



Complete Genome Sequence of *Pasteurella multocida* HuN001, a Capsular Type A Strain from a Human

Lin Lin,^{a,c} Chunhui Li,^b Fei Wang,^{a,c} Xueying Wang,^{a,c} Yue Zhang,^{a,c} Songtao Liu,^{a,c} Wan Liang,^d Lin Hua,^{a,c}  Zhong Peng,^{a,c}  Bin Wu^{a,c}

^aState Key Laboratory of Agricultural Microbiology, College of Veterinary Medicine, Huazhong Agricultural University, Wuhan, China

^bInfection Control Center, Xiangya Hospital of Central South University, Changsha, China

^cMinistry of Science and Technology International Research Center for Animal Disease, The Cooperative Innovation Center for Sustainable Pig Production, Huazhong Agricultural University, Wuhan, China

^dMinistry of Agriculture and Rural Affairs Key Laboratory of Prevention and Control Agents for Animal Bacteriosis, Institute of Animal Husbandry and Veterinary Sciences, Hubei Academy of Agricultural Sciences, Wuhan, China

ABSTRACT Here, we report the complete genome sequence of clinical *Pasteurella multocida* strain HuN001, which was cultured from a sputum sample from a patient with pneumonia. Oxford Nanopore Technologies sequencing provided a complete genome sequence of *P. multocida* HuN001, which contains a 2,287,216-bp circular chromosome with an average G+C content of 40.33%.

Pasteurella multocida is an important zoonotic pathogen that mainly causes respiratory symptoms as well as hemorrhagic septicemia in multiple animal species and even in humans (1, 2). *P. multocida* strains recovered from different host species are classified into five capsular serogroups (A, B, D, E, and F) (3) or genotypes (A, B, D, E, and F) (4) and 16 lipopolysaccharide (LPS) serovars (serovars 1 to 16) (5) or eight LPS genotypes (L1 to L8) (6). Recently, the public availability of increasing numbers of *P. multocida* genome sequences has facilitated great progress in increasing the knowledge of both bacterial typing and the pathogenesis of *P. multocida* (1). However, most of these publicly available genome sequences are those of *P. multocida* strains that originated from animals, and very few of them are from *P. multocida* strains of human origin. Here, we describe the complete genome analysis of a *P. multocida* strain that was isolated from a pulmonary infection.

P. multocida strain HuN001 was isolated from a sputum sample from a patient with pneumonia in Hunan Province, China. Briefly, the sputum sample from the patient was streaked onto tryptic soy agar (TSA) (Becton, Dickinson and Co., Sparks, MD, USA) supplemented with 5% newborn calf serum (Tianhang Biotechnology, Hangzhou, China), which was incubated overnight at 37°C. After that, single colonies were picked for Gram staining and 16S rRNA gene sequencing to determine the genus and species of the bacterial isolates. This process is a routine procedure for diagnosis and has been approved by the Ethics Committee of Xiangya Hospital (Changsha, China). Genomic DNA of the isolate was extracted from a bacterial culture of a single colony at 37°C in tryptic soy broth (TSB) (Becton, Dickinson and Co.) supplemented with 5% bovine serum using the QIAamp DNA minikit (Qiagen, Hilden, Germany). DNA quality and quantity were assessed using electrophoresis on a 0.35% agarose gel and were double checked by NanoDrop spectrophotometry (Thermo Fisher Scientific, USA) and Qubit 3.0 fluorometry (Thermo Fisher Scientific), respectively. Then, 20-kb to 30-kb DNA libraries were generated using an SQK-LSK109 ligation sequencing kit (Oxford Nanopore Technologies [ONT], Oxford, UK) and were sequenced on a PromethION platform (ONT), according to the manufacturer's protocol, at BioMarker Technologies Corp. (Beijing, China). Base calling was performed using the Nanocall package (<https://github.com/mateidavid/nanocall>) with default parameters (7). The strategy yielded 1,582,081,118 bp of raw reads (N_{50} , 16,755 bp; N_{90} , 8,425 bp), and a total of 1,448,813,812 bp of filtered data

Citation Lin L, Li C, Wang F, Wang X, Zhang Y, Liu S, Liang W, Hua L, Peng Z, Wu B. 2021.

Complete genome sequence of *Pasteurella multocida* HuN001, a capsular type A strain from a human. Microbiol Resour Announc 10: e00395-21. <https://doi.org/10.1128/MRA.00395-21>.

Editor David A. Baltrus, University of Arizona

Copyright © 2021 Lin et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Zhong Peng, pengzhong@mail.hzau.edu.cn, or Bin Wu, wub@mail.hzau.edu.cn.

Received 25 April 2021

Accepted 4 June 2021

Published 1 July 2021

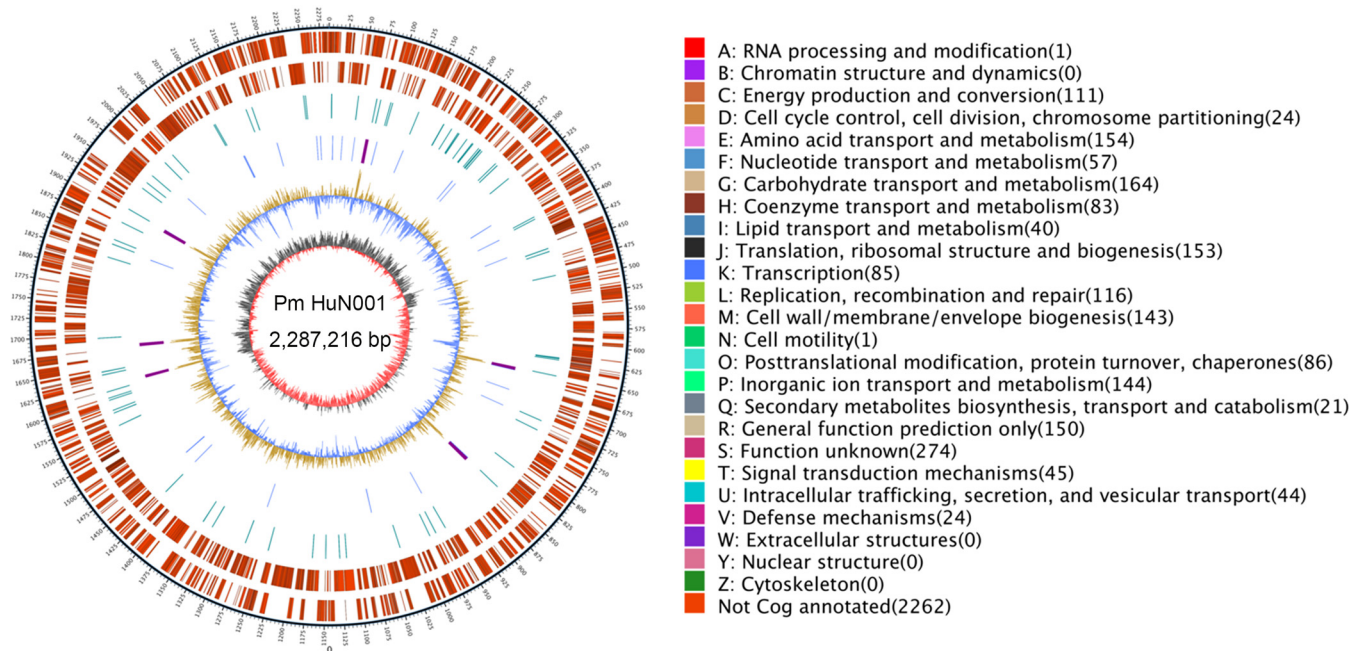


FIG 1 Genomic characteristics of the complete genome sequence of *P. multocida* HuN001. (Left) Circular map of the complete genome sequence. From inside to outside, the circles represent GC skew (circle 1), G+C content (circle 2), positions of tRNAs (in blue) and rRNAs (in purple) (circle 3), repeat sequences (circle 4), genes located on the negative strand (circle 5), and genes located on the positive strand (circle 6). (Right) Clusters of Orthologous Genes (COG) functions of different genes.

(N_{50} , 16,900 bp; N_{90} , 8,815 bp) was finally obtained after removal of reads with mean_qscore_ template of <7 and length of $<2,000$ bp. The Canu v1.5 (8) package was then used to assemble the filtered data, and the quality of the assembly was double checked using Racon v3.4.3 software and Circlator v1.5.5 software with default parameters. Finally, a single circular chromosome 2,287,216 bp in length was generated for the complete genome sequence of *P. multocida* strain HuN001 (Fig. 1 and Table 1).

The public version of the genome was annotated by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (9); through this strategy, we found that the complete genome of *P. multocida* HuN001 contained 2,127 protein-coding genes, 19 rRNAs, and 58 tRNAs. In particular, a number of genes associated with bacterial adherence and invasion,

TABLE 1 General features of the complete genome sequence of *P. multocida* HuN001

Genomic feature	Value
Total length (bp)	2,287,216
G+C content (%)	40.33
Contig N_{90} (bp)	2,287,216
Contig N_{50} (bp)	2,287,216
Avg coverage (\times)	691.7
No. of genes	2,127
Total repetitive sequence length (bp)	3,747
No. of rRNAs	19
No. of 16S rRNAs	6
No. of 23S rRNAs	6
No. of 5S rRNAs	7
No. of tRNAs	58
No. of genomic islands	3
No. of prophages	2
No. of virulence factors (VFDB)	353
No. of resistance factors (CARD database)	3
No. of secreted proteins	197

capsule and LPS biosynthesis, iron acquisition and uptake, sialic acid metabolism, and outer membrane formation were identified in the chromosome using the Virulence Factor Database (VFDB) (10). However, only three antimicrobial resistance determinants, for β -lactams (GE000922) and pulvomycin (GE001297 and GE001710), were identified through the CARD database (11). Genotyping using the PmGT online tool (<http://vetinfo.hzau.edu.cn/PmGT/>) revealed that *P. multocida* HuN001 was assigned as capsular, LPS genotype A, and L1. Strikingly, it was assigned as a novel sequence type because no sequence type was given by the online tool, even though the allele numbers of each of the housekeeping genes used for multilocus sequence typing (*adk*, *aroA*, *deoD*, *gdhA*, *g6pd*, *mdh*, and *pgi*) were obtained.

Data availability. The complete genome sequence of *P. multocida* strain HuN001 has been deposited in GenBank under accession number [CP073238](https://ncbi.nlm.nih.gov/nucl/CP073238). The raw reads are available in the NCBI Sequence Read Archive (SRA) under accession number [SRR14253068](https://ncbi.nlm.nih.gov/sra/SRR14253068). The BioProject accession number is [PRJNA722379](https://ncbi.nlm.nih.gov/bioproject/PRJNA722379), and the BioSample accession number is [SAMN18753491](https://ncbi.nlm.nih.gov/biosample/SAMN18753491).

ACKNOWLEDGMENTS

This study was supported in part by the China Postdoctoral Foundation (grant 2020T130232), the Key Laboratory of Livestock Disease Prevention of Guangdong Province and the Scientific Observation and Experiment Station of Veterinary Drugs and Diagnostic Techniques of Guangdong Province, Ministry of Agriculture and Rural Affairs, China (grant YDWS1901), and the Agricultural Science and Technology Innovation Program of Hubei Province (grant 2018skjcx05).

REFERENCES

1. Peng Z, Wang X, Zhou R, Chen H, Wilson BA, Wu B. 2019. *Pasteurella multocida*: genotypes and genomics. *Microbiol Mol Biol Rev* 83:e00014-19. <https://doi.org/10.1128/MMBR.00014-19>.
2. Wilson BA, Ho M. 2013. *Pasteurella multocida*: from zoonosis to cellular microbiology. *Clin Microbiol Rev* 26:631–655. <https://doi.org/10.1128/CMR.00024-13>.
3. Carter GR. 1955. Studies on *Pasteurella multocida*. I. A hemagglutination test for the identification of serological types. *Am J Vet Res* 16:481–484.
4. Townsend KM, Boyce JD, Chung JY, Frost AJ, Adler B. 2001. Genetic organization of *Pasteurella multocida* cap loci and development of a multiplex capsular PCR typing system. *J Clin Microbiol* 39:924–929. <https://doi.org/10.1128/JCM.39.3.924-929.2001>.
5. Heddleston KL, Gallagher JE, Rebers PA. 1972. Fowl cholera: gel diffusion precipitin test for serotyping *Pasteurella multocida* from avian species. *Avian Dis* 16:925–936. <https://doi.org/10.2307/1588773>.
6. Harper M, John M, Turni C, Edmunds M, St. Michael F, Adler B, Blackall PJ, Cox AD, Boyce JD. 2015. Development of a rapid multiplex PCR assay to genotype *Pasteurella multocida* strains by use of the lipopolysaccharide outer core biosynthesis locus. *J Clin Microbiol* 53:477–485. <https://doi.org/10.1128/JCM.02824-14>.
7. David M, Dursi LJ, Yao D, Boutros PC, Simpson JT. 2017. Nanocall: an open source basecaller for Oxford Nanopore sequencing data. *Bioinformatics* 33:49–55. <https://doi.org/10.1093/bioinformatics/btw569>.
8. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive *k*-mer weighting and repeat separation. *Genome Res* 27:722–736. <https://doi.org/10.1101/gr.215087.116>.
9. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
10. Liu B, Zheng D, Jin Q, Chen L, Yang J. 2019. VFDB 2019: a comparative pathogenomic platform with an interactive web interface. *Nucleic Acids Res* 47:D687–D692. <https://doi.org/10.1093/nar/gky1080>.
11. Alcock BP, Raphenya AR, Lau TTY, Tsang KK, Bouchard M, Edalatmand A, Huynh W, Nguyen AV, Cheng AA, Liu S, Min SY, Miroshnichenko A, Tran HK, Werfalli RE, Nasir JA, Oloni M, Speicher DJ, Florescu A, Singh B, Faltyn M, Hernandez-Koutoucheva A, Sharma AN, Bordeleau E, Pawlowski AC, Zubyk HL, Dooley D, Griffiths E, Maguire F, Winsor GL, Beiko RG, Brinkman FSL, Hsiao WWL, Domselaar GV, McArthur AG. 2020. CARD 2020: antibiotic resistance surveillance with the Comprehensive Antibiotic Resistance Database. *Nucleic Acids Res* 48:D517–D525. <https://doi.org/10.1093/nar/gkz935>.