PLASTOME REPORT

OPEN ACCESS

The complete chloroplast genome of *Orthosiphon aristatus* (Blume) Miq. (Lamiaceae)

Yongjing Su^a* (b), Bing Li^{b*}, Wenwen Liang^a, Haicheng Wen^a (b) and Wei Wei^a

^aFaculty of Zhuang Medicine, Guangxi University of Chinese Medicine, Nanning, China; ^bSchool of Pharmacy, Guangxi University of Chinese Medicine, Nanning, China

ABSTRACT

Orthosiphon aristatus (Blume) Miq. 1858 is a Lamiaceae plant. It is mainly found in southern China. It is an excellent medicinal plant. The complete chloroplast genome of *O. aristatus* is 152,155 bp in length, with an average depth of $287 \times$, and the GC content was 37.86%, a large single-copy (LSC) region of 83,098 bp, a small single-copy (SSC) region of 17,665 bp, and an inverted repeats (IRs) region of 25,696 bp make up the genome's typical tetragonal shape. In addition, the genome consisted of 128 genes, including 85 protein-coding genes, 35 transfer RNA (tRNA), and eight ribosomal RNA (rRNA) genes. A monophyletic group was established by *O. aristatus* and 13 plants from five genera of Lamiaceae, according to the phylogenetic tree. In contrast, an isolated monophyletic group was formed by the alien plant *Cinnamonum aromaticum*. The ML tree bootstrap value was relatively high, and *O. aristatus* was most closely related to *Ocimum tenuiflorum* and *Ocimum basilicum*. This study can help with species identification and phylogenetic analysis within *O. aristatus* and Lamiaceae species.

ARTICLE HISTORY

Received 2 November 2023 Accepted 27 December 2023

Taylor & Francis

Taylor & Francis Group

KEYWORDS

Chloroplast genome; Lamiaceae; Orthosiphon aristatus; phylogenetic tree

1. Introduction

Orthosiphon aristatus (Blume) Miq. 1858 is a Lamiaceae plant. It is primarily found in southern China, where it is frequently used to treat illnesses of the urinary system such as nephritis. cystitis, and urinary calculi (Wen and Wei 2019). It possesses diuretic, uric acid-lowering, antioxidant, and anti-inflammatory effects, according to a recent study (Ameer et al. 2012). Diterpenoids, triterpenoids, phenylpropanoids, and flavonoids are responsible for these pharmacological actions (AbdAziz et al. 2021). There is currently no whole chloroplast (cp) genome information available for Orthosiphon. As a result, we sequenced the whole cp genome of O. aristatus and examined its phylogenetic connection within the Lamiaceae family. Studying and analyzing O. aristatus's cp genome will help with species identification, clinical safe medication usage, and resource protection for O. aristatus. It will also explain O. aristatus's phylogenetic position and genetic link in Lamiaceae plants.

2. Materials and methods

We extracted the genomic DNA of *O. aristatus* from fresh leaves (Figure 1) collected on the campus of Guangxi University of Chinese Medicine (N22°48'14"; E 108°30'4").

The Botanical Herbarium of the Guangxi University of Chinese Medicine houses the gathered specimens (voucher number 202304SCC001, Haicheng Wen, wenhaicheng2015@ gg.com). Whole genome DNA is extracted and purified using the CTAB method from O. aristatus fresh leaves (Liu et al. 2018); the qualifying samples were utilized to build a library, and the qualified library was sequenced on the Illumina HiSeg 2000 platform (Han et al. 2018; Wen et al. 2023). The segmented contig sequences were spliced using NOVOPlasty (version 2.7.2) software (Dierckxsens et al. 2017) and further assembled into scaffold sequences. GeSeg (version 1.78) software is then used to annotate the sequence, after which it has been submitted to the online NCBI database (accession number: OR387684). The genomic circle map of O. aristatus was created using the online CPGView program (http://www. 1kmpg.cn/cpgview) (Liu et al. 2023). The complete cp gene sequences of 13 Lamiaceae-related plants and one outgroup plant were downloaded from the NCBI database, to explore the phylogenetic relationship of O. aristatus. The genome sequences were then compared and corrected using the maximum-likelihood (ML) method and the Tamura-Nei model using MEGA 11 software. The ML tree has been constructed and the bootstrap parameter is set to 1000 repeats to obtain reliable data (Kumar et al. 2018).

CONTACT Haicheng Wen 😒 wenhaicheng2015@qq.com; Wei Wei 😒 65462479@qq.com 🗈 13th Wuhe Avenue, Qingxiu District, Nanning, Guangxi 530200, China

^{*}Both authors contributed equally to this work.

Supplemental data for this article can be accessed online at https://doi.org/10.1080/23802359.2023.2301012.

^{© 2024} The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited. The terms on which this article has been published allow the posting of the Accepted Manuscript in a repository by the author(s) or with their consent.



Figure 1. Reference image of *O. aristatus* taken by Haicheng Wen at the botanical gardens of Guangxi University of Chinese Medicine. (A) Vegetative body. Erect, quadrangular, and pubescent in the back stems. Leaves are opposite; they are ovoid, rhomboid, or ovoid-elliptic, 28.5 cm long and 1.5 cm wide, with a tapered apex, a base that is broadly wedge-shaped or descends to the petiole, a toothed edge or sparse teeth above the base, a tooth tip with small spikes, and pubescent glands on both sides. (B) Blooms. The cymes produce irregular racemes; ovoid-shaped bracts; bell-shaped calyx; pubescent and glandular exteriors; increased following flowering; The corolla is pale purple or white, puberulent on the surface, and has glandular patches on the top lip. It also has a big upper lip that is everted and a three-lobed structure with a broad central lobe. four stamens, somewhat longer than the front pair and extending far beyond the corolla tube; four-lobed ovary, lengthy style, and two-lobed stigma; The disk has a fingerlike shape on the front.

3. Results

O. aristatus has a 152,155 bp long cp genome, with an average depth of $287 \times$ (Supplementary Figure 1), a large singlecopy (LSC) region of 83,098 bp, a small single-copy (SSC) region of 17,665 bp, and an inverted repeats (IRs) region of 25,696 bp make up the genome's typical tetragonal shape (Figure 2). There is 37.86% of GC content overall, 35.90% of GC content in the LSC region, 31.76% of GC content in the SSC region, and 43.12% of GC content in the IR region. There is little doubt that the GC content in the IR regions is higher than in the two single-copy regions (Table 1). The cp genome O. aristatus included 128 genes, including 85 protein-coding genes, 35 transfer RNA (tRNA), and eight ribosomal RNA (rRNA) genes. A monophyletic group was established by O. aristatus and 13 plants from five genera of Lamiaceae, according to the phylogenetic tree, while an isolated monophyletic group was formed by the alien plant Cinnamomum aromaticum (Figure 3). Supplementary Figures 2 and 3 present the results of cis-splicing for correction and annotation by multi-sequence alignment. Of the 14 genes (rps16, atpF, rpoC1, rpl2, ndhB, ndhA, trnG-UCC, trnL-UAA, trnI-GAU, trnA-UGC, trnK-UUU petB, petD, and rp116) all found to contain an intron. two genes (*paf* and *clp*P1) contained two introns, one small-exon gene (rps16), and one trans-splicing gene (rps12).

The complete cp genome map of *O. aristatus*. It was generated using CPGView. The circular cp genome map displays 85 protein-coding genes, 35 tRNA genes, and eight rRNA genes. Different categories of genes are labeled with distinct colors. The transcription directions for the inner and outer genes are clockwise and anticlockwise, respectively; genes are color-coded by their functional classification; the functional classification of the genes is shown in the bottom left corner; from the center outward, the first track shows the dispersed repeats. The dispersed repeats consist of direct (D) and Palindromic (P) repeats, connected with red and green arcs; the second track shows the long tandem repeats as short blue bars; the third track shows the short tandem repeats or microsatellite sequences as short bars with different colors: black: c (complex repeat); green: p1 (repeat unit size = 1); yellow: p2 (repeat unit size = 2); purple: p3 (repeat unit size = 3); blue: p4 (repeat unit size = 4); orange: p5 (repeat unit size = 5); red: p6 (repeat unit size = 6); the SSC, inverted repeat (IRa and IRb), and LSC regions are shown on the fourth track; the GC content along the genome is plotted on the fifth track; the base frequency at each site along the genome will be shown between the fourth and fifth tracks; the genes are shown on the sixth track.

Maximum-likelihood method was used to generate a phylogenetic tree based on 14 representative species. As outgroup taxa, Cinnamomum aromaticum were used the bootstrap support values are shown at the branches after 1000 bootstrap repeats were computed. The following sequences were used: Ocimum tenuiflorum MN687904, Ocimum basilicum KY623639, Orthosiphon aristatus OR387684, Isodon japonicus MW691144 (Wang et al. 2022), Isodon lophanthoides MT317098 (Zhang et al. 2020), Marmoritis complanata OP186462, Mentha canadensis MN047448 (Huaizhu et al. 2019), Mentha spicata MG256495 (Wang et al. 2017), Mentha x piperita MN736963, Scutellaria baicalensis KR233163, Scutellaria barbata MW376479, Clerodendrum japonicum MW222242 (Long et al. 2021), Clerodendrum cyrtophyllum MZ958825, Clerodendrum colebrookianum ON149875, and Cinnamomum aromaticum MN812496.

4. Discussion and conclusions

The genomic structure of *O. aristatus* consists of two IRs, an SSC region, and an LSC region similar to homologous species *Isodon japonicus* (Wang et al. 2022). The ML tree bootstrap



Figure 2. Chloroplast genome map of O. aristatus.

Table 1. Structure and composition of O. aristatus chloroplast genome.

Region	GC (%)	Length (bp)
Chloroplast genome	37.86	152,155
LSC	35.90	83,098
SSC	31.76	17,665
IRA	43.12	25,696
IRB	43.12	25,696

tenuiflorum, and *O. basilicum*, and this work provides new data on the phylogeny, species identification, species resource conservation, and genetic investigations of *O. aristatus*.

Author contributions

HCW and WW designed the experiment and obtained the funding. YJS and BL performed laboratory work (sample collection, DNA extraction, library construction, and sequencing). YJS and BL performed bioinformatics analyses. HCW and WW wrote and revised the manuscript, and all authors reviewed it.

value was quite high, and *O. aristatus* was most closely related to *Ocimum tenuiflorum* and *Ocimum basilicum*, indicating that they might have shared an ancestor. Their revealed genetic information will be beneficial to further refine detailed and in-depth investigations into this species of evolutionary history.

This study is the first to analyze the cp genome of *O. aristatus* and employs phylogenetic trees to examine the connections between similar species. The phylogenetic tree suggests a close relationship between *O. aristatus*, *O.*

Ethics statement

In the present study, *O. aristatus* plants were collected from the campus of Guangxi University of Chinese Medicine, and permission for the collection of samples was obtained from the Nanning Wild Animal and Plant Protection Station. We complied with the regulations of the Guangxi Zhuang Autonomous Region on the protection of wild plants.



0.02

Figure 3. Maximum-likelihood phylogenetic tree composed of the complete gene sequences of 14 related species obtained based on MEGA X analysis.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This work was supported by the Research Fund of Guangxi Key Laboratory of Efficacy Study on Chinese Materia Medica [Grant number 2006538].

ORCID

Yongjing Su (b) http://orcid.org/0000-0002-9364-0984 Haicheng Wen (b) http://orcid.org/0000-0002-5144-2277

Data availability statement

Genomic sequence data supporting the findings of this study are publicly available at the NCBI GenBank at https://www.nibi.nlm.nih.gov under accession number OR387684. The associated BioProject, Bio-Sample, and SRA numbers are PRJNA1000189, SAMN36761994, and SRR25457748, respectively.

References

- AbdAziz NA, Hasham R, Sarmidi MR, Suhaimi SH, Idris MKH. 2021. A review on extraction techniques and therapeutic value of polar bioactives from Asian medicinal herbs: case study on Orthosiphon aristatus, Eurycoma longifolia and Andrographis paniculata. Saudi Pharm J. 29(2): 143–165. doi:10.1016/j.jsps.2020.12.016.
- Ameer OZ, Salman IM, Asmawi MZ, Ibraheem ZO, Yam MF. 2012. Orthosiphon stamineus: traditional uses, phytochemistry, pharmacology, and toxicology. J Med Food. 15(8):678–690. doi:10.1089/jmf.2011. 1973.
- Dierckxsens N, Mardulyn P, Smits G. 2017. NOVOPlasty: de novo assembly of organelle genomes from whole genome data. Nucleic Acids Res. 45(4):e18. doi:10.1093/nar/gkw955.

- Han R, Xie D, Tong X, Zhang W, Liu G, Peng D, Yu N. 2018. Transcriptomic landscape of *Dendrobium huoshanense* and its genes related to polysaccharide biosynthesis. Acta Soc Bot Pol. 87(1):3574.
- Huaizhu L, Lin Y, Bai J, Qiuyan A, Lingling D, Rui-Xue S. 2019. The complete chloroplast genome sequence of *Mentha canadensis* (Labiatae), a traditional Chinese herbal medicine. Mitochondrial DNA B Resour. 5(1):55–56. doi:10.1080/23802359.2019.1687031.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: molecular evolutionary genetics analysis across computing platforms. Mol Biol Evol. 35(6):1547–1549. doi:10.1093/molbev/msy096.
- Liu L, Han R, Yu N, Zhang W, Xing L, Xie D, Peng D. 2018. A method for extracting high-quality total RNA from plant rich in polysaccharides and polyphenols using *Dendrobium huoshanense*. PLOS One. 13(5): e0196592. doi:10.1371/journal.pone.0196592.
- Liu S, Ni Y, Li J, Zhang X, Yang H, Chen H, Liu C. 2023. CPGView: a package for visualizing detailed chloroplast genome structures. Mol Ecol Resour. 23:1–11.
- Long X, Pan Y, Weng Y, Hao Z, Ye D, You Y, Chen J, Shi J. 2021. The complete chloroplast genome of *Clerodendrum japonicum* (Thunb.) Sweet, a traditional Chinese medicinal plant. Mitochondrial DNA B Resour. 6(3):851–852. doi:10.1080/23802359.2021.1885316.
- Wang H, Wang Y, Lu Y, Zhu S, Huang J, Yue C. 2022. Characterization of the complete chloroplast genome sequence of *Isodon japonicus* (N. Burman) H. Hara (Lamiaceae). Mitochondrial DNA B Resour. 7(9): 1713–1715. doi:10.1080/23802359.2022.2123718.
- Wang K, Li L, Hua Y, Zhao M, Li S, Sun H, Lv Y, Wang Y. 2017. The complete chloroplast genome of *Mentha spicata*, an endangered species native to South Europe. Mitochondrial DNA B Resour. 2(2):907–909. doi:10.1080/23802359.2017.1413311.
- Wen H, Su Y, Xie A, Lin C, Wei W. 2023. Characteristics and phylogenetic analysis of the complete chloroplast genome of *Primulina hedyotidea*. Mitochondrial DNA B Resour. 8(9):1007–1011. doi:10.1080/23802359. 2023.2238932.
- Wen HC, Wei W. 2019. Fundamentals of Zhuang Medicine. Nanning: Guangxi Science and Technology Press; p. 296–297.
- Zhang HY, Xia J, Ma WZ. 2020. The complete plastome of a folk medicinal herb *Isodon lophanthoides* var. graciliflorus. Mitochondrial DNA B Resour. 5(3):2219–2221. doi:10.1080/23802359.2020.1768949.