

# Complete Genome Sequences of 11 *Bordetella pertussis* Strains Representing the Pandemic *ptxP3* Lineage

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**Pathogen adaptation has contributed to the resurgence of pertussis. To facilitate our understanding of this adaptation we report here 11 completely closed and annotated *Bordetella pertussis* genomes representing the pandemic *ptxP3* lineage. Our analyses included six strains which do not produce the vaccine components pertactin and/or filamentous hemagglutinin.**

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*Bordetella pertussis* is the causative agent of pertussis or whooping cough, a respiratory disease which is most severe in young unvaccinated infants. After the introduction of vaccination in the 1950s, there was a steep decline in disease incidence. However, in the 1990s pertussis resurged. The increase in notifications was initially attributed to increased awareness and improved diagnostics. However, later it became clear that the pertussis resurgence was mainly due to suboptimal vaccines and pathogen adaptation (1). Large shifts in the *B. pertussis* population resulted in antigenic divergence between circulating strains and vaccine strains (2–4). Further, in the 1980s, strains emerged with a novel allele for the pertussis toxin promoter *ptxP3*, replacing the resident *ptxP1* strains. The *ptxP3* strains produce more pertussis toxin than *ptxP1* strains and therefore may suppress host immunity more efficiently (5–7). More recently, strains deficient in the vaccine components pertactin (Prn) and filamentous hemagglutinin (FHA) were detected (8–15). Loss of one or both of these antigens may confer a selective advantage in vaccinated populations (16, 17). Here we present the completely closed and annotated genome sequences of 11 *B. pertussis* isolates which represent the pandemic *ptxP3* lineage and include six strains deficient in Prn and/or FHA.

Genomic DNA was isolated as described previously (18) and a 10-kb library was prepared. Sequencing was performed using a PacBio RS system with 6 single-molecule real-time (SMRT) cells per genome. The generated sequences were *de novo* assembled with HGAP (19) and trimmed and rotated by hand, resulting in a single circular contig for all genomes. *B. pertussis* genomes are highly similar and therefore RATT (20) was used to transfer annotations from *B. pertussis* Tohama I, CS, and 18323 (21–23). Afterward, sequenced genomes were manually checked for genes not present in the reference genomes.

The genomes comprised 4,100,705 to 4,111,557 bp and were predicted to have between 3,818 and 3,829 genes. Variation in the number of copies of the insertion sequence element IS481, which varied between 249 and 258 copies, was mainly responsible for the difference in gene numbers. The 11 strains were highly similar with respect to single nucleotide polymorphisms (SNPs) ( $n =$

335) and small (up to 1,769 bp) insertions and deletions ( $n = 118$ ). However, significant genome arrangements were observed, most likely mediated by insertion elements. Prn deficiency was caused by insertion of IS481 in the *prn* gene (strains B3582, B3629, and B3640), a 25-bp deletion in the *prn* gene (strain B3621), or C-to-T mutation resulting in a stop codon in the *prn* gene (strain B3658). In one strain (B3582), FHA-deficiency was caused by insertion of a G in a homopolymeric tract of 10 Gs, leading to a premature translational termination. In the other FHA-deficient strain (B3585), no mutations in the *fhaB* gene, its promoter, or genes required for its surface expression were detected.

Comparisons of these strains and already published strains (18, 21, 22, 24) suggest that *B. pertussis* evolves not only by small mutations but also by major genome rearrangements which may affect gene regulation.

**Nucleotide sequence accession numbers.** The whole-genome shotgun projects have been deposited in DDBJ/ENA/GenBank under the accession numbers listed in Table 1. The versions described in this paper are the first versions.

TABLE 1 Characteristics of the 11 *B. pertussis* strains

Strain	Accession no.	Isolation yr	Country	<i>ptxP</i> type	<i>fim3</i> type	Prn <sup>a</sup>	FHA <sup>a</sup>
B1838	<a href="#">CP011440</a>	1999	Netherlands	3	2	+	+
B1865	<a href="#">CP011441</a>	1999	Netherlands	3	2	+	+
B3405	<a href="#">CP011442</a>	2010	Netherlands	3	1	+	+
B3582	<a href="#">CP011443</a>	2009	Sweden	3	2	–	–
B3585	<a href="#">CP011444</a>	2009	Sweden	3	1	+	–
B3621	<a href="#">CP011401</a>	2008	France	3	2	–	+
B3629	<a href="#">CP011400</a>	2009	France	3	2	–	+
B3640	<a href="#">CP011445</a>	2010	Netherlands	3	1	–	+
B3658	<a href="#">CP011446</a>	2009	Norway	3	1	–	+
B3913	<a href="#">CP011447</a>	2012	Netherlands	3	1	+	+
B3921	<a href="#">CP011448</a>	2012	Netherlands	3	1	+	+

<sup>a</sup> +, strain produces Prn and/or FHA; –, strain does not produce Prn and/or FHA.

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## REFERENCES

- Mooi FR, Van Der Maas NAT, De Melker HE. 2014. Pertussis resurgence: waning immunity and pathogen adaptation—two sides of the same coin. *Epidemiol Infect* 142:685–694. <http://dx.doi.org/10.1017/S0950268813000071>.
- Lam C, Octavia S, Bahrame Z, Sintchenko V, Gilbert GL, Lan R. 2012. Selection and emergence of pertussis toxin promoter ptxP3 allele in the evolution of *Bordetella pertussis*. *Infect Genet Evol* 12:492–495. <http://dx.doi.org/10.1016/j.meegid.2012.01.001>.
- Van Gent M, Bart MJ, van der Heide HGJ, Heuvelman KJ, Mooi FR. 2012. Small mutations in *Bordetella pertussis* are associated with selective sweeps. *PLoS One* 7:e46407. <http://dx.doi.org/10.1371/journal.pone.0046407>.
- Bart MJ, Harris SR, Advani A, Arakawa Y, Bottero D, Bouchez V, Cassidy PK, Chiang C-, Dalby T, Fry NK, Gaillard ME, van Gent M, Guiso N, Hallander HO, Harvill ET, He Q, van der Heide HGJ, Heuvelman K, Hozbor DF, Kamachi K, Karataev GI, Lan R, Lutynska A, Maharjan RP, Mertsola J, Miyamura T, Octavia S, Preston A, Quail MA, Sintchenko V, Stefanelli P, Tondella ML, Tsang RS, Xu Y, Yao SM, Zhang S, Parkhill J, Mooi FR. 2014. Global population structure and evolution of *Bordetella pertussis* and their relationship with vaccination. *mBio* 5:e01074. <http://dx.doi.org/10.1128/mBio.01074-14>.
- Mooi FR, van Loo IHM, van Gent M, He Q, Bart MJ, Heuvelman KJ, de Greeff SC, Diavatopoulos D, Teunis P, Nagelkerke N, Mertsola J. 2009. *Bordetella pertussis* strains with increased toxin production associated with pertussis resurgence. *Emerg Infect Dis* 15:1206–1213. <http://dx.doi.org/10.3201/eid1508.081511>.
- King AJ, van der Lee S, Mohangoo A, van Gent M, van der Ark A, van de Waterbeemd B. 2013. Genome-wide gene expression analysis of *Bordetella pertussis* isolates associated with a resurgence in pertussis: elucidation of factors involved in the increased fitness of epidemic strains. *PLoS One* 8:e66150. <http://dx.doi.org/10.1371/journal.pone.0066150>.
- De Gouw D, Hermans PWM, Bootsma HJ, Zomer A, Heuvelman K, Diavatopoulos DA, Mooi FR. 2014. Differentially expressed genes in *Bordetella pertussis* strains belonging to a lineage which recently spread globally. *PLoS One* 9:e84523. <http://dx.doi.org/10.1371/annotation/1b2cfe52-aaea-4148-aa72-a74c78550192>.
- Bouchez V, Brun D, Cantinelli T, Dore G, Njamkepo E, Guiso N. 2009. First report and detailed characterization of *B. pertussis* isolates not expressing pertussis toxin or pertactin. *Vaccine* 27:6034–6041. <http://dx.doi.org/10.1016/j.vaccine.2009.07.074>.
- Otsuka N, Han H, Toyozumi-Ajisaka H, Nakamura Y, Arakawa Y, Shibayama K, Kamachi K. 2012. Prevalence and genetic characterization of pertactin-deficient *Bordetella pertussis* in Japan. *PLoS One* 7:e31985. <http://dx.doi.org/10.1371/journal.pone.0031985>.
- Hegerle N, Paris A-S, Brun D, Dore G, Njamkepo E, Guillot S, Guiso N. 2012. Evolution of French *Bordetella pertussis* and *Bordetella parapertussis* isolates: increase of *Bordetellae* not expressing pertactin. *Clin Microbiol Infect* 18:E340–E346. <http://dx.doi.org/10.1111/j.1469-0691.2012.03925.x>.
- Barkoff A-, Mertsola J, Guillot S, Guiso N, Berbers G, He Q. 2012. Appearance of *Bordetella pertussis* strains not expressing the vaccine antigen pertactin in Finland. *Clin Vaccine Immunol* 19:1703–1704. <http://dx.doi.org/10.1128/CVI.00367-12>.
- Queenan AM, Cassidy PK, Evangelista A. 2013. Pertactin-negative variants of *Bordetella pertussis* in the United States. *N Engl J Med* 368:583–584. <http://dx.doi.org/10.1056/NEJMc1209369>.
- Pawloski LC, Queenan AM, Cassidy PK, Lynch AS, Harrison MJ, Shang W, Williams MM, Bowden KE, Burgos-Rivera B, Qin X, Messonnier N, Tondella ML. 2014. Prevalence and molecular characterization of pertactin-deficient *Bordetella pertussis* in the United States. *Clin Vaccine Immunol* 21:119–125. <http://dx.doi.org/10.1128/CVI.00717-13>.
- Tsang RSW, Shuel M, Jamieson FB, Drews S, Hoang L, Horsman G, Lefebvre B, Desai S, St-Laurent M. 2014. Pertactin-negative *Bordetella pertussis* strains in Canada: characterization of a dozen isolates based on a survey of 224 samples collected in different parts of the country over the last 20 years. *Int J Infect Dis* 28:65–69. <http://dx.doi.org/10.1016/j.ijid.2014.08.002>.
- Zeddeman A, van Gent M, Heuvelman C, van der Heide H, Bart M, Advani A, Hallander H, Wirsing von König C, Riffelman M, Storsaeter J, Vestheim D, Dalby T, Krogfelt K, Fry N, Barkoff A, Mertsola J, He Q, Mooi F. 2014. Investigations into the emergence of pertactin-deficient *Bordetella pertussis* isolates in six European countries, 1996 to 2012. *Euro Surveill* 19. <http://dx.doi.org/10.2807/1560-7917.ES2014.19.33.20881>.
- Martin SW, Pawloski L, Williams M, Weening K, DeBolt C, Qin X, Reynolds L, Kenyon C, Giambrone G, Kudish K, Miller L, Selvage D, Lee A, Skoff TH, Kamiya H, Cassidy PK, Tondella ML, Clark TA. 2015. Pertactin-negative *Bordetella pertussis* strains: evidence for a possible selective advantage. *Clin Infect Dis* 60:223–227. <http://dx.doi.org/10.1093/cid/ciu788>.
- Hegerle N, Dore G, Guiso N. 2014. Pertactin deficient *Bordetella pertussis* present a better fitness in mice immunized with an acellular pertussis vaccine. *Vaccine* 32:6597–6600. <http://dx.doi.org/10.1016/j.vaccine.2014.09.068>.
- Bart MJ, Zeddeman A, van der Heide HGJ, Heuvelman K, van Gent M, Mooi FR. 2014. Complete genome sequences of *Bordetella pertussis* isolates B1917 and B1920, representing two predominant global lineages. *Genome Announc* 2(6):e01301-14. <http://dx.doi.org/10.1128/genomeA.01301-14>.
- Chin C, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.
- Otto TD, Dillon GP, Degraeve WS, Berriman M. 2011. RATT: rapid annotation transfer Tool. *Nucleic Acids Res* 39:e57. <http://dx.doi.org/10.1093/nar/gkq1268>.
- Parkhill J, Sebaihia M, Preston A, Murphy LD, Thomson N, Harris DE, Holden MTG, Churcher CM, Bentley SD, Mungall KL, Cerdeño-Tárraga AM, Temple L, James K, Harris B, Quail MA, Achtman M, Atkin R, Baker S, Basham D, Bason N, Cherevach I, Chillingworth T, Collins M, Cronin A, Davis P, Doggett J, Feltwell T, Goble A, Hamlin N, Hauser H, Holroyd S, Jagels K, Leather S, Moule S, Norberczak H, O'Neil S, Ormond D, Price C, Rabinowitz E, Rutter S, Sanders M, Saunders D, Seeger K, Sharp S, Simmonds M, Skelton J, Squares R, Squares S, Stevens K, Unwin L. 2003. Comparative analysis of the genome sequences of *Bordetella pertussis*, *Bordetella parapertussis* and *Bordetella bronchiseptica*. *Nat Genet* 35:32–40. <http://dx.doi.org/10.1038/ng1227>.
- Zhang S, Xu Y, Zhou Z, Wang S, Yang R, Wang J, Wang L. 2011. Complete genome sequence of *Bordetella pertussis* CS, a Chinese pertussis vaccine strain. *J Bacteriol* 193:4017–4018. <http://dx.doi.org/10.1128/JB.05184-11>.
- Park J, Zhang Y, Buboltz AM, Zhang X, Schuster SC, Ahuja U, Liu M, Miller JF, Sebaihia M, Bentley SD, Parkhill J, Harvill ET. 2012. Comparative genomics of the classical *Bordetella* subspecies: the evolution and exchange of virulence-associated diversity amongst closely related pathogens. *BMC Genomics* 13:545. <http://dx.doi.org/10.1186/1471-2164-13-545>.
- Boinett CJ, Harris SR, Langridge GC, Trainor EA, Merkel TJ, Parkhill J. 2015. Complete genome sequence of *Bordetella pertussis* D420. *Genome Announc* 3(4):e00842-15. <http://dx.doi.org/10.1128/genomeA.00842-15>.