a diuretic, expectorant, and for digestive issues; the whole plant is utilized for irritated wounds [8, 9]. Studies have shown that its phytochemical composition encompasses flavonoids, phenolic compounds, essential oils, fatty acids, tannins, resin, mucilage, saponins, choline, minerals and vitamins [7, 10–12]. Literature indicates that the plant demonstrates antioxidant [7, 12, 13], antimicrobial [7, 8], antifungal, herbicidal [6], antimutagenic; α-amylase and α-glucosidase [7], butyrylcholinesterase [13], tyrosinase, collagenase and, elastase enzyme inhibition, as well as photoprotective activities [14].

Research on the phytochemical and biological activities of the plant is restricted. The plant, traditionally used for intestinal disorders, has demonstrated antioxidant and inhibitory effects on digestive enzymes like α-amylase and α-glucosidase [7, 8]. In addition, rosmarinic acid, danshensu, globoidnan B, and rabdosiin, exhibiting PL inhibitory activity, were isolated from the underground parts [15]. The results indicate that the aerial parts of the plant may be abundant in phytochemicals exhibiting antihyperlipidemic activity via the inhibition of PL and CE. This study aims to assess the PL and CE inhibitory activities as well as the antioxidant capacity of the extracts and major compounds from the aerial parts of *T. orientalis*.

#### **Materials and Methods**

The Materials and Methods section is presented as Supplementary material.

#### **Results and Discussion**

### **Isolation and Elucidation of the Compounds**

In this research, isolation studies were carried out on the subextracts prepared from the methanol extract (TOM), and the biological activities of the extracts and compounds were determined. The isolation studies resulted in the purification of three identified compounds (two phenolic acids and one sterol) by different chromatographic techniques (Figure S1). The structures of the isolated compounds were determined using 1D and 2D nuclear magnetic resonance (NMR) methods, with the spectra are presented in Figures S2-S12.  $\beta$ -sitosterol (1) from the chloroform subextract (TOC), rosmarinic acid (2) from the ethyl acetate subextract (TOE), and the mixture of rosmarinic acid and danshensu (2+3) from the remaining aqueous subextract (TOA) have been purified for the first time of the aerial parts of the plant (Fig. 1). The presence of  $\beta$ -sitosterol and rosmarinic acid in the plant has been demonstrated using HPLC method [12, 16]. Rosmarinic acid and danshensu were previously isolated from the underground parts of the plant [15].

## PL Inhibitory Activities of the Extracts and Compounds

The IC<sub>50</sub> values for the extracts and the standard regarding PL inhibition are displayed in Table 1; Fig. 2. The



Fig. 1 Structures of the compounds. (1)  $\beta$ -sitosterol, (2) Rosmarinic acid, (3) Danshensu

Table 1 PL and CE inhibitory activities of the extracts and compounds

	J	1
Extracts/Compounds	Enzyme inhibitory activities IC <sub>50</sub> (µg/mL)±SD**	
	Pancreatic lipase	Cholesterol
	(PL)	esterase (CE)
TOM	77.255 ± 1.237°	$23.572 \pm 1.283^a$
TOC	nd***	$37.379 \pm 0.966^b$
TOE	$47.577 \pm 0.931^b$	$12.443 \pm 1.233^a$
TOA	$114.035 \pm 7.256^e$	$67.945 \pm 16.177^c$
$\beta$ -sitosterol	$41.698\!\pm\!1.982^b$	$14.249 \pm 1.209^a$
Rosmarinic acid	$48.213\!\pm\!2.490^b$	$21.941 \pm 3.785^a$
Rosmarinic	$88.003 \pm 1.140^d$	$28.491 \pm 5.011^a$
acid+Danshensu		
Positive control*	$22.709 \pm 1.299^a$	$13.981 \pm 2.367^a$

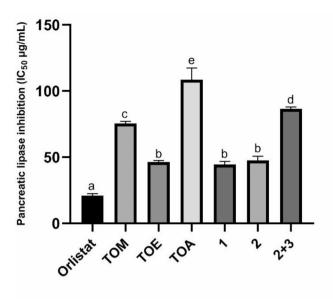
\*Siµvastatin for CE; Orlistat for PL. \*\*: Standard deviation, \*\*\*: not determined. Different letters (a>b>c>d) in the same column indicate significant differences (p<0.05) by Tukey's multiple comparisons test. TOM: The methanol extract of *Trachystemon on orientalis*, TOC: The chloroform subextract of *T. orientalis*, TOE: The ethyl acetate subextract of *T. orientalis*, TOA: The remaining aqueous subextract of *T. orientalis* 

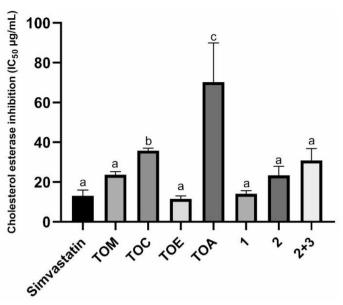
efficacy of the extracts and compounds was weaker than orlistat (p<0.0001). The activity of TOE surpassed that of the other extracts ( $IC_{50}$ =47.577±0.931 µg/mL, p<0.0001). Among the isolated compounds,  $\beta$ -sitosterol ( $IC_{50}$ =41.698±1.982 µg/mL, p<0.0001) and rosmarinic acid ( $IC_{50}$ =48.213±2.490 µg/mL, p<0.0001) exhibited substantially greater efficacy than the combination of rosmarinic acid and danshensu. Rosmarinic acid, an ester of caffeic acid and 3,4-dihydroxyphenyl lactic acid (danshensu), is a significant phenolic acid [17]. Rosmarinic acid, the primary component of TOE, shown comparable activity

to TOE, which may be ascribed to it. A research evaluated the PL inhibitory activity of Rosmarinus officinalis L. extract and pure rosmarinic acid, revealing IC<sub>50</sub> values of 13.8 and 125.2 µg/mL, respectively. It has been suggested that rosmarinic acid and other phenolic compounds may have functioned synergistically [18]. In further investigations, rosmarinic acid shown significant lipase inhibitory activity (IC<sub>50</sub>= $62.8\pm2.7$  µM,  $49.421\pm1.448$  µg/mL), corroborating our findings [15, 19]. Danshensu (salvianic acid A) is a phenolic acid extracted from the roots of Salvia miltiorrhiza Bunge and is present in several Boraginaceae species [15, 20]. Danshensu has demonstrated the ability to suppress PL with an IC<sub>50</sub> of  $65.160 \pm 4.443 \, \mu g/mL$  [15]. This study demonstrated that the combination of rosmarinic acid and danshensu had lower activity than rosmarinic acid alone. It may be inferred that rosmarinic acid and danshensu may exhibit antagonistic effects. The non-competitive binding of these two chemicals to the enzyme's active site, or their simultaneous binding to distinct locations, may diminish their respective inhibitory effects. Furthermore, chemical or steric interactions among the compounds may further reduce their binding affinity to the enzyme.

This study demonstrated that  $\beta$ -sitosterol exhibits significant PL inhibitory activity. Literature has studies investigating the impact of  $\beta$ -sitosterol, a natural sterol, on PL, yielding contradictory results. In one study, similar to our findings (at 100 µg/mL, 68.47% inhibition),  $\beta$ -sitosterol (100 µg/mL) was found to inhibit PL by 79.1±11.3% [21]. In another study, the compound (4 mg/mL) demonstrated considerable inhibition (IC<sub>50</sub>=82.56 mg/mL, 60.67% ± 0.53%) [22]. Other studies indicated that it exhibited little







**Fig. 2** PL and CE inhibitory activity of the extracts and compounds. The values are represented by mean  $\pm$  SD of triplicate measurements. Or listat and simvastatin were used as positive control. Different letters (a>b>c>d) in the same graphic indicate significant differences (p<0.05) by Tukey's multiple comparisons test. (1) β-sitosterol (2)

Rosmarinic acid, (3) Danshensu. TOM: The methanol extract of *Trachystemon orientalis*, TOC: The chloroform subextract of *T. orientalis*, TOE: The ethyl acetate subextract of *T. orientalis*, TOA: The remaining aqueous subextract of *T. orientalis* 

activity (2.99 $\pm$ 0.80%, 8.80 $\pm$ 4.15% inhibition) on PL [23, 24]. The divergent outcomes regarding the PL inhibitory action of  $\beta$ -sitosterol may stem from variations in experimental methodology, solvents, concentration, and the purity of the molecule. It was observed that  $\beta$ -sitosterol efficiently prevented adipogenesis in 3T3-L1 adipocytes, evidenced by a considerable reduction in intracellular triglyceride content and lipid accumulation, without including notable cytotoxicity [25].

# CE Inhibitory Activities of the Extracts and Compounds

All extracts and compounds demonstrated CE inhibitory activity (Table 1; Fig. 2). TOE had greater activity than simvastatin, however without statistical significance (IC $_{50}$ =12.443±1.233 µg/mL). TOM, TOE (p<0.0001), and TOC (p=0.001) shown superior effect compared to TOA.  $\beta$ -sitosterol (IC $_{50}$ =14.249±1.209 µg/mL) exhibited significant action comparable to that of simvastatin. This study is the first publication examining the CE inhibitory action of rosmarinic acid and  $\beta$ -sitosterol.

Studies demonstrates the impact of isolated compounds on hyperlipidemia via several pathways. While direct evidence of  $\beta$ -sitosterol, rosmarinic acid, or danshensu suppressing CE is lacking, their influence on lipid metabolism implies a possible function in regulating cholesterol homeostasis.  $\beta$ -sitosterol was observed to decrease cholesterol absorption and plasma cholesterol levels in hypercholesterolemic

hamsters more effectively than sesamin. It was also discovered to diminish the transfer of cholesterol into micelles, maybe due to its structural similarity to cholesterol [26]. Rosmarinic acid (100 mg/kg) was found to improve cholesterol and triglyceride levels comparably to simvastatin in hypercholesterolemic rats on a high-fat diet [27]. Rosmarinic acid may be effective in combating hyperlipidemia through the regulation of lipid metabolism, antioxidant indicators, and inflammatory pathways [28]. Danshensu was found to reduce lipid accumulation in RAW264.7 macrophage cells by increasing intracellular cholesterol efflux [29].

## **Antioxidant Activity Results**

The antioxidant capacities of the extracts and compounds, as assessed by the FRAP and CUPRAC assays, are presented in Table 2. The calibration curves for FRAP and CUPRAC assays are presented in Figure S13. TOE has the highest activity as indicated by both assays (FRAP=1770.021±4.583, CUPRAC=2189.167±5.401  $\mu$ M TEAC). The combination of rosmarinic acid and danshensu exhibited superior antioxidant activity compared to rosmarinic acid alone (FRAP=1405.067±4.491, CUPRAC=2174.167±9.647  $\mu$ M TEAC).

Oxidative stress is a significant risk factor associated with metabolic disorders, including hyperlipidemia, and may also influence enzyme activity [30]. Ethyl acetate extracts often include a high concentration of polyphenols, including



 Table 2 FRAP and CUPRAC values of the extracts and compounds

Extracts/Compounds	FRAP*	CUPRAC**
TOM	$500.147\!\pm\!2.033$	$489.861 \pm 2.508$
TOC	nd***	$86.528\!\pm\!2.051$
TOE	$1770.021\!\pm\!4.583$	$2189.167\!\pm\!5.401$
TOA	$540.132\!\pm\!7.937$	$107.639\!\pm\!3.987$
$\beta$ -sitosterol	$212.067\!\pm\!5.312$	$91.111 \pm 2.265$
Rosmarinic acid	$1274.400 \pm 1.633$	$1801.389\!\pm\!5.500$
Rosmarinic acid+Danshensu	$1405.067\!\pm\!4.491$	$2174.167\!\pm\!9.647$

\*FRAP value indicates iron reducing antioxidant power (µM Trolox equivalent/gram). \*\*CUPRAC value indicates copper reducing antioxidant power (µM Trolox equivalent/gram), \*: not determined. TOM: The methanol extract of Trachystemon on orientalis, TOC: The chloroform subextract of T. orientalis, TOE: The ethyl acetate subextract of T. orientalis, TOA: The remaining aqueous subextract of T. orientalis

rosmarinic acid. Consequently, they significantly impede free radical processes, mostly owing to their redox characteristics. Biyik et al. [16] asserted that the elevated antioxidant activity of T. orientalis is associated with rosmarinic acid. The combination of rosmarinic acid with danshensu, a significantly more polar molecule, has enhanced antioxidant potential. A research assessing the antioxidant activity of S. miltiorrhiza extracts identified danshensu as the most important marker for antioxidant effect [31]. Another investigation indicated that rosmarinic acid and its metabolite danshensu had radical scavenging ability equivalent to that of quercetin [32]. The TOA from which this mixture was obtained seems to have diminished antioxidant ability. The impact is likely diminished by the presence of other minor compounds in the extract.  $\beta$ -sitosterol and the TOC from which it was isolated had low antioxidant capacity. Ertas et al. [33] determined the antioxidant activity of  $\beta$ -sitosterol to be  $A_{0.5}$ : 22.17±0.98 µg/mL using the CUPRAC assay.  $\beta$ -sitosterol functions as a mild to moderate antioxidant and has demonstrated protective benefits against oxidative damage in in vivo experiments [34].

#### **Conclusions**

This study demonstrated that extracts and isolated compounds from T. orientalis shown inhibitory effects on PL and CE, as well as potential in regulating lipid metabolism. Among the tested extracts, TOE, and pure compounds,  $\beta$ sitosterol and rosmarinic acid exhibited strong enzyme inhibition, suggesting their potential therapeutic application in lipid metabolism disorders. Our findings indicate that T. orientalis possesses considerable promise as a natural resource for the management of hyperlipidemia, principally due to its inhibition of lipid metabolism-related enzymes and its antioxidant capabilities. However, more extensive investigations are necessary to fully clarify its therapeutic potential, and the results of this study offer essential data for future research. Future research should concentrate on the impact of T. orientalis and its bioactive components on distinct lipid metabolism pathways to clarify its precise mechanism of action. Furthermore, in vivo investigations are necessary to evaluate its pharmacokinetic characteristics, bioavailability, and systemic effects. T. orientalis has potential for creating innovative nutraceutical and pharmaceutical products for hyperlipidemia and other lipid metabolism disorders.

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Data Availability No datasets were generated or analysed during the current study.

### **Declarations**

Ethical Approval Not applicable.

Consent to Participate Not applicable.

Consent to Publication Not applicable.

**Competing Interests** The authors declare no competing interests.

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#### References

Wang L, Zheng W, Yang J, Ali A, Qin H (2022) Mechanism of Astragalus Membranaceus alleviating acquired hyperlipidemia



- induced by high-fat diet through regulating lipid metabolism. Nutrients 14(5):954. https://doi.org/10.3390/nu14050954
- Wang S, Du Q, Meng X, Zhang Y (2022) Natural polyphenols: a potential prevention and treatment strategy for metabolic syndrome. Food Funct 13(19):9734–9753. https://doi.org/10.1039/d 2fo01552h
- Zhao Q, Fan Y, Zhao L, Zhu Y, Jiang Y, Gu J, Xue Y, Hao Z, Shen Q (2024) Identification and molecular binding mechanism of novel pancreatic lipase and cholesterol esterase inhibitory peptides from heat-treated Adzuki bean protein hydrolysates. Food Chem 439:138129. https://doi.org/10.1016/j.foodchem.2023.138 129
- He X, Chen L, Pu Y, Wang H, Cao J, Jiang W (2023) Fruit and vegetable polyphenols as natural bioactive inhibitors of pancreatic lipase and cholesterol esterase: Inhibition mechanisms, polyphenol influences, application challenges. Food Biosci 55:103054. h ttps://doi.org/10.1016/j.fbio.2023.103054
- Shen H, Wang J, Ao J, Ye L, Shi Y, Liu Y, Li M, Luo A (2023) The inhibitory mechanism of pentacyclic triterpenoid acids on pancreatic lipase and cholesterol esterase. Food Biosci 51:102341. https://doi.org/10.1016/j.fbio.2022.102341
- Yılar M, Onaran A, Yanar Y, Belguzar S, Kadıoglu I (2014) *Trachystemon orientalis* (L.) G. Don (Kaldırık)'ın herbisidal ve antifungal Potansiyeli. Iğdır Univ J Inst Sci Tech 4(4):19–27
- Ayhan BS, Yalcın E, Cavusoglu K, Acar A (2019) Antidiabetic potential and multi–biological activities of *Trachystemon orien*talis extracts. J Food Meas Charact 13:2887–2893. https://doi.org /10.1007/s11694-019-00209-1
- Uzun E, Sariyar G, Adsersen A, Karakoc B, Otuk G, Oktayoglu E, Pirildar S (2004) Traditional medicine in Sakarya Province (Turkey) and antimicrobial activities of selected species. J Ethnopharmacol 95(2–3):287–296. https://doi.org/10.1016/j.jep.2004.07.013
- Karcı E, Gürbüz I, Akaydın G, Günbatan T (2017) Folk medicines of Bafra (Samsun-Turkey). Turk J Biochem 42(4):381–399. https://doi.org/10.1515/tjb-2017-0172
- Sacan O (2018) Antioxidant activity, total phenol, and total flavonoid contents of *Trachystemon orientalis* (L.) G. Don. Eur J Biol 77:70–75. https://doi.org/10.26650/eurjbiol.2018.18017
- Dimcheva V, Kaloyanov N, Karsheva M (2019) The polyphenol composition of Cistus incanus L., trachystemon orientalis L. and Melissa officinalis L. infusions by HPLC-DAD method. Open J Anal Bioanal Chem 3(1):31–38. https://doi.org/10.17352/ojabc.0 00008
- Demir E (2022) Profile of fatty acids, vitamins, phytosterols and phenolic acids in *Trachystemon orientalis* plant and evaluation of its antioxidant activity. EJOSAT 33:112–118. https://doi.org/10.3 1590/ejosat.1027061
- Boga M, Hacıbekiroglu I, Kolak U (2011) Antioxidant and anticholinesterase activities of eleven edible plants. Pharm Biol 49(3):290–295. https://doi.org/10.3109/13880209.2010.517539
- Kurtulus G, Toksoz O, Sesal NC (2022) Potential application of Trachystemon orientalis L. extracts in the cosmetic industries: skincare, photoprotective and antiaging ingredients. Int J Adv Eng Pure Sci 34(1):147–156. https://doi.org/10.7240/jeps.10562
   74
- Koz AN, Subaş T, Badem M, Kanbolat Ş, Özgen U, Şener SÖ, Demir A, Çalış İ (2024) Isolation of the major compounds and determination of biological activities in the underground parts of *Trachystemon orientalis* D.Don. Turk J Pharm Sci 21(6):544– 550. https://doi.org/10.4274/tjps.galenos.2024.60980
- Bıyık B, Yılmaz Sarıaltın S, Gökbulut A, Çoban T, Çoşkun M (2023) *Trachystemon orientalis* (L.) G. Don as a valuable source of Rosmarinic acid: biological activities and HPLC profiles. Turk J Pharm Sci 20(3):141–148. https://doi.org/10.4274/tjps.galenos. 2022.14265

- Erdoğan U, Oztürk TH, Onder S, Tonguç M (2024) Eco-friendly extraction of six different Rosemary (*Rosmarinus officinalis* L.) genotypes-using natural deep eutectic solvents: optimization and modeling via response surface methodology (RSM). J Mol Liq 407:125167. https://doi.org/10.1016/j.molliq.2024.125167
- Bustanji Y, Issa A, Mohammad M, Hudaib M, Tawah K, Alkhatib H, Almasri I, Al-Khalidi B (2010) Inhibition of hormone sensitive lipase and pancreatic lipase by *Rosmarinus officinalis* extract and selected phenolic constituents. J Med Plant Res 4(21):2235–2242. https://doi.org/10.5897/JMPR10.399
- Lee IC, Bae JS, Kim T, Kwon OJ, Kim TH (2011) Polyphenolic constituents from the aerial parts of *Thymus quinquecostatus* Var. *Japonica* collected on Ulleung Island. J Korean Soc Appl Biol Chem 54(5):811–816. https://doi.org/10.1007/BF03253166
- Zhang J, Zhang Q, Liu G, Zhang N (2019) Therapeutic potentials and mechanisms of the Chinese traditional medicine Danshensu. Eur J Pharmacol 864:172710. https://doi.org/10.1016/j.ejphar.20 19.172710
- Watanabe D, Kerakawati R, Morita T, Nakamura T, Ueno K, Kumamoto T, Nakanishi W, Uzawa J, Ishikawa T, Seki H, Tachi M, Harada K, Higuchi Y, Chaichantipyuth C (2009) Isolation of β-Sitosterol and Digalactopyranosyl-diacylglyceride from *Citrus hystrix*, a Thai traditional herb, as pancreatic lipase inhibitors. Heterocycles 78(6):1497–1505. https://doi.org/10.3987/COM-09-11647
- Likitdacharote P, Chaicharoenpong C (2015) Chemical constituents and lipase inhibitory activity of *Solanum torvum* fruit. Proceedings of the Burapha University International Conference 2015:1035–1042
- Ong SL, Mah SH, Lai HY (2016) Porcine pancreatic lipase inhibitory agent isolated from medicinal herb and inhibition kinetics of extracts from *Eleusine indica* (L.) Gaertner. J Pharm (Cairo) 2016:8764274. https://doi.org/10.1155/2016/8764274
- Shalabi AA, El Halawany AM, Choucry MA, El-Sakhawy FS, Morita H, Ki DW, Abdel-Sattar E (2020) New pregnane glycosides from *Caralluma hexagona* Lavranos and their *in vitro* α-glucosidase and pancreatic lipase inhibitory effects. Phytochem Lett 36:49–57. https://doi.org/10.1016/j.phytol.2020.01.015
- Vasanth K, Minakshi GC, Velu K, Priya T, Kumar RM, Kaliappan I, Dubey GP (2022) Anti-adipogenic β-sitosterol and Lupeol from Moringa oleifera suppress adipocyte differentiation through regulation of cell cycle progression. J Food Biochem 46(8):e14170. https://doi.org/10.1111/jfbc.14170
- Lei L, Zhu H, Zhang C, Wang X, Ma KY, Wang L, Zhao Y, Chen ZY (2017) Dietary β-sitosterol is more potent in reducing plasma cholesterol than Sesamin in hypercholesterolemia hamsters. Eur J Lipid Sci Technol 119(7):1600349. https://doi.org/10.1002/ejlt.2 016003
- Sundaram R, Muthu K (2025) Rosmarinic acid alleviates the cardiac abnormalities in high fat diet induced hypercholesterolemic rats. Prostaglandins Other Lipid Mediat 176:106936. https://doi.org/10.1016/j.prostaglandins.2024.106936
- Bekkouch O, Amssayef A, Elbouny H, Eddouks M, Alem C, Amrani S (2023) Rosmarinic acid as a potential anti-hyperlipidemic agent. Plant Sci Today 10(sp2):106–111. https://doi.org/10. 14719/pst.2362
- Gao H, Li L, Li L, Gong B, Dong P, Fordjour PA, Zhu Y, Fan G (2016) Danshensu promotes cholesterol efflux in RAW264. 7 macrophages. Lipids 51:1083–1092. https://doi.org/10.1007/s11745-016-4178-1
- Kim MH, Lee EJ, Cheon JM, Nam KJ, Oh TH, Kim KS (2016) Antioxidant and hepatoprotective effects of fermented red ginseng against high fat diet-induced hyperlipidemia in rats. Lab Anim Res 32(4):217–223. https://doi.org/10.5625/lar.2016.32.4. 217



- Zhou X, Chan SW, Tseng HL, Deng Y, Hoi PM, Choi PS, Or PM, Yang JM, Lam FF, Lee SM, Leung GP, Kong SK, Ho HP, Kwan YW, Yeung JH (2012) Danshensu is the major marker for the antioxidant and vasorelaxation effects of Danshen (*Salvia miltiorrhiza*) water-extracts produced by different heat water-extractions. Phytomedicine 19(14):1263–1269. https://doi.org/10.1016/j.phymed.2012.08.011
- Adomako-Bonsu AG, Chan SL, Pratten M, Fry JR (2017) Antioxidant activity of Rosmarinic acid and its principal metabolites in chemical and cellular systems: importance of physico-chemical characteristics. Toxicol Vitro 40:248–255. https://doi.org/10.1016/j.tiv.2017.01.016
- Ertas A, Cakirca H, Yener I, Akdeniz M, Fırat M, Topçu G, Kolak U (2021) Bioguided isolation of secondary metabolites from Salvia cerino-pruinosa Rech. f. var. cerino-pruinosa. Rec Nat Prod 15(6):585–592. https://doi.org/10.25135/rnp.248.21.01.1933
- Zhang P, Liu N, Xue M, Zhang M, Liu W, Xu C, Fan Y, Meng Y, Zhang Q, Zhou Y (2023) Anti-Inflammatory and antioxidant properties of β-Sitosterol in copper sulfate-induced inflammation in zebrafish (*Danio rerio*). Antioxidants 12(2):391. https://doi.org/10.3390/antiox12020391

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