

Exploring the Potential of Indigenous Foods to Address Hidden Hunger: Nutritive Value of Indigenous Foods of Santhal Tribal Community of Jharkhand, India

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ABSTRACT

Traditional foods of indigenous communities can be explored as a sustainable means of addressing undernutrition. Our study aimed at identifying indigenous foods of the Santhal tribal community of Godda district of Jharkhand, India, assessing their nutritive value, and appraising their potential role in addressing hidden hunger. A cross-sectional survey using qualitative methods like focus group discussions with women of childbearing age (15–49 years), adult males, and elderly people was conducted for food identification. This was followed by taxonomic classification and quantitative estimate of nutritive value of the identified foods either in a certified laboratory or from secondary data. The community was well aware of the indigenous food resources in their environment. More than 100 different types of indigenous foods including a number of green leafy vegetables were identified. Taxonomic classification was available for 25 food items and an additional 26 food items were sent for taxonomic classification. Many indigenous foods (more than 50% of which were green leafy vegetables) were found to be rich sources of micronutrients like calcium, iron, vitamin A as beta carotene, and folate. Maximizing utilization of indigenous foods can be an important and sustainable dietary diversification strategy for addressing hidden hunger in this indigenous community.

KEYWORDS

Indigenous foods; santhal tribes; micronutrients; hidden hunger

Background

Indigenous people are those who retain knowledge of the land and food resources rooted in historical continuity within their region of residence. The food systems of indigenous people often include “traditional foods”; that is, those that are not purchased but obtained locally from the natural environment. They are chiefly procured either through farming or wild harvesting and utilized based on traditional wisdom and knowledge.¹ It is well

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recognized that traditional foods and dietary diversity within an ecosystem can be powerful sources of nutrients and thus are better for health.² Various ethnobotanical surveys indicate that several species of wild plants have been used for human food at some stage of human history.^{3,4} Use of several species of plants, wild fungi, and edible insects has also been documented.⁵⁻⁸ Bushmeat and fish are reported as providing 20% of protein in many developing countries⁸ and indigenous foods in the Gambia have been shown to be important sources of calcium.⁹ The diversity in wild species augments the variety of family diets and may contribute to household food security.¹⁰ However, the health benefits of many of these indigenous foods have been largely unexplored and research on the nutritive value of underutilized species/local varieties deserves a higher priority in nutrition research.

Dietary diversification is a proven cost-effective strategy to ameliorate malnutrition. The loss of dietary diversity has many implications for the nutrition and health of rural communities including loss of income generation and decreased consumption of diverse foods. The multiple benefits of preservation and promotion of indigenous foods range from a collateral benefit on biodiversity and environmental sustainability to improving micronutrient intakes.¹¹

The tribal communities in India are a good example of indigenous populations with a vast diversity in their cultures, traditions, and environments. The numerous indigenous foods that exist in the Indian tribal environment reflect the rich biodiversity of India that can be potentially used to promote food security, nutrition, and health. Some of these indigenous foods have been analyzed and documented from different regions across India.^{2,5,12-15} However, a comprehensive nationwide database is still to evolve and many indigenous foods are yet to be listed and their nutritive value analyzed.

In addition, despite this wealth of traditional knowledge of natural resources in these tribal communities, challenges of geography, agricultural technology, cultural habits, lack of formal education, poor infrastructure, and poverty may contribute to poor nutrition and health.² A high prevalence of chronic energy deficiency and undernutrition along with micronutrient deficiency among tribal populations is well documented.¹⁶⁻¹⁸ Children belonging to tribal communities are at higher risk of iron-deficiency anaemia and vitamin A deficiency disorders. In addition, in women of certain rural and tribal communities, zinc, vitamin B12, and iron constitute the principal micronutrient deficiencies.^{19,20} Chronic micronutrient deficiency resulting from insufficient intake of vitamins and minerals is often referred to as "hidden hunger." It results from lack of dietary diversity and suboptimal and poor-quality food intake and affects around 2 billion people worldwide.²¹ As the name indicates, the signs and symptoms of undernutrition and hidden hunger are less overtly visible in those affected by it²² compared to the immediate response to lack of adequate food; that is, hunger.

The state of Jharkhand in India is among the states and union territories with a significant tribal population. Jharkhand has a total of 30 Scheduled Tribes (an indigenous group of people officially regarded as socially disadvantaged in India). Out of these, Santhal is the most populous tribal community.²³ Studies have reported a high prevalence of undernutrition, chronic energy deficiency, and iron deficiency in the adults and children of the Santhal community residing in different states of India, including Jharkhand, Orissa, and West Bengal.²⁴⁻²⁸ Studies have also documented a wide variety of indigenous foods that are consumed by this community.¹⁵ However, documentation of the nutritive value of many of these foods is not available.

The present study was undertaken to explore the range of indigenous foods consumed by Santhal tribal community of Jharkhand, India. The specific focus was on analyzing their nutritive values and to appraise the potential of these foods in addressing micronutrient deficiencies.

The study involved listing, identification, and taxonomic classification of indigenous foods, followed by nutrient composition analysis, if their nutritive values were not found to be documented in the Indian Food Composition tables.²⁹

Materials and methods

This was an exploratory cross-sectional study conducted in 4 selected villages inhabited by Santhal tribal community in Godda district of Jharkhand, India. The 4 villages were identified using probability proportional to size sampling³⁰ based on a verified list of villages inhabited by Santhal community.

The data collection was conducted between March 2013 and November 2013; multiple visits were scheduled to capture the diversity of foods that were consumed during each season. In addition to the core research team, the study team also included well-trained non-governmental organization (NGO) workers fluent in the native Santhal dialect.

Study procedures

Participatory rapid assessment

Participatory rapid assessment (PRA) methods were used to elicit information on commonly consumed local foods. Focus group discussions (FGDs) were conducted to assess the range of available foods and the contribution of indigenous wild foods to the regular diets of the Santhal community. The female community health workers or *sahiyyas* in the respective villages were requested to invite community members to participate in the FGDs ahead of the field visits. The participants included women with children, adult men,

and the elderly (men and women). Mothers were especially encouraged to attend because they were mainly responsible for food preparation and feeding their families. The FGDs were held in accessible areas such as the *Anganwadi* centers (community centers for children) or in front of the homes of the *sahiyas*. During the FGDs, a discussion guide was used by the study team to steer the conversation toward the participants' knowledge of food groups, foraging and hunting activities, and rearing of animals for food. The study team with the help of the local NGO workers explained the nature of the study and obtained signed written consents from literate participants. Those who could not read or write gave verbal consents with a third-party signatory. Copies of the translated study information sheets and consent forms were given to participants. All participants were informed that the FGDs were going to be recorded and that no personal information would be used in any of the study reports. Permission was taken for pictures to be taken during the FGDs. The local NGO workers transcribed and translated the discussions to Hindi or English. Personal or identifiable information was not recorded in any reports. The aforementioned description of the study thus adhered to the RATS guidelines for reporting qualitative studies.

During the PRA exercise the following methods were adopted:

- (1) The FGD included a free listing exercise to identify indigenous foods consumed in the community and develop a list of such food items. The participants were then asked to identify indigenous or *deshi* foods gathered from the local environment such as nearby forests (jungle), fields, agricultural fields, gardens (*bari* or kitchen garden), or water resources such as man-made ponds (*pokhar*), creeks, or dams or even those bought from the weekly markets (*haat*). The local names of plants or meat items and their characteristics such as availability, seasonality, and source were documented. The foods identified were then categorized under various food groups based on their edible parts. Ethnographic manuscripts on the tribal populations in the area (formerly Bihar state) were also used to confirm the list of common foods.³¹ The text of the FGDs was reviewed through thematic analysis. Atlas.ti version 7³² was used by 2 researchers who independently coded and analyzed the content of the transcripts. Codes were created to help identify overarching themes or similar items under each theme.
- (2) Pairwise ranking: Pairwise ranking was used to identify perceptions, priority setting, and preferences for local food items. After the free listing exercise, the FGD participants were asked to identify 5 to 6 preferred food items within each food group; for example, green leafy vegetables (GLVs), cereals, vegetables, etc. These preferences were based on criteria of taste and availability of particular food items.

Table 1. Pairwise ranking of traditional rice varieties.^a

	Jadhan	Swarna	Lohna	Bhadai	Jondra
Jadhan	×	Jadhan	Jadhan	Bhadai	Jadhan
Swarna	Jadhan	×	Lohna	Swarna	Swarna
Lohna	Jadhan	Lohna	×	Bhadai	Lohna
Bhadai	Bhadai	Swarna	Bhadai	×	Bhadai
Jondra	Jadhan	Swarna	Lohna	Bhadai	×

^aScores: Jadhan, 6; Swarna, 4; Lohna, 3; Bhadai, 6; Jondra, 0. The participants preferred jadhan and bhadai among the identified traditional varieties of rice.

These were then ranked. This helped in identifying the popular and commonly consumed indigenous foods under different food groups.³³

The most commonly consumed food items as identified under each food group category were then entered into a matrix on a flip chart. An example of pairwise ranking is provided in Table 1. Participants were then asked to compare the first food item in the row with various food items listed in the column one by one. The next step was to ask them to move on to the second food item in the row, keeping that as a constant, and comparing it with the third and the subsequent food items and enter the preference in the relevant grid. The same steps were repeated until all of the food items listed in the row were compared with the subsequent food items listed in the columns pairwise. A score was provided based on the number of times each food item was selected.³² Using this method, a hierarchy of preferred food items in the various food groups was identified.

Identification of food samples

Based on the free listing activity done through FGDs, a list of commonly consumed indigenous food items was compiled (including cereals, roots and tubers, legumes/pulses, vegetables, GLVs, seeds, fruits, and animal foods). A literature search was done to identify the taxonomic classification based on the common names provided by the community. Samples were collected for those food items whose taxonomic names were not found, and these were sent for classification to an expert team at the Department of Botany, Birsa Agricultural University, Ranchi, Jharkhand.

Procedure for sample collection for classification

One sample of the food (around 50–100 g) was collected from the field, wrapped in paper towels, and put in a well-perforated polythene bag and sent to the team of experts for identification of taxonomic classification. The food items being sent for classification were photographed and added to the documentation inventory.

After identification and subsequent verification regarding the availability of the nutritive value of the identified food in the Indian food

composition tables, a list of food items was prepared for the purpose of sample collection for nutrient analysis.

Collection of food samples for analysis

The food samples short listed for nutrient analysis were collected from the field site or procured from the local market (whichever was the usual mode of procurement in the community). Each of the samples was dusted to remove excess soil/dirt taking care to avoid mechanical damage and air dried to remove extraneous moisture. The samples were then weighed, wrapped in clean paper towels, placed in well-perforated polythene bags, and placed in a carrier lined with ice packs before being transported to the site of storage and analysis by train. Five hundred grams of each of the vegetables/fruits/green leafy vegetables/tubers and 500 ml of indigenous alcohol samples were sent to the National Accreditation Board for Testing and Calibration Laboratories (NABL) certified laboratory for analysis.

Nutrient analysis

Nutrient analysis was done according to standard reference protocols (Table 2). The analyte values were reported per 100 g of edible weight. All analyses were performed in duplicates. The raw/uncooked samples were analysed for parameters including energy (Codex Guidelines for Nutritional Labelling _CAC/GL 2-1985),³⁴ protein (IS 7219- 1973),³⁵ total fat (IS-4684-1975),³⁶ total carbohydrates (by calculation),³⁷ sugar (titration; FSSAI manual of methods),³⁸ and dietary fiber (AOAC 991.43).³⁹ The vitamins including vitamin A (as beta carotene)⁴⁰, thiamine (vitamin B1),⁴¹ riboflavin (vitamin B2),⁴¹ niacin (Vitamin B3),⁴¹ were estimated by HPLC based UV-visual detection, vitamin C (as L-ascorbic acid)⁴² by titration (IS 5838-1970) and folates (as folic acid) by the BioRad ELISA kit, MA USA. The minerals i.e. calcium, iron, zinc, sodium were analyzed based on the AOAC 999.10 methodology.⁴³ Three local alcoholic beverages were also analyzed for total ethanol content in addition to the other nutrients. The laboratory followed standard quality control and quality assurance programs (including participation in proficiency testing programs) as part of the analytical methodology.

Ethics approval

Ethical approval was obtained from the Public Health Foundation of India's Institutional Ethics Committee. Adult male respondents participated in the study. The female participants were mostly married and were mothers. Although there was a possibility of inclusion of younger women (less than 18 years) who were married at an early age and had children, we did not seek parental consent because we did not consider them as dependent adolescents in the group. Written informed consent was obtained from all participants



Table 2. List of parameters and relevant specifications for nutrient analysis.^a

No.	Test parameter	Unit	Reported method of testing	Reference method of testing	Instrument used (quality control checks done for all instruments as prescribed)
1	Energy	kcal/100 g	IFS/C/STP/FC/008	Codex Guidelines for Nutritional Labelling _CAC/ GL 2-1985	By calculation
2	Protein ($N \times 6.25$)	%	IS 7219-1973	IS 7219-1973	Kjeldahl digestion apparatus
3	Total fat	%	IFS/C/STP/FC/012	IS-4684-1975 Reaffirmed 1983	Soxhlet apparatus
4	Total carbohydrate	%	IFS/C/STP/FC/013	AOAC 986.25	By calculation
5	Sugar	%	IFS/C/STP/FC/010	FSSAI Manual of methods of Analysis of Food, Lab Manual 4 (32)	Titration
6	Dietary fiber	%	IFS/C/STP/FC/007	AOAC 991.43	Gravimetric
7	Vitamin A (as beta carotene)	mg/100 g	IFS/C/STP/LC/025	International Food Research Journal 19(2): 531-535 (2012)	Thermofisher Scientific HPLC UV-Vis with C-18 column
8	Vitamin B ₁	mg/100 g	IFS/C/STP/LC/002	Food analysis by HPLC (33)	Thermofisher Scientific HPLC UV-Vis with C-18 column
9	Vitamin B ₂	mg/100 g	IFS/C/STP/LC/002	Food analysis by HPLC (33)	Thermofisher Scientific HPLC UV-Vis with C-18 column
10	Vitamin B ₃	mg/100 g	IFS/C/STP/LC/002	Food analysis by HPLC (33)	Thermofisher Scientific HPLC UV-Vis with C-18 column
11	Vitamin C	mg/100 g	IS 5838-1970	IS 5838-1970	Titration
12	Calcium	mg/100 g	IFS/C/STP/AAS/004	AOAC 999.10	Thermofisher Scientific AAS
13	Iron	mg/100 g	IFS/C/STP/AAS/004	AOAC 999.10	Thermofisher Scientific AAS
14	Zinc	mg/100 g	IFS/C/STP/AAS/004	AOAC 999.10	Thermofisher Scientific AAS
15	Sodium	mg/100 g	IFS/C/STP/AAS/004	AOAC 999.10	Thermofisher Scientific AAS
16	Folic acid	µg/kg	IFS/M/STP/027	BioRad ELISA Kit	ELISA reader

HPLC indicates high-performance liquid chromatography; UV-Vis, ultraviolet-visible; AAS, atomic absorption spectrometry; ELISA, enzyme-linked immunosorbent assay.

who were literate. Third-party witnessed verbal consents were obtained from illiterate participants.

Results

The present study was conducted in the villages of Bariyarpur, Kadampur, Tilabad, and Mahuatand of the Sunderpahari block of the Godda district of Jharkhand, India.

The outcome of our study related to indigenous knowledge and nutritive values of various traditional food items consumed by the Santhal tribal community is described below:

Food consumption of Santhal community

Rice was the staple food for the community. The community consumed hybrid rice (varieties like Swarna, Pan patta [pan 819], and Chhabbis number), the seeds for which they bought from the local market. The participants preferred the taste, texture, and smell of traditional rice varieties such as jadhan, bahiyad, and bad but most of them had stopped cultivating it. Some families continued to store the seeds and grew them in smaller plots. These varieties were not for everyday household consumption but were only consumed on special occasions. During the pairwise ranking exercise, hybrid rice (Swarna) emerged as the most important cereal crop not because of taste but because it was readily available and affordable. Although the price of traditional rice varieties when sold were reported to be higher in the market, the yields were quite low. The hybrid rice was consumed mostly with green leafy vegetables. Pulses were consumed but not on a daily basis. The respondents reported consuming fleshy foods, which included wild meat, various birds, rodents, molluscs, and fish. Male members of the community still reported hunting for smaller animals in the surrounding forests but larger game was not hunted because it is prohibited by the government. Consumption of roots and tubers both cultivated and from the wild and fruits, especially wild fruits, was also reported. The availability and consumption of a large variety of GLVs were reported in all discussions. Many of these GLVs were also sundried and preserved for use at other times of the year. The *bari* or kitchen gardens provided a ready supply of GLVs and vegetables such as papaya, pumpkin, beans, gourds, lady finger, tomato, onion, garlic, brinjal, jackfruit, etc. Some households that had vegetables in excess of family requirements reported selling them in the market. Some GLVs were procured from the cultivated lands where they grew as weeds, some GLVs were the new leaves of trees within the village, and some were collected from the forest. The discussions revealed that the local government created artificial ponds that could be rented by individuals who were interested in spawning fish. These

individuals and their families consumed the fish and sold the surplus during the weekly markets. Sometimes a couple of households would add their money and rent the ponds as a group and shared the fish among themselves.

Based on the focus group discussion, a free listing of all of the food items that were considered as indigenous foods by the participants was prepared along with their edible parts. The details are provided in [Table 3](#).

Outcome of pairwise ranking

The commonly consumed indigenous foods under each food group were identified using the exercise of pairwise ranking; one such example for preferred GLVs identified during different FGDs is demonstrated in [Figure 1](#).

In the study villages, among cereals, Jadhan, Bahiyad, and Bad, followed by Lohna, were the most common indigenous variety of rice cultivated and consumed along with the hybrid varieties of rice, which were either the most preferred or equally preferred to indigenous varieties. Among the GLVs, sin-arak, gandhari-arak, mung-arak, susni-arak, matha-arak, kantha-arak, hesak-arak, dhurup-arak, and saru-arak (*arak* means green leafy vegetable) were the most preferred and consumed varieties. Out of these, sin-arak, a seasonal variety of GLV, was the most preferred one, followed by munga-arak, which was available throughout the year. In case of vegetables and roots and tubers, no indigenous variety was commonly consumed except for sem (a kind of bean). Kulthi dal was the most preferred indigenous variety of pulse.

Taxonomic classification of indigenous foods

A literature search was done in order to identify these foods based on their local names. For 25 foods, taxonomic classification was available based on their common names in the Indian food composition tables²⁹ and other secondary data sources. For foods for which data from the literature were not available, samples based on availability ($n = 26$) were collected and sent for identification and taxonomic classification to the Birsa Agricultural University (Botany Department). The scientific names were also corroborated from other literature sources including Indian food composition tables ([Table 4](#)). Some representative photographs of these foods taken by the research team are provided in [Figure 2](#) and [Figure 3](#).

Nutritive value of indigenous foods

In addition to the food items documented in the Indian foods composition tables based on their common names, the nutritive values were available for 7

Table 3. Indigenous foods with edible parts and the source.

Name of the food item	English name	Part consumed	Accessed/ grown
Jondra/desi makka	Maize	Grain	Market
Swarna-bad/bahiyad, sorob, chinabora, jadhan	Rice varieties	Grain	Field, market
Bhadai			
Kulthi, lahar	Pulse varieties	Seed	Market
Sutro, khesari, ghangra			Farm
Barbatti			Jungle
Seem/van murgi	Wild hen	Meat	Forest
Kulhai/khargosh	Rabbit	Meat	Forest
Guddu/moosa	Field rat	Meat	Field
Ghonga	Snail	Meat	Farm
Bir sukri	Wild pig	Meat	Forest
Sigga/tood	Squirrel	Meat	Forest
Saahi/gheenk	Porcupine	Meat	Forest
Mahlah	Cat-like animal		Forest
Rundra/chota siyar/van bilaar	Cat-like animal	Meat	Forest
Panduk/chidiya	Sparrow	Meat	Forest
Jhingi	NA	Meat	Forest
Parwa/kabootar	Pigeon	Meat	Market
makhar	<i>Koel</i>	Meat	Forest
Puthi, ich, chatgoi	Varieties of fish	Meat	Pond
Lindra, doodi, gadai, mangri, sising, bhambui, golden, litoor, chepre, gogli			
Jhinuk	Mussel	Meat	Pond
Taaro, tarop/piyar, mirle/kataar, miraal, tiril/kendu, luya, podo bili, khudi rama	NA	Fruit	Forest
Koot			Kitchen
Ul/desi aam	Mango	Fruit	garden,
Janum/ber	Zizyphus	Fruit	field
Bhelua	Marking nut	Fruit	Forest
Amda, barhu/kusum ka phal, dahu	Ambada, kusum fruit	Fruit	Field, forest
Kwindi bili	Mahua	Fruit	Forest
Taad ka phal	Palm fruit	Fruit	Field
Sin-arak/kondra-arak, munga-arak, suya-arak, taaben- arak, jalibi (sweet tamarind), susni-arak, kantha-arak, garundi-arak, chauri-arak, dhurup-arak, kana-arak, lapong-arak, geetil-arak, Sirgiti-arak/siliary	Varieties of green leafy vegetables	Leaves	Weed, kitchen garden
Pindiya-arak, teeri reeti, thampt-arak, ohoic-arak			Weed, field
Hesak-arak, matha-arak			Weed, forest
But-arak, kaddu-arak			Kitchen garden
Allu-arak, saru-arak, pindiya-arak			Field
Daari Gandhari			Market
Sem (3 varieties), bada ghangra	Field beans	Vegetable	Field
Kukri	Sweet variety of bitter gourd		Kitchen garden, market
Bir karela	Small variety of bitter gourd		Forest, field
Pindra			Field
Pindarkoo			Field
Hoterba	Jute	Stem	Forest, field

(Continued)

Table 3. (Continued).

Name of the food item	English name	Part consumed	Accessed/ grown
Kapu		Tuber	Forest
Busu, butu, machi, turmal, damandi, putka, semhu, jangali chhaati	Varieties of mushroom	Mushroom	Forest, field
Kwindi tel	Mahua oil	Oil (from seed)	Market
Hadiya	Rice alcohol	Fermented rice preparation	Home, market
Taadi	Palm alcohol	Prepared from palm	Home, market
Mahua	Mahua alcohol	Prepared from mahua	Market
Khajur tadi	Date alcohol	Prepared from fresh dates	Home, market

FGD 1			FGD 2			FGD 3		
S. No.	GLVs	Votes	S. No.	GLVs	Votes	S. No.	GLVs	Votes
1	<i>Sin arak</i> (leaves from a tree)	3	1	<i>Munga arak</i> (drumstick, leaves)	5	1	<i>Sin arak</i>	4
2	<i>Gandhari</i> (herb)arak	3	2	<i>Gandhari arak</i>	4	2	<i>Hesak arak</i>	3
3	<i>Munga arak</i>	2	3	<i>Sin arak</i>	1	3	<i>Kantha arak</i>	2
4	<i>Dhurup arak</i>	1	4	<i>Saru arak</i>	1	4	<i>Gandhari arak</i>	1
5	<i>Kantha arak</i>	1	5	<i>Kadu arak</i>	1	5	<i>Munga arak</i>	0
			6	<i>Matha arak</i>	0			
			<i>Sin arak</i> is seasonal and available in March. <i>Munga</i> leaves are available throughout the year					

Figure 1. Outcome of pairwise ranking.

foods in the Indian food composition tables from among those classified by the botanists' team at Birsa University. The nutritive values of both of these groups—that is, 7 foods along with foods identified earlier (whose nutritive values were available in Indian food composition tables, $n = 32$)—were compiled and studied. For the rest, food items were procured as per availability from the locality ($n = 13$), including 3 samples of indigenous alcohol, and sent for nutrient analysis to an NABL-certified laboratory in New Delhi. Table 5 provides the nutritive values of these newly analyzed foods.

Many of the indigenous GLVs analyzed as a part of this study, namely, ohic-arak (*Boerhaavia diffusa*), lapong-arak (*Aerua lanata*), and dhurup-

Table 4. List of indigenous foods collected and sent for classification along with verification from other sources of literature.^a

S. No.	Local name of sample collected	Identification done by the botanist		Verification from other sources
		Genus	Species	
1	Hammal bahiyar (rice)	<i>Oryza</i>	<i>sativa</i>	<i>Oryza sativa</i> ²⁸
2	Swarna (rice)	<i>Oryza</i>	<i>sativa</i>	<i>Oryza sativa</i> ²⁸
3	Layu (dried) (millet)	<i>Lathymus</i>	<i>odoratus</i>	Layo, <i>Panicum antidotale</i> Retz. ³²
4	Sutro (pulse)	<i>Phaseolus</i>	<i>calcaratus</i>	<i>Phaseolus calcaratus</i> (common name, sutri) ²⁸
5	Ghangra dal (pulse)	—	—	<i>Dolichos iat</i> Jang, Ling. ³⁴
6	Gandhari-arak (GLV)	<i>Paederia</i>	<i>foetida</i>	<i>Amaranthus spinosus</i> ²⁸
7	Dhurup-arak (GLV)	<i>Leucas</i>	<i>cephalotes</i>	<i>Leucas cephalotes</i> Spreng. (Labiatae) ³²
8	Sin-arak/kondra (GLV)	<i>Bauhinia</i>	<i>purpurea</i>	<i>Bauhinia purpurea</i> (common name, kohar/konar sag) ²⁸
9	Jalibi (GLV)	<i>Pithecellobium</i>	<i>dulce</i>	—
10	Lotni (GLV)	<i>Brassica</i>	<i>juneca</i>	—
11	Kantha-arak (GLV)	<i>Euphorbia</i>	<i>granulata</i>	<i>Euphorbia granulata</i> Forsk. (Euphorbiaceae) ¹⁴
12	Teeri reeti (GLV)	<i>Vicia</i>	<i>hirsuta</i>	Mentioned as pulse in Nollet and Toldra ³⁴
13	Garundi-arak (GLV)	<i>Alternanthera</i>	<i>sessilis</i>	<i>Alternanthera sessilis</i> R. Br. (Amaranthaceae) ¹⁴ <i>Alternanthera sessilis</i> (common name, Ponnanganni) ²⁸
14	Taaben-arak (GLV)	<i>Portulaca</i>	<i>oleracea</i>	—
15	Susni saag (GLV)	<i>Marsilea</i>	<i>minuta</i>	<i>Marsilea minuta</i> Linn. (Marsiliaceae) ¹⁴
16	Hesa-arak (GLV)	—	—	<i>Ficus religiosa</i> ²⁸
17	Chauri-arak (GLV)	<i>Scoparia</i>	<i>dulcis</i>	—
18	Lapong-arak (GLV)	<i>Aerua</i>	<i>lanata</i>	<i>Aerua lanata</i> Juss. Ex Schult (Amaranthaceae) ¹⁴
19	Kadu-arak (GLV)	<i>Lagenaria</i>	<i>vulgaris</i>	<i>Lagenaria vulgaris</i> ²⁸
20	Layu (GLV)	<i>Lathymus</i>	<i>odoratus</i>	Layo, <i>Panicum antidotale</i> Retz. ³²
21	Ohoic-arak (GLV)	<i>Boerhaavia</i>	<i>diffusa</i>	<i>Boerhaavia diffusa</i> Linn. ¹⁴
22	Lahsun saag (GLV)	<i>Allium</i>	<i>sativum</i>	<i>Allium sativum</i> ²⁸
23	Bada ghangra (vegetable)	—	—	<i>Dolichos iat</i> Jang, Ling ³⁴
24	Kapu (tuber)	<i>Dioscorea</i>	<i>bulbifera</i>	—
25	Jangali chati (mushroom)	<i>Pleurotus</i>	spp.	—
26	Bhelua (fruit)	<i>Semecarpus</i>	<i>anacardium</i>	<i>Semecarpus anacardium</i> (common name, marking nut) ²⁸

^aGLV indicates .

arak (*Leucas cephalotes*), were found to be rich sources of calcium (range 202 to 322 mg/100 g of edible portion), iron (10 to 22.06 mg/100 g), and beta carotene (15,000 to 21,000 µg/100 g). Kantha-arak (*Euphorbia granulata*), another GLV, was found to be exceptionally high in iron (81.09 mg/100 g). One of the indigenous pulses, namely, teeri reeti (*Vicia hirsuta*), was found to



Figure 2. Indigenous foods of Santhal tribal community of Jharkhand.

be a good source of calcium (215.25 mg/100 g). Many indigenous foods for which nutritive values were available in the Indian food composition tables, namely, sin-arak (*Bauhinia purpurea*), garundi-arak (*Alternanthera sessilis*), gandhari-arak (*Amaranthus spinosus*), and matha-arak (*Antidesma diandrum*), have high levels of calcium (300 to 1717 mg/100 g); iron in gandhari-arak is reported as 22.9 mg/100 g, and beta carotene in garundi-arak and gandhari-arak is reported in the range of 1926–3564 μ g/100 g. The vitamin C content of munga-arak is reported as 200 mg/100 g. Among lentils, kulthi dal (*Dolichos biflorus*) and sutro dal (*Phaseolus calcaratus*) are reported to be rich in calcium (200–300 mg/100 g). Fleshy foods like snail and freshwater mussel consumed by the community are rich sources of protein and calcium (592–870 mg/100 g). The majority of indigenous foods identified in the present study were found to be rich sources of calcium, iron, and beta carotene.

Discussion

The search for novel, locally available, high-quality, inexpensive foods has continued to be promulgated as an important strategy for meeting nutritional requirements and addressing hidden hunger within a community.

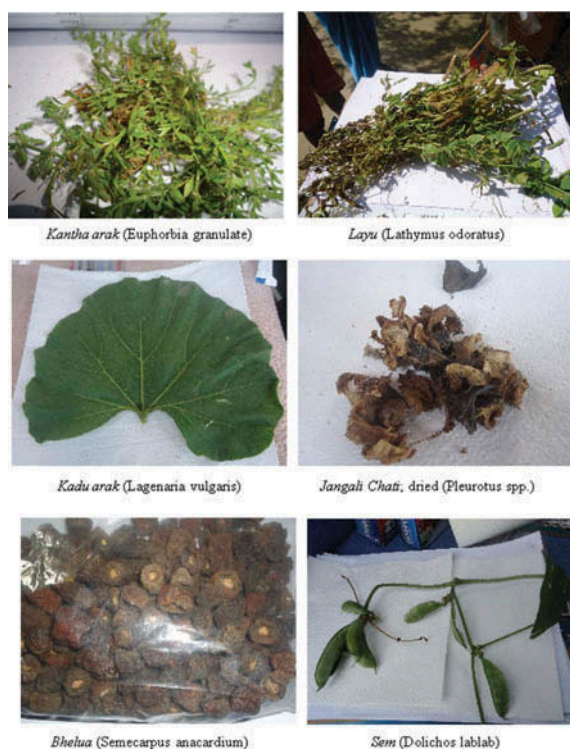


Figure 3. Indigenous foods of Santhal tribal community of Jharkhand.

Maximizing the utilization of indigenous foods can be an important and sustainable dietary diversification strategy for addressing the nutritional needs of an indigenous population. In the present study, a total of 103 types of indigenous foods were identified (Table 1). These included a total of 25 indigenous varieties of GLVs, 6 varieties of pulses, 1 variety of tuber, 7 varieties of vegetable, 17 varieties of fruits, 8 varieties of mushroom, 7 varieties of cereals, 13 varieties of fleshy food items, 13 varieties of fish (plus mussel), and 1 variety of oil. Three varieties of alcohol brewed from locally available plant sources were also identified during the study.⁴⁴

Analysis of nutritive values of the identified indigenous foods showed high levels of micronutrients like calcium, iron, beta carotene, and folate in many of them. A very high level of iron in kantha-arak (*Euphorbia granulate*) as analyzed in the present study has also been reported by Parvez et al.⁴⁵ Studies have documented that indigenous varieties of fruits in communities with normal consumption patterns potentially contributed to fulfilling dietary recommendations of vitamin A.⁴⁶⁻⁵¹ In addition, Ogle et al⁵² found that the daily intake of some naturally occurring vegetables can potentially contribute to fulfill up to 30% and 40% of the recommended allowances of



Table 5. Nutritive values of foods analyzed in the laboratory.

S. No.	Parameter/ food items	Moisture (%)	Energy (kcal/ 100 g)	Protein (g/100 g)	Total fat (g/ 100 g)	Total carbohydrate (g/100 g)	Sugar (g/100 g)	Ethanol (g/100 ml)	Total dietary fiber (g/100 g)	Vitamin A			Vitamin C			Folic acid		
										(beta carotene) (µg/100 g)	B ₁ (mg/ 100 g) ^a	B ₂ (mg/ 100 g) ^a	Vitamin B ₃ (mg/ 100 g)	ascorbic acid (mg/ 100 g)	Calcium (mg/100 g)	Iron (mg/ 100 g)	Zinc (mg/ 100 g)	Sodium (mg/ 100 g)
1	Ohoic-arak (GLV)	82.6	53	3.4	ND	9.8	ND	—	6.1	16 010	ND	ND	12	202	10.68	0.41	39.4	22.3
2	Lapong-arak (GLV)	81.6	56	4.6	ND	9.5	ND	—	5.9	21 760	ND	7.03	19	322	22.06	0.65	10.4	40.5
3	Dhurup-arak (GLV)	80.1	67	5.7	ND	11.1	ND	—	6.7	18 460	ND	ND	8	236	20.02	0.80	10.6	10.7
4	Lahsun saag (GLV)	90.1	34	3.1	ND	5.4	ND	—	4.9	5100	ND	b	7	221	5.95	0.21	10.8	2.9
5	Kantha-arak (GLV)	83.6	46	3.5	ND	8.0	ND	—	7.1	11 680	3.07	8.3	9	425	81.09	1.01	24.9	7.2
6	Hesa-arak (GLV)	65.6	121	2.9	ND	27.3	ND	—	22.3	8200	ND	8.2	ND	295	2.77	0.80	7.5	3.9
7	Ghangra dal (pulse)	7.9	365	24.2	2.3	61.9	1.8	—	28.6	10	ND	ND	5	91	6.07	3.83	4.25	17.89
8	Teeri reeti (pulse)	7.7	361	26.9	1.9	58.9	1.2	—	31.2	550	ND	2.0	23	215	7.78	4.11	33.18	7.11
9	Jadhan (rice)	12.4	351	7.3	0.78	78.7	ND	—	4.9	ND	0.65	5.33	ND	7	1.73	0.97	17.3	5.5
10	Bada ghangra (vegetable)	87.1	49	3.7	ND	8.5	ND	—	4.4	36	ND	ND	9	41	0.95	0.61	1.86	7.0
11	Mahua (alcohol) (per 100 ml) ^c	80.8	134	ND	ND	ND	19.11	—	—	ND	ND	1.55	ND	4	ND	ND	2.1	ND

(Continued)

Table 5. (Continued).

S. No.	Parameter/ food items	Moisture (%)	Energy (kcal/ 100 g)	Protein (g/100 g)	Total fat (g/ 100 g)		Total carbohydrate (g/100 g)	Sugar (g/100 g)	Ethanol (ml)	Total dietary fiber (g/100 g)	Vitamin A (beta carotene)		Vitamin C as L-ascorbic acid			Iron (mg/ 100 g)	Zinc (mg/ 100 g)	Sodium (mg/ 100 g)	Folic acid (µg/ 100 g)
					Energy (kcal/ 100 g)	Protein (g/100 g)					Vitamin B ₁ (mg/ 100 g) ^a	Vitamin B ₂ (mg/ 100 g) ^a	Vitamin B ₃ (mg/ 100 g)	Calcium (mg/100 g)					
12	Hadiya (alcohol) (per 100 ml) ^c	96.5	20	ND	ND	1.6	ND	1.88	—	ND	ND	ND	1.59	ND	12	ND	ND	4.5	0.3
13	Khajur tadi (alcohol) (per 100 ml) ^c	92.0	45	ND	ND	2.7	ND	4.91	—	ND	ND	ND	ND	7	ND	ND	3.3	0.26	

GLV indicates green leafy vegetables; ND, not determined.

^aThe lab used a limit of detection of 0.1 mg/100 g for vitamins B1 and B2. These values are indicative of rich source of these vitamins. Hence, for most of the foods identified the value for these vitamins were not detected.

^bErroneous value.

^cAll parameters are per 100 ml for the alcohol varieties.

vitamin A and calcium, respectively. A study by Singh and Garg⁵³ showed that daily intake of a spice mix can contribute 5%–7% of the recommended daily allowances of some micronutrients (ie, chromium, iron, manganese, zinc, copper, phosphorous, and selenium). These studies strongly support the premise that the indigenous foods identified in the present study, if optimally consumed, can contribute to nutritional security and may address hidden hunger in the Santhal community. The consumption of mostly hybrid varieties of rice observed in the study villages is a cause for concern. This loss of crop diversity may lead to compromised nutrient intake because traditional rice varieties have more fiber and better nutrient composition than high-yield hybrid varieties. In some areas of Jharkhand, specific rice varieties are even used for medicinal purposes and given to lactating mothers or those suffering from dysentery.⁵⁴

The results of our pairwise ranking clearly showed a strong preference for micronutrient-dense indigenous GLVs as part of the daily diet. Daily consumption of local foods is imperative for the food and nutrition security of people living in traditional societies and rural areas.⁵⁵ Further, many of these GLVs were either procured from the wild or grew as weeds in cultivated and noncultivated lands. The local population was also aware of the manifold ways of utilizing these for regular consumption as well as methods of preservation for use during the seasons of the year when they are not available. Additionally, these foods can prove to be an important strategy to complement the routine iron, folate, and calcium supplementation interventions for improving maternal and child micronutrient status in these communities. The neglected and underutilized food resources that are present in indigenous food environments constitute the bedrock of the diversity in traditional and indigenous food systems of developing country communities and would be important in addressing challenges specific to indigenous groups. Studies have indicated a high prevalence of both macro- and micronutrient undernutrition in the Santhal community.^{17–20} This exists amidst a rich knowledge of traditional foods that have potential to contribute to micronutrient intake. Knowledge of the edibility of a wide variety of indigenous flora and fauna exists in the community. What is perhaps missing is the value associated with these foods in terms of their nutritional quality, which could be leading to suboptimal intakes. Thus, there is a need to create awareness about the nutritional quality of these indigenous foods and effectively package the message with promotion of indigenous foods through nutrition education and advocacy. Thus, continuous and sustainable use of indigenous and wild foods can be a cost-effective strategy to address nutritional security and lead to sustainable ecosystem health and nutrition for the Santhal tribal community of Jharkhand.

Limitations

The study was an exploratory work where hitherto undocumented indigenous foods in the Santhal tribal community were listed and analyzed. However, due to logistic reasons including difficult terrain and sample transfer, only a limited number of food items could be analyzed. We believe that there is immense scope for building on this study to expand and consolidate the existing inventory of indigenous foods in the study community.

The laboratory work was outsourced to an NABL-certified lab. The researchers accepted the information provided about the standard procedures and methodology adopted by the laboratory.

Conclusion

The indigenous foods identified in the study were found to be rich sources of micronutrients. These are foods that are adapted to the local agro-ecosystem and do not need any special inputs for their cultivation and sustainability. A substantial contribution to the nutrition security and nutritional status of this indigenous community could be made by promoting the consumption of indigenous foods through creation of an enabling environment for enhancing awareness about their nutritional benefits. Transferring knowledge of these indigenous foods along with their nutritive values to future generations would also facilitate their continued use. The present study may thus pave the path toward further investigations into quantitative consumption estimates of these foods by the community. This would provide information about their contribution to daily micronutrient intake and their potential for alleviating common nutritional deficiencies.

Authors' contributions

SGJ and AS conceived and designed the study with overall supervision from GG. MM and PK developed the qualitative tools and collected and analyzed the qualitative data. SGJ, PK, and AS supervised the collection, identification, and nutrient analysis of food samples. SGJ prepared the first draft of the article. GG, MM, and AS commented on drafts of the article. All authors contributed to critique and modification of the article and read and approved the final version. SGJ had final responsibility for the decision to submit for publication.

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References

1. Kuhnlein HV, Erasmus B, Spigeliski D. *Indigenous Peoples' Food Systems: The Many Dimensions of Culture, Diversity and Environment for Nutrition and Health*. Rome, Italy: Food and Agriculture Organization of the United Nations, Centre for Indigenous Peoples' Nutrition and Environment; 2009.
2. Bhattacharjee L, Kothari G, Priya V, Nandi BK. *The Bhil Food System: Links to Food Security, Nutrition and Health*. Rome, Italy: Food and Agriculture Organization of the United Nations; 2009.
3. Grivetti LE, Ogle BM. Value of traditional foods in meeting macro- and micronutrient needs: the wild plant connection. *Nutr Res Rev*. 2000;13:31–46.
4. Millennium Ecosystem Assessment. *Ecosystems and Human Well-being: Findings of the Sub-global Assessments Working Group of the Millennium Ecosystem Assessment*. Washington, DC: Island Press; 2005.
5. Rathore M. Nutrient content of important fruit trees from arid zone of Rajasthan. *Journal of Horticulture and Forestry*. 2009;1:103–108.
6. DeFoliart G. Insects as human food. *Crop Prot*. 1992;11:395–399.
7. Boa E. *Wild Edible Fungi: A Global Overview of Their Use and Importance to People*. Rome, Italy: Food and Agriculture Organization of the United Nations; 2004.
8. Bennet E, Robinson J. *Hunting of Wildlife in Tropical Forests, Implications for Biodiversity and Forest Peoples*. Washington, DC: The World Bank; 2000. Environment Department Paper No. 76.
9. Prentice A, Laskey MA, Shaw J, et al. The calcium and phosphorus intakes of rural Gambian women during pregnancy and lactation. *Br J Nutr*. 1993;69:885–896.
10. Balemie K, Kebebew F. Ethnobotanical study of wild edible plants in Derashe and Kucha Districts, South Ethiopia. *J Ethnobiol Ethnomed*. 2006;2:53.
11. Bharucha Z, Pretty J. The roles and values of wild foods in agricultural systems. *Philos Trans R Soc B Biol Sci*. 2010;365:2913–2926.
12. Longvah T, Deosthale YG. Compositional and nutritional studies on edible wild mushroom from northeast India. *Food Chem*. 1998;63:331–334.
13. Longvah T, Deosthale YG. Nutrient composition and food potential of *Parkia roxburghii*, a less known tree legume from northeast India. *Food Chem*. 1998;62:477–481.

14. Deosthale YG, Longvah T. Chemical and nutritional studies on Hanshi (*Perilla frutescens*), a traditional oilseed from northeast India. *Journal of Oil & Fat Industries*. 1991;68:781–784.
15. Sinha R, Lakra V. Edible weeds of tribals of Jharkhand, Orissa and West Bengal. *Indian Journal of Traditional Knowledge*. 2007;6:217–222.
16. Mallikharjuna RK, Balakrishna N, Arlappa N, Laxmaiah A, Brahmam GM. Diet and nutritional status of women in India. *J Human Ecol*. 2010;29:165–170.
17. Parimalavalli R. A study of socio-economic and nutritional status of the tribal children. *Studies of Tribes and Tribals*. 2012;10:183–187.
18. Das S, Bose K. Adult tribal malnutrition in India: an anthropometric and socio-demographic review. *Anthropological Review*. 2015;78:47–65.
19. Arlappa N, Laxmaiah A, Balakrishna N, et al. Micronutrient deficiency disorders among the rural children of West Bengal, India. *Ann Hum Biol*. 2011;38:281–289.
20. Menon KC, Skeaff SA, Thomson CD, et al. Concurrent micronutrient deficiencies are prevalent in non-pregnant rural and tribal women from central India. *Nutrition*. 2011;27:496–502.
21. Food and Agriculture Organization. FAO statistical Yearbook, 2012. World Food and Agriculture. Hunger dimensions. Part 2. Available at: <http://www.fao.org/docrep/015/i2490e/i2490e02a.pdf>. Accessed January 20, 2014.
22. Micronutrient Initiative: *Investing in the Future: A United Call to Action on Vitamin and Mineral Deficiencies*. Global Report. Ottawa, ON, Canada: Micronutrient Initiative; 2009. Available at: http://www.unitedcalltoaction.org/documents/Investing_in_the_future.pdf. Accessed March 21, 2014.
23. Office of the Registrar General, India. Jharkand data highlights: the scheduled tribes, Census of India 2001. Available at: http://censusindia.gov.in/Tables_Published/SCST/dh_st_jharkhand.pdf. Accessed March 20, 2015.
24. Dutta CS, Chakraborty T, Ghosh T. Prevalence of under nutrition in Santal children of Puriliya district West Bengal. *Indian Pediatr*. 2008;45:43–46.
25. Chakraborty U, Dutta CS, Dutta G, Ghosh T. A comparative study of physical growth and nutritional status in Santal children of Ghatsila and Bolpur. *Studies of Tribes and Tribals*. 2008;2:79–86.
26. Bose K, Chakraborty F, Mitra K, Bisai S. Nutritional status of adult Santal men in Keonjhar District, Orissa, India. *Food Nutr Bull*. 2006;27:353–356.
27. Chatterjee S, Dhar S, Sengupta B, Sengupta S, Mazumdar L, Chakarabarti S. Coexistence of haemoglobinopathies and iron deficiency in the development of anemias in the tribal population eastern India. *Studies of Tribes and Tribals*. 2011;9:111–121.
28. Rao TVRK, Vijay T. Malnutrition and anemia in tribal pediatric population of Purnia district (Bihar). *Indian Pediatr*. 2006;43:181–182.
29. Gopalan C, Sastri B, Balasubramanian S. *Nutritive Value of Indian Foods*. Hyderabad, India: Indian Council of Medical Research; 2004.
30. Grinnell RM, Unrau YA. *Social Work Research and Evaluation: Foundations of Evidence-based Practice*. New York: Oxford University Press; 2007.
31. Vidyarthi LP. *Changing Dietary Patterns and Habits: A Socio-cultural Study of Bihar*. New Delhi, India: Concept Publishing Company; 1979.
32. Atlas ti software (V7.0). ATLAS.ti Scientific Software Development. London, UK: GmbH.
33. Narayanasamy N. *Participatory Rural Appraisal: Principles, Methods and Application: Principles, Methods and Application*. Sage Publications India Pvt Ltd; 2009.
34. Joint FAO/WHO Food Standards Programme Codex Alimentarius Commission. Food Labelling (5th Ed). FAO; 2007. Available at: <http://www.fao.org/docrep/010/a1390e/a1390e00.htm>
35. BIS-IS. 7219. Method for determination of protein in foods and feeds. 1973 (reaffirmed 2000).

36. Indian Standards 4684–1975; reaffirmed 2000; edition 2.2, appendix F;1.
37. Merrill AL, Watt BK. Energy value of foods, basis and derivation. In *Agriculture Handbook*, No. 74. Washington, DC: United States Department of Agriculture; 1955.
38. FSSAI. Lab Manual 4. Manual of Method of Analysis of Foods. 2014. Available at: <http://www.fssai.gov.in/Portals/0/Pdf/15Manuals/BEVERAGES,%20SUGARS%20&%20CONFECTINERY.pdf>. Accessed May 18, 2015.
39. AOAC International. Official methods of analysis of AOAC International. Official Method 991.43, 17th edition. Gaithersburg, MD, USA; 2000.
40. Norshazila S, Irwandi J, Othman R, Yumi Zuhani H H. Scheme of obtaining β -carotene standard from pumpkin (*Cucurbita moschata*) flesh. *International Food Research Journal* 2012; 19(2):531–535.
41. Pegg RB, Landen WO, Eitenmiller RR. Food Analysis: Vitamin Analysis. In S.S. Nielsen (4th Ed.), *Food Analysis* (pp 179–200). New York: Springer; 2010.
42. BIS-IS. 5838. Methods for estimation of Vitamin C in foodstuffs; 1970 (reaffirmed 2005).
43. Association of Official Analytical Chemists. *Official Methods of Analysis: Lead, Cadmium, Zinc, Copper, and Iron in Foods Sec. 999.10*. 17th edition. Arlington, VA, USA: AOAC; 2002.
44. Bodding PO. Studies in Santal Medicine and Connected Folklore, Part III – How the Santals Live. Kolkata: The Asiatic Society of Bengal; 1925–40 (reprint 2001).
45. Parvez M, Hussain F, Ahmad B, Ali J. *Euphorbia granulata* Forssk as a source of mineral supplement. *Am Eurasian J Agric Environ Sci*. 2013;13:1108–1113.
46. Davey MW, Van den Bergh I, Markham R, Swennen R, Keulemans J. Genetic variability in Musa fruit provitamin A carotenoids, lutein and mineral micronutrient contents. *Food Chem*. 2009;115:806–813.
47. Englberger L, Aalbersberg W, Schierle J, et al. Carotenoid content of different edible pandanus fruit cultivars of the Republic of the Marshall Islands. *J Food Compos Anal*. 2006;19:484–494.
48. Englberger L, Schierle J, Kraemer K, et al. Carotenoid and mineral content of Micronesian giant swamp taro (*Cyrtosperma*) cultivars. *J Food Compos Anal*. 2008;21:93–106.
49. Englberger L, Schierle J, Hofmann P, et al. Carotenoid and vitamin content of Micronesian atoll foods: pandanus (*Pandanus tectorius*) and garlic pear (*Crataeva speciosa*) fruit. *J Food Compos Anal*. 2009;22:1–8.
50. Englberger L, Kuhnlein H, Lorens A, et al. Pohnpei, FSM case study in a global health project documents its local food resources and successfully promotes local food for health. *Pac Health Dialog*. 2010;16:129–136.
51. Englberger L, Lyons G, Foley W, et al. Carotenoid and riboflavin content of banana cultivars from Makira, Solomon Islands. *J Food Compos Anal*. 2010;23:624–632.
52. Ogle BM, Dao HTA, Mulokozi G, Hambraeus L. Micronutrient composition and nutritional importance of gathered vegetables in Vietnam. *Int J Food Sci Nutr*. 2001;52:485–499.
53. Singh V, Garg AN. Availability of essential trace elements in Indian cereals, vegetables and spices using INAA and the contribution of spices to daily dietary intake. *Food Chem*. 2006;94:81–89.
54. Rahman S, Sharma MP, Sahai S. Nutritional and medicinal values of some indigenous rice varieties. *Indian Journal of Traditional Knowledge*. 2006;5:454–458.
55. Penafiel D, Lachat C, Espinel R, et al. A systematic review on the contributions of edible plant and animal biodiversity to human diets. *EcoHealth*. 2011;8:381–399.