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VIP Synthesis, Anti-tubulin and Antiproliferative SAR of Steroidomimetic Dihydroisoquinolinones

Mathew P. Leese,^[a] Fabrice L. Jourdan,^[a] Meriel R. Major,^[a] Wolfgang Dohle,^[a] Mark P. Thomas,^[a] Ernest Hamel,^[b] Eric Ferrandis,^[c] Mary F. Mahon,^[d] Simon P. Newman,^[e] Atul Purohit,^[e] and Barry V. L. Potter^{*[a]}

A SAR translation strategy adopted for the discovery of tetrahydroisoquinolinone (THIQ)-based steroidomimetic microtubule disruptors has been extended to dihydroisoquinolinone (DHIQ)-based compounds. A steroid A,B-ring-mimicking DHIQ core was connected to methoxyaryl D-ring mimics through methylene, carbonyl, and sulfonyl linkers, and the resulting compounds were evaluated against two cancer cell lines. The carbonyl-linked DHIQs in particular exhibit significant in vitro antiproliferative activities (e.g., 6-hydroxy-7-methoxy-2-(3,4,5-trimethoxybenzoyl)-3,4-dihydroisoquinolin-1(2H)-one (**16g**); GI₅₀ 51 nM in DU-145 cells). The broad anticancer activity of DHIQ **16g** was confirmed in the NCI 60-cell line assay giving a mean activity of 33 nM. Furthermore, 6-hydroxy-2-(3,5-dimethoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one

(**16f**) and **16g** and their sulfamate derivatives **17f** and **17g** (2-(3,5-dimethoxybenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one and 7-methoxy-2-(3,4,5-trimethoxybenzoyl)-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one, respectively) show excellent activity against the polymerization of tubulin, close to that of the clinical combretastatin A-4, and bind competitively at the colchicine binding site of tubulin. Compounds **16f** and **17f** were also shown to demonstrate in vitro anti-angiogenic activity. Additionally, X-ray and computational analyses of **17f** reveal that electrostatic repulsion between the two adjacent carbonyl groups, through conformational biasing, dictates the adoption of a "steroid-like" conformation that may partially explain the excellent in vitro activities.

Introduction

In previous studies we optimised sulfamoylated estratrienes **1a–b** (Figure 1) as anticancer agents.^[1–6] These compounds exhibit antiproliferative activity against a range of human cancer cell lines and are also capable of inhibiting angiogenesis. This dual mechanism of action can be ascribed to their ability to inhibit normal microtubule dynamics and, in addition to good oral bioavailability and excellent in vivo activity, they proved

capable of inhibiting the growth of cell lines resistant to existing microtubule disruptors such as the taxanes. To develop further series of compounds that share this mechanism of action we were drawn to investigate whether, by translating key pharmacophoric elements from the steroidal series into non-steroidal motifs, we could generate new microtubule disruptors with further enhanced activity and/or physicochemical properties. In initial studies^[7,8] we used a tetrahydroisoquinoline

[a] Dr. M. P. Leese, Dr. F. L. Jourdan, Dr. M. R. Major, Dr. W. Dohle, Dr. M. P. Thomas, Prof. Dr. B. V. L. Potter
Medicinal Chemistry, Department of Pharmacy & Pharmacology
University of Bath, Bath, BA2 7AY (UK)
E-mail: B.V.L.Potter@bath.ac.uk

[b] Dr. E. Hamel
Screening Technologies Branches, National Cancer Institute
Frederick, MD 21702 (USA)

[c] Dr. E. Ferrandis
Institut de Recherche Henri Beaufour, 91966 Les Ulis Cedex (France)

[d] Dr. M. F. Mahon
X-ray Crystallographic Suite, Department of Chemistry
University of Bath, Bath, BA2 7AY (UK)

[e] Dr. S. P. Newman, Dr. A. Purohit
Oncology Drug Discovery Group, Section of Investigative Medicine
Hammersmith Hospital, Imperial College London, London, W12 0NN (UK)

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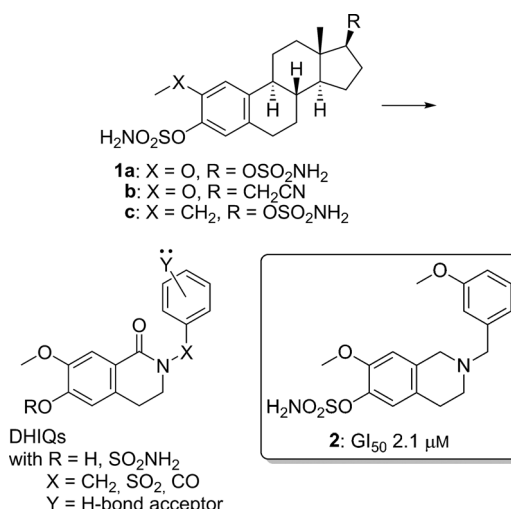


Figure 1. Design of DHIQ-based steroidomimetic microtubule disruptors.

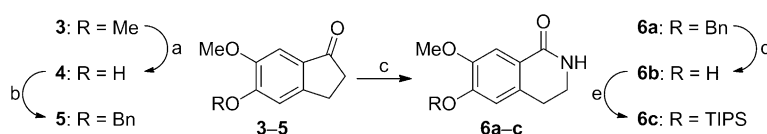
(THIQ) decorated at C6 and C7 to mimic the steroidal A,B-ring of the estratrienes tethered through N2 to a D-ring mimic, initially a benzyl group, which projects a hydrogen bond acceptor into the appropriate region of space to address the pharmacophore for antiproliferative activity in that region. This delivered a series of microtubule disruptors with antiproliferative activity in the micromolar range that could be further optimised by introducing a substituent at C3 to sterically inhibit the free rotation of the *N*-benzyl group and thus favour conformational populations in which the *N*-benzyl group mimics the steroidal D-ring. In this manner compounds displaying nanomolar activity (equivalent to that of the steroid derivatives upon which their design was based) were elaborated.^[9] In tandem, chimeric microtubule disruptors built from the THIQ core and the trimethoxy aryl motif common to many colchicine site binders were constructed.^[10,11]

In the present work we sought to develop new microtubule disruptors through integration of lessons learned in work on the estratrienes and the THIQ-based systems. By using an alternate heterocyclic motif, the dihydroisoquinolinone (DHIQ) to mimic the steroidal A,B-ring system, we envisaged that some conformational pre-organisation could be achieved through electrostatic repulsion. Tethering the D-ring-mimicking aryl group through a carbonyl linkage, repulsion of the two carbonyl groups of the imide should cause adoption of a dihedral angle in which they minimise their electrostatic clash; this should deliver a conformation in which the D-ring mimic projects into a region of space broadly analogous to a steroidal D-ring, thus decreasing rotation to achieve optimal binding. To assess this strategy a series of 2-, 3- and 4-methoxybenzoyl DHIQs was synthesised alongside corresponding *N*-benzyl and *N*-arylsulfonyl DHIQ series for which no positive conformational biasing is envisaged (Figure 1).

Results and Discussion

Chemistry

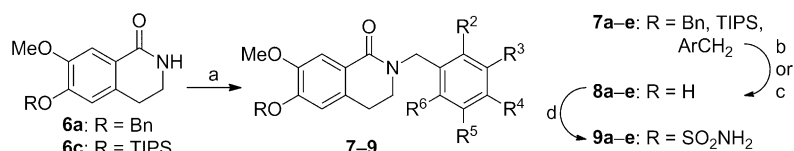
The synthetic strategy to access the dihydroisoquinolinone core structure is outlined below (Scheme 1). 5,6-Dimethoxyindan-1-one **3** was selectively demethylated in aqueous piperidine and the product subsequently reacted with benzyl bro-



Scheme 1. Synthesis of the DHIQ core structure. *Reagents and conditions:* a) piperidine/H₂O, 140 °C; b) BnBr, K₂CO₃, DMF, RT; c) NaN₃, CH₃SO₃H, CH₂Cl₂, 0 °C → RT; d) H₂, Pd/C, THF/MeOH, RT; e) TIPSCl, imidazole, CH₂Cl₂, RT.

mid to furnish compound **5** in good yield. Reaction of **5** with sodium azide and methanesulfonic acid in dichloromethane gave dihydroisoquinolinone **6a**. Some of the material was transformed into the corresponding unprotected phenol **6b**, by treatment with hydrogen and palladium on charcoal, that was subsequently protected with TIPSCl in dichloromethane to afford compound **6c** (Scheme 1).

N-Benzoylation was then performed on compound **6c**. Under the conditions applied in these reactions TIPS-deprotection and subsequent benzylation of the unprotected phenol was also observed to give compounds **7a–c**. In two further experiments the more stable benzyl-protected compound **6a** was reacted in the same manner to afford compounds **7d–e**. These were transformed into the corresponding unprotected phenols **8a–e** either by treatment with hydrogen and Pd/C or with TBAF (only for **7b**) in good yields. Treatment of **8a–e** with sulfamoyl chloride in DMA^[12] gave the corresponding sulfamates **9a–e** (Scheme 2).



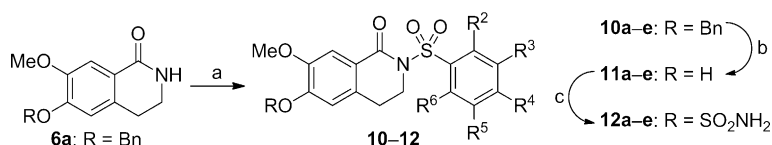
Scheme 2. Synthesis of functionalised *N*-benzyl-substituted DHIQs. *Reagents and conditions:* a) NaH, 0 °C, DMF, then benzyl halide, RT; b) H₂, Pd/C, THF/MeOH, RT; c) TBAF, THF, RT; d) H₂NSO₂Cl, DMA, RT.

A second compound set was produced by *N*-sulfonylating **6a** with various arylsulfonyl chlorides. Compounds **10a–e** were transformed into the corresponding unprotected phenols **11a–e**, by treatment with hydrogen and Pd/C, that were then reacted with sulfamoyl chloride in DMA to afford the corresponding sulfamates **12a–e** (Scheme 3).

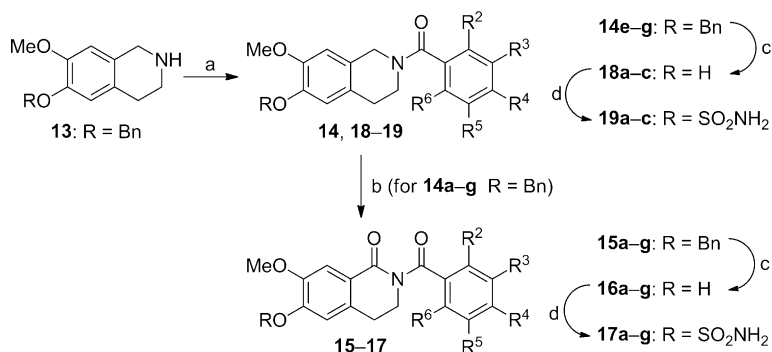
For the synthesis of *N*-benzoylated compounds a direct strategy starting from **6a** proved not very successful. Therefore, an alternate approach using benzyl-protected THIQ **13**^[8] as starting material was adopted. *N*-Benzoylation using various benzoyl chlorides followed by permanganate oxidation gave access to the desired *N*-benzoylated dihydroisoquinolines **15a–g**. These were reacted with hydrogen and Pd/C and the product treated with sulfamoyl chloride in DMA to give sulfamates **17a–g**. Additionally, a set of control compounds without the carbonyl at C1 was achieved by hydrogenation of compounds **14e–g** with Pd/C to give phenols **18a–c**. Treatment with sulfamoyl chloride in DMA then afforded sulfamates **19a–c** (Scheme 4).^[8]

Biology

The compounds were then evaluated *in vitro* for their ability to inhibit the proliferation of DU-145 human prostate cancer cells and MDA MB-231 breast cancer cells. As can be observed in



Scheme 3. Synthesis of functionalised *N*-arylsulfonyl-substituted DHIQs. Reagents and conditions: a) NaH, DMF, 50 °C, then ArSO₂Cl, DMF, RT or 40 °C; b) H₂, Pd/C, THF/EtOH, RT; c) H₂NSO₂Cl, DMA, RT.



Scheme 4. Synthesis of functionalised *N*-benzoyl-substituted DHIQs and THIQs. Reagents and conditions: a) ArCOCl, Et₃N, CH₂Cl₂, RT; b) KMnO₄, 18-crown-6, CHCl₃, RT; c) H₂, Pd/C, THF/MeOH, RT; d) H₂NSO₂Cl, DMA, RT.

Table 1, the *N*-benzyl derivatives, regardless of the nature of substitution of the benzyl motif, prove to be uniformly inactive in contrast to the corresponding THIQ series in which the 3'-methoxy compound **2** (Figure 1) exhibits a GI₅₀ value of 2.1 μM.^[7,8] The addition of a carbonyl at C1 in this case is clearly detrimental, and it seems reasonable to suggest that this group, through steric considerations, disfavours adoption of a conformation in which the benzyl group can potentially mimic the steroid D-ring. Likewise, the series of *N*-arylsulfonyl derivatives prove to be inactive. The corresponding series of *N*-arylsulfonyl THIQs also fails to display significant activity, a fact that we ascribe to the likely unacceptable steric bulk of the sulfonyl group at the site of binding.^[8] The *N*-benzoyl DHIQs, in contrast, provide interesting activity in both the 6-hydroxy and 6-sulfamoyloxy series. Here, both the 2'- and 3'-methoxy compounds give low micromolar GI₅₀ values against the proliferation of both DU-145 and MDA MB-231 cells. It is notable that the 6-hydroxy compounds **16a–b** display superior activity to the 6-*O*-sulfamates **17a–b** and that, as found for the *N*-benzyl THIQs (with which we believed they should act in an analogous manner), the 3'-substituted derivative displays the highest activity. In contrast, as found in the *N*-benzyl THIQ series, the 4'-substituted compounds **16d** and **17d** are inactive. Several polymethoxylated compounds were also evaluated revealing that, although 4'-substitution alone did not afford activity, it is tolerated as the 3',4'-dimethoxy compounds **16e** and **17e** display good activity and are more active than the 3'-methoxy derivatives **16b** and **17b**. The major step forward in activity was realised when the 3',5'-dimethoxy derivative **16f** and its sulfamate **17f** were evaluated, with roughly equivalent activity (GI₅₀ values of 215 and 179 nM for DU-145, respectively). Compounds **16f** and **17f** are >10-fold more active than the 3'-methoxy derivatives **16b** and **17b**. We postulate that

this symmetric substitution adds a further entropic advantage in combination with the pre-organisation we believed would derive from the repulsion of the carbonyls of the imide discussed above. In any case, the activity of **16f** and **17f** is equal to that of both the conformationally biased THIQs and the steroidal derivatives from which the non-steroidal design was conceived.

Finally, 3',4',5'-trimethoxybenzoyl DHIQ derivatives **15g**, **16g** and **17g** were also evaluated. From our understanding drawn from experience with the THIQ systems these compounds could well act as chimeras addressing both the steroidal A-ring binding area and the trimethoxyaryl binding area of the colchicine binding site on tubulin. Phenol **16g** and its sulfamate **17g**

prove to be the most active compounds evaluated, with **16g** displaying respective GI₅₀ values of 51 and 71 nM against the proliferation of DU-145 and MDA MB-231 cells.

Comparing these results with those for related *N*-benzoylated THIQs (Table 2) it is evident that omitting the carbonyl at C1 is in general detrimental towards in vitro antiproliferative activity. Only polymethoxylated compounds are displayed here. The series of 3',4'-dimethoxy THIQ compounds shows a different trend to the corresponding DHIQ series, because here benzyl-protected compound **14e** shows the best, but still relatively modest, activity (GI₅₀ 26.7 μM in DU-145) with phenol **18a** and sulfamate **19a** being surprisingly completely inactive. The 3',5'-dimethoxy and 3',4',5'-trimethoxy derivatives were also evaluated and show a similar trend as their DHIQ counterparts, with phenols **18b–c** showing low micromolar activity that is slightly better than that of the corresponding sulfamates **19b–c**, while benzyl-protected compounds **14f–g** display only modest activity. However, the most active compounds **18c** and **19c** are still about 10- to 20-fold less active than DHIQs **16g** and **17g**, showing the importance of the combination of two carbonyl groups (at C1 and in the linker to the D-ring mimic) to achieve compounds with low nanomolar activity.

The most active compound **16g** was also tested in the NCI 60-cell line assay (Table 3) that allows activity across a wide range of cancer types to be assessed. Data from seven cell lines are presented along with the mean activity across the whole panel (MGM value). Screening was conducted at concentrations ranging from 10 nM upwards, with maximal activity (where a 50% growth inhibition was obtained in all cell lines at a concentration of 10 nM) being indicated by an MGM value of 10 nM. The data obtained in the assay confirm the very high potency of **16g** against a broad range of cancer phenotypes.

Table 1. Antiproliferative activity of DHIQs against DU-145 human prostate cancer cells and MDA MB-231 human breast cancer cells in vitro.

Compd							GI ₅₀ [μM] ^[a]	
	R ¹	X	R ²	R ³	R ⁴	R ⁵	DU-145	MDA MB-231
8a	H	CH ₂	OMe	H	H	H	> 100	ND ^[b]
9a	SO ₂ NH ₂	CH ₂	OMe	H	H	H	> 100	ND ^[b]
8b	H	CH ₂	H	OMe	H	H	> 100	ND ^[b]
9b	SO ₂ NH ₂	CH ₂	H	OMe	H	H	> 100	ND ^[b]
8c	H	CH ₂	H	H	OMe	H	> 100	ND ^[b]
9c	SO ₂ NH ₂	CH ₂	H	H	OMe	H	> 100	ND ^[b]
7d	Bn	CH ₂	H	OMe	H	OMe	> 100	ND ^[b]
8d	H	CH ₂	H	OMe	H	OMe	> 100	ND ^[b]
9d	SO ₂ NH ₂	CH ₂	H	OMe	H	OMe	> 100	ND ^[b]
7e	Bn	CH ₂	H	OMe	OMe	OMe	> 100	ND ^[b]
8e	H	CH ₂	H	OMe	OMe	OMe	> 100	ND ^[b]
9e	SO ₂ NH ₂	CH ₂	H	OMe	OMe	OMe	> 100	ND ^[b]
10a	Bn	SO ₂	OMe	H	H	H	> 100	> 100
11a	H	SO ₂	OMe	H	H	H	> 100	13.4
12a	SO ₂ NH ₂	SO ₂	OMe	H	H	H	> 100	> 100
10b	Bn	SO ₂	H	OMe	H	H	> 100	> 100
11b	H	SO ₂	H	OMe	H	H	> 100	> 100
12b	SO ₂ NH ₂	SO ₂	H	OMe	H	H	> 100	> 100
11c	H	SO ₂	H	H	OMe	H	> 100	> 100
12c	SO ₂ NH ₂	SO ₂	H	H	OMe	H	> 100	> 100
10d	Bn	SO ₂	H	Cl	H	H	> 100	> 100
11d	H	SO ₂	H	Cl	H	H	> 100	> 100
12d	SO ₂ NH ₂	SO ₂	H	Cl	H	H	> 100	> 100
11e	H	SO ₂	CO ₂ Me	H	H	H	> 100	> 100
12e	SO ₂ NH ₂	SO ₂	CO ₂ Me	H	H	H	> 100	13
15a	Bn	CO	OMe	H	H	H	> 100	43.6
16a	H	CO	OMe	H	H	H	6.42	5.6
17a	SO ₂ NH ₂	CO	OMe	H	H	H	9.1	7.1
16b	H	CO	H	OMe	H	H	3.63	1.89
17b	SO ₂ NH ₂	CO	H	OMe	H	H	5.3	3.5
15c	Bn	CO	H	CN	H	H	> 100	> 100
16c	H	CO	H	CN	H	H	70	37.4
17c	SO ₂ NH ₂	CO	H	CN	H	H	25	> 100
15d	Bn	CO	H	H	OMe	H	> 100	ND ^[b]
16d	H	CO	H	H	OMe	H	> 100	ND ^[b]
17d	SO ₂ NH ₂	CO	H	H	OMe	H	> 100	ND ^[b]
15e	Bn	CO	H	OMe	OMe	H	> 100	ND ^[b]
16e	H	CO	H	OMe	OMe	H	2.39	1.6
17e	SO ₂ NH ₂	CO	H	OMe	OMe	H	3.4	2.08
15f	Bn	CO	H	OMe	H	OMe	> 100	ND ^[b]
16f	H	CO	H	OMe	H	OMe	0.215	0.161
17f	SO ₂ NH ₂	CO	H	OMe	H	OMe	0.179	0.118
15g	Bn	CO	H	OMe	OMe	OMe	> 100	77.5
16g	H	CO	H	OMe	OMe	OMe	0.051	0.071
17g	SO ₂ NH ₂	CO	H	OMe	OMe	OMe	0.166	0.275

[a] Results are the mean of three determinations. [b] Not determined.

As with sulfamates in the tetrahydroisoquinoline series, sulfamate **17f** also inhibits carbonic anhydrase (IC₅₀ 890 nM), which may enhance the bioavailability of this compound.^[13] Compounds **16f** and **17f** were also assayed for anti-angiogenic activity in an in vitro model of angiogenesis, wherein endothelial cells co-cultured in a matrix of human dermal fibroblasts are used to assess anti-angiogenic potential (Figure 2). When stimulated with VEGF the endothelial cells proliferate and migrate through the matrix to form tubule-like structures. The extent of tubule formation was quantified after 11 days as de-

Table 2. Antiproliferative activity of polymethoxylated *N*-benzoyl THIQs against DU-145 human prostate cancer cells and MDA MB-231 human breast cancer cells in vitro.

Compd					GI ₅₀ [μM] ^[a]	
	R ¹	R ³	R ⁴	R ⁵	DU-145	MDA MB-231
14e	Bn	OMe	OMe	H	26.7	ND ^[b]
18a	H	OMe	OMe	H	> 100	ND ^[b]
19a	SO ₂ NH ₂	OMe	OMe	H	> 100	ND ^[b]
14f	Bn	OMe	H	OMe	46.8	ND ^[b]
18b	H	OMe	H	OMe	6.4	ND ^[b]
19b	SO ₂ NH ₂	OMe	H	OMe	9	ND ^[b]
14g	Bn	OMe	OMe	OMe	62.7	18.9
18c ^[c]	H	OMe	OMe	OMe	0.921	0.733
19c ^[c]	SO ₂ NH ₂	OMe	OMe	OMe	1.81	1.72

[a] Results are the mean of three determinations. [b] Not determined. [c] Data for **18c** and **19c** are taken from the literature.^[6]

Table 3. GI₅₀ and MGM values of compound **16g** obtained for representative cell lines in the NCI-60 screening panel.

Cell Line	GI ₅₀ [μM] ^[a]	Cell Line	GI ₅₀ [μM] ^[a]
Leukemia CCRF-CEM	0.0171	Melanoma UACC-62	< 0.01
Lung HOP-62	0.0295	Ovarian OVCAR-3	< 0.01
Colon HCT-116	0.0216	Renal SN12-C	0.056
CNS SF-539	0.0102	<i>MGM</i>	0.033

[a] Results are the mean of three determinations. The MGM value (in italics) represents the mean concentration that caused 50% growth inhibition in all 60 cell lines.

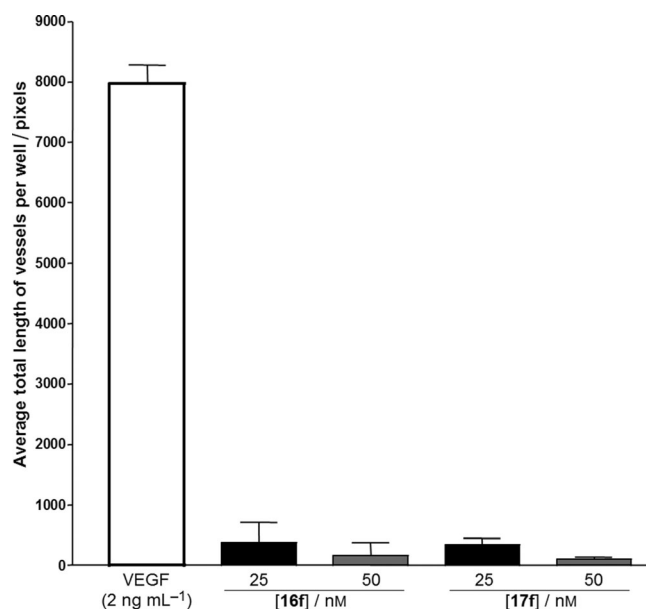


Figure 2. Activity of **16f** and **17f** in an in vitro model of angiogenesis.

scribed elsewhere.^[14] Treatment with 25 nM and 50 nM concentrations of **16f** and **17f** almost completely inhibits the formation of tubule-like structures (Figure 2). Quantification carried

out by calculating total pixel length reflects the lack of tubule formation. Both phenol **16 f** and its sulfamate **17 f** are highly potent.

With these excellent in vitro data in hand, we decided to explore further the microtubule disruptor activity in particular of **16 f–g** and **17 f–g** alongside the established potent agent combretastatin A-4 (CA-4), currently in clinical trials. All the tested novel compounds show excellent activity as inhibitors of tubulin assembly, with IC_{50} values of 1.2–1.5 μM similar to that of CA-4 (IC_{50} 1.1 μM). Ultimately, as observed before, the concentration required in tubulin-based assays far exceeds the antiproliferative dose, and most likely it suffices to disrupt microtubule dynamics to arrest the cell cycle rather than cause a catastrophic depolymerisation event. It should also be stressed that the nominal compound concentration in antiproliferative assays is that of agent added to the culture medium, rather than the actual concentration within cells, and drug transport can also be a key determinant in reaching the concentration required to achieve an effect. We also determined that **16 f–g** and **17 f–g** inhibit colchicine binding to tubulin very effectively. Compound **17 f**, which is the most active (89% inhibition at 5 μM), was also tested at lower concentration and shows 65% inhibition at 1 μM , approaching again the activity of CA-4 (99 and 90%). It is thus reasonable to suggest that the activity of the novel DHIQ derivatives can at least partially be ascribed to their ability to disrupt the normal dynamic polymerisation of tubulin by interaction at, or around, the colchicine binding site (Table 4).

Table 4. Activity of selected DHIQs as inhibitors of tubulin polymerisation (TP) and colchicine binding (CB) to tubulin.

Compd	TP IC_{50} [μM] ^[a]	CB [% inhib.] ^[a]	
		5 μM inhibitor	1 μM inhibitor
CA-4	1.1 \pm 0.1	99 \pm 0.6	90 \pm 0.2
1 a ^[c]	2.2 \pm 0.3	28 \pm 3	ND ^[b]
1 b ^[c]	1.3 \pm 0.08	78 \pm 0.9	ND ^[b]
1 c ^[c]	1.3 \pm 0.01	45 \pm 4	ND ^[b]
16 f	1.5 \pm 0.1	81 \pm 0.7	ND ^[b]
16 g	1.3 \pm 0.1	83 \pm 2	ND ^[b]
17 f	1.3 \pm 0.01	89 \pm 0.1	65 \pm 3
17 g	1.2 \pm 0.03	85 \pm 2	ND ^[b]

[a] Values are the mean \pm SD of at least two determinations. [b] Not determined. [c] Data for **1 a–c** are taken from the literature.^[3]

Finally, an X-ray crystal structure of **17 f** was obtained to explore conformational effects (Figure 3).^[15] The crystal structure shows that in the solid state the molecule adopts a “steroid-like” conformation as planned with a dihedral angle of 148.4° between the two carbonyl groups (Figure 3a). In the single crystal this fixed angle also appears as a result of positive intermolecular interactions (Figure 3b). The sulfamate group plays a key role to stabilise the framework of the lattice structure by acting as a hydrogen donor to the C3'-methoxy group of a second molecular unit that is part of the same string and to the carbonyl group at C1 of a third molecular unit that is part of a second string running in the opposite direction. The

angles in the crystal structure do not necessarily reflect conditions in solution. However, the excellent in vitro antiproliferative activities obtained for this class of compound suggest that adoptable conformations might be very restricted in solution as well, mainly as a result of electrostatic repulsion between the two carbonyl groups, and somewhere around the angle observed in the crystal. Control compounds like **8 d–e** and **9 d–e** without the carbonyl group in the linker to the D-ring mimic and **18 b–c** and **19 b–c** without the carbonyl at C1 do not have this favourable restriction and therefore show far less potency than their counterparts **16 f–g** and **17 f–g** that have both carbonyl groups present.

Molecular structure **17 f** was treated with a computational energy minimisation procedure. These calculations showed, in the minimal energy conformation, that a dihedral angle of 151.0° exists between the two carbonyl groups that is in good agreement with the observed angle of 148.4° in the crystal structure. As we postulated, the 3',5'-dimethoxybenzoyl group is projected into the area of space that is occupied by the steroidal D ring and the C18 methyl group in the estratriene series. This is illustrated in Figure 4 where the minimum energy state of **17 f** is overlaid with energy-minimised **1 a**. Additionally, the D-ring is rotated 47.2° out of plane of the A,B-ring system leading to one methoxy group being projected into the space of the sulfamate group and the other into the space of the C18 methyl group of **1 a**. Although not shown here, the most potent DHIQs were subjected to the same conformational analysis and the results (angles) obtained were unsurprisingly very similar (**16 g**: 149.8° and 46.9°; **17 g**: 150.5° and 47.1°) to the ones for compound **17 f**. These calculations support our postulate that the presence of the imide substructure favours adoption of a “steroid-like” conformation. It seems reasonable to propose that the positive effects on activity can, to some degree, be ascribed to this conformational biasing.

Conclusions

In summary, dihydroisoquinolinone derivatives including their sulfamate esters are compounds with activities in the low micromolar and nanomolar range against the proliferation of prostate and breast cancer cells. The most potent DHIQ derivative synthesised **16 g**, is also confirmed as having very high potency against a broad range of cancer phenotypes in the NCI 60-cell line assay. As anticipated, these compounds also display anti-angiogenic activity in an in vitro angiogenesis assay and appear to function as microtubule disruptors, as evidenced by their ability to inhibit polymerisation of tubulin and compete at the colchicine binding site. The most active DHIQs **16 f–g** and their sulfamates **17 f–g** are nearly equipotent to combretastatin A-4 (CA-4) as inhibitors of tubulin assembly and colchicine binding. Additionally, X-ray analysis of **17 f** reveals that electrostatic repulsion between the two adjacent carbonyl groups might partially explain the excellent in vitro activities of this novel series of microtubule disruptors. This conclusion is further supported by molecular modelling studies. Thus, com-

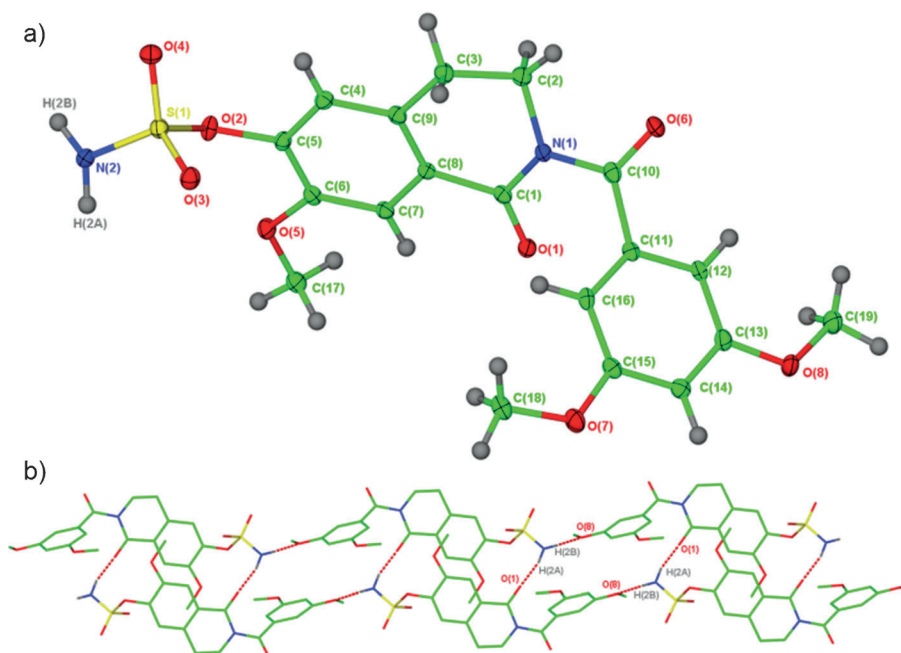


Figure 3. a) Single-crystal X-ray structure of **17 f**. Ellipsoids are depicted at 30% probability. b) Part of the crystal lattice packing diagram of **17 f** to illustrate the hydrogen-bonded linear arrangements present in the gross structure.

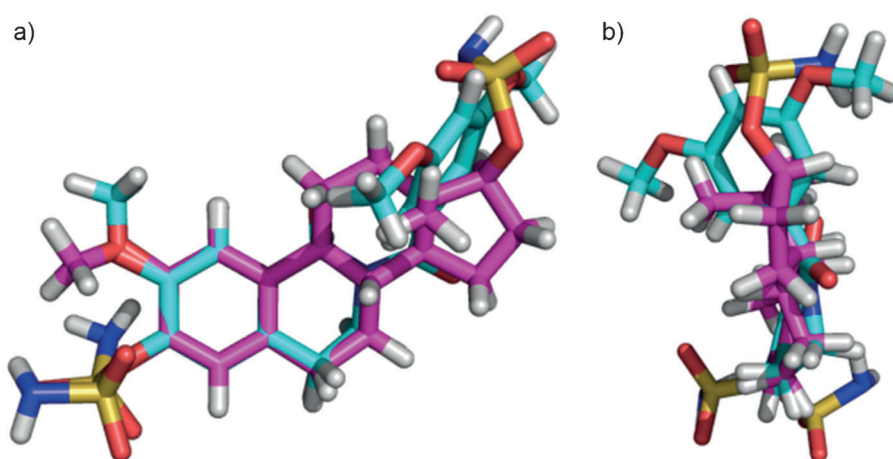


Figure 4. Overlay of the minimum energy conformation of DHIQ **17 f** (cyan) with steroid **1 a** (pink) viewed from two perspectives.

pounds derived from the DHIQ series are worthy of *in vivo* investigation and potential further pre-clinical development.

Experimental Section

In vitro studies

Cell lines: DU-145 (brain metastasis carcinoma of the prostate) and MDA MB-231 (metastatic pleural effusion of breast adenocarcinoma) established human cell lines were obtained from ATCC Global Bioresource Center. Cells were maintained in a 5% CO₂ humidified atmosphere at 37 °C in RPMI-1640 medium, supplemented with 10% fetal bovine serum, penicillin, (100 U mL⁻¹), and streptomycin

(0.1 mg mL⁻¹). The effect of drugs on tubule formation was assessed using an angiogenesis kit (TCS-Cellworks).

Antiproliferative assays: DU-145 and MDA MB-231 cells were seeded into 96-well microtitre plates (5000 cells per well) and treated with compound (10⁻⁹–10⁻⁴ M) or with vehicle control. At 96 h post-treatment, live cell counts were determined by WST-1 cell proliferation assay (Roche, Penzberg, Germany), as per the manufacturer's instructions. Viability results were expressed as a percentage of mean control values resulting in the calculation of the 50% growth inhibition (GI₅₀). All experiments were performed in triplicate.

Anti-angiogenic assays: HUVECs were cultured in a 24-well plate within a matrix of human diploid fibroblasts of dermal origin in optimised medium supplied by the company. The co-cultured cells were incubated throughout the experiment at 37 °C under 5% CO₂ in a humidified incubator. On day 1, the culture medium was removed and replaced with medium containing the drug under investigation. On days 4, 7 and 9 the medium was replaced with fresh medium containing the drugs. Suramin (20 µL) was used as a positive anti-angiogenic control and vascular endothelial growth factor (VEGF, 2 ng mL⁻¹) as a pro-angiogenic control. Each compound was tested in triplicate. On day 11, the cells were washed (PBS) and 70% EtOH (1 mL) added to each well for 30 min to fix the cells. After fixation the cells were washed with blocking buffer (1 mL, PBS 1% BSA; Sigma, UK) and stained for

von Willebrand's factor (manual scoring) for CD31 (computer analysis) in accordance with the manufacturer's instructions (TCS-Cellworks). The extent of tubule formation was then quantified, either manually by eye or using free software available through the National Institutes of Health website (NIH image [Mac version] from <http://rsb.info.nih.gov/nih-image/download.html>). For manual scoring a grid was drawn on the back of the plate using a fine marker pen. A 25-point Chalkley eyepiece graticule (Pyser SGI Ltd, UK) was fitted to the microscope and 10x magnification was used to count the tubules within five viewing fields (where the grid lines intersect), the average and SE were then calculated. For computer analysis, eight fixed fields of view for each well (10x magnification) were digitally photographed using a DC120 camera (Kodak, Rochester, NY, USA) with the Photoshop (Adobe, USA) plug-in MDS

120 software (Kodak). The pictures were copied to the NIH image software using Quicktime (Apple, USA) translation. After correcting for background the software counted the number of pixels representing tubules. The total pixel area within the eight fields was then calculated for each well. The average total pixel area and SE were then calculated. There was a significant correlation ($r=0.94$, $p<0.001$) between manual and computer analysis of tubule formation.

Tubulin assays: Bovine brain tubulin, prepared as described previously,^[16] was used in studies presented here. Assembly IC_{50} values were determined as described in detail elsewhere.^[17] Briefly, 1.0 mg mL^{-1} ($10\text{ }\mu\text{M}$) tubulin was pre-incubated without GTP with varying compound concentrations for 15 min at $30\text{ }^{\circ}\text{C}$. Reaction mixtures were placed on ice, and GTP (final concentration, 0.4 mM) was added. The reaction mixtures were transferred to cuvettes, held at $0\text{ }^{\circ}\text{C}$ in a recording spectrophotometer. Baselines were established at $0\text{ }^{\circ}\text{C}$, and increase in turbidity was followed for 20 min following a rapid ($<30\text{ s}$) jump to $30\text{ }^{\circ}\text{C}$. Compound concentrations required to decrease the turbidity increase by 50% were determined. The method for measuring inhibition of the binding of [^3H]colchicine to tubulin was described in detail previously.^[18] Reaction mixtures contained 0.1 mg mL^{-1} ($1.0\text{ }\mu\text{M}$) tubulin, $5.0\text{ }\mu\text{M}$ [^3H]colchicine, and potential inhibitor at $5.0\text{ }\mu\text{M}$. Compounds were compared with CA-4, a particularly potent inhibitor of the binding of colchicine to tubulin.^[19] Reaction mixtures were incubated for 10 min at $37\text{ }^{\circ}\text{C}$, a time point at which the binding of colchicine in control reaction mixtures is generally 40–60% complete. A minimum of two experiments were performed with each compound.

Chemistry

General: All chemicals were either purchased from Aldrich Chemical Co. (Gillingham, UK) or Alfa Aesar (Heysham, UK). Organic solvents of A.R. grade were supplied by Fisher Scientific (Loughborough, UK) and used as supplied. CHCl_3 , CH_2Cl_2 , DMA, DMF and THF were purchased from Aldrich and stored under a positive pressure of N_2 after use. Compounds **13**, **18c** and **19c** were prepared according to literature procedures.^[8] Sulfamoyl chloride was prepared by an adaptation of the method of Appel and Berger^[20] and was stored in the refrigerator under positive pressure of N_2 as a solution in toluene as described by Woo et al.^[21] An appropriate volume of this solution was freshly concentrated in vacuo immediately before use. Reactions were carried out at room temperature unless stated otherwise. Thin-layer chromatography (TLC) was performed on pre-coated aluminum plates (Merck, silica gel 60 F_{254}). Product spots were visualised either by UV irradiation at 254 nm or by staining with either alkaline KMnO_4 solution or 5% dodecamolybdophosphoric acid in EtOH, followed by heating. Flash column chromatography was performed using gradient elution (solvents indicated in text) on either pre-packed columns (Isolute) on a Flashmaster II system (Biotage, Uppsala, Sweden) or on a CombiFlash R_f Automated Flash Chromatography System (Teledyne Isco, Lincoln, NE, USA) with RediSep R_f disposable flash columns. ^1H and ^{13}C NMR spectra were recorded with either a Delta JMN-GX 270 (Jeol, Peabody, MA, USA) at 270 and 67.5 MHz, respectively, or a Mercury VX 400 NMR spectrometer (Varian, Palo Alto, CA, USA) at 400 and 100 MHz, respectively. Chemical shifts δ are reported in parts per million (ppm) relative to tetramethylsilane (TMS) as internal standard. Coupling constants J are recorded to the nearest 0.1 Hz. Mass spectra were recorded at the Mass Spectrometry Service Centre, University of Bath, UK. FAB-MS was carried out using *m*-nitrobenzyl alcohol (NBA) as the matrix. Melting points were determined using a Stuart SMP3 or a Stanford research systems Optimelt MPA100 melting

point apparatus (Stanford Research Systems, Sunnyvale, CA, USA) and are uncorrected. All compounds were $\geq 98\%$ pure by reversed-phase HPLC run with $\text{CH}_3\text{CN}/\text{H}_2\text{O}$ or $\text{MeOH}/\text{H}_2\text{O}$ (Sunfire C_{18} reversed-phase column, $4.6\times 150\text{ mm}$, $3.5\text{ }\mu\text{m}$ pore size).

Crystallographic methods: Single crystals of compound **17f** were analysed using a Nonius Kappa CCD diffractometer using $\text{Mo}(K\alpha)$ radiation. The structure was solved using SHELXS-97^[22] and refined using full-matrix least squares in SHELXL-97.^[23] The hydrogen atoms attached to N_2 were located and refined subject to being a distance of 0.89 \AA from the parent atom. CCDC 874341 contains the crystallographic data for compound **17f** and can be obtained free of charge from The Cambridge Crystallographic Data Centre via http://www.ccdc.cam.ac.uk/data_request/cif.

Computational methods: The Schrödinger software running under Maestro version 9.2.112 was used for all computational work. The crystal structure of **17f** solved in this work was used. One molecule was taken from this structure and run through a brief geometry optimisation procedure. Both dihedral angles were manually and independently adjusted in steps of 30 ° and the energy of the conformers was calculated. Compounds **16g** and **17g** were built by altering the crystal structure of **17f** to yield the desired structure which was then run through a brief geometry optimisation procedure. Conformers were generated and molecular energies calculated as described above.

5-Hydroxy-6-methoxyindan-1-one (4): 5,6-Dimethoxy-indan-1-one (25.93 g, 135.0 mmol) was placed in a 1 L round-bottom flask. Piperidine (200 mL) and H_2O (50 mL) were added and the reaction mixture was stirred at $140\text{ }^{\circ}\text{C}$ for 8 days. The mixture was concentrated in vacuo and aqueous NaOH (2 M, 400 mL) was added. The mixture was extracted with EtOAc (200 mL) and CH_2Cl_2 (200 mL). The aqueous layer was then acidified with HCl (12 M, 100 mL) and extracted with CH_2Cl_2 ($3\times 200\text{ mL}$). This organic layer system was washed with H_2O (200 mL), dried (MgSO_4), filtered and concentrated in vacuo to give compound **4** as a yellow powder (16.87 g, 70%). ^1H NMR (270 MHz, CDCl_3): $\delta=2.49\text{--}2.60$ (2H, m), 2.86–2.98 (2H, m), 3.83 (3H, s), 6.86 (1H, s), 7.09 (1H, s), 8.03 ppm (1H, s, br).

5-Benzyloxy-6-methoxyindan-1-one (5): Compound **4** (3.0 g, 16.8 mmol), benzyl bromide (2.1 mL, 17.7 mmol) and potassium carbonate (4.70 g, 34.0 mmol) were suspended in DMF (20 mL) and stirred at RT for 3 days. H_2O (50 mL) was added and the mixture was extracted with EtOAc ($2\times 80\text{ mL}$). The combined organics were washed with H_2O and brine, dried (MgSO_4) and concentrated in vacuo. Crystallisation from EtOAc/hexane 2:3 afforded compound **5** as a light-yellow powder (4.10 g, 91%), mp: $139\text{--}140\text{ }^{\circ}\text{C}$. ^1H NMR (270 MHz, CDCl_3): $\delta=2.58\text{--}2.66$ (2H, m), 2.93–3.01 (2H, m), 3.89 (3H, s), 5.22 (2H, s), 6.88 (1H, s), 7.18 (1H, s), 7.27–7.46 ppm (5H, m).

6-Benzyloxy-7-methoxy-3,4-dihydro-2H-isoquinolin-1-one (6a): Compound **5** (2.68 g, 10 mmol) was dissolved in CH_2Cl_2 (10 mL) and methanesulfonic acid (10 mL) and the solution was cooled to $0\text{ }^{\circ}\text{C}$. Sodium azide (1.32 g, 20 mmol) was added portionwise over 0.5 h. The mixture was allowed to warm to RT and stirred for 16 h. Aqueous NaOH (1.5 M, 50 mL) was added dropwise at $0\text{ }^{\circ}\text{C}$. The mixture was extracted with EtOAc ($3\times 80\text{ mL}$). The combined organics were washed with H_2O and brine, dried (MgSO_4) and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 3:1 to EtOAc) and crystallisation from EtOAc afforded compound **6a** as a white solid (1.45 g, 52%), mp: $177\text{--}178\text{ }^{\circ}\text{C}$. ^1H NMR (270 MHz, CDCl_3): $\delta=2.87$ (2H, t, $J=7.6\text{ Hz}$), 3.53 (2H, t, $J=7.6\text{ Hz}$), 3.94 (3H, s), 5.20 (2H, s), 6.03 (1H, s, br), 6.70 (1H, s), 7.29–7.46 (5H, m), 7.60 ppm (1H, s); LC-MS (FAB+): m/z 284.44 $[\text{M}+\text{H}]^+$.

6-Hydroxy-7-methoxy-3,4-dihydro-2H-isoquinolin-1-one (6b): Compound **6a** (1.418 g, 5.0 mmol) was dissolved in MeOH (50 mL) and filtered. The solution was sucked at 1.0 mL min⁻¹ into the H-cube and treated with full hydrogen over Pd/C (10%, 140 mg cartridge) at RT. The resulting solution was concentrated in vacuo to afford compound **6b** as a pale-yellow solid (961 mg, 99%). ¹H NMR (270 MHz, CDCl₃): δ = 2.72 (2H, t, *J* = 6.7 Hz), 3.27–3.31 (2H, m), 3.77 (3H, s), 6.65 (1H, s), 7.33 (1H, s), 7.64 (1H, s, br), 9.65 ppm (1H, s, br); LC–MS (ES⁺): *m/z* 194.2 [M+H]⁺.

7-Methoxy-(6-triisopropoxy)-3,4-dihydro-2H-isoquinolin-1-one (6c): Compound **6b** (3.1 g, 16 mmol), TIPSCI (7.1 mL, 33 mmol) and imidazole (2.4 g, 35 mmol) in DMF (30 mL) was stirred at RT for 16 h. H₂O (30 mL) was added and the mixture was extracted with EtOAc (2 × 80 mL). The combined organic layers were washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. Flash column chromatography (hexane to EtOAc) afforded compound **6c** as a white powder (5.0 g, 93%), mp: 112–113 °C. ¹H NMR (270 MHz, CDCl₃): δ = 1.07 (18H, d, *J* = 6.9 Hz), 1.23 (3H, sept, *J* = 6.9 Hz), 2.85 (2H, t, *J* = 6.7 Hz), 3.51 (2H, dt, *J* = 6.7, 2.9 Hz), 3.82 (3H, s), 5.86 (1H, s, br), 6.66 (1H, s), 7.53 ppm (1H, s); LC–MS (APCI⁺): *m/z* 350.50 [M+H]⁺.

7-Methoxy-2-(2-methoxybenzyl)-6-(2-methoxybenzyloxy)-3,4-dihydroisoquinolin-1(2H)-one (7a): Compound **6c** (505 mg, 1.44 mmol) was dissolved in anhydrous DMF (5 mL) and cooled to 0 °C. Sodium hydride (60% in mineral oil, 115 mg, 2.88 mmol) was added portionwise and the suspension was stirred at 0 °C for 0.5 h. 2-Methoxybenzyl chloride (0.24 mL, 1.73 mmol) was added dropwise and the reaction mixture was stirred at RT for 4 days. Ammonium chloride (saturated, 10 mL) was added and the mixture was extracted with EtOAc (80 mL). The organic layer was washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 10:1 to 2:1) afforded **7a** as a white powder (350 mg, 56%), mp: 133–134 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.81 (2H, t, *J* = 6.7 Hz), 3.50 (2H, t, *J* = 6.7 Hz), 3.83 (3H, s), 3.85 (3H, s), 3.93 (3H, s), 4.78 (2H, s), 5.22 (2H, s), 6.64 (1H, s), 6.84–6.95 (4H, m), 7.18–7.32 (3H, m), 7.44 (1H, dd, *J* = 7.4, 1.5 Hz), 7.65 ppm (1H, s); LC–MS (APCI⁺): *m/z* 434.56 [M+H]⁺; HRMS (ES⁺): *m/z* found 434.1960; C₂₆H₂₈NO₅⁺ [M+H]⁺ requires 434.1962.

7-Methoxy-2-(3-methoxybenzyl)-6-(triisopropylsilyloxy)-3,4-dihydroisoquinolin-1(2H)-one (7b1) and 7-Methoxy-2-(3-methoxybenzyl)-6-(3-methoxybenzyloxy)-3,4-dihydroisoquinolin-1(2H)-one (7b2): Method as for **7a** using compound **6c** (505 mg, 1.44 mmol), sodium hydride (60%, 115 mg, 2.88 mmol) and 3-methoxybenzyl bromide (0.24 mL, 1.73 mmol) in DMF (5 mL) at 0 °C for 0.5 h and at RT for 4 days. Flash column chromatography (hexane/EtOAc 10:1 to 5:1 to 2:1) afforded **7b1** as a colourless oil (120 mg, 18%) and **7b2** as a white powder (110 mg, 18%), mp: 85–86 °C. **7b1:** ¹H NMR (270 MHz, CDCl₃): δ = 1.07 (18H, d, *J* = 6.7 Hz), 1.22 (3H, sept, *J* = 6.7 Hz), 2.79 (2H, t, *J* = 6.7 Hz), 3.44 (2H, t, *J* = 6.7 Hz), 3.77 (3H, s), 3.84 (3H, s), 4.73 (2H, s), 6.61 (1H, s), 6.80 (1H, ddd, *J* = 8.1, 2.5, 0.8 Hz), 6.85–6.92 (2H, m), 7.23 (1H, t, *J* = 7.9 Hz), 7.61 ppm (1H, s); HRMS (ES⁺): *m/z* found 470.2709; C₂₇H₄₀NO₄Si⁺ [M+H]⁺ requires 470.2721. **7b2:** ¹H NMR (270 MHz, CDCl₃): δ = 2.77 (2H, t, *J* = 6.6 Hz), 3.41 (2H, t, *J* = 6.6 Hz), 3.75 (3H, s), 3.77 (3H, s), 3.92 (3H, s), 4.72 (2H, s), 5.14 (2H, s), 6.61 (1H, s), 6.76–6.89 (4H, m), 6.94–7.01 (2H, m), 7.21 (1H, t, *J* = 7.9 Hz), 7.25 (1H, t, *J* = 8.1 Hz), 7.66 ppm (1H, s); LC–MS (APCI⁺): *m/z* 434.56 [M+H]⁺; HRMS (ES⁺): *m/z* found 434.1962; C₂₆H₂₈NO₅⁺ [M+H]⁺ requires 434.1962.

7-Methoxy-6-(triisopropylsilyloxy)-2-(4-methoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (7c): Method as for **7a** using compound **6c** (524 mg, 1.5 mmol), sodium hydride (60%, 64 mg, 1.6 mmol) and 4-methoxybenzyl bromide (0.26 mL, 1.8 mmol) in DMF (5 mL) at 0 °C for 0.5 h and at RT for 6 h. Flash column chromatography (hexane/EtOAc 10:1 to 5:1) afforded compound **7c** as a colourless oil (385 mg, 55%). ¹H NMR (270 MHz, CDCl₃): δ = 1.07 (18H, d, *J* = 6.9 Hz), 1.22 (3H, sept, *J* = 6.9 Hz), 2.77 (2H, t, *J* = 6.7 Hz), 3.42 (2H, t, *J* = 6.7 Hz), 3.78 (3H, s), 3.84 (3H, s), 4.68 (2H, s), 6.60 (1H, s), 6.84 (2H, dt, *J* = 8.6, 2.4 Hz), 7.26 (2H, dt, *J* = 8.5, 2.3 Hz), 7.60 ppm (1H, s); LC–MS (APCI⁺): *m/z* 470.57 [M+H]⁺; HRMS (ES⁺): *m/z* found 470.2706; C₂₇H₄₀NO₄Si⁺ [M+H]⁺ requires 470.2721.

6-Benzyloxy-7-methoxy-2-(3,5-dimethoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (7d): Method as for **7a** using compound **6a** (425 mg, 1.5 mmol), sodium hydride (60%, 120 mg, 3.0 mmol) and 3,5-dimethoxybenzyl bromide (416 mg, 1.8 mmol) in DMF (10 mL) at 0 °C for 0.5 h and at RT for 18 h. Flash column chromatography (hexane/EtOAc 5:1 to 1:1) gave an oil that was stirred in Et₂O (50 mL) and hexane (20 mL), filtered and dried in vacuo to afford compound **7d** as a white powder (440 mg, 68%), mp: 82–83 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.79 (2H, t, *J* = 6.7 Hz), 3.43 (2H, t, *J* = 6.7 Hz), 3.75 (6H, s), 3.93 (3H, s), 4.69 (2H, s), 5.17 (2H, s), 6.35 (1H, t, *J* = 2.5 Hz), 6.45 (2H, d, *J* = 2.5 Hz), 6.62 (1H, s), 7.27–7.45 (5H, m), 7.66 ppm (1H, s); LC–MS (ES⁺): *m/z* 456.03 ([M+Na]⁺, 100%), 434.05 [M+H]⁺; HRMS (ES⁺): *m/z* found 434.1956; C₂₆H₂₈NO₅⁺ [M+H]⁺ requires 434.1962.

6-Benzyloxy-7-methoxy-2-(3,4,5-trimethoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (7e): Method as for **7a** using compound **6a** (425 mg, 1.5 mmol), sodium hydride (60%, 120 mg, 3.0 mmol) and 3,4,5-trimethoxybenzyl chloride (390 mg, 1.8 mmol) in DMF (10 mL) at 0 °C for 0.5 h and at RT for 18 h. The residue was stirred in Et₂O, filtered and dried in vacuo to afford compound **7e** as a white powder (570 mg, 82%), mp: 136–137 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.80 (2H, t, *J* = 6.8 Hz), 3.43 (2H, t, *J* = 6.8 Hz), 3.82 (9H, s), 3.93 (3H, s), 4.68 (2H, s), 5.17 (2H, s), 6.52 (2H, s), 6.63 (1H, s), 7.26–7.43 (5H, m), 7.66 ppm (1H, s); LC–MS (ES⁺): *m/z* 486.26 ([M+Na]⁺, 100%), 464.28 [M+H]⁺; HRMS (ES⁺): *m/z* found 464.2064; C₂₇H₃₀NO₆⁺ [M+H]⁺ requires 464.2068.

6-Hydroxy-7-methoxy-2-(2-methoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (8a): A mixture of compound **7a** (290 mg, 0.67 mmol) and Pd/C (10%, 40 mg) in THF (20 mL) and MeOH (20 mL) was stirred under hydrogen at RT for 4 h. After filtration through Celite and evaporation under reduced pressure the residue was stirred in Et₂O, filtered and dried in vacuo to afford compound **8a** as a white powder (195 mg, 93%), mp: 164–165 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.83 (2H, t, *J* = 6.7 Hz), 3.51 (2H, t, *J* = 6.7 Hz), 3.83 (3H, s), 3.92 (3H, s), 4.78 (2H, s), 6.02 (1H, s), 6.68 (1H, s), 6.87 (1H, d, *J* = 7.4 Hz), 6.91 (1H, dd, *J* = 7.4, 1.0 Hz), 7.23 (1H, dt, *J* = 7.4, 1.5 Hz), 7.30 (1H, dd, *J* = 7.9, 1.5 Hz), 7.63 ppm (1H, s); LC–MS (APCI⁺): *m/z* 314.17 [M+H]⁺; HRMS (ES⁺): *m/z* found 314.1384; C₁₈H₂₀NO₄⁺ [M+H]⁺ requires 314.1387.

6-Hydroxy-7-methoxy-2-(3-methoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (8b): Method as for **8a** using compound **7b2** (85 mg, 0.196 mmol) and Pd/C (10%, 20 mg) in THF (10 mL) and MeOH (10 mL) under hydrogen at RT for 2 h. Flash column chromatography (hexane/EtOAc 1:1) afforded compound **8b** as a white powder (52 mg, 76%), mp: 181–182 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.82 (2H, t, *J* = 6.7 Hz), 3.44 (2H, t, *J* = 6.7 Hz), 3.77 (3H, s), 3.93 (3H, s), 4.74 (2H, s), 6.03 (1H, s), 6.68 (1H, s), 6.80 (1H, dd, *J* = 7.4, 1.7 Hz), 6.84 (1H, d, *J* = 1.7 Hz), 6.90 (1H, d, *J* = 7.7 Hz), 7.23 (1H, t, *J* = 7.9 Hz), 7.64 ppm (1H, s); LC–MS (APCI⁺): *m/z* 314.23 [M+H]⁺;

HRMS (ES+): m/z found 314.1383, $C_{18}H_{20}NO_4^+$ [$M+H$] $^+$ requires 314.1387.

6-Hydroxy-7-methoxy-2-(3-methoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (8b): Compound **7b1** (110 mg, 0.23 mmol) was dissolved in THF (20 mL). TBAF (1.0 M in THF, 0.28 mL, 0.28 mmol) was added dropwise and the reaction mixture was stirred at RT for 2 h. H_2O was added and the mixture extracted with EtOAc. The organic layer was washed with H_2O , brine, dried ($MgSO_4$) and concentrated in vacuo. The resulting yellow solid was stirred in Et_2O , filtered and dried in vacuo to afford compound **8b** as a white powder (60 mg, 82%). Analytical data are identical as shown above.

6-Hydroxy-7-methoxy-2-(4-methoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (8c): Method as for **8b** using compound **7c** (0.24 g, 0.51 mmol) and TBAF (1.0 M in THF, 0.61 mL, 0.61 mmol) in THF (10 mL) at RT for 18 h. Flash column chromatography (hexane/EtOAc 4:1 to 2:1) afforded compound **8c** as a white powder (110 mg, 69%), mp: 171–172 °C. 1H NMR (270 MHz, $CDCl_3$): δ = 2.78 (2H, t, J = 6.7 Hz), 3.41 (2H, t, J = 6.7 Hz), 3.77 (3H, s), 3.90 (3H, s), 4.69 (2H, s), 6.33 (1H, s, br), 6.66 (1H, s), 6.84 (2H, dt, J = 8.7, 2.3 Hz), 7.24 (2H, dt, J = 8.9, 2.3 Hz), 7.63 ppm (1H, s); LC-MS (APCI+): m/z 314.42 [$M+H$] $^+$; HRMS (ES+): m/z found 314.1381; $C_{18}H_{19}NO_4^+$ [$M+H$] $^+$ requires 314.1387.

6-Hydroxy-7-methoxy-2-(3,5-dimethoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (8d): Method as for **8a** using compound **7d** (330 mg, 0.76 mmol) and Pd/C (10%, 40 mg) in THF (20 mL) and MeOH (30 mL) under hydrogen at RT for 18 h. The resulting white solid was stirred in Et_2O , filtered and dried in vacuo to afford compound **8d** as a white powder (240 mg, 92%), mp: 166–167 °C. 1H NMR (270 MHz, $CDCl_3$): δ = 2.82 (2H, t, J = 6.7 Hz), 3.44 (2H, t, J = 6.7 Hz), 3.75 (6H, s), 3.93 (3H, s), 4.70 (2H, s), 6.04 (1H, s), 6.35 (1H, t, J = 2.5 Hz), 6.46 (2H, d, J = 2.5 Hz), 6.68 (1H, s), 7.63 ppm (1H, s); LC-MS (ES-): m/z 342.09 [$M-H$] $^-$; HRMS (ES+): m/z found 344.1477; $C_{19}H_{22}NO_5^+$ [$M+H$] $^+$ requires 344.1492.

6-Hydroxy-7-methoxy-2-(3,4,5-trimethoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (8e): Method as for **8a** using compound **7e** (375 mg, 0.81 mmol) and Pd/C (10%, 50 mg) in THF (20 mL) and MeOH (20 mL) under hydrogen at RT for 18 h. The resulting white solid was stirred in Et_2O , filtered and dried in vacuo to afford compound **8e** as a white powder (260 mg, 86%), mp: 168–169 °C. 1H NMR (270 MHz, $CDCl_3$): δ = 2.82 (2H, t, J = 6.7 Hz), 3.44 (2H, t, J = 6.7 Hz), 3.82 (9H, s), 3.92 (3H, s), 4.69 (2H, s), 6.08 (1H, s, br), 6.53 (2H, s), 6.69 (1H, s), 7.63 ppm (1H, s); LC-MS (ES-): m/z 372.07 [$M-H$] $^-$; HRMS (ES+): m/z found 374.1590; $C_{20}H_{24}NO_6^+$ [$M+H$] $^+$ requires 374.1598.

7-Methoxy-2-(2-methoxybenzyl)-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (9a): Sulfamoyl chloride (0.6 M in toluene, 1.0 mL, 0.6 mmol) was concentrated in vacuo and dissolved in anhydrous DMA (1.0 mL). Compound **8a** (94 mg, 0.3 mmol) was added as a solid and the reaction mixture was stirred at RT for 18 h. H_2O (10 mL) was added and the mixture was extracted with EtOAc (2 × 50 mL). The combined organic layers were washed with H_2O , brine, dried ($MgSO_4$), filtered and concentrated in vacuo. The residue was stirred in Et_2O , filtered and dried in vacuo to afford compound **9a** as a white solid (90 mg, 77%), mp: 142–143 °C. 1H NMR (270 MHz, $CDCl_3$): δ = 2.86 (2H, t, J = 6.7 Hz), 3.54 (2H, t, J = 6.7 Hz), 3.84 (3H, s), 3.93 (3H, s), 4.77 (2H, s), 5.16 (2H, s), 6.87 (1H, d, J = 8.2), 6.91 (1H, dt, J = 7.4, 1.0 Hz), 7.14 (1H, s), 7.21–7.31 (2H, m), 7.77 ppm (1H, s); LC-MS (APCI+): m/z 393.45 [$M+H$] $^+$; HRMS (ES+): m/z found 393.1118; $C_{18}H_{21}N_2O_6S^+$ [$M+H$] $^+$ requires 393.1115.

7-Methoxy-2-(3-methoxybenzyl)-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (9b): Method as for **9a** using compound **8b** (73 mg, 0.23 mmol) and sulfamoyl chloride (0.46 mmol) in DMA at RT for 18 h. Flash column chromatography (hexane/EtOAc 1:1 to 2:3) afforded compound **9b** as a white solid (60 mg, 67%), mp: 139–140 °C. 1H NMR (270 MHz, $CDCl_3$): δ = 2.86 (2H, t, J = 6.7 Hz), 3.47 (2H, t, J = 6.7 Hz), 3.78 (3H, s), 3.94 (3H, s), 4.73 (2H, s), 5.19 (2H, s), 6.79–6.89 (3H, m), 7.15 (1H, s), 7.24 (1H, dt, J = 7.6, 1.0 Hz), 7.78 ppm (1H, s); LC-MS (APCI+): m/z 393.64 [$M+H$] $^+$; HRMS (ES+): m/z found 393.1118; $C_{18}H_{21}N_2O_6S^+$ [$M+H$] $^+$ requires 393.1115.

7-Methoxy-2-(4-methoxybenzyl)-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (9c): Method as for **9a** using compound **8c** (82 mg, 0.26 mmol) and sulfamoyl chloride (0.78 mmol) in DMA (1.0 mL) at RT for 24 h. Flash column chromatography (hexane/EtOAc 3:1 to 1:1) gave a solid that was stirred in Et_2O (10 mL), filtered and dried in vacuo to afford compound **9c** as a white solid (75 mg, 73%), mp: 159–160 °C. 1H NMR (270 MHz, $CDCl_3/[D_4]MeOH$ 10:1): δ = 2.12 (2H, s), 2.81 (2H, t, J = 6.7 Hz), 3.42 (2H, t, J = 6.7 Hz), 3.76 (3H, s), 3.90 (3H, s), 4.67 (2H, s), 6.83 (2H, dt, J = 8.9, 2.5 Hz), 7.14 (1H, s), 7.20 (2H, dt, J = 8.9, 2.5 Hz), 7.72 ppm (1H, s); LC-MS (APCI+): m/z 393.38 [$M+H$] $^+$; HRMS (ES+): m/z found 393.1117; $C_{18}H_{21}N_2O_6S^+$ [$M+H$] $^+$ requires 393.1115.

7-Methoxy-2-(3,5-dimethoxybenzyl)-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (9d): Method as for **9a** using compound **8d** (100 mg, 0.29 mmol) and sulfamoyl chloride (0.87 mmol) in DMA (1.0 mL) at RT for 24 h. The residue was stirred in Et_2O , filtered, washed with Et_2O and dried in vacuo to afford compound **9d** as a white powder (95 mg, 77%), mp: 164–165 °C. 1H NMR (270 MHz, $CDCl_3$): δ = 2.90 (2H, t, J = 6.6 Hz), 3.49 (2H, t, J = 6.6 Hz), 3.74 (6H, s), 3.84 (3H, s), 4.64 (2H, s), 6.40–6.45 (3H, s), 7.26 (1H, s), 7.61 (1H, s), 8.08 ppm (2H, s, br); LC-MS (ES-): m/z 421.13 [$M-H$] $^-$; HRMS (ES+): m/z found 423.1215; $C_{19}H_{23}N_2O_7S^+$ [$M+H$] $^+$ requires 423.1220.

7-Methoxy-6-sulfamoyloxy-2-(3,4,5-trimethoxybenzyl)-3,4-dihydroisoquinolin-1(2H)-one (9e): Method as for **9a** using compound **8e** (100 mg, 0.27 mmol) and sulfamoyl chloride (0.8 mmol) in DMA (1.0 mL) at RT for 24 h. After addition of H_2O (20 mL), EtOAc (100 mL) and THF (50 mL) the organics could not be dissolved. The solvents were evaporated and the resultant solid in the aqueous layer was washed, filtered, washed with H_2O , Et_2O , EtOAc and dried in vacuo to afford compound **9e** as a white powder (90 mg, 74%), mp: 201–203 °C. 1H NMR (270 MHz, $[D_6]DMSO$): δ = 2.90 (2H, t, J = 6.6 Hz), 3.49 (2H, t, J = 6.6 Hz), 3.63 (3H, s), 3.74 (6H, s), 3.84 (3H, s), 4.64 (2H, s), 6.61 (2H, s), 7.26 (1H, s), 7.61 (1H, s), 8.08 ppm (2H, s, br); LC-MS (ES-): m/z 451.18 [$M-H$] $^-$; HRMS (ES+): m/z found 453.1313; $C_{20}H_{25}N_2O_8S^+$ [$M+H$] $^+$ requires 453.1326.

6-Benzyloxy-7-methoxy-2-((2-methoxyphenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (10a): Sodium hydride (60% in mineral oil, 46 mg, 1.9 mmol) was suspended in anhydrous DMF (5 mL). Compound **6a** (300 mg, 1.0 mmol) was added and the reaction mixture was heated at 50 °C for 0.5 h. The reaction mixture was cooled to RT and 2-methoxybenzenesulfonyl chloride (0.15 mL, 1.0 mmol) was added dropwise. The reaction mixture was stirred for 3.5 h and turned from yellow to almost colourless after addition of the sulfonyl chloride. A further 0.5 equiv (0.07 mL) of the sulfonyl chloride was added and the reaction mixture stirred for a further 2 h. The reaction mixture was poured into sodium bicarbonate (sat., 100 mL) and extracted with $CHCl_3$ (3 × 50 mL). The combined organic layers were washed with H_2O (4 × 50 mL) and brine (50 mL), dried ($MgSO_4$) and concentrated in vacuo. Flash column

chromatography (hexane/EtOAc 2:1) afforded the compound **10a** as a colourless foam (314 mg, 65%), mp: 202–203 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.99 (2H, t, *J* = 6.3 Hz), 3.80 (3H, s), 3.88 (3H, s), 4.24 (2H, t, *J* = 6.3 Hz), 5.19 (2H, s), 6.66 (1H, s), 6.96 (1H, d, *J* = 8.2 Hz), 7.14 (1H, dt, *J* = 7.6, 1.0 Hz), 7.31–7.44 (6H, m), 7.52–7.58 (1H, m), 8.20 ppm (1H, dd, *J* = 7.9, 1.7 Hz); LC–MS (ES+): *m/z* 454.46 [M+H]⁺; HRMS (ES+): *m/z* found 476.1124; C₂₄H₂₃NO₆Na⁺ [M+Na]⁺ requires 476.1144.

6-Benzyloxy-7-methoxy-2-((3-methoxyphenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (10b): Method as for **10a** using compound **6a** (300 mg, 1.1 mmol), sodium hydride (60% in mineral oil, 64 mg, 1.9 mmol) and 3-methoxybenzenesulfonyl chloride (0.22 mL, 1.6 mmol) in anhydrous DMF (5 mL) at 50 °C for 0.5 h and at RT for 2 h and at 40 °C for 2 h. Flash column chromatography (hexane/EtOAc 2:1) afforded compound **10b** as a colourless foam (369 mg, 77%), mp: 140–145 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.99 (2H, t, *J* = 6.2 Hz), 3.84 (3H, s), 3.87 (3H, s), 4.18 (2H, t, *J* = 6.2 Hz), 5.18 (2H, s), 6.64 (1H, s), 7.12 (1H, ddd, *J* = 8.2, 2.7, 1.0 Hz), 7.31–7.44 (6H, m), 7.47 (1H, s), 7.58–7.63 ppm (2H, m); LC–MS (ES+): *m/z* 476.50 [M+Na]⁺; HRMS (ES+): *m/z* found 476.1116; C₂₄H₂₃NO₆Na⁺ [M+Na]⁺ requires 476.1144.

6-Benzyloxy-7-methoxy-2-((4-methoxyphenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (10c): Method as for **10a** using compound **6a** (300 mg, 1.0 mmol), sodium hydride (60% in mineral oil, 46 mg, 1.9 mmol) and 3-methoxybenzenesulfonyl chloride (0.22 mL, 1.5 mmol) in anhydrous DMF (5 mL) at 50 °C for 0.5 h and at RT for 6 h. Flash column chromatography (hexane/EtOAc 2:1) afforded compound **10c** as a colourless oil (152 mg, 32%). ¹H NMR (270 MHz, CDCl₃): δ = 2.97 (2H, t, *J* = 6.3 Hz), 3.82 (3H, s), 3.87 (3H, s), 4.16 (2H, t, *J* = 6.3 Hz), 5.17 (2H, s), 6.63 (1H, s), 6.95–6.99 (2H, m), 7.28–7.41 (5H, m), 7.46 (1H, s), 7.98–8.03 ppm (2H, m); LC–MS (ES+): *m/z* 454.60 [M+H]⁺; HRMS (ES+): *m/z* found 454.1320; C₂₄H₂₄NO₆S⁺ [M+H]⁺ requires 454.1324.

6-Benzyloxy-7-methoxy-2-((3-chlorophenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (10d): Method as for **10a** using compound **6a** (300 mg, 1.0 mmol), sodium hydride (60% in mineral oil, 46 mg, 1.9 mmol) and 3-methoxybenzenesulfonyl chloride (0.22 mL, 1.5 mmol) in anhydrous DMF (5 mL) at 50 °C for 0.5 h and at RT for 6 h. Flash column chromatography (hexane/EtOAc 2:1) afforded compound **10d** as a yellow foam (259 mg, 54%), mp: 155–158 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.00 (2H, t, *J* = 6.2 Hz), 3.83 (3H, s), 4.18 (2H, t, *J* = 6.2 Hz), 5.18 (2H, s), 6.65 (1H, s), 7.27–7.49 (7H, m), 7.57 (1H, ddd, *J* = 7.9, 2.0, 1.2 Hz), 7.96–8.01 ppm (2H, m); LC–MS (ES+): *m/z* 458.52 [M+H]⁺; HRMS (ES+): *m/z* found 480.0621; C₂₃H₂₀ClNO₅Na⁺ [M+Na]⁺ requires 480.0643.

Methyl 2-((6-benzyloxy-7-methoxy-1-oxo-3,4-dihydroisoquinolin-2(1H)-yl)sulfonyl)benzoate (10e): Method as for **10a** using compound **6a** (300 mg, 1.0 mmol), sodium hydride (60% in mineral oil, 46 mg, 1.9 mmol) and 3-methoxybenzenesulfonyl chloride (0.22 mL, 1.5 mmol) in anhydrous DMF (5 mL) at 50 °C for 0.5 h and at RT for 6 h. Purification by flash column chromatography (hexane/EtOAc 2:1) afforded compound **10e** as a colourless powder (176 mg, 35%). ¹H NMR (270 MHz, CDCl₃): δ = 3.05 (2H, t, *J* = 6.2 Hz), 3.83 (3H, s), 3.92 (3H, s), 4.19 (2H, t, *J* = 6.2 Hz), 5.18 (2H, s), 6.65 (1H, s), 7.28–7.41 (5H, m), 7.46 (1H, s), 7.61–7.71 (3H, m), 8.55 ppm (1H, dd, *J* = 6.4, 2.0 Hz); LC–MS (APCI-): *m/z* 482.29 [M–H][–]; HRMS (ES+): *m/z* found 504.1075; C₂₅H₂₃NO₇Na⁺ [M+Na]⁺ requires 504.1087.

6-Hydroxy-7-methoxy-2-((2-methoxyphenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (11a): Method as for **8a** using compound **10a** (316 mg, 0.7 mmol) and Pd/C (10%, 32 mg) in THF

(3 mL) and EtOH (3 mL) under hydrogen at RT for 2 h. The residue was crystallised from dichloromethane/hexane to afford compound **11a** as a colourless powder (145 mg, 57%), mp: 222–224 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.02 (2H, t, *J* = 6.3 Hz), 3.81 (3H, s), 3.88 (3H, s), 4.25 (2H, t, *J* = 6.3 Hz), 6.07 (1H, s), 6.72 (1H, s), 6.96 (1H, d, *J* = 8.4 Hz), 7.12 (1H, t, *J* = 7.7 Hz), 7.42 (1H, s), 7.51–7.58 (1H, m), 8.19 ppm (1H, dd, *J* = 7.9, 1.7 Hz); LC–MS (ES–): *m/z* 362.33 [M–H][–]; HRMS (ES+): *m/z* found 386.0651; C₁₇H₁₇NO₆Na⁺ [M+Na]⁺ requires 386.0674.

6-Hydroxy-7-methoxy-2-((3-methoxyphenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (11b): Method as for **8a** using compound **10b** (369 mg, 0.81 mmol) and Pd/C (10%, 37 mg) in THF (5 mL) and EtOH (5 mL) under hydrogen at RT for 2 h. Flash column chromatography (hexane/EtOAc 1:1 to 1:2 to 1:3) afforded compound **11b** as a colourless solid (203 mg, 69%), mp: 195–197 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.02 (2H, t, *J* = 6.3 Hz), 3.84 (3H, s), 3.86 (3H, s), 4.19 (2H, t, *J* = 6.3 Hz), 6.07 (1H, s), 6.71 (1H, s), 7.11 (1H, ddd, *J* = 8.4, 2.6, 1.0 Hz), 7.41 (1H, t, *J* = 8.1 Hz), 7.45 (1H, s), 7.57–7.63 ppm (2H, m); LC–MS (ES–): *m/z* 362.42 [M–H][–]; HRMS (ES+): *m/z* found 386.0657; C₁₇H₁₇NO₆Na⁺ [M+Na]⁺ requires 386.0674.

6-Hydroxy-7-methoxy-2-((4-methoxyphenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (11c): Method as for **8a** using compound **10c** (150 mg, 0.33 mmol) and Pd/C (10%, 15 mg) in THF (5 mL) and EtOH (5 mL) under hydrogen at RT for 3 h. Flash column chromatography (hexane/EtOAc 2:1) afforded compound **11c** as a colourless powder (74 mg, 62%), mp: 202–204 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.00 (2H, t, *J* = 6.3 Hz), 3.84 (3H, s), 3.85 (3H, s), 4.17 (2H, t, *J* = 6.2 Hz), 6.08 (1H, s), 6.70 (1H, s), 6.95–6.99 (2H, m), 7.45 (1H, s), 7.99–8.03 ppm (2H, m); LC–MS (ES–): *m/z* 362.33 [M–H][–]; HRMS (ES+): *m/z* found 386.0651; C₁₇H₁₇NO₆Na⁺ [M+Na]⁺ requires 386.0674.

6-Hydroxy-7-methoxy-2-((3-chlorophenyl)sulfonyl)-3,4-dihydroisoquinolin-1(2H)-one (11d): Method as for **8a** using compound **10d** (240 mg, 0.53 mmol) and Pd/C (10%, 24 mg) in THF (5 mL) and EtOH (5 mL) under hydrogen at RT for 1 h. Flash column chromatography (hexane to EtOAc) afforded compound **11d** as a colourless solid (158 mg, 82%), mp: 178–181 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.03 (2H, t, *J* = 6.2 Hz), 3.85 (3H, s), 4.19 (2H, t, *J* = 6.3), 6.11 (1H, s), 6.72 (1H, s), 7.44 (1H, s), 7.44–7.50 (1H, m), 7.54–7.59 (1H, m), 7.96–8.00 (1H, m), 8.02 ppm (1H, t, *J* = 1.5 Hz); LC–MS (ES–): *m/z* 366.46 [M–H][–]; HRMS (ES+): *m/z* found 390.0160; C₁₆H₁₄ClNO₅Na⁺ [M+Na]⁺ requires 390.0173.

Methyl 2-((6-hydroxy-7-methoxy-1-oxo-3,4-dihydroisoquinolin-2(1H)-yl)sulfonyl)benzoate (11e): Method as for **8a** using compound **10e** (160 mg, 0.33 mmol) and Pd/C (10%, 90 mg) in THF (5 mL) and EtOH (5 mL) under hydrogen at RT for 3 h. Flash column chromatography (hexane to EtOAc) afforded compound **11e** as a colourless solid (99 mg, 76%), mp: 236–239 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.08 (2H, t, *J* = 6.2 Hz), 3.83 (3H, s), 3.93 (3H, s), 4.19 (2H, t, *J* = 6.2 Hz), 6.08 (1H, s), 6.71 (1H, s), 7.44 (1H, s), 7.60–7.70 (3H, m), 8.54 ppm (1H, dd, *J* = 6.2, 1.7 Hz); LC–MS (APCI-): *m/z* 390.08 [M–H][–]; HRMS (ES+): *m/z* found 414.0609; C₁₈H₁₇NO₇Na⁺ [M+Na]⁺ requires 414.0618.

7-Methoxy-2-((2-methoxyphenyl)sulfonyl)-6-(sulfamoyloxy)-3,4-dihydroisoquinolin-1(2H)-one (12a): Method as for **9a** using compound **11a** (126 mg, 0.35 mmol) and sulfamoyl chloride (0.69 mmol) in anhydrous DMA (1.0 mL) at RT for 22 h. Flash column chromatography (hexane to EtOAc) afforded compound **12a** as a white powder (115 mg, 75%), mp: 195–197 °C. ¹H NMR (270 MHz, [D₆]DMSO): δ = 3.09 (2H, t, *J* = 6.2 Hz), 3.78 (3H, s), 3.89

(3H, s), 4.18 (2H, t, $J=6.2$ Hz), 7.18 (1H, t, $J=7.7$ Hz), 7.24 (1H, d, $J=8.4$ Hz), 7.37 (1H, s), 7.44 (1H, s), 7.68 (1H, td, $J=8.4, 1.6$ Hz), 7.79 (1H, dd, $J=7.8, 1.6$ Hz), 8.14 ppm (2H, s, br); LC-MS (ES⁻): m/z 441.17 [M-H]⁻; HRMS (ES⁺): m/z found 465.0394; C₁₇H₁₈N₂O₈S₂Na⁺ [M+Na]⁺ requires 465.0402.

7-Methoxy-2-((3-methoxyphenyl)sulfonyl)-6-(sulfamoyloxy)-3,4-dihydroisoquinolin-1(2H)-one (12b): Method as for **9a** using compound **11b** (200 mg, 0.55 mmol) and sulfamoyl chloride (2.2 mmol) in anhydrous DMA (3.0 mL) at RT for 24 h. Flash column chromatography (hexane to EtOAc) afforded compound **12b** as a colourless powder (118 mg, 49%), mp: 164–166 °C. ¹H NMR (270 MHz, [D₆]DMSO): $\delta=3.12$ (2H, t, $J=6.1$ Hz), 3.78 (3H, s), 3.84 (3H, s), 4.21 (2H, t, $J=6.1$ Hz), 7.30 (1H, dt, $J=7.6, 2.1$ Hz), 7.36 (1H, s), 7.47 (2H, s, br), 7.52–7.61 (2H, m), 8.15 ppm (2H, s, br); LC-MS (ES⁻): m/z 441.38 [M-H]⁻; HRMS (ES⁺): m/z found 465.0382; C₁₇H₁₈N₂O₈S₂Na⁺ [M+Na]⁺ requires 465.0402.

7-Methoxy-2-((4-methoxyphenyl)sulfonyl)-6-(sulfamoyloxy)-3,4-dihydroisoquinolin-1(2H)-one (12c): Method as for **9a** using compound **11c** (45 mg, 0.13 mmol) and sulfamoyl chloride (0.5 mmol) in anhydrous DMA (1.0 mL) at RT for 18 h. Flash column chromatography (hexane to EtOAc) gave a colourless solid which was stirred in hexane/CH₂Cl₂ to afford compound **12c** as a colourless solid (30 mg, 56%), mp: 177–180 °C. ¹H NMR (400 MHz, [D₆]DMSO): $\delta=3.09$ (2H, t, $J=6.0$ Hz), 3.79 (3H, s), 3.86 (3H, s), 4.17 (2H, t, $J=6.2$ Hz), 7.12–7.16 (2H, m), 7.34 (1H, s), 7.46 (1H, s), 7.94–7.97 (2H, m), 8.13 ppm (2H, s, br); LC-MS (ES⁻): m/z 441.31 [M-H]⁻; HRMS (ES⁺): m/z found 465.0392; C₁₇H₁₈N₂O₈S₂Na⁺ [M+Na]⁺ requires 465.0402.

2-((3-Chlorophenyl)sulfonyl)-7-methoxy-6-(sulfamoyloxy)-3,4-dihydroisoquinolin-1(2H)-one (12d): Method as for **9a** using compound **11d** (121 mg, 0.33 mmol) and sulfamoyl chloride (0.66 mmol) in anhydrous DMA (1.0 mL) at RT for 22 h. Flash column chromatography (hexane to EtOAc) afforded compound **12d** as a colourless solid (132 mg, 90%), mp: 170–173 °C. ¹H NMR (270 MHz, [D₆]DMSO): $\delta=3.14$ (2H, t, $J=6.2$ Hz), 3.79 (3H, s), 4.24 (2H, t, $J=6.2$ Hz), 7.37 (1H, s), 7.48 (1H, s), 7.68 (1H, t, $J=8.0$ Hz), 7.82–7.85 (1H, m), 8.00 (1H, d, $J=8.2$ Hz), 8.06 (1H, t, $J=1.8$ Hz), 8.17 ppm (2H, s); LC-MS (ES⁻): m/z 445.30 [M-H]⁻; HRMS (ES⁺): m/z found 468.9894; C₁₆H₁₅ClN₂O₇S₂Na⁺ [M+Na]⁺ requires 468.9901.

Methyl 2-((7-methoxy-1-oxo-6-(sulfamoyloxy)-3,4-dihydroisoquinolin-2(1H)-yl)sulfonyl)benzoate (12e): Method as for **9a** using compound **11e** (68 mg, 0.17 mmol) and sulfamoyl chloride (0.52 mmol) in anhydrous DMA (1.0 mL) at RT for 20 h. Flash column chromatography (hexane to EtOAc) afforded compound **12e** as a colourless powder (49 mg, 60%), mp: 181–186 °C. ¹H NMR (270 MHz, [D₆]DMSO): $\delta=3.14$ (2H, t, $J=5.8$ Hz), 3.80 (3H, s), 3.88 (3H, s), 4.13 (2H, t, $J=5.8$ Hz), 7.37 (1H, s), 7.49 (1H, s), 7.72–7.87 (3H, m), 8.16 (2H, s, br), 8.32–8.35 ppm (1H, m); LC-MS (APCI⁺): m/z 390.02 [M-SO₂NH₂]⁻; HRMS (ES⁺): m/z found 493.0343; C₁₈H₁₈N₂O₉S₂Na⁺ [M+Na]⁺ requires 493.0346.

6-Benzyloxy-2-(2-methoxybenzoyl)-7-methoxy-1,2,3,4-tetrahydroisoquinoline (14a): Compound **13** (325 mg, 1.2 mmol) was dissolved in CHCl₃ (20 mL) and Et₃N (1.0 mL, 7.2 mmol). 2-Methoxybenzoyl chloride (239 mg, 1.4 mmol) was added portionwise. The reaction mixture was stirred at RT for 16 h and washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 3:1 to 2:3) afforded compound **14a** as a white powder (405 mg, 84%), mp: 92–93 °C. ¹H NMR (270 MHz, CDCl₃): $\delta=2.58$ –2.70 and 2.76–2.86 (2H, m), 3.38–3.48 and 3.74–3.94 (2H, m), 3.72, 3.76, 3.81 and 3.88 (6H, s),

4.21–4.42 and 4.76–4.93 (2H, m), 5.11 (2H, s), 6.38 and 6.61 (1H, s), 6.67 and 6.69 (1H, s), 6.92 (1H, d, $J=8.2$ Hz), 6.99 (1H, t, $J=7.4$ Hz), 7.22–7.46 ppm (7H, m); LC-MS (APCI⁺): m/z 402.51 (M⁺-H), m/z 404.46 [M+H]⁺; HRMS (ES⁺): m/z found 404.1856; C₂₅H₂₆NO₄⁺ [M+H]⁺ requires 404.1856.

6-Benzyloxy-2-(3-methoxybenzoyl)-7-methoxy-1,2,3,4-tetrahydroisoquinoline (14b): Method as for **14a** using compound **13** (404 mg, 1.5 mmol), 3-methoxybenzoyl chloride (0.22 mL, 1.65 mmol) and Et₃N (0.42 mL, 3.0 mmol) in CHCl₃ (10 mL) at RT for 18 h. The reaction mixture was diluted with EtOAc (80 mL) and washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 10:1 to 1:1) afforded compound **14b** as a thick colourless oil (500 mg, 83%). ¹H NMR (270 MHz, CDCl₃): $\delta=2.66$ –2.76 and 2.76–2.88 (2H, m), 3.52–3.64 and 3.72–3.98 (2H, m), 3.80 (6H, s), 4.49 and 4.79 (2H, s), 5.11 (2H, s), 6.40 and 6.69 (1H, br), 6.64 (1H, br), 6.92–7.05 (2H, m), 6.97 (1H, s), 7.26–7.46 ppm (6H, m); LC-MS (APCI⁺): m/z 402.45 (M⁺-H), m/z 404.46 [M+H]⁺; HRMS (ES⁺): m/z found 404.1858; C₂₅H₂₆NO₄⁺ [M+H]⁺ requires 404.1856.

6-Benzyloxy-2-(3-cyanobenzoyl)-7-methoxy-1,2,3,4-tetrahydroisoquinoline (14c): Method as for **14a** using compound **13** (300 mg, 1.1 mmol), 3-cyanobenzoyl chloride (202 mg, 1.22 mmol) and Et₃N (1.0 mL, 7.2 mmol) in CHCl₃ (20 mL) at RT for 2 h. Flash column chromatography (hexane/EtOAc 2:1 to 1:2) afforded compound **14c** as a white powder (325 mg, 74%), mp: 156–157 °C. ¹H NMR (270 MHz, CDCl₃): $\delta=2.70$ –2.87 (2H, br), 3.52–3.57 and 3.91–3.95 (2H, br), 3.79 and 3.97 (3H, s), 4.46 and 4.80 (2H, br), 5.12 (2H, s), 6.40, 6.65 and 6.68 (2H, br), 7.27–7.45 (5H, m), 7.52–7.58 (1H, m), 7.66–7.75 ppm (3H, m); LC-MS (ES⁺): m/z 421.54 [M+Na]⁺.

6-Benzyloxy-2-(4-methoxybenzoyl)-7-methoxy-1,2,3,4-tetrahydroisoquinoline (14d): Method as for **14a** using compound **13** (406 mg, 1.5 mmol), 4-methoxybenzoyl chloride (318 mg, 1.8 mmol) and Et₃N (0.42 mL, 3.0 mmol) in CHCl₃ (30 mL) at RT for 18 h. The reaction mixture was diluted with EtOAc (80 mL) and washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 10:1 to 1:1) afforded compound **14d** as a white powder (520 mg, 85%), mp: 126–127 °C. ¹H NMR (270 MHz, CDCl₃): $\delta=2.77$ (2H, br), 3.67 (2H, br), 3.83 (6H, s), 4.73 (2H, br), 5.11 (2H, s), 6.46 and 6.69 (1H, br), 6.65 (1H, s), 6.92 (2H, dt, $J=8.6, 2.3$ Hz), 7.26–7.45 ppm (7H, m); LC-MS (APCI⁺): m/z 404.53 [M+H]⁺; HRMS (ES⁺): m/z found 404.1858; C₂₅H₂₆NO₄⁺ [M+H]⁺ requires 404.1856.

6-Benzyloxy-2-(3,4-dimethoxybenzoyl)-7-methoxy-1,2,3,4-tetrahydroisoquinoline (14e): Method as for **14a** using compound **13** (300 mg, 1.1 mmol), 3,4-dimethoxybenzoyl chloride (240 mg, 1.2 mmol) and Et₃N (0.5 mL, 3.6 mmol) in CHCl₃ (20 mL) at RT for 18 h. Flash column chromatography (hexane/EtOAc 2:1 to 1:3) afforded compound **14e** as a white powder (370 mg, 77%), mp: 126–127 °C. ¹H NMR (270 MHz, CDCl₃): $\delta=2.77$ (2H, br), 3.83 (2H, br), 3.88 (3H, s), 3.89 (3H, s), 3.90 (3H, s), 4.61 and 4.74 (2H, br), 5.11 (2H, s), 6.65 (1H, s), 6.86 (1H, d, $J=8.9$ Hz), 7.01 (1H, s), 7.03 (1H, dd, $J=8.9, 2.0$ Hz), 7.23–7.46 ppm (6H, m); LC-MS (APCI⁺): m/z 432.48 (M⁺-H), m/z 434.50 [M+H]⁺; HRMS (ES⁺): m/z found 434.1966; C₂₆H₂₈NO₅⁺ [M+H]⁺ requires 434.1962.

6-Benzyloxy-2-(3,5-dimethoxybenzoyl)-7-methoxy-1,2,3,4-tetrahydroisoquinoline (14f): Method as for **14a** using compound **13** (404 mg, 1.5 mmol), 3,5-dimethoxybenzoyl chloride (331 mg, 1.65 mmol) and Et₃N (0.42 mL, 3.0 mmol) in CHCl₃ (20 mL) at RT for 18 h. The reaction mixture was diluted with EtOAc (80 mL) and washed with H₂O and brine, dried (MgSO₄), filtered and concentrated

ed in vacuo. Flash column chromatography (hexane/EtOAc 10:1 to 1:1) afforded compound **14f** as a white powder (520 mg, 85%), mp: 171–172 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.70 and 2.81 (2H, br), 3.58 and 3.92 (2H, br), 3.83 (9H, s), 4.49 and 4.78 (2H, br), 5.11 (2H, s), 6.42, 6.63, 6.66 and 6.68 (2H, br), 6.50 (1H, t, *J* = 2.2 Hz), 6.55 (2H, d, *J* = 2.2 Hz), 7.26–7.45 ppm (5H, m); LC–MS (APCI+): *m/z* 434.43 [*M*+H]⁺; HRMS (ES+): *m/z* found 434.1962; C₂₆H₂₈NO₅⁺ [*M*+H]⁺ requires 434.1962.

6-Benzyloxy-7-methoxy-2-(3,4,5-trimethoxybenzoyl)-1,2,3,4-tetrahydroisoquinoline (14g): Method as for **14a** using compound **13** (808 mg, 3.0 mmol), 3,4,5-trimethoxybenzoyl chloride (765 mg, 3.3 mmol) and Et₃N (2.0 mL, 14.4 mmol) in CHCl₃ (20 mL) at RT for 18 h. The reaction mixture was diluted with CHCl₃ (80 mL) and washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 3:1 to 1:2) afforded compound **14g** as a white powder (1.10 g, 79%), mp: 63–64 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.80 (2H, br), 3.60 and 3.80 (2H, br), 3.85 (6H, s), 3.86 (6H, s), 4.50 and 4.77 (2H, br), 5.12 (2H, s), 6.44 and 6.65 (4H, br), 7.25–7.44 ppm (5H, m); LC–MS (ES+): *m/z* 496.26 ([*M*+Na]⁺, 100%), 464.28 [*M*+H]⁺; HRMS (ES+): *m/z* found 464.2060; C₂₇H₃₀NO₆⁺ [*M*+H]⁺ requires 464.2068.

6-Benzyloxy-2-(2-methoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (15a): Compound **14a** (400 mg, 1.0 mmol), KMnO₄ (0.79 g, 5.0 mmol) and 18-crown-6 (50 mg, 0.19 mmol) were mixed in dichloromethane (50 mL) and the reaction mixture was stirred at RT for 8 h. The reaction mixture was diluted with CHCl₃ and sodium metabisulfite (sat.) was added. The mixture was washed with H₂O and brine, dried with MgSO₄, filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 3:1) afforded compound **15a** as a white powder (170 mg, 40%), mp: 163–164 °C. ¹H NMR (270 MHz, CDCl₃): δ = 2.98 (2H, t, *J* = 6.2 Hz), 3.63 (3H, s), 3.83 (3H, s), 4.19 (2H, t, *J* = 6.2 Hz), 5.22 (2H, s), 6.72 (1H, s), 6.86 (1H, d, *J* = 8.4 Hz), 7.01 (1H, dt, *J* = 8.4, 0.8 Hz), 7.28–7.46 (7H, m), 7.56 ppm (1H, s); LC–MS (ES+): *m/z* 440.49 [*M*+Na]⁺; LC–MS (ES–): *m/z* 416.44 [*M*–H][–]; HRMS (ES+): *m/z* found 418.1635; C₂₅H₂₄NO₅⁺ [*M*+H]⁺ requires 418.1649.

6-Benzyloxy-7-methoxy-2-(3-methoxybenzoyl)-3,4-dihydroisoquinolin-1(2H)-one (15b): Method as for **15a** using compound **14b** (560 mg, 1.38 mmol), KMnO₄ (1.1 g, 6.9 mmol) and 18-crown-6 (70 mg, 0.26 mmol) in dichloromethane (50 mL) at RT for 16 h. Flash column chromatography (hexane/EtOAc 3:1) afforded compound **15b** as a white powder (235 mg, 41%), mp: 141–143 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.03 (2H, t, *J* = 6.2 Hz), 3.81 (3H, s), 3.85 (3H, s), 4.08 (2H, t, *J* = 6.2 Hz), 5.23 (2H, s), 6.73 (1H, s), 7.01 (1H, ddd, *J* = 8.2, 2.7, 1.2 Hz), 7.11–7.15 (2H, m), 7.26–7.45 (6H, m), 7.56 ppm (1H, s); LC–MS (ES+): *m/z* 440.43 [*M*+Na]⁺; LC–MS (ES–): *m/z* 416.38 [*M*–H][–]; HRMS (ES+): *m/z* found 418.1636; C₂₅H₂₄NO₅⁺ [*M*+H]⁺ requires 418.1649.

6-Benzyloxy-2-(3-cyanobenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (15c): Method as for **15a** using compound **14c** (315 mg, 0.79 mmol), KMnO₄ (0.62 g, 3.8 mmol) and 18-crown-6 (40 mg, 0.15 mmol) in dichloromethane (40 mL) at RT for 16 h. Flash column chromatography (hexane/EtOAc 3:2) afforded compound **15c** as a white powder (100 mg, 31%), mp: 195–196 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.05 (2H, t, *J* = 6.2 Hz), 3.82 (3H, s), 3.86 (2H, t, *J* = 6.2 Hz), 5.24 (2H, s), 6.74 (1H, s), 7.30–7.45 (5H, m), 7.49–7.55 (2H, m), 7.74 (1H, ddd, *J* = 7.9, 1.7, 1.5 Hz), 7.79 (1H, ddd, *J* = 7.9, 1.7, 1.2 Hz), 7.82 ppm (1H, d, *J* = 1.5, 1.2 Hz); LC–MS (ES+): *m/z* 435.58 [*M*+Na]⁺; HRMS (ES+): *m/z* found 413.1496; C₂₅H₂₁N₂O₄⁺ [*M*+H]⁺ requires 413.1496.

6-Benzyloxy-2-(4-methoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (15d): Method as for **15a** using compound **14d** (403 mg, 1.0 mmol), KMnO₄ (0.79 g, 5.0 mmol) and 18-crown-6 (53 mg, 0.2 mmol) in dichloromethane (50 mL) at RT for 16 h. Flash column chromatography (hexane/EtOAc 4:1 to 1:1 to EtOAc) afforded compound **15d** as a white powder (188 mg, 45%), mp: 151–152 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.02 (2H, t, *J* = 6.1 Hz), 3.83 (3H, s), 3.86 (3H, s), 4.04 (2H, t, *J* = 6.1 Hz), 5.23 (2H, s), 6.73 (1H, s), 6.88 (2H, d, *J* = 8.7 Hz), 7.30–7.46 (5H, m), 7.58 (1H, s), 7.62 ppm (2H, d, *J* = 8.7 Hz); HRMS (ES+): *m/z* found 418.1651; C₂₅H₂₄NO₅⁺ [*M*+H]⁺ requires 418.1649.

6-Benzyloxy-2-(3,4-dimethoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (15e): Method as for **15a** using compound **14e** (450 mg, 1.04 mmol), KMnO₄ (0.82 g, 5.2 mmol) and 18-crown-6 (50 mg, 0.19 mmol) in dichloromethane (50 mL) at RT for 8 h. Flash column chromatography (hexane/EtOAc 3:1) afforded compound **15e** as a white powder (170 mg, 37%), mp: 200–201 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.04 (2H, t, *J* = 6.2 Hz), 3.87 (3H, s), 3.90 (6H, s), 4.04 (2H, t, *J* = 6.2 Hz), 5.23 (2H, s), 6.74 (1H, s), 6.81 (1H, d, *J* = 8.4 Hz), 7.20 (1H, dd, *J* = 8.4, 2.0 Hz), 7.26 (1H, d, *J* = 2.0 Hz), 7.28–7.46 (5H, m), 7.58 ppm (1H, s); LC–MS (ES+): *m/z* 470.52 [*M*+Na]⁺; LC–MS (ES–): *m/z* 446.47 [*M*–H][–].

6-Benzyloxy-2-(3,5-dimethoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (15f): Method as for **15a** using compound **14f** (433 mg, 1.0 mmol), KMnO₄ (0.79 g, 5.0 mmol) and 18-crown-6 (53 mg, 0.2 mmol) in dichloromethane (50 mL) at RT for 16 h. Flash column chromatography (hexane/EtOAc 10:1 to 4:1) afforded compound **15f** as a white powder (193 mg, 43%), mp: 180–181 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.02 (2H, t, *J* = 6.2 Hz), 3.78 (6H, s), 3.85 (3H, s), 4.06 (2H, t, *J* = 6.2 Hz), 5.23 (2H, s), 6.56 (1H, t, *J* = 2.2 Hz), 6.70 (2H, d, *J* = 2.2 Hz), 6.73 (1H, s), 7.29–7.46 (5H, m), 7.56 ppm (1H, s); LC–MS (APCI+): *m/z* 448.54 [*M*+H]⁺; HRMS (ES+): *m/z* found 448.1756; C₂₆H₂₆NO₆⁺ [*M*+H]⁺ requires 448.1755.

6-Benzyloxy-7-methoxy-2-(3,4,5-trimethoxybenzoyl)-3,4-dihydroisoquinolin-1(2H)-one (15g): Method as for **15a** using compound **14g** (0.85 g, 1.83 mmol) and 18-crown-6 (48 mg, 0.18 mmol) in CHCl₃ (30 mL) at 0 °C for 2 h then at RT 16 h. The reaction mixture was diluted with CHCl₃ and sodium bisulfite (sat.) and HCl (2 M, 1.0 mL, 2.0 mmol) were added. The mixture was washed with H₂O and brine, dried with MgSO₄, filtered and concentrated in vacuo. Flash column chromatography (hexane/EtOAc 3:1 to 1:1) afforded compound **15g** as a white powder (220 mg, 25%), mp: 195–196 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.05 (2H, t, *J* = 6.2 Hz), 3.83 (6H, s), 3.87 (3H, s), 3.87 (3H, s), 4.05 (2H, t, *J* = 6.2 Hz), 5.23 (2H, s), 6.75 (1H, s), 6.84 (2H, s), 7.29–7.46 (5H, m), 7.56 ppm (1H, s); LC–MS (ES+): *m/z* 500.18 ([*M*+Na]⁺, 100%), 478.20 [*M*+H]⁺; HRMS (ES+): *m/z* found 478.1844; C₂₇H₂₈NO₇⁺ [*M*+H]⁺ requires 478.1860.

6-Hydroxy-2-(2-methoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (16a): Method as for **8a** using compound **15a** (145 mg, 0.35 mmol) and Pd/C (10%, 20 mg) in THF (10 mL) and MeOH (10 mL) under hydrogen at RT for 2 h. The residue was stirred in EtOAc (5 mL), filtered and dried in vacuo to afford compound **16a** as a white powder (105 mg, 92%), mp: 199–200 °C. ¹H NMR (270 MHz, [D₆]DMSO): δ = 2.94 (2H, t, *J* = 6.2 Hz), 3.57 (3H, s), 3.74 (3H, s), 4.05 (2H, t, *J* = 6.2 Hz), 6.76 (1H, s), 6.93–7.00 (2H, m), 7.27 (1H, dd, *J* = 7.4, 1.7 Hz), 7.32 (1H, s), 7.34–7.41 (1H, m), 10.21 ppm (1H, s); LC–MS (ES–): *m/z* 326.51 [*M*–H][–]; HRMS (ES+): *m/z* found 328.1167; C₁₈H₁₈NO₅⁺ [*M*+H]⁺ requires 328.1179.

6-Hydroxy-2-(3-methoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (16b): Method as for **8a** using compound **15b** (200 mg, 0.48 mmol) and Pd/C (10%, 25 mg) in THF (10 mL) and MeOH (10 mL) under hydrogen at RT for 1 h. The residue was stirred in EtOAc (5 mL), filtered and dried in vacuo to afford compound **16b** as a white powder (125 mg, 80%), mp: 198–199 °C. ¹H NMR (270 MHz, [D₆]DMSO): δ = 3.03 (2H, t, *J* = 5.9 Hz), 3.75 (3H, s), 3.76 (3H, s), 3.95 (2H, t, *J* = 5.9 Hz), 6.77 (1H, s), 7.05–7.10 (3H, m), 7.32 (1H, dt, *J* = 6.7, 1.0 Hz), 7.34 (1H, s), 10.21 ppm (1H, s); LC–MS (ES–): *m/z* 326.58 [M–H][–]; HRMS (ES+): *m/z* found 328.1168; C₁₈H₁₈NO₅⁺ [M+H]⁺ requires 328.1179.

2-(3-Cyanobenzoyl)-6-hydroxy-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (16c): Method as for **8a** using compound **15c** (155 mg, 0.38 mmol) and Pd/C (10%, 20 mg) in THF (10 mL) and MeOH (10 mL) under hydrogen at RT for 1 h. The residue was dissolved in hot EtOAc (5 mL), filtered and concentrated then stirred in Et₂O (20 mL), filtered and dried in vacuo to afford compound **16c** as a white powder (71 mg, 59%), mp: 197–198 °C. ¹H NMR (270 MHz, [D₆]acetone): δ = 3.14 (2H, t, *J* = 6.2 Hz), 3.85 (3H, s), 4.08 (2H, t, *J* = 6.2 Hz), 6.83 (1H, s), 7.42 (1H, s), 7.65 (1H, dd, *J* = 8.2, 7.6 Hz), 7.86–7.91 (2H, m), 8.00–8.06 (1H, m), 8.71 ppm (1H, s, br); LC–MS (ES–): *m/z* 321.47 [M+H]⁺; HRMS (ES+): *m/z* found 323.1012; C₁₈H₁₅N₂O₄⁺ [M+H]⁺ requires 323.1026.

6-Hydroxy-2-(4-methoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (16d): Method as for **8a** using compound **15d** (96 mg, 0.23 mmol) and Pd/C (10%, 20 mg) in THF (10 mL) and EtOH (10 mL) under hydrogen at RT for 0.5 h. The residue was stirred in EtOAc (10 mL) and hexane (2 mL), filtered and dried in vacuo to afford compound **16d** as a white powder (72 mg, 96%), mp: 224–225 °C. ¹H NMR (270 MHz, [D₆]DMSO): δ = 3.01 (2H, t, *J* = 5.7 Hz), 3.77 (3H, s), 3.82 (3H, s), 3.90 (2H, t, *J* = 5.7 Hz), 6.77 (1H, s), 6.95 (2H, d, *J* = 8.6 Hz), 7.36 (1H, s), 7.55 (2H, d, *J* = 8.6 Hz), 10.19 ppm (1H, s, br); LC–MS (APCI+): *m/z* 328.46 [M+H]⁺; HRMS (ES+): *m/z* found 328.1181; C₁₈H₁₈NO₅⁺ [M+H]⁺ requires 328.1179.

6-Hydroxy-2-(3,4-dimethoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (16e): Method as for **8a** using compound **15e** (165 mg, 0.37 mmol) and Pd/C (10%, 20 mg) in THF (150 mL) under hydrogen at RT for 2 h. Crystallisation from EtOAc (5 mL) afforded compound **16e** as a white powder (115 mg, 87%), mp: 199–200 °C. ¹H NMR (400 MHz, [D₆]DMSO): δ = 3.02 (2H, t, *J* = 5.9 Hz), 3.75 (3H, s), 3.76 (3H, s), 3.81 (3H, s), 3.90 (2H, t, *J* = 5.9 Hz), 6.77 (1H, s), 6.96 (1H, d, *J* = 9.2 Hz), 7.15–7.18 (2H, m), 7.36 (1H, s), 10.20 ppm (1H, s, br); LC–MS (ES–): *m/z* 356.60 [M–H][–]; HRMS (ES+): *m/z* found 358.1273; C₁₉H₂₀NO₆⁺ [M+H]⁺ requires 358.1285.

6-Hydroxy-2-(3,5-dimethoxybenzoyl)-7-methoxy-3,4-dihydroisoquinolin-1(2H)-one (16f): Method as for **8a** using compound **15f** (200 mg, 0.45 mmol) and Pd/C (10%, 30 mg) in THF (15 mL) and MeOH (15 mL) under hydrogen at RT for 3 h. The residue was stirred in Et₂O, filtered and dried in vacuo to afford compound **16f** as a white powder (130 mg, 81%), mp: 198–199 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.05 (2H, t, *J* = 6.2 Hz), 3.78 (6H, s), 3.86 (3H, s), 4.07 (2H, t, *J* = 6.2 Hz), 6.16 (1H, s), 6.56 (1H, t, *J* = 2.2 Hz), 6.71 (2H, d, *J* = 2.2 Hz), 6.79 (1H, s), 7.54 ppm (1H, s); LC–MS (APCI+): *m/z* 358.25 [M+H]⁺; HRMS (ES+): *m/z* found 358.1285; C₁₉H₂₀NO₆⁺ [M+H]⁺ requires 358.1285.

6-Hydroxy-7-methoxy-2-(3,4,5-trimethoxybenzoyl)-3,4-dihydroisoquinolin-1(2H)-one (16g): Method as for **8a** using compound **15g** (180 mg, 0.38 mmol) and Pd/C (10%, 40 mg) in THF (20 mL) and MeOH (20 mL) under hydrogen at RT for 24 h. The residue was stirred in Et₂O, filtered, washed with Et₂O and dried in vacuo to

afford compound **16g** as a white powder (140 mg, 96%), mp: 175–176 °C. ¹H NMR (270 MHz, CDCl₃): δ = 3.07 (2H, t, *J* = 6.4 Hz), 3.83 (6H, s), 3.88 (3H, s), 3.89 (3H, s), 4.06 (2H, t, *J* = 6.4 Hz), 6.18 (1H, s), 6.80 (1H, s), 6.85 (2H, s), 7.55 ppm (1H, s); HRMS (ES+): *m/z* found 388.1380; C₂₀H₂₂NO₇⁺ [M+H]⁺ requires 388.1391; LC–MS (ES–): *m/z* 386.24 [M–H][–].

2-(2-Methoxybenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17a): Method as for **9a** using compound **16a** (70 mg, 0.21 mmol) and sulfamoyl chloride (0.43 mmol) in DMA (1.0 mL) at RT for 16 h. The residue was stirred in Et₂O (30 mL), filtered and dried in vacuo to afford compound **17a** as a white powder (65 mg, 75%), mp: 165–166 °C. ¹H NMR (400 MHz, [D₆]DMSO): δ = 3.05 (2H, t, *J* = 6.2 Hz), 3.60 (3H, s), 3.80 (3H, s), 4.11 (2H, t, *J* = 6.2 Hz), 6.96–7.03 (2H, m), 7.31 (1H, dd, *J* = 7.7, 1.7 Hz), 7.39 (1H, s), 7.41 (1H, dt, *J* = 8.4, 1.7 Hz), 7.53 (1H, s), 8.18 ppm (2H, s, br); LC–MS (ES–): *m/z* 405.50 [M–H][–]; HRMS (ES+): *m/z* found 407.0898; C₁₈H₁₉N₂O₇S⁺ [M+H]⁺ requires 407.0907.

2-(3-Methoxybenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17b): Method as for **9a** using compound **16b** (80 mg, 0.24 mmol) and sulfamoyl chloride (0.48 mmol) in DMA (1.0 mL) at RT for 16 h. The residue was stirred in Et₂O (30 mL), filtered and dried in vacuo to afford compound **17b** as a white powder (75 mg, 77%), mp: 173–174 °C. ¹H NMR (400 MHz, [D₆]DMSO): δ = 3.15 (2H, t, *J* = 5.9 Hz), 3.77 (3H, s), 3.81 (3H, s), 4.01 (2H, t, *J* = 5.9 Hz), 7.08–7.18 (3H, m), 7.31–7.36 (1H, m), 7.41 (1H, s), 7.57 (1H, s), 8.18 ppm (2H, s, br); LC–MS (ES–): *m/z* 405.43 [M–H][–]; HRMS (ES+): *m/z* found 407.0895; C₁₈H₁₉N₂O₇S⁺ [M+H]⁺ requires 407.0907.

2-(3-Cyanobenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17c): Method as for **9a** using compound **16c** (50 mg, 0.16 mmol) and sulfamoyl chloride (0.40 mmol) in DMA (0.5 mL) at RT for 16 h. Crystallisation from EtOAc afforded compound **17c** as a white powder (45 mg, 70%), mp: 171–172 °C. ¹H NMR (270 MHz, [D₆]acetone): δ = 3.23 (2H, t, *J* = 6.2 Hz), 3.88 (3H, s), 4.14 (2H, t, *J* = 6.2 Hz), 7.24 (2H, s, br), 7.40 (1H, s), 7.58 (1H, s), 7.69 (1H, dt, *J* = 7.9, 0.8 Hz), 7.91–7.98 (2H, m), 8.06 ppm (1H, dt, *J* = 1.8, 0.8 Hz); LC–MS (ES–): *m/z* 400.53 [M–H][–]; HRMS (ES+): *m/z* found 402.0739; C₁₈H₁₆N₃O₆S⁺ [M+H]⁺ requires 402.0754.

2-(4-Methoxybenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17d): Method as for **9a** using compound **16d** (90 mg, 0.28 mmol) and sulfamoyl chloride (0.55 mmol) in DMA (1.0 mL) at RT for 18 h. Flash column chromatography (hexane/EtOAc 10:1 to 4:1) afforded compound **17d** as a white powder (80 mg, 71%), mp: 171–172 °C. ¹H NMR (270 MHz, CDCl₃/[D₄]MeOH 5:1): δ = 3.08 (2H, s, br), 3.10 (2H, t, *J* = 6.4 Hz), 3.83 (3H, s), 3.85 (3H, s), 4.03 (2H, t, *J* = 6.4 Hz), 6.88 (2H, dt, *J* = 8.9, 2.5 Hz), 7.31 (1H, s), 7.61 (2H, dt, *J* = 8.9, 2.5 Hz), 7.67 ppm (1H, s); LC–MS (APCI+): *m/z* 407.42 [M+H]⁺; HRMS (ES+): *m/z* found 407.0904; C₁₈H₁₉N₂O₇S⁺ [M+H]⁺ requires 407.0907.

2-(3,4-Dimethoxybenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17e): Method as for **9a** using compound **16e** (71 mg, 0.20 mmol) and sulfamoyl chloride (0.40 mmol) in DMA (1.0 mL) at RT for 16 h. After addition of H₂O (5 mL) the reaction mixture was extracted into EtOAc (2 × 50 mL), the organic layers washed with H₂O and brine, then dried (MgSO₄) and evaporated. The residue was stirred in EtOAc (5 mL), filtered and dried in vacuo to afford compound **17e** as a white powder (55 mg, 77%), mp: 180–181 °C. ¹H NMR (270 MHz, [D₆]acetone): δ = 3.19 (2H, t, *J* = 6.2 Hz), 3.81 (3H, s), 3.87 (3H, s), 3.88 (3H, s), 4.03 (2H, t, *J* = 6.2 Hz), 6.96 (1H, d, *J* = 8.9 Hz), 7.21 (2H, s, br), 7.28 (1H, s), 7.30

(1H, d, $J=8.9$ Hz), 7.39 (1H, s), 7.62 ppm (1H, s); LC-MS (ES⁻): m/z 435.44 [M-H]⁻; HRMS (ES⁺): m/z found 437.1011; C₁₉H₂₁N₂O₉S⁺ [M+H]⁺ requires 437.1013.

2-(3,5-Dimethoxybenzoyl)-7-methoxy-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17f): Method as for **9a** using compound **16f** (80 mg, 0.22 mmol) and sulfamoyl chloride (0.45 mmol) in DMA (1.0 mL) at RT for 18 h. The residue was stirred in Et₂O, filtered and dried in vacuo to afford compound **17f** as a white powder (76 mg, 78%), mp: 169–170 °C. ¹H NMR (270 MHz, (CDCl₃/[D₄]MeOH 10:1): $\delta=2.46$ (2H, s, br), 3.08 (2H, t, $J=6.2$ Hz), 3.75 (6H, s), 3.82 (3H, s), 4.05 (2H, t, $J=6.2$ Hz), 6.56 (1H, t, $J=2.3$ Hz), 6.68 (2H, d, $J=2.3$ Hz), 7.29 (1H, s), 7.65 ppm (1H, s); LC-MS (APCI⁺): m/z 437.39 [M+H]⁺; HRMS (ES⁺): m/z found 437.1011; C₁₉H₂₁N₂O₉S⁺ [M+H]⁺ requires 437.1013.

7-Methoxy-2-(3,4,5-trimethoxybenzoyl)-6-sulfamoyloxy-3,4-dihydroisoquinolin-1(2H)-one (17g): Method as for **9a** using compound **16g** (80 mg, 0.21 mmol) and sulfamoyl chloride (0.6 mmol) in DMA (1.0 mL) at RT for 24 h. H₂O (20 mL) was added and the mixture extracted with EtOAc (50 mL) and THF (50 mL). The organic layer was washed with H₂O and brine, dried (MgSO₄), filtered and concentrated in vacuo. The residue was stirred in Et₂O (5 mL) and EtOAc (5 mL), filtered and dried to afford compound **17g** as a white powder (80 mg, 82%), mp: 196–197 °C. ¹H NMR (270 MHz, [D₆]DMSO): $\delta=3.16$ (2H, t, $J=5.7$ Hz), 3.72 (3H, s), 3.75 (6H, s), 3.81 (3H, s), 3.98 (2H, t, $J=5.7$ Hz), 6.90 (2H, s), 7.40 (1H, s), 7.57 (1H, s), 8.18 ppm (2H, s, br); LC-MS (ES⁻): m/z 465.22 [M-H]⁻; HRMS (ES⁺): m/z found 467.1109; C₂₀H₂₃N₂O₉S⁺ [M+H]⁺ requires 467.1119.

2-(3,4-Dimethoxybenzoyl)-6-hydroxy-7-methoxy-1,2,3,4-tetrahydroisoquinoline (18a): Method as for **8a** using **14e** (403 mg, 0.93 mmol) and Pd/C (10%, 40 mg) in THF (15 mL) and MeOH (15 mL) under hydrogen at RT for 1.5 h. The resulting solid was stirred in Et₂O, filtered and dried in vacuo to afford compound **18a** as a white solid (300 mg, 94%), mp: 132–133 °C. ¹H NMR (270 MHz, CDCl₃): $\delta=2.80$ (2H, br), 3.83 (2H, br), 3.88 (3H, s), 3.90 (6H, s), 4.61 and 4.72 (2H, br), 5.62 (1H, s), 6.60 (1H, br), 6.69 (1H, br), 6.86 (1H, d, $J=8.9$ Hz), 7.02 (1H, s), 7.03 (1H, d, $J=8.9$ Hz), 8.84 ppm (1H, s); LC-MS (APCI⁺): m/z 344.27 [M+H]⁺; HRMS (ES⁺): m/z found 344.1493; C₁₉H₂₂NO₅⁺ [M+H]⁺ requires 344.1492.

2-(3,5-Dimethoxybenzoyl)-6-hydroxy-7-methoxy-1,2,3,4-tetrahydroisoquinoline (18b): Method as for **8a** using **14f** (390 mg, 0.90 mmol) and Pd/C (10%, 40 mg) in THF (15 mL) and MeOH (15 mL) under hydrogen at RT for 2 h. The resulting solid was stirred in Et₂O, filtered and dried in vacuo to afford compound **18b** as a white solid (290 mg, 94%), mp: 136–137 °C. ¹H NMR (270 MHz, CDCl₃): $\delta=2.73$ and 2.84 (2H, br), 3.59 and 3.93 (2H, br), 3.79 (9H, s), 4.48 and 4.77 (2H, br), 5.58 (1H, s), 6.38 and 6.64 (1H, br), 6.50 (1H, t, $J=2.2$ Hz), 6.55 (2H, d, $J=2.2$ Hz), 6.68 ppm (1H, br); LC-MS (APCI⁺): m/z 344.33 [M+H]⁺; HRMS (ES⁺): m/z found 344.1493; C₁₉H₂₂NO₅⁺ [M+H]⁺ requires 344.1492.

2-(3,4-Dimethoxybenzoyl)-7-methoxy-6-sulfamoyloxy-1,2,3,4-tetrahydroisoquinoline (19a): Method as for **9a** using **18a** (75 mg, 0.24 mmol) and sulfamoyl chloride (0.48 mmol) in anhydrous DMA (1.0 mL) at RT for 24 h. The residue was stirred in Et₂O, filtered and dried in vacuo to afford compound **19a** as a white powder (80 mg, 85%), mp: 155–156 °C. ¹H NMR (270 MHz, CDCl₃/[D₄]MeOH 5:1): $\delta=2.73$ (2H, br), 3.59 and 3.72 (2H, br), 3.77 (3H, s), 3.79 (6H, s), 4.52 and 4.65 (2H, br), 6.61 and 6.70 (1H, br), 6.80 (1H, d, $J=8.2$ Hz), 6.87 (1H, d, $J=2.0$ Hz), 6.90 (1H, dd, $J=8.2, 2.0$ Hz), 7.03 ppm (1H, s); LC-MS (APCI⁺): m/z 423.42 [M+H]⁺; HRMS

(ES⁺): m/z found 423.1220; C₁₉H₂₃N₂O₇S⁺ [M+H]⁺ requires 423.1220.

2-(3,5-Dimethoxybenzoyl)-7-methoxy-6-sulfamoyloxy-1,2,3,4-tetrahydroisoquinoline (19b): Method as for **9a** using **18b** (75 mg, 0.24 mmol) and sulfamoyl chloride (0.48 mmol) in anhydrous DMA (1.0 mL) at RT for 24 h. The residue was stirred in Et₂O, filtered and dried in vacuo to afford compound **19b** as a white powder (80 mg, 85%), mp: 170–171 °C. ¹H NMR (270 MHz, CDCl₃/[D₄]MeOH 5:1): $\delta=2.70$ and 2.80 (2H, t, $J=5.3$ Hz), 3.55 and 3.82 (2H, t, $J=5.3$ Hz), 3.63 (2H, s), 3.71 (9H, s), 4.46 and 4.72 (2H, br), 6.44 (2H, s), 6.46 and 6.74 (1H, br), 7.05 ppm (1H, br); LC-MS (APCI⁺): m/z 423.35 [M+H]⁺; HRMS (ES⁺): m/z found 423.1224; C₁₉H₂₃N₂O₇S⁺ [M+H]⁺ requires 423.1220.

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