

Article

The Potential Role of *Citrus limon* Powder as a Natural Feed Supplement to Boost the Productive Performance, Antioxidant Status, and Blood Biochemistry of Growing Rabbits



Hamada A. M. Elwan ¹, Dawood Hosni Dawood ², Sally Mohamed Abd El-Aziz El-Shafei ³, Atef Abd El-Mohsen Abd El-Rahman ³, Shaker A. Abdel-Latif ¹, Mohamed Mohany ⁴,*, Faleh Algahtani ⁴, Saeed Algahtani ⁴ and Salim S. Al-Rejaie ⁴

¹ Animal and Poultry Production Department, Faculty of Agriculture, Minia University, El-Minya 61519, Egypt

- ² Agricultural Chemistry Department, Faculty of Agriculture, Mansoura University, Mansoura 35516, Egypt
- ³ Department of Agricultural Chemistry, Faculty of Agriculture, Minia University, El-Minya 61519, Egypt
- ⁴ Department of Pharmacology and Toxicology, College of Pharmacy, King Saud University, Riyadh 11451, Saudi Arabia
- * Correspondence: mmohany@ksu.edu.sa; Tel.: +966-508157791

Received: 20 May 2019; Accepted: 4 July 2019; Published: 7 July 2019



Simple Summary: Although some studies have suggested that the flavonoids in *Citrus limon* possess numerous biological functions, including antioxidant, anti-inflammatory, antiallergic, antiviral, antiproliferative, antimutagenic, and anticarcinogenic activities, a rabbit model has not been used to establish the beneficial effects and possible antioxidant activity of dried *C. limon*. Therefore, we investigated the hemato-biochemical alteration, thyroid activity, and antioxidant responses of dried *C. limon* as a powder in growing rabbits. By employing New Zealand White rabbits, we demonstrated that rabbits fed a diet supplemented with 1% dry lemon powder (DLP) or 2% DLP had an increase in their productive performance, better feed conversion, enhanced hematological and biochemical values, improved antioxidant enzyme activity, and inhibited lipid peroxidation. These results suggest that dry lemon supplementation might play a role as a growth enhancer for rabbits when administered at a maximum of 2% in the rabbit diet. In general, the addition of *Citrus limon* improved growth performance, physiology, and antioxidant status in serum and liver of the growing rabbit.

Abstract: The current study examined the influence of *Citrus limon* (dry lemon) on the hemato-biochemical profiles, and antioxidant indices of growing rabbits. Forty-eight growing New Zealand White rabbits (age, eight weeks; weight, 1543.33 ± 25 g) were allocated into three groups (16 animals each), the first group was (control) fed a basal diet, whereas the second and third groups were supplemented with dried lemon, 1% or 2% DLP, respectively. A GC-MS analysis of more than 27 active constituents was performed. Feed conversion efficiency was (p < 0.05) better with diets containing 1% or 2% dry lemon, compared to the control group. Hematological indexes were increased significantly with the addition of DLP compared to those in the control group. Adding 1% or 2% dry lemon to rabbit diet increased (p < 0.05) enzymatic and non-enzymatic antioxidant activities (TAC, SOD, GSH, GST, and CAT) in serum and liver tissues. Taken together, these data reveal the advantages and antioxidant effects of dry lemon supplementation for growing rabbits once supplemented at a maximum of 2% in their daily diet.

Keywords: Citrus limon; oxidative biomarkers; GC-MS analysis; rabbits

1. Introduction

Nutritional factors play an important role in the protection against the deleterious consequences of free radicals. Natural antioxidants-rich diets can significantly affect the enhancement of reactive antioxidants and reduce the risk of some diseases induced by free radicals generation. A high enough level of antioxidants in a diet is considered to encourage immunological processes and correctly elevate the defensive abilities of the cell [1].

Over the years, plants have served as precious sources of bioactive compounds for humans and animals, thereby possessing great potential for use in the discovery of new drugs [2]. *Citrus* fruit has the highest antioxidant activity [3] as it contains many flavonoids, vitamin C, and carotenoids [4]. There are numerous bioflavonoids in *Citrus limon* such as hesperidin, rutin, and naringin, which exhibit many pharmacological actions in the body [5]. Citrus fruits, especially *C. limon*, have been scientifically proven to display many health benefits as well as limiting the risk of diseases due to their richness in vitamins and minerals and other active constituents [6].

Numerous studies have proposed that *C. limon* may display antifungal [7] and anti-cancer [8] activities. Also, some studies have suggested that its flavonoids possess numerous biological functions, including anti-inflammatory, antioxidant, antiviral, antiallergic, antiproliferative, antimutagenic, and antitumor activities [9–12]. Essential oils of *C. limon* has also been found to prevent damage to the brain, lung, and intestine induced by aspirin toxicity [13].

Several progenies of farm rabbits were known to produce a higher amount of meat at a rate greater than 10–15 times of their weight [14]. Rabbit meat is preferred for its nutritional value compared with other meat products because of its low calorie content and high digestion [15]. Proper care to maintain the physiological status of rabbits is an essential factor in the production of high-quality products that meat consumers expect. Nobakht [16] demonstrated that different concentrations of dried lemon pulp in diets had marked influence on broilers performance. Azra et al. [17] found that consuming lime peel or fresh lime juice induced the highest changes in serum antioxidant capacity. Flavonoids in *Citrus* was found to protect against hyperglycemia via increasing liver glycolysis and lowering liver gluconeogenesis [18]. Hesperidin and naringin, both *C. limon* active compounds, were also found to significantly lowered the blood glucose level [19]. Moreover, dietary dried citrus pulp supplementation may improve laying hens performance by their positive effects on lipid profile and blood glucose levels [20].

Previous studies demonstrated the use of citrus pulp, juice, and peel meal as an animal feed additive without any accompanying side effects [21]. In addition, dietary fresh or dry fruit of *C. limon* was demonstrated to contribute to the maintenance of optimum health due to its health-promoting phytochemicals. To date, however, impactful investigations on dried *C. limon* have not been performed to establish its beneficial effect and possible antioxidant activity with a rabbit model. Hence, this experiment was employed to explore the hemato-biochemical profiles, thyroid activity, and antioxidant responses of dried *C. limon* as a powder in growing rabbits.

2. Materials and Methods

This study was in accordance with the guidelines of Egyptian Research Ethics Committee, Animal and Poultry Production Department, Faculty of Agriculture, Minia University, and approved the guidelines contained in the Guide for the Care and Use of Laboratory Animals (2011). Project identification MU-FA-APPD/1/M/321/2018.

2.1. In Vitro Studies on Dry Lemon Powder (DLP)

2.1.1. Preparation of Dry Lemon

Lemon was obtained from a private commercial market in El-Minya Governorate, Egypt and dried at 40 °C until constant weight. After the drying process, the obtained dried lemon was ground,

sieved (1 mm mesh) and finally stored in air-tight dark polyethylene bags at 25 °C. The composite sample of the lemon powder was placed in a plastic sample bag for chemical and nutritional analysis.

2.1.2. Preparation and Extraction of Methanol Extract

One hundred g of DLP was macerated with methanol at 25 °C for 24 h, then was filtered using a filter paper (Whatman No. 1), the residual underwent an extraction process twice by the same manner. Methanol was removed at 40 °C in a rotary evaporator. To obtain the methanol extract, the samples were dried in desiccators [22].

2.1.3. Total Phenolic Compounds Content

Total phenolic compounds content was measured by the Folin-Ciocalteu method with little modifications [23]. In brief, 50 μ L of the extract was mixed with 3 mL of distilled water, 250 μ L of Folin-Ciocalteu reagent, and 750 μ L of 7.5% Na₂CO₃, which was then vigorously shaken for two minutes and incubated for 30 min at 25 °C. Absorbance values were estimated at 760 nm (UV-VIS Spectrophotometer, Shimadzu, Kyoto, Japan). Quantification of total phenolic compounds in the extracts was obtained by gallic acid standard curve. Results are expressed as mg gallic acid equivalent/g dried sample. All measurements were done in triplicate.

2.1.4. Assessment of Total Flavonoid Content

Total flavonoid content was measured by the aluminum chloride colorimetric assay method [24]. In brief, 0.5 mL of the extract was added to 1.5 mL of absolute ethanol, 0.1 mL of 10% aluminum chloride, 0.1 mL of 1 M potassium acetate, and 3 mL of distilled water. The yield mixture was mixed by vortex, incubated at 25 °C for 30 min; the absorbance was measured at 415 nm (UV-VIS Spectrophotometer, Shimadzu). The concentration of total flavonoid content in the methanolic extract was calculated by quercetin standard curve. Data are presented as mg quercetin equivalent/g dried sample.

2.1.5. Determination of Antioxidant Activity

Antioxidant activity was determined using 2, 2-biphenyl 1-picrylhydrazyl (DPPH) as reported by Oliveira et al. [25]. In brief, 100 μ L of methanolic extract was mixed with 3.9 mL DPPH (0.004%) methanolic solution. Absorbance was measured at 517 nm after incubation at 25 °C in the dark for 60 min. The reference compound was ascorbic acid. The following formula was used to estimate % inhibition:

Inhibition (%) = (A control – A test)/A control
$$\times$$
 100 (1)

where; A control = the absorbance of the control reaction and A test = the absorbance of the sample (methanol extract).

2.1.6. Gas Chromatography/Mass Spectrometry (GC/MS) Analysis

Mass spectra of chemical compounds were conducted using GC/MS (Agilent 6890 (19091S-433) series GC-MS) equipped with an HP-5MS column (diameter length 30 m; diameter 0.25 mm; film thickness 0.25 μ m) and mass spectrometer programmed to operate at temperature of 30 °C—280/300 °C, withhold time of 5 min, and rate of 10 °C/min. The conditions of chromatography were injection mode, column flow rate (1 mL/min), split; and carrier gas, helium (purity 99.999%). Mass library search (NIST based AMDIS software) with their relative retention indices was used to identify GC-MS spectra compounds [26].

2.1.7. Nutritional Analysis of Rabbit Diets and Citrus Limon

Using A.O.A.C., standard methods, dry matter (DM), crude fiber (CF), crude protein (CP) and ether extract (EE) were determined [27]. By difference, nitrogen-free extract (NFE) was calculated. According to the method of Van Soest et al. [28], fiber fractions of neutral detergent fiber (NDF),

acid detergent fiber (ADF) and acid detergent lignin (ADL) were assessed. Whereas, hemicellulose was calculated by the difference between NDF and ADF and cellulose by the difference between ADF and ADL. Analysis of amino acid in the basal diet and DLP was performed by hydrolyzing the samples by HCl (6 M) at 110 °C for 24 h using HPLC (Hitachi L-8900 Amino Acid Analyzer, Tokyo, Japan; Table 1).

Flow on to	Dry Lomon *	Experimental Groups			
Elements	Dry Lemon	Control	1% DLP	2% DLP	
Yellow corn (%)	-	44.00	44.00	44.00	
Wheat bran (%)	-	40.50	40.50	40.50	
Soybean mail (44% crude protein) (%)	-	13.50	13.50	13.50	
Lime (%)	-	0.5	0.5	0.5	
Sodium chloride (%)	-	1.00	1.00	1.00	
Vitamin and mineral premix ^{**} (%)	-	0.50	0.50	0.50	
Dry lemon powder (%)	-	0	1	2	
Analyzed nutritional composition (%)					
Moisture	6.51	5.92	5.90	5.91	
OM	88.97	89.20	89.36	89.17	
CP	9.40	18.65	18.86	18.87	
CF	12.62	10.42	10.83	10.78	
EE	4.93	2.94	2.95	3.01	
NFE	62.02	57.19	56.72	56.51	
Ash	4.52	4.88	4.74	4.92	
NDF	60.58	41.45	42.02	42.69	
ADF	37.48	24.25	26.09	25.17	
ADL	5.64	5.44	5.51	5.81	
Hemicellulose	23.10	17.20	15.93	17.52	
Cellulose	31.84	18.81	20.98	19.36	
Calculated gross energy, kcal/g	4.10	4.16	4.17	4.17	
Analyzed amino acids (g/100 g)					
Alanine	0.746	0.849	0.868	0.876	
Arginine	0.365	0.827	0.842	0.847	
Aspartic acid	0.265	0.780	0.794	0.798	
Cysteine	1.032	0.250	0.272	0.283	
Glutamic acid	0.465	0.644	0.660	0.665	
Glycine	0.323	0.600	0.615	0.619	
Histidine	0.265	0.370	0.384	0.388	
Isoleucine	0.142	0.628	0.641	0.644	
Leucine	0.365	1.162	1.177	1.182	
Lysine	0.266	0.672	0.686	0.689	
Methionine	0.365	0.389	0.404	0.409	
Phenylalanine	0.613	0.616	0.634	0.641	
Proline	0.511	0.679	0.696	0.702	
Serine	0.323	0.683	0.698	0.702	
Threonine	0.412	0.611	0.626	0.632	

Table 1. Ingredients, amino acids and nutritional analysis of rations and dry lemon.

* Percentage of dry weight content of fresh lemon fruit peels = 24.05 ± 0.52 ; Gross energy was calculated according to NRC [29], where g CP = 5.72 kcal, EE = 9.50 kcal CF = 4.79 kcal and NFE = 4.03. SEM = standard error of means; DM = dry matter; OM = organic matter; CP = crude protein; CF = crude fiber; EE = ether extract; NFE = nitrogen free extract; NDF = neutral detergent fiber; ADF = acid detergent fiber; ADL = acid detergent lignin.

2.2. In Vivo Studies on DLP

2.2.1. Experimental Animals

The experiments were conducted at the faculty farm (Animal and Poultry Production Department, Faculty of Agriculture, Minia University). The rabbits were housed individually in galvanized wire cages ($40 \times 50 \times 35$ cm) that were provided with feeders and an automatic drinking water system.

They were housed in a semi-closed building and maintained under the same managerial, hygienic and environmental conditions. A period of 14–16 h of daylight was also provided.

2.2.2. Rabbits Ration and Dry Lemon Supplementation

Experimental rations of the dry lemon were prepared in the form of pellets (0.3 cm) and mixed with other feed ingredients for each batch at 0, 1 or 2% for control, and 1% DLP and 2% DLP, respectively.

2.2.3. Animals Grouping and Treatments

Forty-eight growing New Zealand White (NZW) rabbits (1 Male:1 Female), aged 8 weeks, (average weight, 1543.33 ± 25 g) were allocated by weight into three groups (16 animals each); the first group was fed a basal diet free of dry lemon (control), while the second and third groups recieved the same basal diet containing 1% or 2% DLP, respectively, as a supplement for 8 weeks. Body weight and feed intake were recorded every week.

2.2.4. Physiological Response

At the end of the experiment, rabbits (n = 8 per group) were fasted for 12 h, weighed and handily slaughtered. Then, the slaughtered animals were de-skinned, dressed out, and their carcass weight was recorded. Also, weights of edible offal's (liver, heart, and kidneys) were recorded.

2.2.5. Hematological Parameters

At the end of the experiment (16 weeks), blood was collected from the marginal ear vein in clean tubes with or without anticoagulant substance. Coagulated samples were centrifuged at $2515.5 \times g$ for 15 min to separate the serum, which was stored at -80 °C until analysis. Whole blood was analyzed after sampling for physical blood characteristics using a veterinary hematology analyzer (VetScan HM5 Hematology System Abaxis Europe, UK).

2.2.6. Serum Total Antioxidant Capacity and Lipid Profiles

Serum total antioxidant capacity was determined, according to Trachootham et al. [30]. Concentrations of total cholesterol (TC) and triacylglycerols were measured after hydrolysis, followed by oxidation of the sample. HDL-cholesterol levels were assessed using the methods of Lopes-Virella et al. [31]. Serum LDL-c concentration was calculated using Friedewald equation: LDL (mg/dL) = TC – HDL – (TG/5) which was modified by Ahmadi et al. [32] as:

$$LDL (mg/dL) = TC/1.19 + TG/1.9 - HDL/1.1 - 38$$
(2)

The estimated very low-density lipoprotein cholesterol (VLDL-c), was obtained by the equation of Friedewald as follows:

$$VLDL-c = Triacylglyceroles/5$$
(3)

2.2.7. Thyroid Hormones

The levels of triiodothyronine (T3) and thyroxin (T4) were determined by an ELISA technique (Boston, MA, USA).

2.2.8. Serum and Hepatic Oxidative Stress Biomarkers

Serum and liver tissue concentrations of superoxide dismutase (SOD), catalase (CAT), glutathione-S-transferase (GST), glutathione peroxidase (GSH-Px) and malondialdehyde (MDA) were evaluated by kits (BioMed chemical company, Cairo, Egypt).

2.3. Statistical Analysis

The experiment was carried out with a completely randomized design (CRD) of three treatments and 16 replicates per group. The obtained results were analyzed with the statistical analysis system software (SAS, Version 9.1.3, 2003, Cary, NV, USA) using the generalized linear model (GLM) procedure [33]. Significant differences among treatment means for each trait in the experiment were measured using Duncan's multiple range test. The following mathematical model was used:

$$Xij = \mu + Ti + Eij$$
(4)

where; Xij = value observed in each experimental unit, μ = mean population, Ti = the effect of each treatment, and Eij = the effect of experimental errors.

3. Results

3.1. Dry Lemon In Vitro Studies

3.1.1. Determination of Chemical Profile, Total Phenolics, Total Flavonoids Content, and Total Antioxidant Capacity of Dry Lemon

The chemical profile of dry lemon based on DM basis was tabulated in Table 1; 9.40% CP, 12.62% CF, 4.93% EE, 62.02% NFE, 4.52% ash, 60.58% NDF, 37.48% ADF, 5.64% ADL. Temporarily, the hemicellulose and cellulose were 23.10% and 31.84%, respectively. Also, amino acids content in DL powder were analyzed after hydrolyzing using HPLC Amino Acid Analyzer (Table 1). The respective total phenolics, total flavonoids, and total antioxidant capacity in the methanolic extract were 120.57 mg/g gallic acid equivalent, 31.28 mg/g quercetin equivalent, and 49.12% based on the percentage of DPPH radicals inhibition.

3.1.2. GC-MS Analysis

GC-MS was used to analyze dry lemon extract major metabolites. More than 27 phytochemicals (Figure 1, Table 2) were present in the methanolic extract of dried lemon after separation based on retention times. The chemical constituents obtained were characterized by different classes of compounds viz: alkaloids, phenols, terpenes, glycosides, coumarins, fatty acids, and phytosterols. Based on the percentage area, the analysis demonstrated the existence of 5-(hydroxymethyl)-2-(methoxymethyl)furan (24.49%), imidazole (18.3%), p-limonene (8.40%), β -methoxy-(s)-2-furanethanol (8.20%), β -p-glucopyranoside, methyl (7.10%), 5,7-dimethoxy-3-nitro-2H-1-benzopyran-2-one (5.39%), quinic acid (4.66%), 6-methoxychroman-2-one (2.84%), scoparone (2.81%), and squalene (2.61%) as the major phytochemicals. Table 3 displays the phytochemicals identified in the lemon methanol extract along with their formula and compound nature.





Retention Time (min)	Compound Name	Mol. Formula	Compound Nature	Peak Area (%)
7.198	3-Carene	C ₁₀ H ₁₆	Bicyclic monoterpenes	1.13
7.530	Imidazole	$C_3H_4N_2$	Alkaloids	18.3
7.769	β-methoxy-(s)-2-furanethanol	C7H10O3	Glycosides	8.20
9.166	D-Limonene	$C_{10}H_{16}$	Monoterpenes	8.40
11.312	3-Methyl-4-propyl-2,5-furandione	$C_8H_{10}O_3$	Glycosides	0.43
12.899	Geraniol	C ₁₀ H ₁₈ O	Monoterpenes	1.79
13.540	5-(Hydroxymethyl)-2-(dimethoxymethyl)furan	$C_8H_{12}O_4$	Glycosides	24.49
13.828	2-Methoxy-5-vinylphenol	$C_9H_{10}O_2$	Phenols	1.82
14.420	Geranyl Acetate	$C_{12}H_{20}O_2$	Monoterpenes	1.66
15.304	Caryophyllene	$C_{15}H_{24}$	Sesquiterpenes	0.44
15.487	2,6-Di-Tert-butylphenol	C ₁₄ H ₂₂ O	Phenols	0.14
16.259	β-D-Glucopyranose, 1,6-anhydro-	$C_{6}H_{10}O_{5}$	Glycosides	0.53
16.348	β-Bisabolene	$C_{15}H_{24}$	Sesquiterpenes	1.59
17.809	Quinic acid	$C_7 H_{12} O_6$	Organic acids	4.66
18.005	6-Methoxychroman-2-one	$C_{10}H_{10}O_3$	Coumarins	2.84
18.115	β-D-Glucopyranoside, methyl	C7H14O6	Glycosides	7.10
19.260	Methyl 3-(4-hydroxy-3-methoxyphenyl) propanoate	$C_{11}H_{14}O_4$	Phenols	1.89
21.229	Hexadecanoic acid	$C_{16}H_{32}O_2$	Saturated Fatty acid	0.38
21.523	Scoparone	$C_{11}H_{10}O_4$	Coumarins	2.81
21.618	5,7-Dimethoxy-3-nitro-2H-1-benzopyran-2-one	$C_{11}H_9O_6$	Coumarins	5.39
22.944	Cis-13-Octadecenoic acid	C ₁₈ H ₃₄ O ₂	Long-chain Fatty acids	0.13
27.469	Imperatorin	C ₁₆ H ₁₄ O ₄	Coumarins	1.34
28.400	Squalene	C ₃₀ H ₅₀	Triterpenes	2.61
29.128	Geranyl linolenate	$C_{28}H_{48}O_2$	Triterpenes	0.08
31.846	Campesterol	C ₂₈ H ₄₈ O	Phytosterol	0.43
32.185	Stigmasterol	C ₂₉ H ₄₈ O	Phytosterol	0.21
32.855	γ -Sitosterol	C ₂₉ H ₅₀ O	Phytosterol	1.22

Table 2. Phytochemical active constituents of dried lemon methanolic extract.

Table 3. The effects of dried lemon supplementation on productive performance of growing rabbits.

Parameters	E	n-Vəluo		
r arameters	Control	1% DLP	2% DLP	<i>p</i> -value
Initial weight, g	1518.33 ± 8.46	1548.33 ± 8.30	1563.33 ± 8.33	0.11
Final weight, g	2567.66 ± 6.49 ^b	2959.00 ± 5.44 ^a	2966.33 ± 5.42 ^a	< 0.01
Body weight gain, g	1049.33 ± 6.25 ^b	1396.33 ± 6.12 ^a	1418 ± 6.15 ^a	< 0.01
Average daily gain, g	18.73 ± 0.04 ^b	24.93 ± 0.02^{a}	25.32 ± 0.03^{a}	0.04
Feed intake g/day	123 ± 1.89	126 ± 2.01	130 ± 2.02	0.06
Feed conversion ratio (g feed/g gain)	6.56 ± 0.09^{a}	5.05 ± 0.05 ^b	5.13 ± 0.05 ^b	0.03

^{a,b} Values within a row with different letters differ significantly (p < 0.05). Control = Lemon free diet; 1% DLP = 1 % dry lemon powder; 2% DLP = 2% dry lemon powder.

3.2. In Vivo Study of DLP

3.2.1. Effect of Dry Lemon Dietary Levels on Productive Performance of Growing Rabbits

The effect of dietary treatments on the growth performance of growing rabbits is shown in Table 3. Final weight, weight gain, and average daily gain were significantly (p < 0.05) increased as affected by dietary dry lemon levels compared with the control group. Animals fed on control diet (0% DLP) recorded the highest feed conversion ratio (p < 0.05) compared to other groups.

3.2.2. The Influences of Dietary Treatments on Growing Rabbit Carcass Traits

Rabbits fed 1% or 2% DLP had the heavier (p < 0.05) carcass weight compared with the control group; however, there was no significant difference between the two treatments. Values for liver weight, liver percentage, kidney weight, and kidney percentage were significantly (p < 0.01) decreased when rabbits were fed 1% and 2% DLP compared with the control group. Moreover, rabbits supplemented

with 1% DLP did not have any significant (p > 0.05) changes in liver percentage, kidney weight, and kidney percentage in comparison with that fed 2% DLP. Also, significant differences were not found, except for body weight, among all groups, especially for heart weight and heart % (Table 4).

Davam atava	I	" Value		
rarameters	Control	1% DLP	2% DLP	<i>p</i> -value
Carcass Weight (including head) (g)	1194.50 ± 17.50 ^b	1798.00 ± 24.74 ^a	1819.00 ± 24.10^{a}	< 0.01
Carcass %	46.45 ± 0.61 ^c	61.26 ± 0.59 ^b	63.20 ± 0.51 ^a	< 0.01
Dressing %	49.29 ± 0.53 ^c	63.53 ± 0.49 ^b	65.44 ± 0.50 ^a	< 0.01
Liver weight (g)	54.20 ± 6.30^{a}	51.0 ± 6.90 ^b	47.90 ± 8.90 ^c	< 0.01
Liver %	2.10 ± 0.11^{a}	$1.73 \pm 0.10^{\text{ b}}$	1.66 ± 0.11 ^b	< 0.01
Kidney Weight (g)	13.15 ± 3.05^{a}	10.52 ± 4.31 ^b	11.00 ± 4.29 ^b	< 0.01
Kidney %	0.51 ± 0.05 ^a	0.36 ± 0.05 ^b	0.38 ± 0.05 ^b	< 0.01
Heart Weight (g)	5.40 ± 0.10	5.24 ± 0.11	5.50 ± 0.10	0.66
Heart %	0.20 ± 0.02	0.18 ± 0.02	0.19 ± 0.02	0.32

Table 4. Carcass traits of growing rabbits after dietary dried lemon supplementation.

^{a,b,c} Values within a row with different letters differ significantly (p < 0.05). Control = Lemon free diet; 1% DLP = 1% dry lemon powder; 2% DLP = 2% dry lemon powder.

3.2.3. Rabbit Physiological Changes after Dried Lemon Supplementation

(1) Hematological Alterations

As shown in Table 5, the results indicated that rabbits fed on 2% DLP had the higher (p < 0.05) RBC's count, PCV, Hb, MCV, and MCH compared to the other dietary treatments; however, there were no significant (p > 0.05) differences between 1% or 2% DLP on MCH. Instead, a significant decrease in MCHC value (p < 0.05) was observed when rabbits were fed on 2% DLP compared to that in those fed to other groups. Rabbits fed on 1% DLP did not display any significant (p > 0.05) differences in RBC's count and MCV compared to those fed the control diet. In addition, they had the highest MCHC value for all groups. Also, there were no significant differences (p > 0.05) among all groups on platelets number (PLT) and its indices (MPV, PCT, PDW-cv, and PDW-SD).

Table 5. The effects of dried lemon supplementation of different hematological parameters.

Devene	E	n-Value		
rarameters	Control	1% DLP	2% DLP	<i>p</i> -value
RBC's (10 ⁶ /µL)	5.87 ± 0.02 ^b	5.89 ± 0.02 ^b	6.22 ± 0.02 ^a	0.03
PCV (%)	35.00 ± 0.02 ^b	34.00 ± 0.02 ^c	38.00 ± 0.02 ^a	0.02
Hb (g/dL)	11.67 ± 0.02 ^c	11.98 ± 0.02 ^b	12.67 ± 0.02 ^a	0.02
MCV (fL)	59.62 ± 1.55 ^b	57.72 ± 1.57 ^b	61.09 ± 1.56 ^a	0.02
MCH (pg)	19.88 ± 0.13 ^b	20.33 ± 0.12 ^a	20.36 ± 0.15^{a}	0.04
MCHC (g/dL)	33.34 ± 0.01 ^b	35.23 ± 0.01 ^a	33.34 ± 0.01 ^b	0.02
RDW-cv (%)	16.6 ± 0.11	16.5 ± 0.10	16.54 ± 0.10	0.06
RDW-SD (fL)	41.2 ± 1.58	40.98 ± 1.57	41.02 ± 1.59	0.07
PLT (10 ³ /µL)	170 ± 1.55	169.9 ± 1.57	170.01 ± 1.55	0.06
MPV (fL)	6.99 ± 0.34	6.78 ± 0.36	6.89 ± 0.33	0.06
PCT (%)	0.11 ± 0.01	0.10 ± 0.01	0.10 ± 0.01	0.06
PDW-cv (%)	29.35 ± 0.54	29.25 ± 0.58	29.45 ± 0.56	0.07
PDW-SD (fL)	6.89 ± 0.47	6.80 ± 0.46	6.92 ± 0.44	0.06

^{a,b,c} Values within a row with different letters differ significantly (p < 0.05). RBC's = red blood cells; PCV = peaked cell volume; Hb = hemoglobin; MCV = mean corpuscular volume; MCH = mean corpuscular hemoglobin; MCHC= mean corpuscular hemoglobin concentration; RDW-cv = coefficient of variation of red blood cell distribution width; RDW-SD = standard deviation of red blood cell distribution width; PLT= platelets; MPV = mean platelet volume; PCT = procalcitonin; PDW-cv = coefficient of variation of platelet distribution width; PDW-SD = standard deviation of platelet distribution width; Control = lemon free diet; 1% DLP = 1 % dry lemon powder; 2% DLP = 2% dry lemon powder.

(2) Total Antioxidant Capacity and Lipid Profiles

The effect of dietary treatments on triacylglycerol, cholesterol, HDL-c, LDL-c, VLDL-c, levels is shown in Table 6. Rabbits fed a diet containing a high amount of dried lemon supplementation (2% DLP) had the highest values (p < 0.05) for HDL-c, and TAC compared to others fed diets containing the lowest level DLP (1%), and the control diet. Also, rabbits fed on the control diet had the highest (p < 0.05) triacylglycerol, cholesterol, and LDL-c levels compared to those fed supplemented diets.

Table 6. Changes in serum levels of lipid profiles and total antioxidant capacity in rabbits treated with dietary dried lemon.

					_
Denemotors	E	n-Valuo			
ralameters	Control	1% DLP	2% DLP	<i>p</i> -varue	
Triacylglycerol (mg/dL)	181.17 ± 1.22 ^a	179.01 ± 1.24 ^b	171.60 ± 1.23 ^c	0.04	
Cholesterol (mg/dL)	238.53 ± 1.44 ^a	184.40 ± 1.46 ^b	166.97 ± 1.48 ^c	0.02	
HDL-Cholesterol (mg/dL)	94.97 ± 0.47 ^b	95.62 ± 0.48 ^b	117.51 ± 0.45 ^a	0.04	
LDL-Cholesterol (mg/dL)	171.46 ± 1.05^{a}	120.34 ± 1.09 ^b	89.69 ± 1.07 ^c	0.03	
VLDL-Cholesterol (mg/dL)	36.23 ± 0.26 ^a	35.80 ± 0.24 ^b	34.32 ± 0.22 ^c	0.02	
TAC (mM/L)	68.02 ± 0.02 ^c	69.02 ± 0.02 ^b	70.25 ± 0.02 ^a	0.02	

 a,b,c Values within a row with different letters differ significantly (p < 0.05). HDL-Cholesterol = high-density lipoprotein cholesterol; LDL-Cholesterol = low-density lipoprotein cholesterol; VLDL-Cholesterol = very low-density lipoprotein cholesterol; TAC = total antioxidant capacity. Control = lemon free diet; 1% DLP = 1 % dry lemon powder; 2% DLP = 2% dry lemon powder.

(3) Thyroid Hormones as Affected by Dry Lemon Supplementation

Table 7 shows that DLP supplementation in the diet of growing rabbits significantly (p < 0.05) increased their T3 level by 20.26 and 26.92% (1% and 2% DLP, respectively) compared to those fed the control diet, respectively. There is no significant (p > 0.05) between DLP levels on the T3. The concentrations of T3 were 0.8343 and 0.8805 (ng/mL) for 1% and 2% DLP, consecutively. Thyroxin, T4 levels significantly (p < 0.05) increased by 14.33% and 19.05% for rabbits fed on DLP (1% or 2%) in comparison with those fed the control diet, respectively. While the T4 level was insignificantly (p > 0.05) greater for rabbits fed diet contains 2% DLP when compared to those fed 1% DLP. Also, T4/T3 ratio significantly (p < 0.01) increased when rabbits fed on 2% DLP were compared with the control group. At the same time, results indicated that the (T4/T3 ratio) did not significantly (p > 0.05) differ between rabbits fed on diets containing 1% or 2% DLP levels.

\mathbf{T} 11 = \mathbf{T} (()	C 1 · 11	11	• 1 1	· ·	11.4
Iable / Httpcts	of dried lemor	n diet on th	wrold hormone	c of orowin	no raphite
Table 7. Lifetto	of unicu icilior	i ulci oli ul		S OI gIOWI	ig rabbito.
			2	0	0

Itoms	E	n-Value		
Items	Control	1% DLP	2% DLP	<i>p</i> -value
Triiodothyronine (ng/mL)	$0.6937 \pm 0.02^{\text{ b}}$	0.8343 ± 0.02 ^a	0.8805 ± 0.02 ^a	0.02
Thyroxin (µg/dL)	11.44 ± 1.66 ^b	13.08 ± 1.68 ^a	13.62 ± 1.65 ^a	0.02
Triiodothyronine/Thyroxin ratio	0.0607 ± 0.01 ^b	0.0637 ± 0.01 ^{ab}	0.0647 ± 0.01 ^a	0.02

^{a,b,c} Values within a row with different letters differ significantly (p < 0.05). Control = lemon free diet; 1% DLP = 1 % dry lemon powder; 2% DLP = 2% dry lemon powder.

(4) Serum and Hepatic Oxidative Stress Biomarkers as Affected by Dry Lemon Supplementation

The levels of serum oxidative stress biomarkers (Table 8) such as GST, CAT, SOD, GSH, and GSH-Px were significantly (p < 0.05) increased when 1% or 2% DLP was administrated to growing rabbits compared to those of the control group. Whereas GST activity in rabbit serum significantly (p < 0.05) increased by 3.89% and 5.09% compared to that in the control group, a significant difference was not found between treated groups 1 or 2% DLP. Additionally, data indicated that the catalase activity (CAT) did not significantly differ with 1% or 2% DLP treated levels. In contrast, lipid peroxidation,

MDA level, significantly (p < 0.05) decreased when rabbits fed on DLP (1% or 2%) were compared with the control group, as shown in Table 8. Moreover, the highest level of 2% DLP had the lowest MDA values compared with other groups. As shown in Table 8, the results revealed that the values of SOD, GSH and GSH-Px activity of treated rabbits 1% or 2% DLP, increased significantly (p < 0.05) compared to the levels for those fed the control diet. Additionally, hepatic oxidative biomarkers displayed the same trend in serum.

Table 8. Effects of dried lemon diet on serum and hepatic oxidative stress biomarkers of growing rabbits.

Itome	E	<i>n</i> -Vəluo		
items	Control	1% DLP	2% DLP	<i>p</i> -value
Serum				
GST (U/L)	67.58 ± 3.04 ^b	70.21 ± 3.06 ^a	71.02 ± 3.04 ^a	0.02
CAT (U/L)	20.21 ± 1.15 ^b	22.01 ± 1.16^{a}	22.10 ± 1.14 ^a	0.03
SOD (U/mL)	1.90 ± 0.05 ^c	2.98 ± 0.05 ^b	3.25 ± 0.05^{a}	0.02
GSH (mg/dL)	0.65 ± 0.01 ^c	0.72 ± 0.01 ^b	0.78 ± 0.01 ^a	0.02
GSH-Px activity (mU/mL)	11.37 ± 1.30 ^b	13.38 ± 1.26 ^a	13.35 ± 1.28 ^a	0.02
MDA (nmol/ mL)	2.90 ± 0.20^{a}	2.41 ± 0.18 ^b	2.25 ± 0.16 ^c	0.02
Liver				
GST (U/g tissue)	123.02 ± 4.05 ^b	135.56 ± 4.04 ^a	136.58 ± 4.07 ^a	< 0.01
CAT (U/g tissue)	30.12 ± 2.24 ^c	37.29 ± 2.26 ^b	41.00 ± 2.22 ^a	0.03
SOD (U/mg tissue)	134.20 ± 3.56 ^c	150.29 ± 3.58 ^b	158.57 ± 3.54 ^a	< 0.01
GSH (mg/g tissue)	11.23 ± 0.58 ^b	12.96 ± 0.54 ^a	13.31 ± 0.56 ^a	0.02
GSH-Px activity (U/g tissue)	142.73 ± 3.16 ^c	155.83 ± 3.17 ^b	160.53 ± 3.13^{a}	0.02
MDA (nmol/g protein)	11.22 ± 1.80^{a}	7.13 ± 1.78 ^b	6.18 ± 1.79 ^b	0.02

^{a,b,c} Values within a row with different letters differ significantly (p < 0.05). GST = glutathione S transferase; CAT = catalase; SOD = superoxide dismutase; GSH = glutathione; GSH-Px = glutathione peroxidase. Control = lemon free diet; 1% DLP = 1 % dry lemon powder; 2% DLP = 2% dry lemon powder.

4. Discussion

Dietary antioxidant supplements can enhance the physiological and productive statuses of animals by modifying metabolic processes. The current study aimed to investigate the influences of dry lemon on the hemato-biochemical alterations, and antioxidant indices of growing rabbits. Firstly, we sought to investigate in vitro phytochemical compounds of dry lemon (total phenolics, total flavonoids content) as well as total antioxidant capacity. The presence of phenolic compounds and flavonoids in the diet play an important role in cancer chemoprevention. Additionally, the health benefits of phenolic compounds depend on their consumption and bioavailability [34]. Phenolic compounds have a wide range of biological and chemical activities, including radical scavenging properties. Total phenolic content and total flavonoids content were 120.57 mg/g and 31.28 mg/g, respectively. In contrast, Sultana et al. [35] reported respective values of 158.79 mg/g and 29.33 mg/g. The differences between these findings and our data may be attributed to several factors including the regions and seasons of the sample collection, sections (peels, fruits), used for analysis, and the extraction procedure.

Secondly, the DPPH method was employed to assess the antioxidant activity of the dried lemon methanolic extract.

Through phytochemical analysis, the percentage of DPPH radicals inhibition of *C. limon* extract was found to be 49.12%. A similar, antioxidant activity (43.60%) of lemon peel was reported by Reyhan et al. [36] and may have been due to its high phenolic content. Our results also demonstrated that *C. limon* extracts possess a high concentration of total polyphenols (120.57 mg/g) and flavonoids (31.28 mg/g).

The antioxidant activity of lemon fruits might be due to the occurrence of ascorbic acid content and biologically active secondary metabolites, which have been known to display significant antioxidant activity [37]. Data of the present study showed that dry lemon methanolic extract contains marked

content of total phenolics, flavonoids, and antioxidant activity. Several phytochemical molecules were identified by dry lemon extract GC analysis.

Generally, terpenoids, glycosides, and coumarins give the dry lemon extract their antioxidant and antimicrobial properties [38]. The significance and activities of these molecules attribute to their lipophilic character of hydrocarbon skeleton and functional groups [39]. GC analysis revealed that dry lemon methanolic extract had a relatively high content of 5-(Hydroxymethyl)-2-(dimethoxymethyl) furan, imidazole, and limonene, which are known to possess crucial antioxidant activity. Interactions among the components of the dry lemon methanolic extract could also affect their activity because of the synergistic effect of their main constituents [40]. Increasing terpenes levels makes essential oils more effective [41]. A substantial number of studies reported that phenolic compounds not only cause the antioxidant activity, but that all the phytochemical molecules are responsible for this activity [42].

Data revealed that adding both levels of DLP (1% or 2%) to the diet of growing rabbit diets significantly increased their body weight, weight gain, and carcass weight. The improvement of rabbit performance by adding DLP to their diets may be due to the use of lemon as whole fruit, which led to an increase in the appetite and a higher consumption of feed. Also, lemon fruits were enriched with the most essential amino acids, potent antioxidant activity, total phenolic and total flavonoids compounds including anthocyanins, coumarins, flavonoids, flavones, isoflavones, catechins, isocatechins, and lignans, which might be represent the optimum antioxidant activity of dry lemons. A similar study conducted by Chaudry et al. [43] revealed that adding 5% of citrus pulp into broiler diets did not have any significant effects on feed consumption. This has been confirmed in the present study. Furthermore, our results concerning feed conversion differed from those of Mourao et al. [44] who found that boilers fed 5% or 10% citrus pulp had significantly higher FCR than the control birds. However, Wang et al. [21] added 0%, 4%, 8%, 12%, or 16% DCP to a geese diet until 70 d of age, and they found that geese that were fed on diets containing 4% DCP resulted in higher average daily gain than other groups. Additionally, diets containing 16% DCP led to an elevation in the average daily feed intake and a higher FCR compared to control and 12%, respectively. Diet of geese supplement with DCP showed no effects on carcass traits across the treatment groups [45]. However, rabbits fed the control diet had the highest values for the absolute and proportions of the liver, kidney, and heart compared to treated groups. The highest values of carcass and dressing percentages were observed when rabbits were fed the higher amount of DLP (2%) than the lower amount (1% DLP). No effect was detected in the absolute and proportion of the kidney based on addition amount. The previous results disagree with the study conducted by Petracci et al. [15], which showed that gizzard weight and abdominal fat percentages were significantly affected by the addition of DLP in broiler diets. However, numerically, DLP had beneficial effects on some carcass traits.

Following DLP supplementation, a significant improvement in all hematological parameters was found. Moreover, 2% DLP led to the highest values for RBCs, PCV, Hb, MCV, and MCH (5.96%, 8.57%, 8.56%, 2.46% and 2.41%, respectively) compared to the control group. The enhancement in most hematological parameters might be due to the presence of many active compounds such as vitamin C, flavonoids, iron, and pyridoxine. Azra et al. [17] reported similar results, as they found that different *C. limon* doses (0.2, 0.4, and 0.6 mL/kg) could significantly affect different hematological parameters in healthy rabbits.

The results of the current study showed that *Citrus limon* significantly increased the thyroid hormone levels (T3, T4, and T3/T4 ratio) compared to those found for the control group. This finding is supported by Miler et al. [45], who found that *Citrus* flavanones increased serum TSH without changing T4 levels. Moreover, by supplementing rabbit diets with 1% or 2% DLP showed a significant reduction in serum triglycerides, cholesterol, LDL-c, and VLDL-c were significantly reduced, while the level of HDL-c level statistically increased compared to the level in control animals. The hypolipidemic effect of *C. limon* may be due to its flavonoids content, which can reduce blood cholesterol, triglycerides, and LDL-c levels based on its antioxidant effect [46]. Complete methoxyylation of ring A with Citrus flavonoids seems to be an optimal structural change that exhibits substantial effects on the modulation

of hepatic lipid metabolism via the suppressed secretion of lipoproteins with apoB in HepG2 cells [47]. Other optimal molecular structures, such as tangeretin and nobiletin, exhibited a lower effect on blood cholesterol levels and triglyceride concentration. Also, *Citrus* flavonoids, such as hesperidin and naringin content lacking a fully methoxylated ring, can virtually exhibit little or no reduction in the effects of lipids in humans [48].

In the current study, a significant elevation in the levels of serum HDL was detected [49], suggesting that *C. limon* may serve as a valuable supplement in rabbit diet. Other compounds, such as pectin, a component of the plant cell wall, was reported to play a role in reducing cholesterol. This is due to the structure of flavonoids containing numerous OH groups, which can supply H atoms to quench free radicals, ultimately contributing to its potent antioxidant property [50].

Three major levels of defense mechanisms were found in the antioxidant system of living cells. The activities of SOD, GSH-PX, and CAT is the first, which act to prevent free radical formation [51]. The second level act by chain-breaking antioxidants (vitamin E, C, carotenoids) that disrupt chain multiplication and formation. The third level restores or eliminates damaged molecules from the cell by the help of specific enzymes [52].

More importantly, this study showed that the activities of antioxidant enzymes such as GST CAT, SOD, and GSH were increased, while that of MDA was markedly decreased in the rabbits of the DLP treatment group compared to those in the control group. The increase in activities of antioxidant enzymes may be related to active DLP constituents such as ascorbic acid, flavonoids, and phenolic compounds and agree with many previous studies [53–55], where citrus extracts, such as citrus peel, *C. karna*, and *C. limetta* were demonstrated to potentially have antioxidant bioactivity. The most important defense mechanism of dry lemon phenolic compounds is based on the absorption and neutralization of free radicals [56]. However, previous reports have placed more emphasis on the antioxidant capacity of *C. limon* [4,57,58]. *C. limon* also contains several constituents such as 3-carene, imidazole, D-limonene, geraniol, and squalene as well as other compounds reported as potent antioxidants that can prevent microsomal lipid peroxide and protein degradation of ROS [59–63], These data are in accordance with our phytochemical study results which highlighted *C. limon* as a powerful scavenging agent.

5. Conclusions

The present study demonstrated that rabbits fed a diet supplemented with 1% DLP or 2% DLP had an increase in their productive performance, better feed conversion, enhanced hematological and biochemical values, improved antioxidant enzyme activity, and inhibited lipid peroxidation. These results suggest that DLP would be effective in improving their physiological status and antioxidant capacity during growth.

Author Contributions: Conceptualization, H.A.M.E.; data curation, H.A.M.E. and M.M.; formal analysis, H.A.M.E., D.H.D., A.A.E.-M.A.E.-R., S.M.A.E.-A.E.-S., F.A., and S.A.; funding acquisition, M.M.; investigation, H.A.M.E. and M.M.; methodology, H.A.M.E., D.H.D., A.A.E.-M.A.E.-R., S.M.A.E.-A.E.-S., and M.M.; software, H.A.M.E., D.H.D., A.A.E.-M.A.E.-R., and S.M.A.E.-A.E.-S.; writing—original draft, H.A.M.E.; writing—review editing, H.A.M.E., S.M.A.E.-A.E.-S., A.A.E.-M.A.E.-R., S.A.A.-L., and S.S.A.-R.

Funding: This work was funded by the Deanship of Scientific Research at King Saud University (research group No. RG-1440-085).

Acknowledgments: The authors extend their appreciation to the Deanship of Scientific Research at King Saud University for funding this work through research group No. (RG-1440-085).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Sikora, E.; Ewa, C.; Topolska, K. The sources of natural antioxidants. *Acta Sci. Pol. Technol. Aliment.* 2008, 7, 5–17.
- Pellegrini, N.M.; Serafini, B.O.; Colombi, D.D.; Rio, S.T.; Salvotore, R.A.; Bionchi, W.O. Total antioxidant capacity of plant foods, beverages and oils consumed in italy assessed by three different in vitro assays. *J. Nutr.* 2007, 2812–2819. [CrossRef] [PubMed]
- 3. Chun, O.K.Y.; Chung, S.J.; Claycombe, K.J.; Song, W.O. Serum C-reactive protein concentrations are inversely associated with dietary flavonoid intake in U.S. Adults. *Nutrition* **2008**, *138*, 753–760. [CrossRef] [PubMed]
- 4. Xu, G.; Liu, D.; Chen, J.; Ye, X.; Ma, Y.; Shi, J. Juice component and antioxidant capacity of citrus varieties cultivated in China. *Food Chem.* **2008**, *106*, 545–551. [CrossRef]
- 5. Tripoli, E.; Guardia, M.L.; Giammanco, S.; Di, M.D.; Giammanco, M. Citrus flavonoids: Molecular structure, biological activity and nutritional properties: A review. *Food Chem.* **2007**, *104*, 466–479. [CrossRef]
- Peterson, O.A.; Marian, O.A.; Marian, A.E.; Leblong, P.S. Synthesis of complex carbohydrates in the golgi region as shown by autoradiography after injection of labelled glucose. *J. Cell Biol.* 2006, 21, 143–148. [CrossRef] [PubMed]
- Viuda, M.M.; Ruiz, N.Y.; Fernandez, L.J.; Perez, A.J. Antifungal activity of lemon (*Citrus lemon* L.), mandarin (*Citrus reticulata* L.), grapefruit (*Citrus paradisi* L.) and orange (*Citurs sinensis* L.) essential oils. *Food Control* 2008, 19, 1130–1138. [CrossRef]
- 8. Arias, B.A.; Ramon, L.L. Pharmacological properties of citrus and their ancient and their medical use in the mediteraanean region. *J. Ethnopharmacol.* **2005**, *97*, 89–95. [CrossRef]
- 9. Ezzat, S.M. In vitro inhibition of *Candida albicans* growth by plant extracts and essential oils. *World J. Microbiol. Biotechnol.* **2000**, *17*, 757–759. [CrossRef]
- Gonzalez, A.; Robles-Sa'nchez, G.; Martinez-Tellez, R.M.; Olivas, M.A.; Alvarez-Parrilla, G.I.; De La Rosa, L.A. Bioactive compounds in fruits: Health benefits and effect of storage conditions. *Stewart Postharvest Rev.* 2008, 4, 1–10.
- 11. Pennington, J.A.T.; Fisher, R.A. Food component profiles for fruit and vegetable subgroups. *J. Food Compos. Anal.* **2010**, *23*, 411–418. [CrossRef]
- 12. Dembitsky, V.M.; Poovarodom, S.; Leontowicz, H. The multiple nutrition properties of some exotic fruits: Biological activity and active metabolites. *Food Res. Int.* **2011**, *44*, 1671–1701. [CrossRef]
- Bouzenna, H.; Samout, N.; Dhibi, S.; Mbarki, S.; Akermi, S.; Khdhiri, A.; Elfeki, A.; Hfaiedh, N. Protective effect of essential oil from *Citrus limon* against aspirin-induced toxicity in rats. *Hum. Exp. Toxicol.* 2019, *38*, 499–509. [CrossRef] [PubMed]
- 14. Banerjee, G.C. A Textbook of Animal Husbandry; Oxford and IBH Publishing: New Delhi, India, 2005.
- 15. Petracci, M.; Bianchi, M.; Cavani, C. Development of rabbit meat products fortified with n-3 polyunsaturated fatty acids. *Nutrients* **2009**, *1*, 111–118. [CrossRef] [PubMed]
- 16. Nobakht, A. Effects of different levels of dried lemon (*Citrus aurantifulia*) pulp on performance, carcass traits, blood biochemical and immunity parameters of broilers. *Iran. J. Appl. Anim. Sci.* **2013**, *3*, 145–151.
- 17. Azra, R.; Rafeeq, A.K.; Talat, M.; Tazeen, M.; Mansoor, A. In vitro/in vivo effect of *Citrus limon* (*L. burm*. F.) juice on blood parameters, coagulation and anticoagulation factors in rabbits. *Pak. J. Pharm. Sci.* **2014**, 27, 907–915.
- 18. Shen, W.; Xu, Y.; Lu, Y.H. Inhibitory effects of *Citrus* flavonoids on starch digestion and antihyperglycemic effects in HepG2 cells. *J. Agric. Food Chem.* **2012**, *60*, 9609–9619. [CrossRef]
- Jung, U.J.; Lee, M.K.; Jeong, K.S.; Chol, M.S. The hypoglycemic effects of hesperidin and naringin are partly mediated by hepaticglucose regulating enzymes in C57 BL/KsJ-db/db Mice. *J. Nutr.* 2004, 134, 2499–2503. [CrossRef]
- 20. Nazok, A.; Rezaei, M.; Sayyahzadeh, H. Effect of different levels of dried citrus pulp on performance, egg quality, and blood parameters of laying hens in early phase of production. *Trop. Anim. Health Prod.* **2010**, *42*, 737–742. [CrossRef]
- 21. Wang, C.G.L.; Gao, J.X.; Huang, K.S.; Zhang, H.; Zhong, H.W.; Wang, J.; Su, M.; Xie, Q.; Wang, G. Nutritive value of dry citrus pulp and its effect on performance in geese from 35 to 70 days of age. *J. Appl. Poult. Res.* **2017**, *26*, 253–259. [CrossRef]

- 22. Al-Fatimi, M.; Wurster, M.; Schroder, G.; Lindequist, U. Antioxidant, antimicrobial and cytotoxic activities of selected medicinal plants from Yemen. *J. Ethnopharmacol.* **2007**, *111*, 657–666. [CrossRef] [PubMed]
- 23. Wolfe, K.; Wu, X.; Liu, R.H. Antioxidant activity of apple peels. J. Agric. Food Chem. 2003, 51, 609–614. [CrossRef] [PubMed]
- 24. Djeridane, A.; Yousfi, M.; Nadjemi, B.; Boutassouna, D.; Stocker, P.; Vidal, N. Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. *Food Chem.* **2006**, *97*, 654–660. [CrossRef]
- 25. Oliveira, I.; Sousa, A.; Ferreira, I.; Bento, A.; Estevinho, L.; Pereira, J.A. Total phenols, antioxidant potential and antimicrobial activity of walnut (*Juglans regia* L.) green husk. *Food Chem. Toxicol.* **2008**, *46*, 2326–2331. [CrossRef] [PubMed]
- 26. Kulkarni, A.; Govindappa, M.; Ramachandra, Y.L.; Koka, P. GC-MS analysis of methanol extract of *Cassia fistula* and its in vitro anticancer activity on human prostate cancer cell line. *Ind. Am. J. Pharm. Res.* **2015**, *5*, 937–944.
- 27. AOAC International. Official Methods of Analysis, 18th ed.; AOAC International: Washington, DC, USA, 2006.
- 28. VanSoest, P.J.; Robertson, B.; Lewis, B.A. Methods for dietary fiber, neutral detergent fiber and non-starch polysaccharides in relation to animal nutrition. *J. Dairy Sci.* **1991**, *74*, 3583–3597. [CrossRef]
- 29. National Research Council (NRC). *Nutrient Requirements of Beef Cattle*, 8th ed.; The National Academies Press: Washington, DC, USA, 1984.
- Trachootham, D.; Lu, W.; Ogasawara, M.A.; Nilsa, R.D.; Huang, P. Redox regulation of cell survival. *Antioxid. Redox Signal.* 2008, 10, 1343–1374. [CrossRef] [PubMed]
- 31. Lopes-Virella, M.F.; Binzafar, N.; Rackley, S.; Takei, A.; LaVia, M.; Virella, G. The uptake of LDL-IC by human macrophages: Predominant involvement of the Fc gamma RI receptor. *Atherosclerosis* **1997**, *135*, 161–170. [CrossRef]
- 32. Ahmadi, S.A.; Mohammad, A.B.; Katayoun, G.M.; Parvin, T.; Seyed, M.D. The impact of low serum triglycerides on LDL-cholesterol estimation. *Arch. Iran. Med.* **2008**, *11*, 318–321.
- 33. SAS Institute Inc. SAS/STAT User's Guide: Statistics, Version 6, 4th ed.; SAS Institute, Inc.: Cary, NC, USA, 2003.
- 34. Rodríguez, H.; Gómez-Cordovés, C.; Curiel, J.A.; Mancheño, J.M.; Landete, J.M.; Rivas, B.D.L.; Muñoz, R. Food phenolics and lactic acid bacteria. *Int. J. Food Microbiol.* **2009**, 132, 79–90. [CrossRef]
- 35. Sultana, B.; Anwar, F.; Mushtaq, M.; Alim, M. *Citrus* residues: A potential source of phenolics with high antioxidant values. *Int. Food Res. J.* **2015**, *22*, 1163–1168.
- 36. Reyhan, I.; Serap, D.; Nurcan, D.; Mehmet, E.D.; Metin, G. Phenolic content, antioxidant activities and stimulatory roles of citrus fruits on some lactic acid bacteria. *Arch. Biol. Sci. Belgrade* **2015**, *67*, 1313–1321.
- 37. Barile, E.; Bonanomi, G.; Antignani, V.; Zolfaghari, B.; Sajjadi, E.; Scala, F.; Lanzotti, V. Phytochemical screening and antimicrobial assessment of *Abutilon mauritianum*, *Bacopa monifera* and *Datura stramonium*. *Phytochemistry* **2007**, *68*, 596–603. [CrossRef] [PubMed]
- 38. Buchanan, B.B.; Gruissem, W.; Jones, R.L. *Biochemistry & Molecular Biology of Plants*; American Society of Plant Physiologists: Rockville, MA, USA, 2000.
- 39. Gonny, M.; Bradesi, P.; Casanova, J. Identification of the components of the essential oil from wild Corsican *Daucus carota* L. using 13C-NMR spectroscopy. *Flavour Fragr. J.* **2004**, *19*, 424–433. [CrossRef]
- 40. Lambert, R.J.; Skandamis, P.N.; Coote, P.J.; Nychas, G.J. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *J. Appl. Microbiol.* **2001**, *91*, 453–462. [CrossRef] [PubMed]
- 41. Pinto, E.; Pina-Vaz, C.; Salgueiro, L.; Jose Goncalves, M.; Costa-de-Oliveira, S.; Cavaleiro, C.; Palmeira, I.; Rodrigues, I.; Martinez-de-Oliveira, J. Antifungal activity of the essential oil of *Thymus pulegioides* on Candida, *Aspergillus* and dermatophyte species. *J. Med. Microbiol.* **2006**, *55*, 1367–1373. [CrossRef]
- 42. Cosentino, S. In vitro antimicrobial activity and chemical composition of Sardinian *Thymus* essential oils. *Lett. Appl. Microbiol.* **1999**, *29*, 130–135. [CrossRef] [PubMed]
- 43. Chaudry, M.A.; Badshan, A.; Bibi, N.; Zeb, A.; Ahmed, T.; Ali, S.; Termeulen, U. Citrus waste in poultry rations. *Arch. Geflugelkunde* **2004**, *68*, 206–210.
- 44. Mourao, J.L.; Pinheiro, V.M.; Prates, A.M.; Bessa, R.J.B.; Ferreira, L.M.A. Effect of dietary dehydrated pasture and citrus pulp on the performance and meat quality of broiler chickens. *Poult. Sci.* **2008**, *87*, 733–743. [CrossRef]
- 45. Miler, M.; Jarić, I.; Živanović, J.; Ajdžanović, V.; Tanić, N.; Milošević, V.; Šošić-Jurjević, B. Citrus flavanones mildly interfere with pituitary-thyroid axis in old-aged male rats. *Acta Histochem.* **2017**, *119*, 292–301. [CrossRef]

- 46. Khan, Y.; Khan, R.A.; Afroz, S.; Siddiq, A. Evaluation of hypolipidemic effect of Citrus lemon. *J. Basic Appl. Sci.* **2010**, *6*, 39–43.
- 47. Lin, Y.; Vermeer, M.A.; Bos, W.; Van Buren, L.; Schuurbiers, E.; Miret-Catalan, S.; Trautwein, E.A. Molecular structures of citrus flavonoids determine their effects on lipid metabolism in HepG2 cells by primarily suppressing apoB secretion. *J. Agric. Food Chem.* **2011**, *59*, 4496–4503. [CrossRef] [PubMed]
- 48. Demonty, I.; Lin, Y.; Zebregs, Y.E.; Vermeer, M.A.; Van der Knaap, H.C.; Jäkel, M.; Trautwein, E.A. The citrus flavonoids hesperidin and naringin do not affect serum cholesterol in moderately hypercholesterolemic men and women. *J. Nutr.* **2010**, *140*, 1615–1620. [CrossRef] [PubMed]
- 49. Stein, O.; Stein, Y. A thero-protective mechanism of HDL. Atherosclerosis 1999, 144, 285–301. [CrossRef]
- 50. Shahelian, R.M.D. Decreased pro-inflammatory cytokine production by LPS-stimulated PBMC upon in vitro incubation with the flavonoids apigenin, luteolin or chrysin, due to selective elimination of monocytes/macrophages. *Biochem. Pharm.* **2005**, *69*, 241–248. [CrossRef] [PubMed]
- 51. Birben, E.; Sahiner, U.M.; Sackesen, C.; Erzurum, S.; Kalayci, O. Oxidative stress and antioxidant defense. *World Allergy Organ. J.* **2012**, *5*, 9. [CrossRef] [PubMed]
- 52. Ojabo, L.D.; Oluremi, O.I.A.; Carew, S.N.; Uza, D.V. Heamatology and serum biochemistry of pullet grower chickens fed sweet orange (*Citrus sinensis*) fruit peel meal-based diets. *Res. Opin. Anim. Vet. Sci.* 2013, *3*, 252–256.
- 53. Akbarian, A.; Golian, A.; Gilani, A.; Kermanshahi, H.; Zhaleh, S.; Akhavan, A.; De Smet, S.; Michiels, J. Effect of feeding citrus peel extracts on growth performance, serum components, and intestinal morphology of broilers exposed to high ambient temperature during the finisher phase. *Livest. Sci.* **2013**, 157, 490–497. [CrossRef]
- 54. Surai, P.F. Natural Antioxidants in Avian. Nutrition and Reproduction; Nottingham University Press: Nottingham, UK, 2002.
- 55. Jagdeep, S.; Shailja, S.; Arunachalam, M. In-vitro evaluation of bioactive compounds, anti-oxidant, lipid peroxidation and lipoxygenase inhibitory potential of *Citrus karna* L. peel extract. *J. Food Sci. Technol.* **2014**, *51*, 67–74.
- 56. Impellizzeri, D.; Bruschetta, G.; Di Paola, R.; Ahmad, A.; Campolo, M.; Cuzzocrea, S.; Esposito, E.; Navarra, M. The anti-inflammatory and antioxidant effects of bergamot juice extract (BJe) in an experimental model of inflammatory bowel disease. *Clin. Nutr.* **2014**, *27*, 1146–1154. [CrossRef]
- 57. Padilla-Camberos, E.; Lazcano-Díaz, E.; Flores-Fernandez, J.M.; Owolabi, M.S.; Allen, K.; Villanueva-Rodríguez, S. Evaluation of the inhibition of carbohydrate hydrolyzing enzymes, the antioxidant activity, and the polyphenolic content of *Citrus limetta* peel extract. *Sci. World J.* **2014**, *2014*, 1–4. [CrossRef] [PubMed]
- Osawa, T. Novel natural antioxidants for utilization in food and biological systems. In *Uritani I;* Garcia, V.V., Mendoza, E.M., Eds.; Postharvest biochemistry of plant food-materials in the tropics; Japan Scientific Societies Press: Tokyo, Japan, 1994; pp. 241–251.
- 59. Aazza, S.; Lyoussi, B.; Miguel, M.G. Antioxidant and antiacetylcholinesterase activities of some commercial essential oils and their major compounds. *Molecules* **2011**, *16*, 7672–7690. [CrossRef] [PubMed]
- 60. Tansini, C.M.; Durigon, K.; Testa, C.G.; Belló-Klein, A.; Wajner, M.; Wannmacher, C.M.; Wyse, A.T.; Dutra-Filho, C.S. Effects of histidine and imidazolelactic acid on various parameters of the oxidative stress in cerebral cortex of young rats. *Int. J. Dev. Neurosci.* **2004**, *22*, 67–72. [CrossRef] [PubMed]
- Bacanlı, M.; Anlar, H.G.; Aydın, S.; Çal, T.; Arı, N.; Ündeğer-Bucurgat, Ü.; Başaran, A.A.; Başaran, N. D-limonene ameliorates diabetes and its complications in streptozotocin-induced diabetic rats. *Food Chem. Toxicol.* 2017, 110, 434–442. [CrossRef] [PubMed]
- 62. Ozkaya, A.; Sahin, Z.; Gorgulu, A.O.; Yuce, A.; Celik, S. Geraniol attenuates hydrogen peroxide-induced liver fatty acid alterations in male rats. *J. Intercult. Ethnopharmacol.* **2016**, *6*, 29–35. [CrossRef] [PubMed]
- 63. Ravi-Kumar, S.; Narayan, B.; Kizawa, Y.; Hosokawa, M.; Miyashita, K. Does squalene alter the antioxidant potential of astaxanthin and fucoxanthinol? In vitro evidence in RAW 264.7 cells, a murine macrophage. *J. Food Sci. Technol.* **2016**, *53*, 2139–2143. [CrossRef] [PubMed]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).