

was derived from a patient with no prior CAZ-AVI exposure. Whole-genome sequencing will be performed to identify other genes or mutations that may confer resistance.

Disclosures. All authors: No reported disclosures.

622. The Accessory Genome in Enterococcal Bacteremia: Results from the Vancomycin-Resistant Enterococcal Bacteremia Outcomes Study (VENOUS)

Shelby Simar, MPH¹; Blake Hanson, PhD¹; German Contreras, MD²; Katherine Reyes, MD, MPH³; Pranoti V. Sahasrabhojane, MS⁴; Helina Misikir, MPH⁵; Catherine Liu, MD³; Yohei Doi, MD, PhD⁶; Fernanda Barberis, MD⁷; Lilian Abbo, MD, FIDSA⁸; An Q. Dinh, BS⁹; Maria Spencer, BSc, MSc^{10,11}; Marcus Zervos, MD³; Samuel L. Aitken, PharmD³; Samuel L. Aitken, PharmD⁴; David van Duin, MD, PhD¹²; Samuel A. Shelburne, MD, PhD⁵; Samuel A. Shelburne, MD, PhD⁵; Truc T. Tran, PharmD¹⁰; Jose M. Munita, MD¹³; Cesar A. Arias, MD, MSc, PhD, FIDSA^{14,15}; Maria de los Angeles Spencer, Program Coordinator; ¹School of Public Health, University of Texas Health Science Center at Houston, Houston, Texas; ²McGovern Medical School, University of Texas Health Science Center, Houston, Texas; ³Henry Ford Health System, Detroit, Michigan; ⁴The University of Texas MD Anderson Cancer Center, Houston, Texas; ⁵Fred Hutchinson Cancer Research Center, Seattle, Washington; ⁶School of Medicine, University of Pittsburgh, Pittsburgh, Pennsylvania; ⁷SADI, Buenos Aires, Ciudad Autonoma de Buenos Aires, Argentina; ⁸Miller School of Medicine, University of Miami, Miami, Florida; ⁹Center for Antimicrobial Resistance and Microbial Genomics, University of Texas Health, Houston, Texas; ¹⁰Genomics and Resistant Microbes (GeRM), Instituto de Ciencias e Innovación en Medicina, Facultad de Medicina Clínica Alemana, Universidad del Desarrollo, Chile; ¹¹Millennium Initiative for Collaborative Research on Bacterial Resistance (MICROB-R), Santiago, Region Metropolitana, Chile; ¹²School of Medicine, University of North Carolina, Chapel Hill, North Carolina; ¹³Genomics and Resistant Microbes (GeRM) Group, Millennium Initiative for Collaborative Research On Bacterial Resistance (MICROB-R), Santiago, Region Metropolitana, Chile; ¹⁴CARMiG, University of Texas Health and Center for Infectious Diseases, University of Texas Health School of Public Health, Houston, Texas; ¹⁵Molecular Genetics and Antimicrobial Resistance Unit and International Center for Microbial Genomics, Universidad El Bosque, BOG, COL, Houston, Texas

Session: 66. Molecular and Genomic Epidemiology of Resistant Pathogens
Thursday, October 3, 2019: 12:15 PM

Background. Vancomycin-resistant enterococci (VRE) are a major cause of nosocomial bloodstream infections. Enterococci exhibit remarkable genomic plasticity and can recombine through the acquisition of genetic material via mobile genetic elements (MGEs), including resistance genes. The accessory genome plays a major role in the evolution of enterococci within the human host. Thus, dissecting the entire genome (pan-genome) is of paramount importance to characterize the population structure of enterococci causing disease.

Methods. VENOUS is an ongoing prospective, observational study of adults with enterococcal bacteremia. From September 2016 to March 2018, *E. faecalis* (*Efs*) and *E. faecium* (*Efm*) were collected in 14 hospitals of a single hospital system and a major cancer center in Houston, TX, and a general hospital in Detroit, MI. Short- and long-read genomic sequencing were performed with Illumina MiSeq and Oxford Nanopore Technologies GridION X5, respectively. A proprietary bioinformatics pipeline was utilized for genome assembly and further analyses.

Results. 156 *Efs* and 98 *Efm* isolates from single patients were analyzed. The average proportion of core genes in each genome was 64.6% (53.0–74.1) and 49.1% (45.2–51.0) for *Efs* and *Efm*, respectively. The *vanA* gene cluster was identified in 5.1% (8/157) of *Efs* and 57.1% (56/98) of *Efm*. The plasmid-encoded *aac(6)-Ie-aph(2)-Ia* gene conferring high-level resistance to aminoglycosides was found in 37.6% (59/157) *Efs*, seven of which also possessed *vanA*. Long-read sequencing of *vanA*-harboring plasmids from a subset of VRE revealed that the *vanA* cluster was carried in plasmids ranging from 31.7 to 132.3 kb. Although the *vanA* operon was fairly conserved, insertions of MGE were identified in the intergenic regions of *vanS/vanH* and *vanX/vanY*. Furthermore, a variety of MGE insertions mediated integration of the *vanA* operon, including IS1216 and IS256 (figure).

Conclusion. Accessory genes, including AMR genes, comprise a significant proportion of the enterococcal pan-genome, indicating major genetic plasticity within these organisms. Acquired resistance genes seem to have a high degree of recombination and play a substantial role in the expansion of the genomic repertoire in clinical isolates.

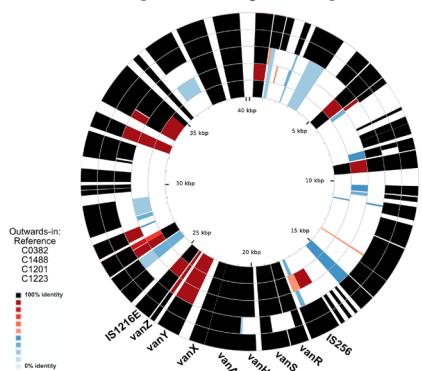


Figure. Composite view of homology within the coding sequences of plasmids containing the *vanA* operon obtained through long-read sequencing. The outermost black ring denotes a reference plasmid containing a conventional *vanA* operon, and similarity to the reference decreases in an inwards direction.

Disclosures. Samuel L. Aitken, PharmD, Melinta Therapeutics: Grant/Research Support, Research Grant; Merck, Sharpe, and Dohme: Advisory Board; Shionogi: Advisory Board.

623. Antimicrobial Resistance in Non-Typhoidal Salmonella from Retail Poultry Meat by Antibiotic Usage-related Production Claims—Pennsylvania, 2008–2017

Xin Yin, MPH¹; Nkuchia M. M'ikanatha, DrPH, MPH²; Lisa Dettinger, Medical Technologist³; Melinda Johnston³; William Eckroth³; Brigitte Husband²; James Tait²; Epiphany Nyirabazhi, PhD³; Heather Tate, PhD⁴; ¹Penn State College of Medicine, Hershey, Pennsylvania; ²Pennsylvania Department of Health, Harrisburg, Pennsylvania; ³Food and Drug Administration, Laurel, Maryland; ⁴US Food and Drug Administration, Laurel, Maryland

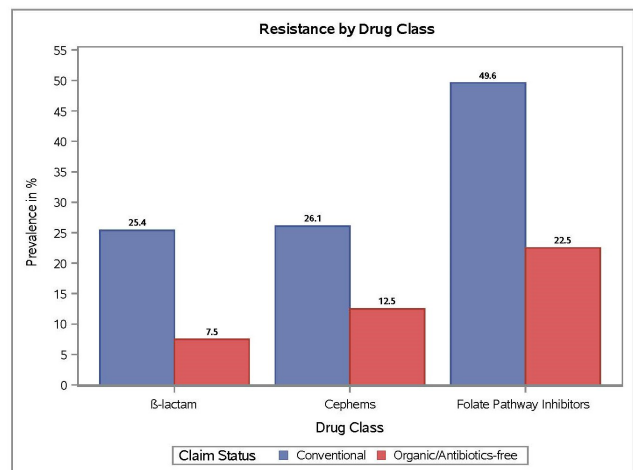
Session: 66. Molecular and Genomic Epidemiology of Resistant Pathogens
Thursday, October 3, 2019: 12:15 PM

Background. Antimicrobial-resistant (AMR) nontyphoidal Salmonella infections are a public health concern. Injudicious use of antimicrobials fuels emergence of resistance. The National Antimicrobial Resistance Monitoring System (NARMS) tracks AMR in Salmonella from humans, animals and foods. There is limited evidence regarding antimicrobial use in food animals and AMR bacteria in retail meat.

Methods. We reviewed antimicrobial susceptibility and whole-genome sequencing data from 320 Salmonella isolated from poultry meat in Pennsylvania as part of NARMS activities. Salmonella strains were isolated from 3,481 samples purchased from randomly selected retail outlets during 2008–2017. Antibiotic usage claims on meat packages were used to compare AMR Salmonella from conventional and antibiotic-free/organic (Abx-free) samples. Genetic mechanisms for AMR were investigated in a subset of isolates.

Results. The prevalence of Salmonella in conventional poultry meat 10.2% (280/2,733) was significantly higher than the prevalence in poultry meat labeled as Abx-free (5.3%, 40/748; $P < 0.0001$). Salmonella from conventional poultry meat was more likely to be resistant to 3 or more drugs (55.0%, 154/280) compared with poultry meat labeled as Abx-free (27.5%, 11/40; $P = 0.0011$). Salmonella from conventional poultry exhibited significantly higher resistance to 4 drug classes including β -lactams ($P = 0.006$) (figure). One hundred isolates from conventional poultry meat and 8 isolates from antibiotic-free/organic samples harbored a gene conferring resistance to the β -lactam class; 24.3% (68/280) of isolates from conventional and 7.5% (3/40) of isolates from Abx-free samples ($P = 0.0145$) contained the extended-spectrum β -lactamase (ESBL) gene blaCMY-2.

Conclusion. Meat samples from conventionally-raised poultry were more likely to be contaminated with AMR Salmonella strains and have genes that reduce the effectiveness of antimicrobial drugs recommended for treatment of severe infections. Contamination of poultry with AMR Salmonella strains is concerning as is the presence of genes that decrease the power of critical antibiotics such as β -lactams. These findings highlight the importance of judicious use of antibiotics in food-producing animals.



Disclosures. All authors: No reported disclosures.

624. Molecular Characterization of Baseline Enterobacteriaceae and Pseudomonas aeruginosa from a Phase 3 Nosocomial Pneumonia (ASPECT-NP) Clinical Trial

Mariana Castanheira, PhD¹; Matthew G. Johnson, MD²; Brian Yu, PharmD²; Jennifer A. Huntington, PharmD²; Patricia Carmelitano, MSc²; Christopher Bruno, MD²; Elizabeth G. Rhee, MD²; Mary Motyl, PhD²; JMI Laboratories, North Liberty, Iowa; ²Merck & Co., Inc., Kenilworth, New Jersey

Session: 66. Molecular and Genomic Epidemiology of Resistant Pathogens
Thursday, October 3, 2019: 12:15 PM

Background. ASPECT-NP, a phase 3, randomized, double-blind, multicenter trial, evaluated ceftolozane/tazobactam (C/T) 3 g q8h vs. meropenem 1 g q8h for

8–14 days in adults for treatment of ventilated nosocomial pneumonia. Baseline Gram-negative (GN) isolates from patients were tested for mechanisms of resistance.

Methods. Lower respiratory tract (LRT) isolates were sent to a central laboratory for organism identification and susceptibility. Of 664 total Enterobacteriaceae (ENT) and *Pseudomonas aeruginosa* (PsA) isolates, 351 (53%) were nonsusceptible to broad-spectrum cephalosporins and/or carbapenems and underwent whole-genome sequencing, quantitative RT-PCR, and western blot analysis. ENT isolates were tested for the presence of acquired β -lactamase genes and AmpC levels (selected species). PsA isolates were tested for acquired β -lactamase genes, AmpC (PDC) levels, efflux pump expression, and OprD loss.

Results. Of 262 ENT isolates, 114 (44%) were susceptible to C/T (MIC ≤ 2 μ g/mL). An extended-spectrum β -lactamase (ESBL) gene was carried by 89 (78%) of the C/T-susceptible isolates. Of 148 C/T-nonsusceptible (C/T-NS) isolates, 87 (59%) were carbapenemase negative, and the majority 135 (91%) also carried an ESBL gene. The most common ESBL was *bla*_{CTX-M15} with *bla*_{OXA-1} and *bla*_{OXA-30}. *Klebsiella pneumoniae* often displayed higher C/T MICs compared with other species carrying the same resistance genes. Among all C/T-NS isolates, 61 (41%) were carbapenemase positive, most commonly *K. pneumoniae* carrying *bla*_{OXA-48}, *bla*_{NDM-1}, and *bla*_{NDM-5}. Of 89 PsA isolates, 58 (65%) were susceptible to C/T (MIC ≤ 4 μ g/mL), despite elevated AmpC expression, efflux pumps, or loss of OprD; only 5 isolates had an acquired β -lactamase. Of the 31 C/T-NS PsA isolates, only 12 (39%) were carbapenemase positive and carried *bla*_{VIM} or *bla*_{GES}; isolates carrying *bla*_{GES} had lower C/T MICs (8–32 μ g/mL) compared with *bla*_{VIM} (MIC > 128 μ g/mL). PDC alleles were similar in isolates with high and low C/T MICs.

Conclusion. In baseline GN LRT isolates from ASPECT-NP, the most common ESBL detected in ENT was *bla*_{CTX-M15}; carbapenemases were uncommon. There was no correlation of ESBL phenotype to C/T susceptibility among ENT, nor of PDC allele to C/T susceptibility among PsA.

Disclosures. All authors: No reported disclosures.

625. Genomic Epidemiology of Carbapenem-Resistant Enterobacteriaceae from Colombia: A Prospective Multicenter Study

Jinnette Reyes, MSc, PhD^{1,2}; Lorena Diaz, PhD^{1,3}; Lina P. Carvajal, PhD Student^{1,4}; Rafael Rios, MSc^{1,5}; Lina V. Millan, MSc¹; Aura M. Echeverri, MSc^{1,5}; Angie K. Hernandez, BSc¹; Sandra Vargas, BSc^{1,6}; Soraya Salcedo, Physician⁷; Adriana Marin, Microbiologist⁸; Laura Mora, MD⁹; Karen M. Ordóñez Diaz, MD¹⁰; Edilberto Cristancho Quintero¹⁰; Sandra Valderrama, MD, ID¹¹; Beatriz Elena Ariza, Bacteriologist¹²; Gloria Cortes, MSc¹²; Laura J. Rojas, PhD¹³; Henry F. Chambers, MD¹⁴; Vance G. Fowler, Jr., MD, MHS¹⁵; Barry Kreiswirth, PhD¹⁶; María Virginia Villegas, MD¹⁷; Robert A. Bonomo, MD¹⁸; Blake Hanson, PhD¹⁹; David van Duin, MD, PhD²⁰; Cesar A. Arias, MD, MSc, PhD, FIDSA^{21,22}; ¹Molecular Genetics and Antimicrobial Resistance Unit, Universidad El Bosque, Bogotá, Colombia; ²International Center for Microbial Genomics, Universidad El Bosque, Bogotá, Distrito Capital de Bogotá, Colombia; ³International Center for Microbial Genomics, Universidad El Bosque, BOG, COL; MICROB-R, Bogotá, Distrito Capital de Bogotá, Colombia; ⁴International Center for Microbial Genomics, Universidad El Bosque Bogotá, Distrito Capital de Bogotá, Colombia; ⁵International Center for Microbial Genomics, Universidad El Bosque, Bogotá, Distrito Capital de Bogotá, Colombia; ⁶International Center of Microbial Genomics, Universidad El Bosque, Bogotá, Distrito Capital de Bogotá, Colombia; ⁷Clinica General del Norte. Universidad Simón Bolívar, Barranquilla, Atlantico, Colombia; ⁸Clinica General del Norte, Barranquilla, Atlantico, Colombia; ⁹Clinica General del Norte, Barranquilla, Atlantico, Colombia; ¹⁰Grupo de Investigación Hospital Universitario San Jorge, Pereira, Risaralda, Colombia; ¹¹Hospital Universitario San Ignacio, Pontificia Universidad Javeriana, Bogotá, Distrito Capital de Bogotá, Colombia; ¹²Hospital Universitario San Ignacio, Bogotá, Distrito Capital de Bogotá, Colombia; ¹³Case Western Reserve University, Cleveland, Ohio; ¹⁴UCSF, San Francisco, California; ¹⁵Duke University Medical Center, Durham, North Carolina; ¹⁶Hackensack Meridian Health, Hackensack, New Jersey; ¹⁷Universidad El Bosque, Bogotá, Distrito Capital de Bogotá, Colombia; ¹⁸Louis Stokes Cleveland VA Medical Center, Cleveland, Ohio; ¹⁹University of Texas Health Science Center School of Public Health, Houston, Texas; ²⁰UNC School of Medicine, Chapel Hill, North Carolina; ²¹CARMiG, UTHealth and Center for Infectious Diseases, UTHealth School of Public Health, Houston, Texas; ²²Molecular Genetics and Antimicrobial Resistance Unit and International Center for Microbial Genomics, Universidad El Bosque, BOG, COL, Houston, Texas

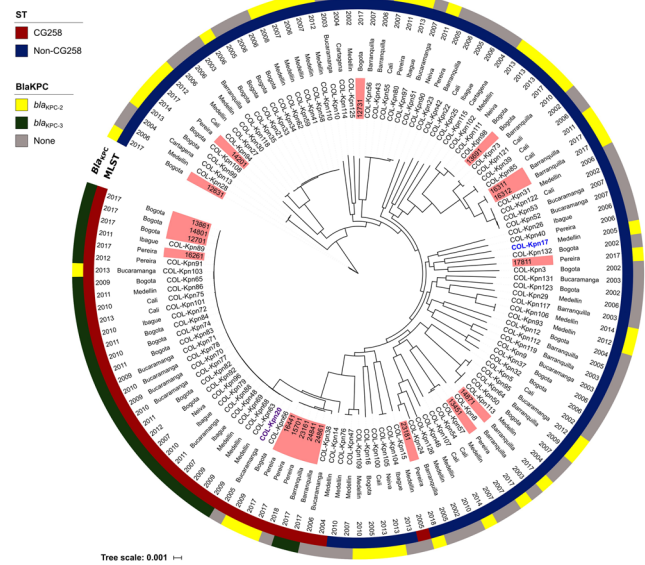
Session: 66. Molecular and Genomic Epidemiology of Resistant Pathogens
Thursday, October 3, 2019: 12:15 PM

Background. Carbapenem-resistant Enterobacteriaceae (CRE) is a serious public health threat. A major epidemic of carbapenemase-producing *Klebsiella pneumoniae* has occurred in Colombia through complex mechanisms of *bla*_{KPC} dissemination. In the framework of a prospective, observational cohort study (CRACKLE-2), we aimed to characterize the genomic epidemiology of CRE circulating in Colombia.

Methods. We performed whole-genome sequencing of 52 isolates collected from the same number of patients (July 2017–April 2018) in 5 Colombian hospitals. Species confirmation and sequence type were determined using Strain Seeker and MLST database. Resistance genes were detected using ResFinder and CARD databases. Phylogenetic reconstruction included additional 108 isolates of carbapenem-resistant *K. pneumoniae* from a previous study.

Results. *K. pneumoniae* (36%), *Escherichia coli* (17%), and *Enterobacter cloacae* complex (17%) were the most frequent species. Genes conferring resistance to carbapenems were detected in 93% of isolates. *bla*_{KPC-2/3}, *bla*_{NDM-1}, and *bla*_{VIM-2/24} were identified in 81%, 15%, and 5% of isolates, respectively. Phylogenetic reconstructions of *K. pneumoniae* showed that clonal group 258 (CG258) were the predominant genetic lineage (Figure 1). Among CG258, ST11 was the most common comprising ca. 26% of isolates. Of note, ST11 had been extremely rare in previous surveillance studies in Colombia. The non-CG258 were from 9 different STs and exhibited high genomic diversity. Among *E. coli* isolates 33% belonged to the high-risk clone ST131 harboring *bla*_{KPC-2} and we detected both *bla*_{KPC-2} and *bla*_{VIM-24} in 1 *E. coli* ST131 isolate. ST510 *E. cloacae* complex harboring *bla*_{KPC-2} was the most common (44%) lineage.

Conclusion. *K. pneumoniae* and *E. coli* are the most frequent CRE isolated from patients in Colombian hospitals. Dissemination of *bla*_{KPC} through horizontal gene transfer to several species of Enterobacteriaceae continues to be a common mechanism of spread. Although KPC continues to be the most common carbapenemase, a rise in high-risk clonal lineages harboring metallo-carbapenemases, in particular NDM-1 is worrisome. Our results indicate emergence of virulent genetic lineages of *K. pneumoniae* ST11 and *E. coli* ST131 carrying carbapenemases in Colombia.



Disclosures. All authors: No reported disclosures.

626. Mobile Genetic Element Dynamics of Co-Circulating Klebsiella pneumoniae Sequence Types Carrying blaKPC in Houston, Texas

William C. Shropshire, MPH¹; An Q. Dinh, BS²; William R. Miller, MD²; Heather Ecklund, BS³; Audrey Wanger, PhD⁴; Truc T. Tran, PharmD²; Cesar A. Arias, MD, MSc, PhD, FIDSA^{5,6}; Blake Hanson, PhD¹; ¹School of Public Health, University of Texas Health Science Center, Houston, Texas; ²Center for Antimicrobial Resistance and Microbial Genomics, University of Texas Health, Houston, Texas; ³University of Texas Health, Houston, Texas; ⁴University of Texas Health Science Center, Houston, Texas; ⁵CARMiG, University of Texas Health and Center for Infectious Diseases, University of Texas Health, School of Public Health, Houston, Texas; ⁶Molecular Genetics and Antimicrobial Resistance Unit and International Center for Microbial Genomics, Universidad El Bosque, BOG, COL, Houston, Texas

Session: 66. Molecular and Genomic Epidemiology of Resistant Pathogens
Thursday, October 3, 2019: 12:15 PM

Background. Carbapenem-resistant *Klebsiella pneumoniae* (CR-Kpn) are a significant cause of hospital-associated infections. Class A β -lactamases, e.g., *Klebsiella pneumoniae* carbapenemases (KPCs), are major contributors to carbapenem resistance. Sequence type 258 (ST258) is the most common genetic lineage of CR-Kpn associated with *bla*_{KPC} carriage. Recently, a newly emergent lineage ST307 has been identified within the Houston metropolitan region. The transmission of *bla*_{KPC} and other antimicrobial resistance (AMR) genes is driven largely by exchange of mobile genetic elements (MGEs). We sought to describe the dynamics of horizontal gene transfer (HGT) in particular between co-circulating strains of ST307 and ST258.

Methods. Long-read sequencing technologies allow us to resolve plasmid sequences and their associated AMR genes as well as characterize a comprehensive range of MGEs enabling transmission of these clinically important resistance mechanisms. CR-Kpn isolates were collected as part of a study to describe CRE burden within a Houston metropolitan hospital system. The Oxford Nanopore Technology (ONT) GridION X5 was used for long-read sequencing with Illumina short-read data used to refine and generate high-quality, consensus assemblies. A custom bioinformatic pipeline was used to resolve plasmid structures and identify the genomic context of plasmids carrying *bla*_{KPC} variants.