RESEARCH ARTICLE

■ WILEY

Changes in demography and geographic distribution in the weeping pinyon pine (*Pinus pinceana*) during the Pleistocene

Laura Figueroa-Corona^{1,2} | Alejandra Moreno-Letelier³ | Diego Ortega-Del Vecchyo⁴ | Pablo Peláez⁵ | David S. Gernandt⁶ | Luis E. Eguiarte² | Jill Wegrzyn⁷ | Daniel Piñero²

¹Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

²Departamento de Ecología Evolutiva, Instituto de Ecología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

³Jardín Botánico, Instituto de Biología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

⁴Laboratorio Internacional de Investigación sobre el Genoma Humano, Universidad Nacional Autónoma de México, Juriquilla, Mexico

⁵Centro de Ciencias Genómicas, Universidad Nacional Autónoma de México, Cuernavaca, Morelos, Mexico

⁶Departamento de Botánica, Instituto de Biología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

⁷Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, Connecticut, USA

Correspondence

Laura Figueroa-Corona, Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Ciudad de México, Mexico.

Email: laurafc25@gmail.com

Funding information

Consejo Nacional de Ciencia y Tecnología, Grant/Award Number: 449615; PAPIIT-DGAPA, UNAM, Grant/Award Number: IA200620 and IN209816; UC-MEXUS CONACYT, Grant/Award Number: CN-19-29

Abstract

Climate changes, together with geographical barriers imposed by the Sierra Madre Oriental and the Chihuahuan Desert, have shaped the genetic diversity and spatial distribution of different species in northern Mexico. Pinus pinceana Gordon & Glend. tolerates extremely arid conditions. Northern Mexico became more arid during the Quaternary, modifying ecological communities. Here, we try to identify the processes underlying the demographic history of *P. pinceana* and characterize its genetic diversity using 3100 SNPs from genotyping by sequencing 90 adult individuals from 10 natural populations covering the species' entire geographic distribution. We inferred its population history and contrasted possible demographic scenarios of divergence that modeled the genetic diversity present in this restricted pinyon pine; in support, the past distribution was reconstructed using climate from the Last Glacial Maximum (LGM, 22kya). We inferred that P. pinceana diverged into two lineages ~2.49 Ma (95% CI 3.28-1.62), colonizing two regions: the Sierra Madre Oriental (SMO) and the Chihuahuan Desert (ChD). Our results of population genomic analyses reveal the presence of heterozygous SNPs in all populations. In addition, low migration rates across regions are probably related to glacial-interglacial cycles, followed by the gradual aridification of the Chihuahuan Desert during the Holocene.

KEYWORDS

Chihuahuan Desert, demographic contraction, historical isolation, Pinus pinceana, Pleistocene

TAXONOMY CLASSIFICATION Genomics, Global change ecology

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. © 2022 The Authors. *Ecology and Evolution* published by John Wiley & Sons Ltd.

1 | INTRODUCTION

Chihuahuan Desert (ChD) communities have a long history of high variability in their spatial dimensions, driven primarily by water input, which has fluctuated over time (Noy-Meir, 1973; Zavala-Hurtado & Jiménez, 2020).

Geological and climatic events during the Pleistocene affected the diversity patterns of ChD communities (Scheinvar et al., 2020). Several phylogeographic studies have reported dynamic scenarios where populations migrated and expanded to the south through the Central Mexican Plateau during the interglacial (15–20 glacial cycles), followed by events of recolonization of the northern ChD (Van Devender & Burgess, 1985). Such is the case for *Ephedra compacta* (Loera et al., 2017), *Agave lechuguilla* (Scheinvar et al., 2017), and *Pinus remota*, which retreated to Pleistocene refugia in the Bolsón de Mapimí (Lanner & Van Devender, 1981).

The ChD is a high-elevation desert ranging from 600 to 1675 ma.s.l. and receives more rain than other deserts (235 mm of mean annual precipitation; National Park Service, https://www.nps.gov/im/chdn/ecoregion.htm). The ChD is the third-largest desert in the American continent, considered the most diverse in the Western Hemisphere and one of the most diverse arid regions in the world (Beck & Gibbens, 1999). It also has both a high species diversity and richness of endemism (Rzedowski & Calderón de Rzedowski, 1993). The ChD landscape maintains different vegetation types, such as grass steppes, xeric shrubs in the intermountain plains and valleys, and open woodlands at higher elevations (Czaja et al., 2014; Van Devender & Burgess, 1985).

The Sierra Madre Oriental (SMO) is a high mountain range (1480–3720 ma.s.l.) dating to the Late Cretaceous and early Tertiary Laramide (80–40 Ma; Eguiluz de Antuñano et al., 2000). With a high climatic diversity due to its complex physiographic heterogeneity and meteorological phenomena (Hernandez-Cerda & Carrasco-Anaya, 2004), the SMO receives moisture from the Gulf of Mexico; while the eastern slopes have tropical and temperate forests, the western slope is much drier with xeric scrubs and pine-oak-juniper woodlands (Morrone et al., 2017; Rzedowski, 1978).

The ChD biota has experienced climate fluctuations throughout geologic time. Some of these fluctuations occurred during the Late Quaternary. Paleoecological reconstructions suggest that a woodland corridor covered the area between the SMO and the Sierra Madre Occidental in Mexico (Lanner & Van Devender, 1981; Metcalfe, 2006). One of several reductions in pinyon-juniper and oak woodland occurred during the last 2.5 Myears (Pleistocene), which included intense environmental and climatic changes. During the Pleistocene, 11 climatic cycles of growth and reduction in the polar ice cap occurred in North America, affecting the global climatic conditions and consequently the composition of species associated with this woodland corridor. In particular, during the Late Wisconsin glaciation, the vegetation that covered most of the ChD during the Pleistocene was composed of pinyon pines, junipers, and oaks, which started to decrease in extension ca. 11 kya (Holocene), when the reduction in the paleolakes formed during the interglacial period

(~115-117kya) turned this corridor into a warmer and drier area, resulting in a shift to xeric vegetation (Castiglia & Fawcett, 2006; Czaja et al., 2014; Elias & Van Devender, 1990; Lanner & Van Devender, 1981; Ortega-Ramírez et al., 1998; Ramírez et al., 2003; Van Devender & Burgess, 1985).

Pinus pinceana is an endangered conifer, considered a paleorelict (Perry, 1991; SEMARNAT 059, 2010) that inhabits rocky soils with extreme aridity (Passini, 1985). It is locally restricted to the slopes of ChD in the western slopes of the SMO (1480–3000 ma.s.l.; Farjon et al., 1997) with a mean precipitation of 300–400 mm over the summer (Perry, 1991). The dominant trees in these communities are *Pinus cembroides, Juniperus* spp., and *Yucca* spp. Morphological differences, like differential needle wax cover, have been reported across the distribution of *P. pinceana* (Martiñón-Martínez et al., 2010; Ortiz-Medrano et al., 2016). Previous population genetic studies have shown that *P. pinceana* has a high genetic diversity within and high differentiation among populations (Escalante, 2001; Ledig et al., 2001; Molina-Freaner et al., 2001).

In this study, we aim to identify how phylogeography, historical demography, and the climatic transformation during the Quaternary influenced the current genetic variation and geographic distribution of *P. pinceana* by addressing the following four goals: (1) characterize its genetic diversity using genome-wide single nucleotide polymorphisms (SNPs) data, (2) identify phylogeographic patterns across its natural distribution range, (3) infer its past demographic history during the Pleistocene and Holocene, and (4) reconstruct its potential distribution during the LGM to infer the principal climatic limitations of the species.

2 | METHODS

2.1 | Sampling and genetic diversity characterization

The 10 sampled localities were distributed across the entire geographic range of *P. pinceana* (Figure S1). Individuals were chosen over the whole area. Usually, these populations are formed by 50–100 trees and we tried to sample the whole ecological area. Needles from nine healthy, regularly distributed adult trees were sampled. Figure 1 shows the regionalization in biogeographic provinces following Morrone et al. (2017).

DNA was extracted using the CTAB protocol (Doyle & Doyle, 1987). Samples with a concentration over 100 ng/ml were double-digested with the restriction enzymes *Pstl/Mspl*, which were tested in silico in the genome of *Pinus lambertiana* to generate fragments along different regions across the genome. The fragments were single-end sequenced with an Illumina Hi-Seq 2500 at the Wisconsin Biotechnology Center.

The read adapters were removed using TRIMMOMATIC (v0.36; Bolger et al., 2014). Only sequences with a minimum length of 80 bp and a minimum Phred quality score of 30 were retained. The read quality was visualized with FASTQC (v0.11.7; https://www.bioin

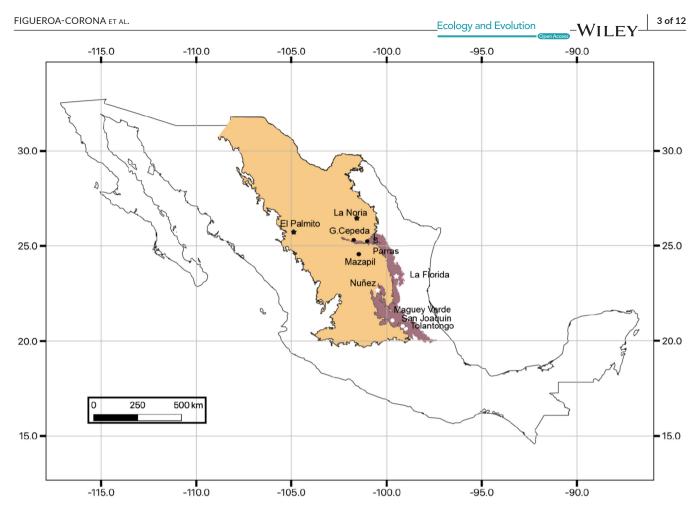


FIGURE 1 Geographic locations of the 10 populations sampled of *Pinus pinceana* analyzed genetically. The biogeographic province that encompasses the Chihuahuan Desert is located inside the sand color area, and the sampled populations from this province are indicated by a black dot; black stars correspond to Chw. The Sierra Madre Oriental is located inside the purple area, and the sampled populations from this area are shown with white dots; white stars correspond to SMOn.

formatics.babraham.ac.uk/projects/fastqc/) and MULTIQC (v1.9; Ewels et al., 2016). High-quality reads were mapped to the *P. lambertiana* genome assembly (Stevens et al., 2016; v1.5) using the BWA-MEM algorithm (v0.7.10; Li et al., 2009). Unmapped reads were discarded. The MarkDuplicates feature from the Picard package (http://broadinstitute.github.io/picard/) was used to correct artifacts of PCR duplication.

Finally, we used HaplotypeCaller and VariantFiltration tools from the Genome Analysis Toolkit (GATK v3.7; McKenna et al., 2010) to filter SNPs by quality-by-depth (QD 2.0), mapping quality (MQ 40.0), Fisher strand bias test (to determine differences in the number of sequences that support the reference and alternate alleles on each strand; FS > 60.0), rank-sum test for the evaluation of the mapping qualities supporting the reference or alternate alleles (MQRankSum <-12.5), rank sum test for assessment of bias in the position of alleles within the sequences (ReadPosRankSum <-8.0), and strand bias between forward and reverse strands by the symmetric odds ratio test (SOR > 4.0).

We obtained a set of curated SNPs that were filtered prior to analysis using VCFtools (v0.1.16; Danecek et al., 2011). The filtering process removed SNPs with differences compared with the reference but fixed them in *P. pinceana*, followed by keeping biallelic alleles with a maximum of 25% maximum missing data and individuals with <0.2 of missing data, and at least 15× of coverage.

To remove putative paralogs, we used HDplot (v0.5-7; McKinney et al., 2017) to filter polymorphisms with H (heterozygosity) larger than 0.6 and D (deviation from even read ratios in heterozygous) outside the range of -10 to +10. These parameters were chosen according to the proportion of heterozygous individuals within a population and allelic ratios within heterozygous individuals. A filter based on linkage disequilibrium (LD) was applied using PLINK v1.9 (with a window size of 50 bp, LD < 0.5, and window shift of 5; Purcell et al., 2007). An additional filter was applied to eliminate polymorphisms out of Hardy-Weinberg equilibrium (Figure S2). Based on these filtering parameters we expected to get a better representation of neutral loci, reducing the artifact incorporated by the abundance in paralogs and pseudogenes, which are abundant in conifers (Prunier, Verta, & MacKay, 2016), and giving more confidence in the accuracy of variant calls to the coalescent models. We retained a total of 3100 SNPs in 88 individuals.

We estimated the genetic diversity for the 10 populations using observed heterozygosity (H_{o}), within-population gene diversity (H_{s}),

and the inbreeding coefficient (F_{IS}) using the *R* package *Hierfstat* (Goudet, 2005). Additionally, DNAsp (v6; Rozas et al., 2017) was used to calculate allelic richness, per site nucleotide diversity (π), and pairwise population differentiation (F_{ST} ; Weir & Cockerham, 1984).

2.2 | Phylogeographic and population structure

To estimate the clustering among samples without any prior geographic provenance, we ran a discriminant analysis of principal components (DAPC) with the *r* package *Adegenet* (Jombart & Ahmed, 2011). Then, we conducted an ADMIXTURE analysis (v1.3.0; Alexander & Lange, 2011) using the genotypes that were previously converted to ordinary PLINK files (.ped) using PLINK (v1.9; Purcell et al., 2007) to estimate the population structure among individuals for *K* values from 2 to 10 admixture proportions. We selected the replicate with the lowest log-likelihood value for each value of *K*. Then, we chose the value of *K* with the lowest cross-validation error.

2.3 | Hypotheses of demographic history

The demographic history of *P. pinceana* was inferred by testing four different scenarios via the composite likelihood approach of fastsimcoal (v2.6; Excoffier & Foll, 2011; Excoffier et al., 2013), using the site (allele) frequency spectrum (SFS) of 3100 SNPs filtered without bias of frequency computed by *easySFS.py* (Figure S4; https://github. com/isaacovercast/easySFS), to infer the divergence times between and within biogeographic regions (Figure 1).

Following the clustering of DAPC, our four scenarios modeled included two scenarios without migration: Divergence, to estimate divergence times between ChD and SMO populations (T_{DIV}), and effective population sizes (N_e). Contraction, to estimate the divergence

times between the ChD (T_{DIV}) and SMO populations, followed by the independent contraction of both groups during the Holocene. We also modeled two scenarios M, including migrations, based on the former two scenarios with the same parameter ranges but adding independent migration rates among groups (Figure 2).

For each scenario, we estimated divergence times converted from generations to years using a generation time of 40 years. This generation time represents an approximation of the average age of reproductive individuals in a population. It is based on seed production and the proportion of viable seeds through the lifespan in another pinyon pines species (*Pinus nelsonii*) related to *Pinus pinceana* (Suzan-Azpiri, Sanchez-Ramos, Martínez-Avalos, Villa-Melgarejo, & Franco, 2002). The mutation rate was set at 7.28×10^{-10} per base pair per year, as reported for Pinaceae (i.e., 2.91×10^{-8} per base per generation; De la Torre et al., 2017).

The Divergence without migration model was set using search ranges for the effective population sizes (N_e) from 100 to 1000 individuals in the SMO and ChD lineages, and $T_{\rm DIV}$ between 275 and 62,500 generations ago. The Contraction model was set using search ranges for the effective population sizes (N_e) of SMO and ChD from 100 to 1000 individuals. $T_{\rm DIV}$ was set to date the divergence event within ChD and SMO between 275 and 62,500 generations ago, and $T_{\rm CON SMO}$ was set to date a contraction event in the SMO group and $T_{\rm CON ChD}$ using the range of 40 and 275 generations ago. Finally, for the two migration models, we used the same range adding between 1×10^{-9} and 1×10^{-4} , per base per generation, drawn from a log uniform prior distribution.

For each of the four models, we simulated 100 replicates, 150,000 iterations, and 30 ECM cycles. To select the best scenario, we estimated the Akaike Information Criterion AIC value (Akaike, 1974) using the maximum composite likelihood estimated from the best replicate and compared the models using the weighted Akaike information criterion (wAIC). Finally, we

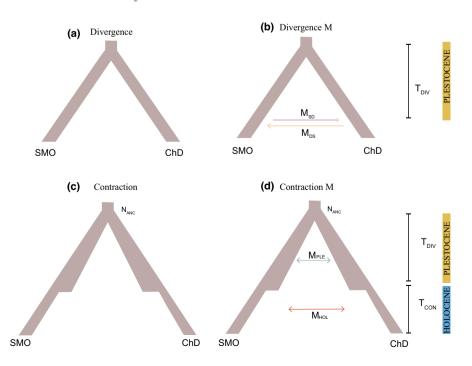


FIGURE 2 Demographic histories modeled for *Pinus pinceana*. (a) Divergence, the model draw for the divergence between SMO and ChD. (b) Divergence M includes asymmetrical and bidirectional migration among regions.

VII FY

calculated confidence intervals of parameter estimates from 100 parametric bootstrap replicates by simulating SFS from the maximum composite likelihood estimates and re-estimating the parameters each time (Excoffier et al., 2013).

2.4 | Ecological niche modeling and refugia hypotheses

We compiled 37 records verified from Escalante (2001), Figueroa-Corona (2012) and Figueroa-Corona, Delgado, Wegrzyn, & Piñero (2021) and specimen records from the National Herbarium of Mexico (MEXU; Figure S1). Each presence point was separated by at least 1 km from other presence points, to avoid spatial autocorrelation. We extracted the values for 19 bioclimatic variables with a resolution of 30 arc-seconds from CHELSEA (Krager et al., 2017) using QGIS (v3.10; https://qgis.org/es/site/) and performed an outlier analysis using Mahalanobis distances using the package JMP (v9; SAS Institute Inc., 2010).

To reduce model overfitting, we selected independent bioclimatic variables before constructing the ENMs with a Principal Component Analysis using JMP (v9; SAS Institute Inc., 2010). We retained the variables that had the higher loading values for the first three principal components (>0.7) and had a correlation coefficient of <0.8: Annual Mean Temperature (Bio1), Isothermality (Bio3), Temperature Seasonality (Bio4), Annual Precipitation (Bio12), and Precipitation in the Driest Month (Bio14) from PC1; Precipitation Seasonality (Bio15), and Precipitation of Warmest Quarter (Bio18) from PC2; and Minimum Temperature of Coldest Month (Bio6) from PC3.

To construct a potential niche model, we used Maxent (v3.3.3e; Phillips & Dudík, 2008) implemented in the suite Wallace (Kass et al., 2018; Appendix S1), testing linear, quadratic, product, and hinge features and regularization multipliers from 0.5 to 2 with 0.5 intervals. The best fitting model was selected with the Δ AlC (ENMeval implemented by Wallace). The details of the analyses can be found in the R script available in the Appendix S1. We processed the selected model (LQHP Rm = 1.5) by removing the 10th percentile training probability, as described by Liu et al. (2005).

The projection to the Last Glacial Maximum bioclimatic layers from CHELSEA on the PMIP3 data (Krager et al., 2021) was performed using the same parameters for the current distribution. Areas for each model (current and LGM) were estimated by counting pixels with the package *raster* (Hijmans & van Etten, 2012), where each pixel corresponds to approximately 1 km².

3 | RESULTS

3.1 | Genotyping and genetic characterization

The single-end sequences produced an average of 2.8 M reads per individual, which was reduced to 2.54 M reads after quality control. The datasets obtained with GATK produced 605,489 variants. VCFtools was used to obtain the final processed data with 3100 biallelic SNPs, with a moderate amount of missing data by allele (mean = 0.0912%), and a mean depth per site of 64×.

Globally, the observed heterozygosity (H_o) was 0.0663, the within-population gene diversity (H_s) was 0.0541, and the inbreeding coefficient (F_{1S}) was -0.2247 (Table 1). Levels of within-population gene diversity (H_s) were similar among populations and ranged from 0.0388 (Maguey Verde, SMO) to 0.0686 (General Cepeda; ChD). Levels of H_o were highest for the ChD location, General Cepeda (0.0824), and the lowest for Maguey Verde (0.0463) in SMO. Estimates for the inbreeding coefficient F_{1S} were negative for all populations, ranging from -0.0874 (Maguey Verde; SMO) to -0.1439 (Sierra de Parras; ChD), while the highest nucleotide diversity was found in Sierra de Parras (0.002), and the lowest in La Florida (0.0004).

Population		H _s	H _o	F _{IS}	π	Tajima D	θ_{W}
SMO	Tolantongo	0.049	0.058	-0.0881	0.0009	-0.6791 _{NS}	0.0811
	San Joaquín	0.0453	0.055	-0.1117	0.0011	-0.5995 _{NS}	0.0767
	Maguey Verde	0.0388	0.0463	-0.0874	0.0009	-0.7144 _{NS}	0.0611
	Nuñez	0.0512	0.0636	-0.1158	0.0012	-0.6266 _{NS}	0.08
	La Florida	0.0469	0.0569	-0.1073	0.0004	-0.6251 _{NS}	0.0748
ChD	El Palmito	0.0494	0.0607	-0.1023	0.0011	-0.6596 _{NS}	0.0744
	Mazapil	0.0612	0.0748	-0.1201	0.001	-0.5852 _{NS}	0.0892
	La Noria	0.0638	0.0792	-0.1328	0.0017	-0.5478 _{NS}	0.0919
	G. Cepeda	0.068	0.0848	-0.139	0.0015	-0.4724 _{NS}	0.0943
	Sierra Parras	0.0654	0.0824	-0.1439	0.002	-0.4688 _{NS}	0.096
Global		0.0541	0.0663	-0.2247	0.0006	-0.7457 _{NS}	0.147

 TABLE 1
 Measures of genetic diversity for 88 individuals of P. pinceana from ten populations calculated from 3100 single nucleotide polymorphism loci

Abbreviations: H_0 , observed heterozygosity; H_E , expected heterozygosity within populations; F_{IS} , inbreeding coefficient; π , nucleotide diversity.

3.2 | Genetic structure and phylogeography pattern

The global F_{ST} value was 0.097 (Table 2). Within SMO, the populations had a mean $F_{ST} = 0.0144$, compared to 0.012 within the ChD populations. Between biogeographic regions, the mean value was 0.0536.

The discriminant analysis of principal components (DAPC) showed two clear population clusters divided by PC1, describing 17.14% of the total variance while PC2 describes 4.3% of the total variance. PC2 helps to divide the SMO cluster with one subcluster including Tolantongo, San Joaquín, and Maguey Verde and another including La Florida and Nuñez (Figure 3a).

The ADMIXTURE analysis showed that K = 2 supports the clustering in two biogeographic areas. For K = 2, 42 of the 45 samples from SMO were classified 100% in one cluster (Figure 3b). Forty-three of the 45 samples from ChD were classified in the first cluster, while two samples from the northwest population of El Palmito were found with a level of admixture between both groups. The higher values of *K* tested (3–9) showed sub fragmentation within these two biogeographic areas but with less likelihood.

3.3 | Demographic history

We used the entire 3100 SNP dataset to model the population divergence time hypotheses from the demographic simulations. Three of the four scenarios resulted in a divergence time estimated during the Early Pleistocene while the Contraction M scenario estimated the divergence between clusters during the Miocene (Tables S1 and S2).

The best-supported model was the divergence with migration according to the Δ AIC that considers the split into two lineages according to the biogeographical regions (Figure 4; Table S3; Appendix S1). The dated divergence (T_{DIV}) between them was estimated in the Early Pleistocene (2.49 Ma; 95% CI: 3.28–1.62). Effective population sizes (Ne) for groups were similar between groups, SMO with 4367 individuals (95% CI: 4958.59–3775.41), and ChD with 6740 individuals (95% CI: 7531.42–5948.58). Migration rates per generation (*m*) were estimated to be significantly lower 3.93×10⁻⁶ (95% CI: $1.364 \times 10^{-5} - 5.78 \times 10^{-6}$) from SMO to ChD, than in the opposite direction (3×10^{-5} ; 95% CI: $3.86 \times 10^{-5} - 3.8 \times 10^{-5}$).

3.4 | Historical distribution projections

The LGM model showed a reduced suitable area compared with the current model (Figure 5), centered in the northern part of the Sierra Madre Oriental and the Northern Plateau. The LGM covered an area 56% smaller than the current distribution (80,445 pixels in LGM vs. 182,294 pixels for current prediction). There was no suitable area predicted for the Sierra Madre Occidental foothills and the southern part of the Sierra Madre Oriental. Flatlands and basins seem to be a barrier to the distribution of *P. pinceana*, as the areas identified as current or past water bodies have not been occupied by the species. The areas that remained stable, that is, with suitable conditions for *P. pinceana* since the LGM include the current populations of General Cepeda and Mazapil, covering an area of 55,582 pixels (1 pixel is around 1 km²).

4 | DISCUSSION

4.1 | Phylogeography

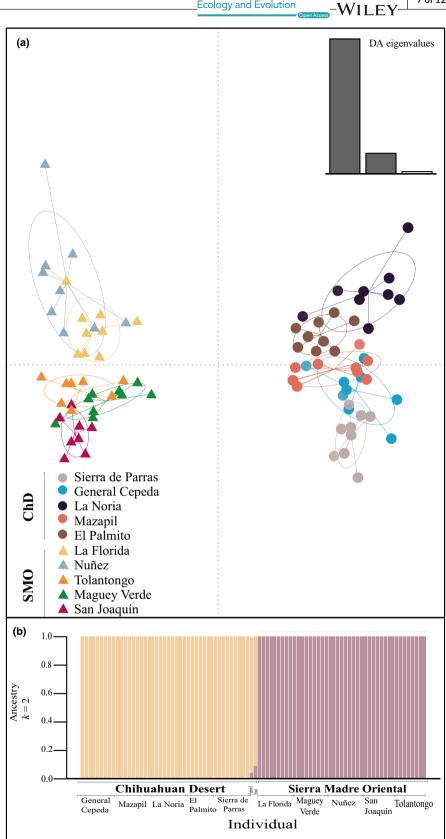
We detected a clear geographic structure of genetic diversity of *P. pinceana* between the Chihuahuan Desert and the Sierra Madre Oriental. A similar differentiation has been described in several desert scrub plant species in this area, as we discuss below. This structure is apparently related to climate dynamics during the Quaternary (Castiglia & Fawcett, 2006; Czaja et al., 2014; Ramírez et al., 2003). These changes highlight the importance of two distinct processes in the Chihuahuan Desert: first, genetic drift due to reductions in effective population sizes in different areas; and second, isolation of populations due to low migration rates.

The genetic diversity $(H_{\rm O})$ patterns using SNPs were similar to the values described in conifer species with broad geographic ranges in Mexico, like *Pinus patula*, *Abies religiosa*, and *Juniperus blancoi* (Giles-Pérez et al., 2022; Peláez et al., 2020; Reyes, 2016). By contrast, the genetic diversity values previously reported for

TABLE 2 Pairwise differentiation (F_{ST}) among the 10 populations sampled of P. pinceana with 3100 SNPs

	Tolantongo									
SMO	San Joaquín	0.0078								
	Maguey Verde	0.0099	0.01							
	Nuñez	0.01	0.0185	0.02						
	La Florida	0.0121	0.021	0.0214	0.0128					
ChD	El Palmito	0.0439	0.0471	0.0445	0.0473	0.0453				
	Mazapil	0.0469	0.0494	0.0526	0.0546	0.0536	0.0162			
	La Noria	0.0482	0.051	0.0541	0.0532	0.0525	0.0124	0.01		
	G. Cepeda	0.0536	0.0575	0.0596	0.061	0.0589	0.0184	0.008	0.008	
	Sierra Parras	0.0574	0.0577	0.0626	0.065	0.0626	0.0219	0.008	0.013	0.005

FIGURE 3 Population structure and nucleotide diversity based on 3100 genome-wide SNPs in Pinus pinceana. (a) Scatterplots resulting from discriminant analysis of principal components (DAPC) for 88 samples, and the eigenvalues of discriminant functions portrayed in the bottom right. (b) Admixture proportions of all the individuals for ancestral populations (k) for K = 2. Each bar represents one individual and shows its admixture proportions.



microsatellite (cpDNA) SSR loci in P. pinceana (Escalante, 2001; Figueroa-Corona, 2012) were substantially higher than the values found here. This is expected because microsatellites have higher mutation rates (Hamblin et al., 2007).

In accordance with previous reports in P. pinceana (Ledig et al., 1986, 2001; Molina-Freaner et al., 2001), we found a high presence of heterozygous alleles and consequently negative F_{IS} within populations. Similar situations with high heterozygosities

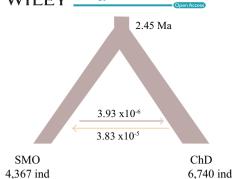


FIGURE 4 Divergence with migration hypothesis selected as the best-fit model along comparisons with resulting values for each parameter. Parameter values for this and additional models are provided in Tables S1 and S2.

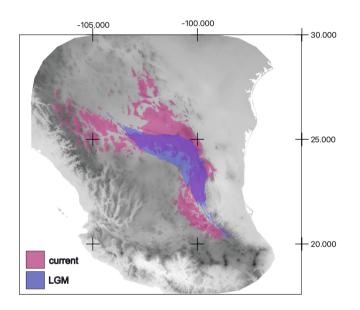


FIGURE 5 Ecological niche modeling predictions of *Pinus pinceana* current climate change during the last interglacial (LIG), and present.

and in some cases negative F_{IS} averages have been found in some conifers like *Cryptomeria japonica* var. *sinensis*, and *Pinus patula* (Cai et al., 2020; Peláez et al., 2020), while in *P. albicaulis*, the estimates have been positive (Liu et al., 2016).

We attribute these negative F_{IS} values to two possibilities, first the presence of paralogous SNPs that were not filtered with the approach of McKinney et al. (2017), as has been suggested in view of the complexities of conifer genomes (De la Torre et al., 2014; Leitch & Leitch 2012). The second possibility is the existence of natural selection as described in *P. patula* by Peláez et al. (2020). Disentangling these possibilities would require a different experimental design to analyze parental heritance or survival of seedings, in addition to better recognition of the organization of the *Pinus pinceana* genome.

The differentiation of the SMO and ChD was attributed to isolation by distance in a previous study based on cpDNA SSR loci (Escalante, 2001). Our results of genetic structure clearly identified two lineages, corresponding to the same two biogeographic regions. Furthermore, these lineages present relatively high levels of genetic differentiation between them ($F_{ST} = 0.097$), like that typically reported for conifer taxa with wide distribution ranges (Cobo-Simón et al., 2020; Jaramillo-Correa et al., 2020; Peláez et al., 2020). Nevertheless, the discriminant components analysis shows that SMO and ChD groups have significant levels of admixture between them.

The structured phylogeographic pattern between the ChD and SMO that we detected has been reported in several desert scrub plant species from the ChD (e.g., Sosa et al., 2009; Scheinvar et al., 2017; Vásquez-Cruz & Sosa, 2016). In all cases, a strong phylogeographic structure found within the SMO was correlated with geologic and paleoclimatic changes. As an example of this pattern, Vásquez-Cruz & Sosa (2016) identified a main phylogeographic pattern in species of Rosaceae where the ChD was the ancestral area, followed by contraction of suitable habitat during the Last Interglacial (~120,000-140,000 years), then an expansion during the LGM (~22,000 years) to the SMO and the Central Mexican Plateau and finally recolonization of the ChD during the Mid-Holocene (~6000 years). Loera et al. (2017) argued that this structured pattern between SMO and ChD in Ephedra compacta was reinforced by the lack of dispersal and changes in elevation due to biogeographic barriers within this region.

4.2 | Demographic history

With a patchy geographic distribution, *P. pinceana* is not considered an endangered species by the IUCN, but it is considered endangered by Mexican law based on more detailed ecological, geographic, and genetic information (SEMARNAT 059, 2010). Demographic data can help us to apply historical context to this question. The current genetic differentiation patterns of the ChD of the xeric scrubs *Ephedra compacta* and *Agave lechuguilla* (Loera et al., 2017; Scheinvar et al., 2017) can be explained by an initial expansion, followed by processes of isolation between lineages. In the case of *P. pinceana* the isolation is related to the present very reduced zones of secondary contact, and limited migration evidencing the biotic transformation during the Quaternary (Elias & Van Devender, 1990;Lanner & Van Devender, 1981; Van Devender & Burgess, 1985).

The results of the four scenarios that we analyzed give support consistent with changes in climate and the genetic structure found in *P. pinceana* in the ChD during the Middle Pleistocene. We suggest that this allopatric fragmentation of two lineages, together with an expansion of the xeric scrubs and pine-oak forests after the end of the glacial inter-cycles, was crucial in shaping the present distribution of the genetic diversity in *P. pinceana*.

Quaternary dynamics in the ChD have been described with geological and biogeographic evidence in the xeric scrub and nonarboreal species in the region. These results agree with the pattern of the geographic distribution of genetic diversity found in this study. In particular, for *Agave lechugilla*, Scheinvar et al. (2020) described contrasting levels of genetic diversity between the northern and

WILEY

southern parts of the ChD. On the other hand, genetic structure and demographic history during the Pleistocene were found in different species with a diversity of life history traits, reproductive strategies, and demographic dynamics, for example, in xeric scrubs *Agave lechuguilla*, *A. striata*, and *A. stricta* (Martínez-Ainsworth, 2013; Scheinvar et al., 2017; Trejo et al., 2016), the rodents *Thomomys umbrinus* and *Perognathus avus*, and the turtle *Kinosternon avescens* (Mathis et al., 2014; Neiswenter & Riddle, 2010; Serb et al., 2001). The estimated migration rates in our study show an asymmetric flux between regions (Table S2). Nevertheless, we still cannot establish a cause for the percentage of admixture in individuals from El Palmito given the low connectivity with nearby populations that was suggested by the ADMIXTURE analysis (Figure 3).

4.3 | Genetic diversity in a temporal context

Phylogenetic divergence within *Pinus* subsection *Cembroides* is estimated to have occurred during the Miocene (11 Mya; Gernandt et al., 2008; Saladin et al., 2017) or the Oligocene (Jin et al., 2021). The fossil evidence for pinyon pines records large changes during the Pliocene in their distribution throughout the Central Mexican Plateau (Lanner & Van Devender, 1981; Van Devender & Burgess, 1985). Conifers had a wider distribution during the Pleistocene in Mexico in the ChD, and a complex dynamic around the Holocene that diminished or extremely reduced their populations (Jaramillo-Correa et al., 2006; Quiñones-Pérez et al., 2017).

Morpho-physiological studies in *P. pinceana* detected morphological differentiation patterns, where the individuals from the ChD had thicker epicuticular waxes on the needles, faster growth, larger primary roots, and greater volume of secondary roots in Chihuahuan Desert individuals than in individuals from the Sierra Madre Oriental, differences that suggest local morphological adaptation to aridity exposure (Córdoba-Rodríguez et al., 2011; Martiñón-Martínez et al., 2010). We could not ascribe any adaptive importance to the differentiation of the genetic clusters, probably because of the absence of candidate genes adapted to climatic factors across regions. An independent analysis using comparative transcriptomics for this species in both biogeographic regions detected a possible plastic response to biotic and abiotic factors present in the populations (Figueroa-Corona et al., 2021).

4.4 | Historical distribution

Based on our analyses, we propose that *P. pinceana* differentiated into two lineages in the Middle Pleistocene (~2.49 Ma; 95% Cl: 3.28–1.62), followed by the division of both biogeographic regions (~127.7–~539.2 kya) during the interglacial cycles. The climatic conditions in the habitat of *P. pinceana* are more suitable at present than they were during the LGM; this is in contrast to what has been described in other conifers like *Picea chihuahuana*, which has become more restricted due to the disappearance of a suitable habitat

making it susceptible to the loss of genetic variability via genetic drift, inbreeding depression and strong selection (Jaramillo-Correa et al., 2006; Quiñonez-Perez et al., 2017).

The recent climatic changes in the ChD, the increment in aridification, and the reduction in water bodies (Czaja et al., 2014) promoted the geographic expansion of *P. pinceana*, and the phylogeographic patterns inferred from our data, involving a gradual expansion over the ChD since the LGM, in particular to the southern and western part of the distribution. This pattern of expansion has been described in the ChD in other scrub plant species like *A. lechuguilla* and *E. compacta* (Loera et al., 2012; Scheinvar et al., 2017). The lack of compatible environmental layers to other points in the past prevents us from exploring the distribution closer to the divergence of the two lineages.

Mazapil and General Cepeda are two of the three most genetically diverse populations but also represent, according to the estimation of the habitat, the most stable region over the climate dynamics since the interglacial cycles. Thus we interpreted that this region was a refuge for *P. pinceana*. Indeed this region has not been previously described as a refuge for the ChD (Gámez et al., 2017; Loera et al., 2017; Scheinvar et al., 2017, 2020; Vásquez-Cruz & Sosa, 2020).

5 | CONCLUSIONS

The genetic diversity in *P. pinceana* was modified by dynamics during interglacial cycles in the Pleistocene. The demographic scenarios studied resulted in a ranking of models that were useful in gauging relative support for competing hypotheses.

In particular, the best model involved the divergence into two lineages in the Middle Pleistocene (~2.49 Ma; 95% CI: 3.28–1.62). These lineages later colonized two biogeographic regions, the SMO and the ChD, while interglacial cycles modified the Chihuahuan Desert as the aridification increased and paleolakes shrank. This division probably occurred during the climatic changes of the Pleistocene and was related to the glacial-interglacial cycles.

Thus, the phylogeographic history of *P. pinceana* is likely explained by climate dynamics that left perceptible marks in the patterns of genetic diversity and structure observed in the species today.

AUTHOR CONTRIBUTIONS

Laura Figueroa-Corona: Data curation (lead); formal analysis (lead); investigation (lead); methodology (equal); resources (equal); software (equal); visualization (equal); writing – original draft (equal); writing – review and editing (equal). Alejandra Moreno-Letelier: Formal analysis (equal); methodology (equal); visualization (equal); writing – review and editing (equal). Ortega-Del Vecchyo Diego: Data curation (equal); formal analysis (equal); methodology (equal). Pablo Pelaez: Data curation (equal); formal analysis (equal); software (equal). David S. Gernandt: Formal analysis (equal); funding acquisition (equal); investigation (equal); writing – original draft (equal); I FV_Ecology and Evolution

writing – review and editing (equal). Luis E. Eguiarte: Formal analysis (equal); investigation (equal); supervision (equal); writing – review and editing (equal). Jill L. Wegrzyn: Formal analysis (equal); methodology (equal); software (equal); writing – review and editing (equal). Daniel Piñero: Conceptualization (equal); data curation (equal); formal analysis (supporting); funding acquisition (lead); investigation (equal); project administration (lead); resources (equal); supervision (lead); writing – original draft (lead); writing – review and editing (lead).

ACKNOWLEDGMENTS

To the memory of Dr. Miguel Angel Soto Arenas, in recognition for the discovery of El Palmito locality. We particularly thank Velia and Miriam Soto Arenas for showing us the population in 2009. We also thank Carmen Bautista, Patricia Delgado, Celestino Flores, Socorro González, Azalea Guerra, Ulises Manzanilla, Rosa Maricel Portilla, Rodolfo Salas-Lizana, Alejandra Vázquez-Lobo, Alfredo Villarruel, and Ana Wegier for fieldwork assistance, and Sumaira Zaman for bioinformatic assistance in variant calling. We also acknowledge the Institute for Systems Genomics Computational Biology Core, UCONN, for providing access to software and hardware. Fieldwork funding was provided by PAPIIT-DGAPA, UNAM Grant IN209816 to D.S.G., and sequencing funding was provided by IE (UNAM) to D.P. D.O.D. is supported by PAPIIT-UNAM IA200620 and a UC-MEXUS CONACYT fellowship (CN-19-29). This work was supported by Consejo Nacional de Ciencia y Tecnología through PhD Scholarship 449615 and constitutes a partial fulfillment of the Ph.D. degree (Posgrado de Ciencias Biológicas, Universidad Nacional Autónoma de México, UNAM) to L.F.C.

CONFLICT OF INTEREST

All authors contributed to the manuscript writing and revisions, and declared no conflict of interest.

OPEN RESEARCH BADGES

0 😳

This article has earned Open Data and Open Materials badges. Data and materials are available at https://www.ncbi.nlm.nih.gov/biopr oject/PRJNA719106/.

DATA AVAILABILITY STATEMENT

Raw reads are available in the SRA NCBI database (https://www.ncbi. nlm.nih.gov/sra/) deposited under the PRJNA719106 BioProject.

ORCID

Laura Figueroa-Corona [®] https://orcid.org/0000-0003-2872-5428 Alejandra Moreno-Letelier [®] https://orcid. org/0000-0001-7524-7639 Diego Ortega-Del Vecchyo [®] https://orcid. org/0000-0003-4054-3766 Pablo Peláez [®] https://orcid.org/0000-0001-7270-5155 David S. Gernandt [®] https://orcid.org/0000-0002-3592-994X Luis E. Eguiarte b https://orcid.org/0000-0002-5906-9737 Jill Wegrzyn b https://orcid.org/0000-0001-5923-0888 Daniel Piñero b https://orcid.org/0000-0002-2509-2445

REFERENCES

- Akaike, H. (1974). A new look at the statistical model identification. *IEEE Transactions on Automatic Control*, *19*, 716–723.
- Alexander, D. H., & Lange, K. (2011). Enhancements to the ADMIXTURE algorithm for individual ancestry estimation. BMC Bioinformatics, 12, 246.
- Beck, R. F., & Gibbens, R. P. (1999). The Chihuahuan desert ecosystem. New Mexico Journal of Science, 45.
- Bolger, A. M., Lohse, M., & Usadel, B. (2014). Trimmomatic: A flexible trimmer for Illumina sequence data. *Bioinformatics*, 15, 2114–2120.
- Córdoba-Rodríguez, D., Vargas-Hernández, J. J., López-Upton, J., & Muñoz-Orozco, A. (2011). Crecimiento de la raíz en plantas jóvenes de *Pinus pinceana* Gordon en respuesta a la humedad del suelo. *Agrociencia*, 45, 493–506.
- Cai, M., Wen, Y., Uchiyama, K., Onuma, Y., & Tsumura, Y. (2020). Population genetic diversity and structure of ancient tree populations of *Cryptomeria japonica* var. *sinensis* Based on RAD-seq data. *Forests*, 11(11), 1192.
- Castiglia, P., & Fawcett, J. (2006). Large Holocene lakes and climate change in the Chihuahuan Desert. *Geology*, *34*, 113–116.
- Cobo-Simón, I., Méndez-Cea, B., Jump, A. S., Seco, J., Gallego, F. J., & Linares, J. C. (2020). Understanding genetic diversity of relict forests. Linking long-term isolation legacies and current habitat fragmentation in Abies pinsapo Boiss. Forest Ecology Managment, 461, 117947.
- Czaja, A., Palacios-Fest, M. R., Estrada-Rodríguez, J. L., Romero Méndez, U., & Alba Ávila, J. A. (2014). Inland dunes fauna and flora from Paleolake Irritila in the Comarca Lagunera, Coahuila, northern Mexico. Boletin de la Sociedad Geológica Mexicana, 66, 541–551.
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., Handsaker, R. E., Lunter, G., Marth, G. T., Sherry, S. T., McVean, G., Durbin, R., & 1000 Genomes Project Analysis Group. (2011). The variant call format and VCFtools. *Bioinformatics*, 27, 2156–2158.
- De la Torre, A. R., Birol, I., Bousquet, J., Ingvarsson, P. K., Jansson, S., Jones, S. J., Keeling, C. I., MacKay, J., Nilsson, O., Ritland, K., & Street, N. (2014). Insights into conifer giga-genomes. *Plant Physiology*, *166*, 1724–1732.
- De la Torre, A. R., Li, Z., Van de Peer, Y., & Ingvarsson, P. K. (2017). Contrasting rates of molecular evolution and patterns of selection among gymnosperms and flowering plants. *Molecular Biology and Evolution*, 34, 1363–1377.
- Doyle, J. J., & Doyle, J. L. (1987). A rapid DNA isolation procedure from small quantities of fresh leaf tissue. *Phytochemistry Bulletin*, 19, 11–15.
- Eguiluz de Antuñano, S., Aranda-García, M., & Marrett, R. (2000). Tectónica de la Sierra Madre Oriental, México. *Boletín de la Sociedad Geológica Mexicana*, 53, 1–26.
- Elias, S. A., & Van Devender, T. R. (1990). Fossil insect evidence for Late Quaternary climatic change in the Big Bend region, Chihuahuan Desert, Texas. *Quaternary Research*, *34*, 249–261.
- Escalante AE (2001) Estructura genética de poblaciones de *Pinus pinceana* usando como marcadores moleculares microsatélites de cloroplasto (cpSSR's). B. Sc. Dissertation Universidad Nacional Autonoma de Mexico.
- Ewels, P., Magnusson, M., Lundin, S., & Käller, M. (2016). MultiQC: Summarize analysis results for multiple tools and samples in a single report. *Bioinformatics*, 32, 3047–3048.
- Excoffier, L., Dupanloup, I., Huerta-Sánchez, E., Sousa, V. C., & Foll, M. (2013). Robust demographic inference from genomic and SNP data. *PLoS Genetics*, 9, e1003905.

- Excoffier, L., & Foll, M. (2011). Fastsimcoal: A continuous-time coalescent simulator of genomic diversity under arbitrarily complex evolutionary scenarios. *Bioinformatics*, *27*, 1332–1334.
- Farjon, A., de la Rosa, J. A. P., & Styles, B. T. (1997). A field guide to the pines of Mexico and Central American. Royal Botanic Gardens.
- Figueroa-Corona, L. (2012). Genética de la conservación en pinos piñoneros de las subsecciones *Cembroides* y *Nelsoniae* de México. B. Sc. Dissertation Universidad Nacional Autónoma de México, Mexico.
- Figueroa-Corona, L., Delgado, P., Wegrzyn, J., & Piñero, D. (2021). Transcriptome of weeping pinyon pine, *Pinus pinceana*, shows differences across heterogeneous habitats. *Trees*, 35, 1351–1365.
- Gámez, N., Nihei, S. S., Scheinvar, E., & Morrone, J. J. (2017). A temporally dynamic approach for cladistic biogeography and the processes underlying the biogeographic patterns of North American deserts. Journal of Zoological Systematics and Evolutionary Research, 55, 11–18.
- Gernandt, D. S., Magallón, S., Geada-López, G., Zerón Flores, O., Willyard, A., & Liston, A. (2008). Use of simultaneous analyses to guide fossil-based calibrations of *Pinaceae phylogeny*. *International Journal of Plant Sciences*, 169, 1086–1099.
- Giles-Pérez, G. I., Aguirre-Planter, E., Eguiarte, L. E., & Jaramillo-Correa, J. P. (2022). Demographic modelling helps track the rapid and recent divergence of a conifer species pair from Central Mexico. *Molecular Ecology*, 1–15.
- Goudet, J. (2005). Hierfstat, a package for *R* to compute and test hierarchical F-statistics. *Molecular Ecology Notes*, 5, 184–186.
- Hamblin, M. T., Warburton, M. L., & Buckler, E. S. (2007). Empirical comparison of simple sequence repeats and single nucleotide polymorphisms in assessment of maize diversity and relatedness. *PLoS One*, 2, e1367.
- Hernandez-Cerda, M., & Carrasco-Anaya, G. (2004). Climatología. In I. Luna, J. J. Morrone, & D. Espinosa (Eds.), *Biodiversidad de La Sierra Madre Oriental. Prensas de Ciencias*. Facultad de Ciencias, UNAM.
- Hijmans, R., & van Etten, J. (2012). Geographic analysis and modeling with raster data. R Package version. 2. 1–25.
- Jaramillo-Correa, J. P., Bagnoli, F., Grivet, D., Fady, B., Aravanopoulos, F. A., Vendramin, G. G., & González-Martínez, S. C. (2020). Evolutionary rate and genetic load in an emblematic Mediterranean tree following an ancient and prolonged population collapse. *Molecular Ecology*, 29, 4797–4811.
- Jaramillo-Correa, J. P., Beaulieu, J., Ledig, F. T., & Bousquet, J. (2006). Decoupled mitochondrial and chloroplast DNA population structure reveals Holocene collapse and population isolation in a threatened Mexican-endemic conifer. *Molecular Ecology*, 15, 2787–2800.
- Jin, W. T., Gernandt, D. S., Wehenkel, C., Xia, X. M., Wei, X. X., & Wang, X. Q. (2021). Phylogenomic and ecological analyses reveal the spatiotemporal evolution of global pines. Proceedings of the National Academy of Sciences of the United States of America, 118, e2022302118.
- Jombart, T., & Ahmed, I. (2011). Adegenet 1.3-1: New tools for the analysis of genome-wide SNP data. *Bioinformatics*, *27*, 3070–3071.
- Karger, D. N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza,
 R. W., Zimmermann, N. E., Linder, H. P., & Kessler, M. (2017).
 Climatologies at high resolution for the earth's land surface areas.
 Scientific Data, 4, 170122.
- Karger, D. N., Wilson, A. M., Mahony, C., Zimmermann, N. E., & Jetz, W. (2021). Global daily 1 km land surface precipitation based on cloud cover-informed downscaling. *Scientific Data*, 8, 307.
- Kass, J. M., Aiello-Lammens, M. E., Muscarella, R., Merow, C., & Anderson, R. P. (2018). Wallace: A flexible platform for reproducible modeling of species niches and distributions built for community expansion. *Methods in Ecology and Evolution*, 9, 1151–1156.
- Lanner, R. M., & Van Devender, T. R. (1981). Late Pleistocene piñon pines in the Chihuahuan Desert. *Quaternary Research*, 15, 278–290.

- Ledig, F. T. (1986). Conservation strategies for forest gene resources. Forest Ecology and Managenent, 14, 77–90.
- Ledig, F. T., Capó-Arteaga, M. A., Hodgskiss, P. D., Sbay, H., Flores-López, C., Thompson Conkle, M., & Bermejo-Velázquez, B. (2001). Genetic diversity and the mating system of a rare Mexican pinon, *Pinus pinceana*, and a comparison with *Pinus maximartinezii* (Pinaceae). *American Journal of Botany*, 88, 1977–1987.
- Leitch, A. R., & Leitch, I. J. (2012). Ecological and genetic factors linked to contrasting genome dynamics in seed plants. *New Phytologist*, 194, 629–646.
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., Durbin, R., & 1000 Genome Project Data Processing Subgroup. (2009). The sequence alignment/map format and SAMtools. *Bioinformatics*, *25*, 2078–2079.
- Liu, J. J., Sniezko, R., Murray, M., Wang, N., Chen, H., Zamany, A., Sturrock, R. N., Savin, D., & Kegley, A. (2016). Genetic diversity and population structure of whitebark pine (*Pinus albicaulis* Engelm.) in Western North America. *PLoS One*, 11, e0167986.
- Liu, C., Berry, P. M., Dawson, T. P., & Pearson, R. G. (2005). Selecting thresholds of occurrence in the prediction of species distributions. *Ecography*, 28(3), 385-393. https://doi. org/10.1111/j.0906-7590.2005.03957.x
- Loera, I., Ickert-Bond, S. M., & Sosa, V. (2017). Pleistocene refugia in the Chihuahuan Desert: The phylogeographic and demographic history of the gymnosperm *Ephedra compacta. Journal of Biogeography*, 44, 2706–2716.
- Loera, I., Sosa, V., & Ickert-Bond, S. M. (2012). Diversification in north American arid lands: Niche conservatism, divergence and expansion of habitat explain speciation in the genus *Ephedra*. *Molecular Phylogenetics and Evolution*, 65, 437–450.
- Martínez-Ainsworth, N. (2013). Genética de poblaciones de Agave stricta Salm-Dyck, especie endémica al Valle de Tehuacán-Cuicatlán. México. B. Sc. Dissertation Universidad Nacional Autónoma de México, Mexico.
- Martiñón-Martínez, R. J., Vargas-Hernández, J., López-Upton, J., Gómez-Guerrero, A., & Vaquera-Huerta, H. (2010). Respuesta de *Pinus pinceana* Gordon a estrés por sequía y altas temperaturas. Revista *Fitotecnia Mexicana*, 33, 239–248.
- Mathis, V. L., Hafner, M. S., & Hafner, D. J. (2014). Evolution and phylogeography of the *Thomomys umbrinus* species complex (Rodentia: Geomyidae). Journal of Mammalogy, 95, 754–771.
- McKenna, A., Hanna, M., Banks, E., Sivachenko, A., Cibulskis, K., Kernytsky, A., Garimella, K., Altshuler, D., Gabriel, S., Daly, M., & DePristo, M. A. (2010). The Genome Analysis Toolkit: A MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Research*, 20, 1297–1303.
- McKinney, G. J., Waples, R. K., Seeb, L. W., & Seeb, J. E. (2017). Paralogs are revealed by proportion of heterozygotes and deviations in read ratios in genotyping-by-sequencing data from natural populations. *Molecular Ecology Resources*, 17, 656–669.
- Metcalfe, S. E. (2006). Late quaternary environments of the northern deserts and central transvolcanic belt of Mexico. *Annals of the Missouri Botanical Garden*, 93, 258–273.
- Molina-Freaner, F., Delgado, P., Piñero, D., Perez-Nasser, N., & Alvarez-Buylla, E. (2001). Do rare pines need different conservation strategies? Evidence from three Mexican species. *Canadian Journal of Botany*, 79, 131–138.
- Morrone, J. J., Escalante, T., & Rodríguez-Tapia, G. (2017). Mexican biogeographic provinces: Map and shapefiles. *Zootaxa*, 4277, 277–279.
- Neiswenter, S. A., & Riddle, B. R. (2010). Diversification of the *Perognathus* avus species group in emerging arid grasslands of western North America. *Journal Mammalogy*, *91*, 348–362.
- Noy-Meir, I. (1973). Desert ecosystems: Environment and producers. Annual Review of Ecology and Systematics, 4, 25–51.
- Ortega-Ramírez, J., Valiente-Banuet, A., Urrutia-Fucugauchi, J., Mortera-Gutiérrez, C. A., & Alvarado-Valdez, G. (1998). Paleoclimatic

changes during the Late Pleistocene - Holocene in Laguna Babícora, near the Chihuahuan Desert, México. *Canadian Journal of Earth Sciences*, 35, 1168–1179.

- Ortiz-Medrano, A., Scantlebury, D. P., Vázquez-Lobo, A., Mastretta-Yanes, A., & Piñero, D. (2016). Morphological and niche divergence of pinyon pines. *Ecology and Evolution*, 6, 2886–2896.
- Passini, M. (1985). Les forêts de Pinus cembroides Zucc. de la Sierra de Urica, réserve de la biosphère «La Michilia» (Etat de Durango, Mexique). I: Présentation générale.
- Peláez, P., Ortiz-Martínez, A., Figueroa-Corona, L., Montes, J. R., & Gernandt, D. S. (2020). Population structure, diversifying selection, and local adaptation in *Pinus patula*. *American Journal of Botany*, 107, 1555–1566.
- Perry, J. P. (1991). The pines of Mexico and central America (No. 04; QK494. 5. P66, P4.). Timber Press.
- Phillips, S. J., & Dudík, M. (2008). Modeling of species distributions with Maxent: New extensions and a comprehensive evaluation. *Ecography*, 31, 161–175.
- Prunier, J., Verta, J. P., & MacKay, J. J. (2016). Conifer genomics and adaptation: At the crossroads of genetic diversity and genome function. *The New Phytologist*, 209, 44–62.
- Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M. A. R., Bender, D., Maller, J., Sklar, P., de Bakker, P. I. W., Daly, M. J., & Sham, P. C. (2007). PLINK: A tool set for whole-genome association and population-based linkage analyses. *American Journal Human Genetics*, 81, 559–575.
- Quiñones-Pérez, C. Z., González-Elizondo, M., & Wehenkel, C. (2017). Ruling out genetic erosion in *Picea chihuahuana* Martínez. *New Forests*, 48, 201–215.
- Ramírez, J. O., Maillol, J. M., Bandy, W., Banuet, A. V., Gutiérrez, C. M., Sánchez, J. M., & Cruz, G. C. (2003). Late Quaternary evolution of alluvial fans in the Playa, El Fresnal region, northern Chihuahua desert, Palaeoclimatic implications. *Geofísica Internacional*, 43(3), 445–466.
- Reyes, G.V. (2016). Genómica de poblaciones de *Juniperus blancoi* y J. *monticola*: Diversidad genética y altitudinal. B. Sc. Dissertation Universidad Nacional Autónoma de México, Mexico.
- Rozas, J., Ferrer-Mata, A., Sánchez-Del Barrio, J. C., Guirao-Rico, S., Librado, P., Ramos-Onsins, S. E., & Sánchez-Gracia, A. (2017). DnaSP 6: DNA sequence polymorphism analysis of large datasets. *Molecular Biology and Evolution*, 34, 3299–3302.

Rzedowski, J. (1978). Vegetación de México. Limusa.

- Rzedowski, J., & Calderón de Rzedowski, G. (1993). Datos sobre la dinámica de la flora fanerogámica del Valle de México, con énfasis en especies narrativas raras, en peligro de extinción y aparentemente extintas. Acta Botanica Mexicana, 25, 81-108.
- Saladin, B., Leslie, A. B., Wüest, R. O., Litsios, G., Conti, E., Salamin, N., & Zimmermann, N. E. (2017). Fossils matter: Improved estimates of divergence times in *Pinus* reveal older diversification. *BMC Ecology* and Evolution, 17, 95.
- Scheinvar, E., Gámez, N., Castellanos-Morales, G., Aguirre-Planter, E., & Eguiarte, L. E. (2017). Neogene and Pleistocene history of Agave lechuguilla in the Chihuahuan Desert. Journal of Biogeography, 44, 322–334.
- Scheinvar, E., Gámez, N., Moreno-Letelier, A., Aguirre-Planter, E., & Eguiarte, L. E. (2020). Phylogeography of the Chihuahuan Desert: Diversification and evolution over the Pleistocene. In M. Mandujano, I. Pisanty, & L. Eguiarte (Eds.), Plant diversity and ecology in the Chihuahuan desert. Cuatro Ciénegas Basin: An endangered hyperdiverse oasis. Springer.
- SEMARNAT 059. (2010). Modificación a la Norma Oficial Mexicana NOM-059-ECOL-2001, Protección ambiental-Especies nativas de

México de flora y fauna silvestres- Categorías de riesgo y especificaciones para su inclusión, exclusión o cambio-Lista de especies en riesgo. *Diario Oficial de la Federación*, 6, 158–169. Segunda sección. https://www.profepa.gob.mx/innovaportal/file/435/1/nom_059_ semarnat_2010.pdf

- Serb, J. M., Phillips, C. A., & Iverson, J. B. (2001). Molecular phylogeny and biogeography of *Kinosternon avescens* based on complete mitochondrial control region sequences. *Molecular Phylogenetics and Evolution*, 18, 149–162.
- Sosa, V., Ruiz-Sanchez, E., & Rodriguez-Gomez, F. C. (2009). Hidden phylogeographic complexity in the Sierra Madre Oriental: The case of the Mexican tulip poppy *Hunnemannia fumariifolia* (Papaveraceae). *Journal of Biogeography*, 36, 18–27.
- Stevens, K. A., Wegrzyn, J. L., Zimin, A., Puiu, D., Crepeau, M., Cardeno, C., Paul, R., Gonzalez-Ibeas, D., Koriabine, M., Holtz-Morris, A. E., Martínez-García, P. J., Sezen, U. U., Marçais, G., Jermstad, K., McGuire, P. E., Loopstra, C. A., Davis, J. M., Eckert, A., de Jong, P., ... Langley, C. H. (2016). Sequence of the Sugar Pine Megagenome. *Genetics*, 204(4), 1613–1626.
- Suzán-Azpiri, H., Sanchez-Ramos, G., Martínez-Avalos, J. G., Villa-Melgarejo, S., & Franco, M. (2002). Population structure of *Pinus* nelsonii Shaw, an endemic pinyon pine in Tamaulipas, Mexico. Forest Ecology and Management, 165, 193–120.
- Trejo, L., Alvarado-Cárdenas, L. O., Scheinvar, E., & Eguiarte, L. E. (2016). Population genetic analysis and bioclimatic modeling in Agave striata in the Chihuahuan Desert indicate higher genetic variation and lower differentiation in drier and more variable environments. American Journal of Botany, 103, 1020–1029.
- Van Devender, T. R., & Burgess, T. L. (1985). Late Pleistocene woodlands in the Bolson de Mapimi: A refugium for the Chihuahuan Desert Biota? *Quaternary Research*, *24*, 346–353.
- Vásquez-Cruz, M., & Sosa, V. (2016). New insights on the origin of the woody flora of the Chihuahuan Desert: The case of Lindleya. *American Journal of Botany*, 103(9), 1694–1707.
- Vásquez-Cruz, M., & Sosa, V. (2020). Assembly and origin of the flora of the Chihuahuan Desert: The case of sclerophyllous *Rosaceae*. *Journal of Biogeography*, 47, 445–459.
- Weir, B. S., & Cockerham, C. C. (1984). Estimating f-statistics for the analysis of population structure. *Evolution*, 38, 1358–1370.
- Zavala-Hurtado, J. A., & Jiménez, M. (2020). Diversity and uniqueness at its best: Vegetation of the Chihuahuan Desert. In M. Mandujano,
 I. Pisanty, & L. Eguiarte (Eds.), Plant diversity and ecology in the Chihuahuan Desert. Cuatro Ciénegas Basin: An endangered hyperdiverse oasis. Springer.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Figueroa-Corona, L., Moreno-Letelier, A., Ortega-Del Vecchyo, D., Peláez, P., Gernandt, D. S., Eguiarte, L. E., Wegrzyn, J., & Piñero, D. (2022). Changes in demography and geographic distribution in the weeping pinyon pine (*Pinus pinceana*) during the Pleistocene. *Ecology and Evolution*, *12*, e9369. https://doi.org/10.1002/ece3.9369