Original Article





*Maryam NIYYATI, Alireza NAGHAHI, Hamed BEHNIAFAR, Zohreh LASJERDI

Dept. of Medical Parasitology and Mycology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

*Corresponding Author: Email: maryamniyati@sbmu.ac.ir

(Received 09 Apr 2017; accepted 16 Sep 2017)

Abstract

Background: The presence of potentially pathogenic Free Living Amoebae (FLA) in hospital environment could be a health hazard for high-risk patients such as immunosuppressed patients. This study was carried out to investigate the presence of potentially pathogenic FLAs in the environment and medical instruments of different hospital wards, and nasal swabs of immunosuppressed patients of a hospital in Tehran, Iran.

Methods: In this cross-sectional study, 60 environmental (26 samples) and nasal swab (34 samples) samples were collected between Dec 2015 and Feb 2016. The samples were assessed using culturing, staining and morphological methods based on page key. To decrease the bacterial and fungal contamination and better identification of FLAs, cloning was performed.

Results: Overall, 17 (28%) samples, including 13 environmental samples and 4 nasal swabs samples, were found positive for FLAs. The most frequent amoebae were *Acanthamoeba* spp. and two plates had mix contamination of *Acanthamoeba* spp. and Vahlkampfiids/*Vermamoeba*. Overall, *Acanthamoeba* species (58%), Vahlkampfiids (26%) and *V. vermiformis* (15%) were identified in clinical and environmental samples.

Conclusion: The occurrence of these FLAs in environmental and clinical samples of hospital may threat health status of patients directly, particularly in immunosuppressed patients, and can transmit other pathogens. Thus, the increasing awareness of clinical setting staffs about FLAs and improvement of disinfection methods in hospitals is needed.

Keywords: Immunosuppression, Hospital, Iran

Introduction

Free-living amoebae (FLA) are the opportunistic, amphizoic and ubiquitous protozoan. FLAs have global distribution and were isolated from various resources such as water, soil, air, wastewater and other environmental resources, and clinical samples (1). Some genera of FLAs such as *Naegleria, Acanthamoeba, Balamuthia*, and *Sappinia* are medically important and are causative agents of opportunistic and non-opportunistic infections in humans (1-3). Recently, researchers have reported *Vahlkampfia*, *Paravahlkamfia*, and *Vermamoeba* responsible for FLA-related diseases (4-7).

There is an increasing trend regarding *Acanthamoeba* keratitis (AK) in Iran (8). However, no researches have been done to detect FLAencephalitis in this region. There is a single report presenting *N. fowleri* occurrence in a 5-month infant in Iran. This patient recovered using drug therapy (9).

The immunological status of the host is the main risk factor for developing *Acanthamoeba* encepha-

litis and the raised number of immunodeficient hosts has resulted in an increase of the infection incidence (10). The majority of GAE cases have occurred in immune-suppressed patients such as HIV, graft receivers, steroid users, leukemia, and cirrhotic and hepatitis patients (10, 11). The main routes of GAE acquisition are inhalation of cysts and entrance of the agent through skin wound (2). The presence of free-living amoeba cysts in hospital environment is a health threat. Moreover, these amoebas can transport and transmit pathogenic bacteria (12). Cyst of FLA including Acanthamoeba spp. are very resistant to disinfectants and they can resist harsh environment (13). Acanthamoeba could present in oxygen mask of an isolated room (14).

There is limited data regarding the presence of free-living amoeba in both hospitalized patients and their clinical setting environment. Indeed, few studies have been conducted in Iran regarding the occurrence of *Acanthamoeba* spp. in cancer patients and immunosuppressed patients (15).

Intensive care unit and critical care unit admit patients with severe condition and any contamination of these wards to pathogenic microorganisms could be a healthy treat for patients. Occurrence of FLAs in the medical instruments and hospital environment can be a risk factor to the health of patients (16).

Therefore, the main aim of the present study was the determination of the occurrence of FLAs in immune-suppressed patients and in dust from ICU and CCU hospital ward in Tehran, Iran, using culturing and microscopic methods. To the best of our knowledge, this is the first study that investigates the presence of FLAs in environmental sources of hospital wards, medical instruments, and patients, simultaneously.

Materials and Methods

Samples

In this cross-sectional study, 60 samples were collected between Dec 2015 and Feb 2016, from intensive care units and critical care units of a hospital in Tehran, Iran. In this study, various locations were selected including surgical, general and heart ICU and CCU randomly for taking samples, and sterile swabs were used for sample collection. All samples were transferred within 24 h to the Protozoology Laboratory of Shahid Beheshti University of Medical Science, Tehran, Iran.

Clinical samples

Overall, 34 nasal swabs were collected from hospitalized patients. Due to corticosteroid therapy, all of the selected patients were immunesuppressed. Selected patients were hospitalized in CCU (16), surgical ICU (5), general ICU (8) and open heart surgery ICU (5) wards.

All patients were informed regarding the study procedure and they were all satisfied to participate in the present research. Ethics Committee of the university approved the study.

Environmental samples

26 dust samples were collected from CCU (16), surgical ICU (4), general ICU (4) and open heart surgery ICU (2) wards. Samples were taken from medical instruments, central air conditioners, windows, and doors. Because of isolation in ICU wards, we collected our samples only from central air conditioners.

Isolation and identification of FLAs based on page key

All of environmental swabs were washed in approximately 200 mL sterile water. Water samples were then filtered through cellulose nitrate membranes (pore size, 1.6 µm) (17). Central part of each membrane was incubated on 1.5% nonnutrient agar (NNA) plate overlaid with a monolaver of heat-inactivated Escherichia coli (18). Nasal swabs were cultured on plates directly after sampling. The plated were then sealed and incubated at 28 °C. One week after cultivation the plates were investigated (using 100X magnification of microscope) daily for out-growth of FLAs up to two months. Positive samples were investigated using page key. Morphological characteristics of trophozoites and cysts were used to identify FLAs according to page key (19). To decrease the bacterial and fungal contamination and better identification of FLAs, cloning was performed. To this end, a small part of agar containing amoebae cysts was cut and placed in fresh medium. This procedure was done to achieve a pure plate.

Results

Overall, 17 (28%) samples out of 60 samples were found positive for FLAs and 19 isolates were detected. Thirteen dust samples (50%) from ICU and CCU wards and 4 nasal swap samples (12%) from severe immunosuppressed patients were positive for outgrowth of free-living amoebae (Tables 1 and 2). Medical instrument of CCU wards was the most contaminated source. Accordingly, nasal swaps of hospitalized patients in the same CCU wards were also showed highest contaminated sources. Acanthamoeba spp. was detected in 30.7% and 8.8% of dust and nasal swabs, respectively according to double-walled cyst and flat shaped trophozoites (Fig. 1). According to page key, all isolates belonged to morphological group 2. Vahlkampfiids were detected by characteristic round cysts measuring 10 µ and elongated shape trophozoites (Fig. 1). V. vermiformis were also characterized using its wormy shape trophozoites and round cysts (Fig. 1). In this study, the most frequent amoeba was Acanthamoeba spp. and two plates had mix con-Acanthamoeba tamination of spp. and Vahlkampfiids/ Vermamoeba (Table 3).

Collection site	No. of samples	<i>No. of positive samples</i> (%)	Isolated FLA
CCU	16	11 (69)	Acanthamoeba spp. Vahlkampfiids Vermamoeba vermiformis
Surgical ICU	4	1 (25)	Acanthamoeba spp.
General ICU	4	1 (25)	Acanthamoeba spp.
Open heart ICU	2	00 (00)	-
Total	26	13 (50)	Acanthamoeba spp. Vahlkampfiids Vermamoeba vermiformis



Fig. 1: A: Acanthamoeba cysts (x 100), B: Vahlkampfiids cyst (x1000), C: Vermamoeba cysts (x1000)

Collection site	No. of samples	No. of positive sample (%)	Isolate FLA
CCU	16	4 (25)	Acanthamoeba spp.
			Vahlkampfiids Vermamoeba
			vermiformis
Surgical ICU	5	00 (00)	_
General ICU	8	00 (00)	-
Open heart ICU	5	00 (00)	-
Total	34	4 (12)	Acanthamoeba spp.
			Vahlkampfiids Vermamoeba
			vermiformis

Table 2: Data from FLAs isolated from nasal swaps of hospitalized patients in CCU and ICU wards

Table 3: Frequency of isolated FLA from patients and hospital environment

Isolated FLA	No.	Percentage
Acanthamoeba spp.	11	58
Vahlkampfiids	5	26
Vermamoeba vermiformis	3	15

Discussion

This study was the first investigations of the occurrence of FLAs in dust from ICU and CCU wards and hospitalized immune-suppressed patients (high-risk people) simultaneously in Iran. Detected FLAs belonged to three genera, including Acanthamoeba (11), Vahlkampfiids (5) and Vermamoeba (3). FLAs were isolated from various environmental sources and patients in Iran (20-23). Occurrence of FLAs was reported 52.9% and 42.86% in dust and biofilm samples of immunodeficiency and ophthalmology wards of hospital in Iran, respectively (14, 24). There are various study on occurrence of FLAs in therapeutic pools and water system of hospital (7, 16, 25-27), and some studies were conducted to investigate FLAs contamination of other sources in hospital worldwide (14, 28, 29).

Dust was previously reported as potential source for *Acanthamoeba*, *Balamuthia* and Vahlkampfiid amoebae in Iran (14, 30, 31). Due to heavy air pollution and airborne dust in Tehran, cyst form of FLAs can easily be transferred to visitors and patients of hospital wards (28, 32, 33). In addition to dust samples, the present study evaluated the presence of FLAs in nasal swabs of hospitalized patients. In Peru, 21 (28.4%) samples out of 74 nasal swabs positive for *Acanthamoeba* species in healthy individuals were found (34). However, no study was conducted to evaluate the presence of all of medically important FLAs in nasal swabs of patients or healthy individuals. As expected, among three detected amoebas *Acanthamoeba* belonging to group 2 has the highest occurrence in both of the dust and nasal swabs. This can be due to high resistance of *Acanthamoeba* cysts to usual chlorine-based disinfectants used in the most of the Tehran hospitals (10, 28).

In this study, all of the nasal swabs were taken from patients' immune-suppressed by corticosteroid therapy. Deficient or suppressed immune system is an important risk factor for most of the FLAs induced infections, such as GAE.

Conclusion

The high occurrence of FLAs in environmental samples of the selected hospital wards, and it may result in life-threating infections. On the other hand, all of the FLAs detected in dust samples, were isolated from CCU patients too, thus dust may be source of infection for patients. Increase awareness of hospital staff about FLAs, improvement of disinfection methods and using efficient disinfectants in hospital, particularly in wards which immunodeficient or immunesuppressed patients hospitalized is necessary.

Ethical considerations

Ethical issues (Including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and/or submission, redundancy, etc.) have been completely observed by the authors.

Acknowledgements

Dr. Maryam Niyyati has been supported by National Elite Foundation.

Conflict of interest

The authors declare that there is no conflict of interests.

References

- Visvesvara GS, Moura H, Schuster FL (2007). Pathogenic and opportunistic free-living amoebae: Acanthamoeba spp., Balamuthia mandrillaris, Naegleria fowleri, and Sappinia diploidea. *FEMS Immunol Med Microbiol*, 50(1):1-26.
- Marciano-Cabral F, Cabral G (2003). Acanthamoeba spp. as agents of disease in humans. *Clin Microbiol Rev*, 16(2):273-307.
- 3. Qvarnstrom Y, da Silva AJ, Schuster FL et al (2009). Molecular confirmation of Sappinia pedata as a causative agent of amoebic encephalitis. *J Infect Dis*, 199(8):1139-42.
- Aitken D, Hay J, Kinnear FB et al (1996). Amebic keratitis in a wearer of disposable contact lenses due to a mixed Vahlkampfia and Hartmannella infection. *Ophthalmology*, 103(3):485-94.

- Niyyati M, Lorenzo-Morales J, Rezaie S et al (2010). First report of a mixed infection due to Acanthamoeba genotype T3 and Vahlkampfia in a cosmetic soft contact lens wearer in Iran. *Exp Parasitol*, 126(1):89-90.
- 6. Lorenzo-Morales J, Martínez-Carretero E, Batista N et al (2007). Early diagnosis of amoebic keratitis due to a mixed infection with Acanthamoeba and Hartmannella. *Parasitol Res*, 102(1):167-9.
- Visvesvara GS, Sriram R, Qvarnstrom Y et al (2009). Paravahlkampfia francinae n. sp. masquerading as an agent of primary amoebic meningoencephalitis. J Eukaryot Microbiol, 56(4):357-66.
- Hajialilo E, Behnia M, Tarighi F et al (2016). Isolation and genotyping of Acanthamoeba strains (T4, T9, and T11) from amoebic keratitis patients in Iran. *Parasitol Res*, 115(8): 3147-51.
- Movahedi Z, Shokrollahi MR, Aghaali M, Heydari H (2012). Primary amoebic meningoencephalitis in an Iranian infant. *Case Rep Med*, 2012: 782854.
- 10. Khan NA (2009). *Acanthamoeba: biology and pathogenesis:* Caister Academic Press, Norfolk, Great Britain.
- Walochnik J, Scheikl U, Haller- Schober EM (2015). Twenty years of Acanthamoeba diagnostics in Austria. J Eukaryot Microbiol, 62(1):3-11.
- Winiecka-Krusnell J, Linder E (2001). Bacterial infections of free-living amoebae. *Res Microbiol*, 152(7):613-9.
- Thomas V, Bouchez T, Nicolas V et al (2004). Amoebae in domestic water systems: resistance to disinfection treatments and implication in Legionella persistence. J Appl Microbiol, 97(5):950-63.
- Lasjerdi Z, Niyyati M, Haghighi A et al (2011). Potentially pathogenic free-living amoebae isolated from hospital wards with immunodeficient patients in Tehran, Iran. *Parasitol Res*, 109(3):575-80.
- 15. Memari F, Niyyati M, Haghighi A et al (2015). Occurrence of pathogenic Acanthamoeba genotypes in nasal swabs of cancer patients in Iran. *Parasitol Res*, 114(5):1907-12.

- Trabelsi H, Dendana F, Neji S et al (2016). Morphological and molecular identification of free living amoeba isolated from hospital water in Tunisia. *Parasitol Res*, 115(1):431-5.
- Rezaeian M, Niyyati M, Farnia S, Haghi AM (2008). Isolation of Acanthamoeba spp. from different environmental sources. *Iran J Parasitol*, 3(1):44-7.
- Khan NA (2006). Acanthamoeba: biology and increasing importance in human health. *FEMS Microbiol Rev*, 30(4):564-95.
- Schroeder JM, Booton GC, Hay J (2001). Use of subgenic 18S ribosomal DNA PCR and sequencing for genus and genotype identification of acanthamoebae from humans with keratitis and from sewage sludge. J Clin Microbiol, 39(5):1903-11.
- Behniafar H, Niyyati M, Lasjerdi Z, Dodangeh S (2015). High occurrence of potentially pathogenic free living amoebae in water bodies of kaleybar and khodaafarin, east azerbaijan province. *Curr World Environ*, 10(Special Issue 1):727-31.
- Niyyati M, Lasjerdi Z, Zarein-Dolab S et al (2015). Morphological and Molecular Survey of Naegleria spp. in Water Bodies Used for Recreational Purposes in Rasht city, Northern Iran. *Iran J Parasitol*, 10(4):523-9.
- Niyyati M, Karamati SA, Morales JL, Lasjerdi Z (2016). Isolation of Balamuthia mandrillaris from soil samples in North-Western Iran. Parasitol res, 115(2):541-5.
- 23. Rezeaian M, Farnia S, Niyyati M, Rahimi F (2007). Amoebic keratitis in Iran (1997-2007). *Iran J Parasitol*, 2(3):1-6.
- 24. Lasjerdi Z, Niyyati M, Lorenzo-Morales J et al (2015). Ophthalmology hospital wards contamination to pathogenic free living Amoebae in Iran. *Acta Parasitol*, 60(3):417-22.
- 25. Rohr U, Weber S, Michel R et al (1998). Comparison of free-living amoebae in hot water systems of hospitals with isolates from moist sanitary areas by identifying genera and determining temperature tolerance. *Appl Emviron Microbiol*, 64(5):1822-4.

- Thomas V, Herrera-Rimann K, Blanc DS, Greub G (2006). Biodiversity of amoebae and amoeba-resisting bacteria in a hospital water network. *Appl Environ Microbiol*, 72(4):2428-38.
- Muchesa P, Leifels M, Jurzik L et al (2017). Coexistence of free-living amoebae and bacteria in selected South African hospital water distribution systems. *Parasitol Res*, 116(1):155-165.
- Costa AO, Castro EA, Ferreira GA et al (2010). Characterization of acanthamoeba isolates from dust of a public hospital in Curitiba, Parana, Brazil. J Eukaryot Microbiol, 57(1):70-5.
- 29. Carlesso AM, Artuso GL, Caumo K, Rott MB (2010). Potentially pathogenic Acanthamoeba isolated from a hospital in Brazil. *Curr Microbiol*, 60(3):185-90.
- Niyyati M, Lorenzo-Morales J, Rahimi F et al (2009). Isolation and genotyping of potentially pathogenic Acanthamoeba strains from dust sources in Iran. *Trans R Soc Trop Med Hyg*, 103(4):425-7.
- Niyyati M, Lorenzo-Morales J, Rezaeian M et al (2009). Isolation of Balamuthia mandrillaris from urban dust, free of known infectious involvement. *Parasitol Res*, 106(1):279-81.
- 32. Teixeira LH, Rocha S, Pinto RMF et al (2009). Prevalence of potentially pathogenic freeliving amoebae from Acanthamoeba and Naegleria genera in non-hospital, public, internal environments from the city of Santos, Brazil. Braz J Infect Dis, 13(6):395-7.
- Chan L-L, Mak J-W, Low Y-T et al (2011). Isolation and characterization of Acanthamoeba spp. from air-conditioners in Kuala Lumpur, Malaysia. *Acta Trop*, 117(1):23-30.
- 34. Cabello-Vílchez AM, Martín-Navarro CM, López-Arencibia A et al (2014). Genotyping of potentially pathogenic Acanthamoeba strains isolated from nasal swabs of healthy individuals in Peru. Acta Trap, 130:7-10.