



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



Available online at
ScienceDirect
www.sciencedirect.com

Elsevier Masson France
EM|consulte
www.em-consulte.com/en



REVIEW

Risks of viral contamination in healthcare professionals during laparoscopy in the Covid-19 pandemic



J. Veziant^a, N. Bourdel^b, K. Slim^{a,*}

^a *Digestive surgery department, University Hospital of Clermont-Ferrand, 63003 Clermont-Ferrand, France*

^b *Gynaecology Department, University Hospital of Clermont-Ferrand, 63003 Clermont-Ferrand, France*

Available online 17 April 2020

KEYWORDS

Virus;
Viral contamination;
Laparoscopy;
SARS-CoV-2

Summary The Covid-19 pandemic has markedly changed our practices. This article analyses the risks of contamination among healthcare professionals (HCPs) during laparoscopic surgery on patients with Covid-19. Harmful effects of aerosols from a pneumoperitoneum, with the virus present, have not yet been quantified. Measures for the protection of HCPs are an extrapolation of those taken during other epidemics. They must still be mandatory to minimise the risk of viral contamination. Protection measures include personal protection equipment for HCPs, adaptation of surgical technique (method for obtaining pneumoperitoneum, filters, preferred intracorporeal anastomosis, precautions during the exsufflation of the pneumoperitoneum), and organisation of the operating room.

© 2020 Elsevier Masson SAS. All rights reserved.

Introduction

Contamination of healthcare professionals (HCPs) during surgery has been known for many years and is well described. Before the laparoscopy era, the source was essentially surgical fumes produced by heat-producing techniques such as electrosurgery (mono- or bipolar), lasers or ultrasound [1]. These peroperative risks have been abundantly addressed in the literature, which we need not detail here. Depending on the case, surgical smoke can contain water vapour (95%), inorganic pollutants (CO, CO₂), organic pollutants (hydrocarbons, hydrocyanic acid, aldehydes), and

biological pollutants such as cells (some cancerous), bacteria and fragments of viral DNA. Laparoscopy has added a further source of airborne pollution, namely aerosols generated by pneumoperitoneum gas flow. This article specifically concerns the risk of viral contamination during laparoscopy in the setting of the current Covid-19 epidemic. Our purpose here is to analyse the literature data on the viral contamination risk for HCPs by the SARS-CoV-2 coronavirus responsible for the Covid-19 epidemic and discuss means of protection and prevention.

Risk of viral contamination during surgical care

Several viruses have been implicated to different degrees in the contamination of HCPs during surgery: human immunodeficiency virus (HIV), hepatitis B virus (HBV), bovine

* Corresponding author at: Digestive surgery department, University Hospital of Clermont-Ferrand, CHU Estaing, 1, place Lucie-et-Raymond-Aubrac, 63003 Clermont-Ferrand, France.

E-mail address: kslim@chu-clermontferrand.fr (K. Slim).

papilloma virus and human papilloma virus (HPV). Most of the studies published on the risk of transmission have used *in vitro* analyses. Several studies have focused on HPV (during the treatment of warts or condylomas) with cases of laryngeal papillomatosis, recognised as an occupational illness in one nurse [2]. In the 2000s, it was difficult for researchers to measure the biological activity of viral DNA in order to assess its infectiousness [3]. However, viable HIVs have been found in cell cultures, especially when aerosol-generating devices are used, although the viability of HIVs is still being debated, and the potential risk of contamination by smoke has not been quantifiable. [4,5].

For both HPV and HIV, most authors conclude, however, that it is preferable to take all necessary precautions against surgical smoke when operating on infected patients.

There is to our knowledge no large-scale epidemiological study that proves a direct link between a patient's viral load and the contamination of HCPs by emitted surgical smoke. It should also be noted that the published studies have mainly been on surgical smoke from operations on skin or genital lesions. Few studies have evaluated the risk of contamination of HCPs during laparoscopy.

One small-sample study ($n=11$) analysed the presence of HBV in surgical smoke during conventional and robot-assisted laparoscopies [6]. The aerosol from the pneumoperitoneum was trapped in a filter, where the authors found HBV in 10 instances out of 11. This study thus showed the presence of HBV in pneumoperitoneum gas during surgical laparoscopy. The subject matter of this study comes closest to the issue addressed in this article.

Special features of laparoscopy

Laparoscopic surgery requires the creation and maintenance of an efficient pneumoperitoneum. There is thus a permanent risk of an aerosol effect through gas leaks or at exsufflation. Moreover, the ultrasonic systems often used do not produce enough heat to deactivate the virus. These concerns are confirmed by a recent experimental study showing that after 10 minutes of laparoscopic dissection by electrosurgery or ultrasound, the concentration of particles measuring 0.3–0.5 μm was higher with laparoscopy than with laparotomy [7]. Owing to the low rate of replacement of the pneumoperitoneum gas, leaked aerosol may thus contain high concentrations of suspended viruses [8]. This suggests that the risk of contamination of HCPs may be greater in laparoscopy than in laparotomy, particularly if accidental gas leakage occurs or exsufflation is poorly controlled. Other authors [9] have claimed the opposite, arguing that the closed surgical site (relative to laparotomy) lowers the risk of contamination, and that there is no hard evidence yet that the viruses are viable or that they are actually transmitted during laparoscopy.

All these studies have focused on HIV and HBV and have shown rather low peritoneal contamination. No study to date has considered SARS-CoV-2. Studies have evaluated contamination risk for similar earlier viruses (Middle East coronavirus MERS CoV) but their results cannot be extrapolated to laparoscopy [10]. There is no expert consensus on the actual or extrapolated presence of ambient SARS-CoV-2 in the pneumoperitoneum as factual evidence is lacking [11].

Properties of the SARS-CoV-2 coronavirus

General

The SARS-CoV-2 virus responsible for the Covid-19 pandemic is an RNA virus 0.06–0.14 μm in size. In comparison, the HIV measures 0.12 μm , the HBV measures 0.04 μm , and bacteria about 0.30 μm . The RNA of SARS-CoV-2 is very long (30 kb), making it the largest RNA virus known so far, larger than flu and previous SARS viruses [12]. In infected patients it is found in the oropharynx, the respiratory tract and the whole of the digestive tract. It is not found in urine or cerebrospinal fluid.

Ultrastructure

Like other coronaviruses, SRAS-CoV-2 is made up of four proteins [13,14].

Spike (S) protein forms the namesake crown-like protuberances that are essential to attach the virus to the host cell and determine which cells it targets. Envelope (E) protein forms the envelope. Membrane (M) protein forms the membrane. Nucleocapsid (N) protein protects the viral RNA. Proteins S, E and M together enclose the virus.

Once the virus has penetrated the host cell, its RNA is released, and copies are produced. The resulting virions burst out and spread to other cells, where the process is repeated.

Survival of the virus outside an infected organism [15,16]

Coronaviruses remain infectious for 2–3 h in air, and up to 9 days on smooth non-porous surfaces (stainless steel, plastic, ceramics, glass). By contrast, they do not survive on copper, brass or bronze, which are biocidal.

SARS-CoV-2 has a half-life (50% of viruses inactivated) of 13 h on stainless steel and 16 h on polypropylene. The virus is short-lived on paper and most non-waterproof textiles.

SRAS-CoV-2 is broken down by soap, ethanol-containing gels and solutions (62–71% ethanol), and household disinfectants such as hypochlorite bleach (diluted to 0.1%).

No contamination of HCPs by SARS-CoV-2 during a laparoscopy has yet been reported.

Although the strength of evidence is still low, cautionary measures are in order. All the available scientific and clinical data must be used to ensure the protection of HCPs and prevent viral contamination.

Means to protect HCPs

Means to protect HCPs must be implemented whenever the laparoscopy is performed in a patient infected with SARS-CoV-2, or in the context of the Covid-19 epidemic in a patient with fever or a recent cough, or who has spent time in the last two weeks in a location where the epidemic is active, or who has been in contact with infected persons [17]. Some authors even advocate extending testing for SARS-CoV-2 to all patients awaiting surgery whether or not they have Covid-19 symptoms [8,18]. Measures must include personal protection equipment (PPE) for HCPs, laparoscopy equipment, surgical technique and operating room organisation.

Table 1 Technical measures for preventing contamination of HCPs by SARS-CoV-2.

- Prefer the "closed" technique for obtaining pneumoperitoneum
- Reduce the pneumoperitoneum pressure as much as possible (without compromising safety)
- Reduce the power of electrosurgery and ultrasonic dissection
- Systematically use laparoscopic smoke aspiration systems
- Systematically use particle filters
- Prefer intracorporeal anastomosis
- Extract excised tissue after complete emptying of the pneumoperitoneum
- Fully aspirate the pneumoperitoneum before removing the last trocar

Table 2 Organisational measures for preventing contamination of HCPs by SRAS-CoV-2.

- Train HCPs in protection measures
- Set up separate circuits for patients with Covid-19 or suspected of having Covid-19 and for certainly non-Covid-19 patients
- Have as few HCPs as possible in operating rooms
- Avoid movement or changing of HCPs during laparoscopy
- Ventilate the operating room
- Manage waste appropriately during and after laparoscopy
- Encourage the surgical team to leave the operating room during the intubation and extubation phases
- Take general hygiene measures (hand-washing, cleaning of furniture and instruments, etc.)

Personal protection equipment for HCPs

The surgery team (surgeon, assistant, scrub nurse or circulating nurse) seem to be less exposed than the anaesthesia team (anaesthetist and anaesthetist's nurse), who are in direct contact with the patient's upper airways, which are the main source of contamination [19]. PPE is only one component of the protection measures. It must be used by every member of the HCP team according to the World Health Organisation (WHO) guidelines [20]. PPE includes long-sleeved fluid protection gowns, lined gloves, goggles (or visors, especially for the anaesthesia teams) [19], and masks.

Conventional surgical masks do not provide sufficient protection in an operating room with a risk of aerosols rich in viral droplets. The N95 respirator masks regulated by the US National Institute for Occupational Safety and Health (NIOSH) are not EC-approved. The FFP2 and FFP3 filtering facepiece masks are more efficient as regards the filtration of aerosols (of saline solution or paraffin oils), with an efficiency of 94% for FFP2 and 99% for FFP3. FFP2 masks meet the filter efficiency criteria of the NIOSH [21]. To be effective, these masks must, however, be used correctly and be properly fitted.

Laparoscopy equipment and surgical technique

The risk of contamination of HCPs is highest during the insertion of trocars, extraction of the excised tissues and removal of trocars at the end of the operation. Technical measures are summarised in **Table 1**.

As the virus can be present in the digestive tract [22], intracorporeal anastomosis (particularly in colorectal surgery) is to be preferred because it reduces the risk of contamination of the HCPs.

Lastly, there is no data in the literature suggesting that laparoscopy should be systematically replaced by laparotomy during the Covid-19 epidemic.

Organisational measures

The first measure is to raise awareness among HCPs of the risk of contamination and train them in implementing prevention measures. We need not dwell on the now well-known general sanitising measures: frequent hand-washing, perma-

gent wearing of masks, and frequent washing of furniture and floors according to the general rules of hospital hygiene.

The main organisational measures are summarised in **Table 2**. These measures are to be implemented jointly with the hygiene and administration departments of each hospital.

Disclosure of interest

NB declares competing interest with Gedeon Richter, Storz. KS has competing interest with Sanofi, MSD, FSK, B-Braun, Baxter. JV declares that he has no competing interest.

References

- [1] https://ww1.issa.int/sites/default/files/documents/prevention/1-Fumes_chirurgicales.fr-36173.pdf. (retrieved 7 April 2020).
- [2] Calero L, Brusis T. Laryngeal papillomatosis – first recognition in Germany as an occupational disease in an operating room nurse. *Laryngorhinootologie* 2003;82:790–3.
- [3] Garden JM, O'Banion MK, Bakus AD, Olson C. Viral disease transmitted by laser-generated plume (aerosol). *Arch Dermatol* 2002;138:1303–7.
- [4] Johnson GK, Robinson WS. Human immunodeficiency virus-1 (HIV-1) in the vapours of surgical power instruments. *J Med Virol* 1991;33:47–50.
- [5] Hensman C, Baty D, Willis RG, Cuschieri A. Chemical composition of smoke produced by high-frequency electrosurgery in a closed gaseous environment. An in vitro study. *Surg Endosc* 1998;12:1017–9.
- [6] Kwak HD, Kim SH, Seo YS, Song KJ. Detecting hepatitis B virus in surgical smoke emitted during laparoscopic surgery. *Occup Environ Med* 2016;73:857–63.
- [7] Li CI, Pai JY, Chen CH. Characterisation of smoke generated during the use of surgical knife in laparotomy surgeries. *J Air Waste Manag Assoc* 2020;70:324–32.
- [8] Zheng MH, Boni L, Fingerhut A. Minimally invasive surgery and the novel Coronavirus outbreak: lessons learned in China and Italy. *Ann Surg* 2020, <http://dx.doi.org/10.1097/SLA.0000000000003924>.
- [9] <https://esge.org/wp-content/uploads/2020/03/Covid19StatementESGE.pdf>. (retrieved 7 April 2020).
- [10] de Wit E, van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: Recent insights into emergent coronaviruses. *Nat Rev Microbiol* 2016;5:523–34.
- [11] Lewis D. Is the coronavirus airborne? Experts can't agree. *Nature* 2020, <http://dx.doi.org/10.1038/d41586-020-00974-w>.

- [12] Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel Coronavirus from patients with pneumonia in China, 2019. *N Engl J Med* 2020;382:727–33.
- [13] Walls AC, Park YJ, Tortorici MA, Wall A, McGuire AT, Veesler D. Structure, function, and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell* 2020, <http://dx.doi.org/10.1016/j.cell.2020.02.058>.
- [14] Kannan S, Shaik Syed Ali P, Sheeza A, Hemalatha K. COVID-19 (Novel Coronavirus 2019) – recent trends. *Eur Rev Med Pharmacol Sci* 2020;24:2006–11.
- [15] <http://www.inrs.fr/actualites/coronavirus-SARS-CoV-2-COVID-19.html> (accessed on 7 avril 2020).
- [16] Kampf G, Todt D, Pfaender S, Steinmann E. Persistence of coronaviruses on inanimate surfaces and their inactivation with biocidal agents. *J Hosp Infect* 2020;104:246–51.
- [17] Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020;395:497–506.
- [18] <https://www.sages.org/recommendations-surgical-response-covid-19/> (accessed on 7 avril 2020).
- [19] Wax RS, Christian MD. Practical recommendations for critical care and anesthesiology teams caring for novel coronavirus (2019-nCoV) patients. *Can J Anaesth* 2020, <http://dx.doi.org/10.1007/s12630-020-01591-x>.
- [20] <https://apps.who.int/iris/bitstream/handle/10665/331498/WHO-2019-nCoV-ICPPE-use-2020.2-eng.pdf> (accessed on 7 avril 2020).
- [21] <https://www.cdc.gov/niosh/index.htm> (accessed on 7 avril 2020).
- [22] Yeo C, Kaushal S, Yeo D. Enteric involvement of coronaviruses: is faecal-oral transmission of SARS-CoV-2 possible? *Lancet Gastroenterol Hepatol* 2020;5:335–7.