Original Article

Clinical Chemistry



Ann Lab Med 2016;36:223-229 http://dx.doi.org/10.3343/alm.2016.36.3.223 ISSN 2234-3806 eISSN 2234-3814

ANNALS OF LABORATORY MEDICINE

Plasma Levels and Diagnostic Utility of Macrophage Colony-Stimulating Factor, Matrix Metalloproteinase-9, and Tissue Inhibitor of Metalloproteinases-1 as New Biomarkers of Breast Cancer

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Background: Macrophage colony-stimulating factor (M-CSF), matrix metalloproteinase-9 (MMP-9), and its specific tissue inhibitor - tissue inhibitor of metalloproteinases-1 (TIMP-1) may play an important role in the pathogenesis and spread of cancer. We investigated the plasma levels of M-CSF, MMP-9, and TIMP-1 in comparison with a commonly accepted tumor marker CA 15-3 in breast cancer patients and in control groups.

Methods: The cohort included 110 breast cancer patients in groups at stages I-IV. The control group consisted of 50 healthy volunteers and 50 benign tumor patients. Plasma levels of M-CSF, MMP-9, and TIMP-1 were determined by using ELISA, while CA 15-3 concentrations were determined by using chemiluminescent microparticle immunoassay (CMIA).

Results: The results showed significant differences in concentrations of the analyzed parameters and in levels of CA 15-3 between the groups of breast cancer patients and the two control groups. Diagnosis using these markers was equal to that using CA 15-3 in terms of sensitivity, predictive values of positive and negativetest results (PPV, NPV) and area under the ROC curve (AUC) in the studied groups. The diagnostic specificities of MMP-9, TIMP-1, M-CSF, and CA 15-3 showed equally high values (95%). The combined use of all tested parameters with CA 15-3 resulted in increased sensitivity, NPV, and AUC, especially in the combination of M-CSF with tumor markers (76%, 64%, and 0.8653).

Conclusions: These findings suggest the tested parameters are useful in the diagnosis of breast cancer patients (except stage I), when combined with CA 15-3.

Key Words: Breast cancer, CA 15-3, M-CSF, MMP-9, TIMP-1

Received: September 1, 2015 Revision received: January 26, 2016 Accepted: January 29, 2016

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INTRODUCTION

Breast cancer is the most common malignant disease among women. In 2014 in the United States alone, 232,670 women were diagnosed as having this type of cancer, and 40,000 of them died [1]. Breast tumors can appear at any age but the incidence increases drastically after the age of 50, in connection with hormonal changes [2].

The most effective way to prevent mortality caused by breast cancer is early diagnosis. Biochemical detection of this cancer is based on the determination of markers such as CA 15-3, CEA, and CA 27.29 [3]. However, these are still neither fully specific nor sensitive; hence the researchers attempt to find the biomarkers whose levels would correlate with the presence and stage of breast cancer. We assumed that these factors might include matrix metalloproteinase (MMP), tissue inhibitors of matrix metalloproteinase (TIMPs), and cytokines, for example macrophage colony-stimulating factor (M-CSF). The diagnostic role of the analyzed parameters will involve determination of breast tumor stage based on the following relationship: the more advanced tumor stage, the higher the concentration of these substances.

MMPs form a large group of enzymes responsible for the remodeling of the extracellular matrix (ECM) in tumor invasion [4]. One of them – MMP-9 – also known as a gelatinase B, plays an important role in cancer growth and invasion with its unique ability to destroy collagen type IV and other ECM components. Furthermore, overexpression of this factor correlates with poor prognosis and patient survival [5].

TIMPs are natural inhibitors of MMPs. Only four proteins are included in this group [6]. TIMP-1 can be detected in a variety of body fluids and tissues. This factor stimulates cancer invasion by apoptosis inhibition, promotion of tumor cell growth, and regulation of angiogenesis [7]. The relationship between high levels of TIMP-1 and poor prognosis of breast tumor patients had been confirmed [8].

Previous studies revealed an autologous synthesis of M-CSF in various human cancer cell lines *in vitro* and by tumors *in vivo*. This glycoprotein, which belongs to the group of hematopoietic growth factors (HGFs), is responsible for monocyte production and function; yet in pathological conditions, it stimulates cancer cell growth [9].

The aim of this study was to evaluate the plasma levels of MMP-9, TIMP-1, M-CSF, and CA 15-3 (comparative marker) in breast tumor patients in respect to two control groups: benign breast cancer patients and healthy subjects. The data obtained in this work may be helpful in both the determination of the use-fulness of the analyzed parameters (separately and together) in diagnosis of breast cancer and the differentiation of its subtypes.

METHODS

1. Patients

Table 1 shows the tested groups. The study included 110 breast cancer patients whose conditions were diagnosed by the Oncology Group and treated in the Department of Oncology, Medical University, Białystok, Poland, between 2013 and 2015. Tumor classification and staging were carried out in accordance with the International Union Against Cancer Tumor-Node-Metastasis (UICC-TNM) classifications [10]. The breast cancer histopathol-

Table 1. Characteristics of breast cancer patients and co	ontrol
groups: benign breast tumor patients and healthy subjects	

Study group		N of patients
Breast cancer patients	Adenocarcinoma	110
Туре	ductale	110
Median age (range)		54 (35-76)
	$I-T_1N_0M_0$	22
	$IIA-T_2N_0M_0$	10
	$IIB-T_2N_1M_0$	14
Tumor stage	$T_3N_0M_0$	10
	$IIIA-T_2N_2M_0$	9
	$T_3N_1M_0$	7
	$IIIB-T_4N_2M_0$	8
	$T_4N_3M_0$	7
	IV (metastases)	23
Menopausal status:		
-Premenopausal		52
-Postmenopausal		58
Benign breast tumor		50
Туре	Adenomapapilloma	21
	Intraductale	6
	Fibroadenoma	12
	Mastopatia	11
Median age (range) Menopausal status:		43 (23-71)
-Premenopausal		16
-Postmenopausal		34
Healthy subjects		50
Median age (range) Menopausal status:		41 (20-72)
-Premenopausal		20
-Postmenopausal		30

ogy was established in all cases by tissue biopsy of mammary tumor or based on tumor cancer tissues collected during surgery (all patients with adenocarcinoma ductale). Pretreatment staging procedures included physical and blood examinations, mammography, mammary ultrasound scanning, breast core biopsies, and chest X-ray. In addition, radioisotopic bone scans, examination of bone marrow aspirates, and brain or chest computerized tomography (CT) scans were performed when necessary. None of the patients had received chemo- or radiotherapy before blood sample collection.

The control groups included 50 patients with benign breast tumors and 50 healthy untreated women who had undergone mammary gland examination performed by a gynecologist prior to blood collection. Moreover, mammary ultrasound scanning was performed when necessary. The benign breast tumor histopathology was established in all cases with the use of tissue biopsy of mammary tumor or after surgery.

The study was approved by the local Ethics Committee in Medical University in Białystok no R-I-002/239/2014/. All the patients gave their informed written consent for the examination.

2. Biochemical analyses

Venous blood samples were collected from each patient into a heparin sodium tube, centrifuged for 15 min at 1,000g to obtain plasma samples, and stored at -85°C until assayed. The tested cytokines (MMP-9, TIMP-1, and M-CSF) were measured by using ELISA (Quantikine Human HGFs Immunoassay, R&D systems, Minneapolis, MN, USA), according to the manufacturer's protocols. Duplicate samples were assessed for each patient.

The intra-assay CV of MMP-9 is reported to be 1.9% at a mean concentration of 2.04 ng/mL, SD=0.039; TIMP-1-3.9% at a mean concentration of 1.27 ng/mL while SD=0.05; M-CSF-3.4% at mean concentration of 227 pg/mL, SD=7.7.

The inter-assay CV of MMP-9 amounted to 7.8% at a mean concentration of 2.35 ng/mL, SD=0.184; TIMP-1-3.9% at a mean concentration of 1.28 ng/mL while SD=0.05; and M-CSF-3.1% at mean concentration of 232 pg/mL, SD=7.3. The assay showed no significant cross-reactivity or interference with numerous human cytokines and other growth factors.

The plasma concentrations of CA 15-3 were measured with chemiluminescent microparticle immunoassay (CMIA) (Abbott, Abbott Park, IL, USA). The intra-assay CV for CA 15-3 is reported to be 2.2% at mean concentration of 27.0 U/mL, SD=0.6 while the inter-assay CV for CA 15-3 is reported to be 2.6% at mean concentration of 27.0 U/mL, SD=0.7.

The value of intra- and and inter- assay CVs were calculated by the manufacturers and enclosed in the reagent kits.

3. Statistical analysis

Statistical analysis was performed by using STATISTICA 8.0 PI (StatSoft, Tulsa, OK, USA). A preliminary statistical analysis (Chisquare test) revealed that the cytokine and tumor marker levels did not follow normal distribution. Consequently, the Mann-Whitney U test was used for statistical analysis between cancer patients and control groups. Additionally, statistical analysis between the groups with different stages of breast cancerwas performed by using the Kruskal-Wallis test and a multivariate analysis of various data by the post-hoc Dwass-Steele-Crichlow-Flinger test. The data were presented as a median and a range [11].

Diagnostic sensitivity, specificity, and the predictive values of positive and negative test results (PPV, NPV) were calculated by using the following cut-off value: 95th percentile from the control

group (calculated from healthy blood donors). This value for each of the substances was as follows: MMP-9-382.00 ng/mL; TIMP-1-119.22 ng/mL; M-CSF-396.94 pg/mL; and CA 15-3-20.85 U/mL. The construction of the ROC curves was performed using GraphRoc program for Windows (Windows,Royal, AR, USA) and the areas under the ROC curve (AUC) were calculated.

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In the analyses of both diagnostic performance (sensitivity, specificity, PPV, and NPV) and ROC curve, only healthy subjects were used as a control group.

RESULTS

Table 2 presents the median and the range of plasma levels of the investigated parameters and CA 15-3 in tested groups. The median values for MMP-9 (286.00 ng/mL) and M-CSF (437.05 pg/mL), similar to those of commonly accepted tumor marker CA 15-3 (26.90 U/mL) in all breast cancer patients (stages I-IV) were significantly higher compared with the values in healthy subjects (181.00 ng/mL; 281.20 pg/mL; 15.20 U/mL, respectively) (P<0.001). Furthermore, the median level of TIMP-1 in BC total group (155.74 ng/mL) was significantly higher than that in the benign breast tumor group (74.75 ng/mL). We also noticed significantly higher concentrations of M-CSF and CA 15-3 at stage III or IV of cancer compared with stage I or II (M-CSF: I vs. III P<0.001, I vs. IV P<0.001 and II vs. III P=0.005, II vs. IV P=0.001; CA 15-3: I vs. III P=0.001, I vs. IV P<0.001 and II vs. III P=0.023, II vs. IV P=0.001). Furthermore, we detected significantly higher plasma levels of TIMP-1 in the comparison of stage III to stage I (P=0.018) and to stage II (P=0.042).

Additionally, we observed significantly higher concentrations of CA 15-3 in all the analyzed groups at all stages of breast cancer (with the exception of stage I) in comparison with healthy women (*P* values were: 0.004 at stage I and below 0.001 at stages II and III).

Moreover, M-CSF, TIMP-1, and CA 15-3 concentrations showed a statistical difference between the two control groups (Table 2).

Table 3 shows the diagnostic criteria: sensitivity, specificity, PPV, and NPV in breast cancer patients. The sensitivity of the tested parameters in the total cancer group was higher for M-CSF (57%) than for MMP-9 (38%) and TIMP-1 (18%), and slightly lower than for CA 15-3 (64%). The combined use of tested factors with CA 15-3 resulted in an increase in sensitivity. A maximum value for the BC total group was obtained for the combination of four studied parameters (84%). Among all tested factors, CA 15-3 showed the highest sensitivity at almost

15-3 (U/mL) 5 0-34.20* 0-38*.‡
0-34.20*
J_38* ^{,‡}
5 50
0-167.50* ^{,†,‡}
0-250.00*
0-250.00 [§]
0-48.30
0-27.20
0- 0-

*Statistically significant when compared breast cancer patients with healthy subjects (P < 0.05); ¹Statistically significant when compared breast cancer patients with benign breast tumor group (P < 0.05); ¹Statistically significant when compared breast cancer patients stage III or IV with breast cancer patients stage I or II (P < 0.05); [§]Statistically significant when compared benign breast tumor patients with healthy subjects (P < 0.05); [§]Statistically significant when compared benign breast tumor patients with healthy subjects (P < 0.05); [§]Statistically significant when compared benign breast tumor patients with healthy subjects (P < 0.05); [§]Statistically significant when compared benign breast tumor patients with healthy subjects (P < 0.05).

Abbreviations: MMP-9, matrix metalloproteinase-9; TIMP-1, tissue inhibitor of metalloproteinases-1; M-CSF, macrophage colony-stimulating factor.

every stage of cancer (with the exception of stage III-equal to M-CSF-and stage IV- the highest value of M-CSF). The sensitivity of the analysis using the tested parameters distinctly rose with the increase in the stage of cancer (with the exception of MMP-9 and TIMP-1 at stage IV), in an identical manner to CA 15-3. The combined use of the analyzed factors and antigen CA 15-3 resulted in an increase in sensitivity at every stage of breast cancer. Higher values were obtained for the combination of MMP-9, TIMP-1, or M-CSF with CA 15-3 (stage III-97%, 87%, 97%, stage IV-87%, 87%, 96%; respectively). A maximum range was observed for the combination of all studied parameters at stageIII and IV of breast cancer (100%).

The diagnostic specificity for M-CSF, MMP-9, TIMP-1, and CA 15-3 showed very high values (95% in all cases).

The PPVs in the total group of breast cancer patients was the highest for M-CSF and CA 15-3 (97%) compared with MMP-9 (95%) and TIMP-1 (91%). The combined use of the tested parameters with CA 15-3 resulted in a decrease in the PPV range (Table 3).

The NPVs in the total group of breast cancer was repeatedly higher for CA 15-3 (48%) than for M-CSF (45%), MMP-9 (36%), and TIMP-1 (29%). The combined use of the tested factors and CA 15-3 resulted in an increase in the NPV in all cases, for example with M-CSF-to 59 %. A maximum range was obtained for

the combination of CA 15-3, M-CSF, MMP-9, and TIMP-1-64%. The combined use of the analyzed parameters and CA 15-3 resulted in an increase in the NPV at every cancer stage. Maximum equal ranges (100%) were obtained for the combination of M-CSF, MMP-9, TIMP-1, and CA 15-3 at stage III and IV of BC (Table 3).

The relationship between the diagnostic sensitivity and specificity is illustrated by the ROC curve. The AUC indicates the clinical usefulness of a tumor marker and its diagnostic power. We noticed that the M-CSF AUC (0.7955) in the total breast cancer group was larger than the area of MMP-9 (0.6827) and TIMP-1 (0.6250) and slightly lower than the AUC of CA 15-3 (0.8548). In case of MMP-9 and M-CSF, the AUC was significantly larger in comparison with AUC=0.5, which is similar to the case of CA 15-3 (borderline of the diagnostic usefulness of the test) (P < 0.001; in all cases). The combined analysis of AUC tested parameters with CA 15-3 resulted in an increase in the AUCs (MMP-9-0.8851, TIMP-1-0.8547, and M-CSF-0.8840), but a maximum range in the BC total group was obtained for the combination of all studied parameters (0.9125). Additionally, the AUCs in combined analysis of MMP-9, TIMP-1 or M-CSF with CA 15-3 were significantly larger in comparison with AUC=0.5 (Fig. 1) (Table 4).



 Table 3. Diagnostic criteria of tested parameters and in combined analysis with CA 15-3 in breast cancer patients

Parameters tested	Diagnostic	Breast cancer				
	criteria (%)	Stage I	Stage II	Stage III	Stage IV	Total group
MMP-9	Sensitivity	27	35	45	43	38
	Specificity	95	95	95	95	95
	PPV	75	86	88	83	95
	NPV	70	75	69	75	36
TIMP-1	Sensitivity	9	8	35	17	18
	Specificity	95	95	95	95	95
	PPV	50	60	85	67	91
	NPV	66	55	66	64	29
M-CSF	Sensitivity	18	38	84	87	57
	Specificity	95	95	95	95	95
	PPV	67	87	93	91	97
	NPV	68	64	88	93	45
CA 15-3	Sensitivity	32	50	84	83	63
	Specificity	95	95	95	95	95
	PPV	78	89	93	90	97
	NPV	72	81	88	90	48
MMP-9+	Sensitivity	41	65	97	87	74
CA 15-3	Specificity	90	90	90	90	90
	PPV	69	85	88	83	95
	NPV	73	75	97	92	55
TIMP-1+ CA 15-3	Sensitivity	36	50	87	87	65
	Specificity	90	90	90	90	90
	PPV	67	81	87	83	95
	NPV	72	68	90	92	49
M-CSF+	Sensitivity	45	68	97	96	77
CA 15-3	Specificity	90	90	90	90	90
	PPV	71	85	88	85	96
	NPV	75	77	97	97	59
MMP-9+	Sensitivity	55	76	100	100	84
TIMP-1+	Specificity	83	83	83	83	83
M-CSF+	PPV	60	76	79	74	92
CA 15-3	NPV	76	80	100	100	64

Abbreviations: MMP-9, matrix metalloproteinase-9; TIMP-1, tissue inhibitor of metalloproteinases-1; M-CSF, macrophage colony-stimulating factor; PPV, predictive value of positive test result; NPV, predictive value of negative test result.

DISCUSSION

The group of HGFs is composed of cytokines such us stem cell factor (SCF), interleukin 3 (IL-3), granulocyte-macrophage-col-

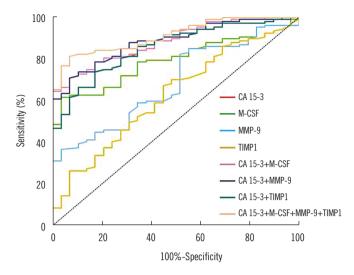


Fig. 1.Diagnostic criteria of ROC curve for tested parameters and in combination with CA 15-3 in total breast cancer group. Abbreviations: MMP-9, matrix metalloproteinase-9; TIMP-1, tissue inhibitor of metalloproteinases-1; M-CSF, macrophage colony-stimulating factor.

Table 4. Diagnostic criteria of the ROC curve for tested parameters and CA 15-3 $\,$

Parameters tested –	ROC criteria in breast cancer total group				
	AUC	Sensitivity	95% CI	Р	
M-CSF	0.7955	0.0381	0.721-0.870	< 0.001	
MMP-9	0.6827	0.0511	0.582-0.783	< 0.001	
TIMP-1	0.6250	0.0574	0.513-0.738	0.0294	
CA 15-3	0.8548	0.0346	0.787-0.923	< 0.001	
M-CSF + CA 15-3	0.8840	0.0287	0.828-0.940	< 0.001	
MMP-9 + CA 15-3	0.8851	0.0289	0.829-0.942	< 0.001	
TIMP-1 + CA 15-3	0.8547	0.0346	0.787-0.922	< 0.001	
M-CSF + MMP-9 + TIMP-1 + CA 15-3	0.9125	0.0241	0.865-0.960	< 0.001	

P-statistically significantly larger AUCs compared with AUC=0.5. Abbreviations: AUC, area under the ROC curve; CI, confidence interval;

MMP-9, matrix metalloproteinase-9; TIMP-1, tissue inhibitor of metalloproteinases-1; M-CSF, macrophage colony-stimulating factor.

ony-stimulating factor (GM-CSF), granulocyte-colony stimulating factor (G-CSF), and M-CSF. Their main function is regulation of hematopoietic progenitor cell growth and differentiation. Furthermore, HGFs have the ability to stimulate the proliferation of non-hematopoietic cells *in vitro*, for instance cancer cells [12].

Significant plasma overexpression of M-CSF was detected in patients suffering from many types of tumors, for instance endometrial [13], ovarian [14], or breast tumors [15]. Moreover, scientists who examined this factor demonstrated that M-CSF is a good candidate as a marker of pancreatic tumor [16] or non-

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small-cell lung cancer [17]. McDermond *et al.* [18] found that serum levels of this marker were elevated in breast cancer patients but the highest concentrations of M-CSF were detected by Kaciński [19] in the ascites and pleural effusion of the patients in the advanced stages of this cancer. The high grade and poor prognosis correlated with elevated levels of M-CSF [20].

MMPs and TIMPs play a crucial role in ECM and basement membrane homeostasis [21]. Giannelli *et al.* [22] proved that in physiological conditions, the MMP: TIMP ratio is highly regulated. The imbalance between concentrations and activities of those factors may lead to the disturbances in the production and degradation of ECM components. They also demonstrated that the MMP and TIMP dysfunction might lead to faster invasion and metastasis of the cancer. Moreover, Jinga *et al.* [23] affirmed that high ratios of MMP-9: TIMP-1 signify strong proteolytic ability of MMPs (especially gelatinases) in malignant tumors. The correlation between the levels of MMP-9 and TIMP-1 and breast tumor sizes was notpreviously confirmed [24].

According to our research, plasma concentrations of MMP-9, M-CSF, and CA 15-3 were significantly higher in the whole breast cancer group than in healthy subjects. The same observations on MMP-9 were reported previously [23-25]. The over-expression of M-CSF in similar groups was confirmed by Kaciński [19]. We also found comparable data in our previous studies [26, 27].

Contrary to the other parameters analyzed in this study, there was no difference between TIMP-1 levels in healthy subjects and breast cancer patients. According to other researchers, the concentrations of TIMP-1 were either significantly lower [28] or significantly higher [29] in carcinomas versus fibroadenomas.

Interestingly, the levels of TIMP-1, M-CSF, and CA 15-3 were significantly higher in total breast cancer patients than in benign breast tumor cases. These observations about M-CSF and CA 15-3 are in accordance with both the data found in our previous studies, in which we analyzed a smaller benign breast cancer group [14] and the study by Wu *et al.* [24]. Concurrently, Jingla *et al.* [23] found no statistical differences between TIMP-1 serum levels in benign and malignant cases.

In this work we found that MMP-9, TIMP-1, and M-CSF present high levels at stage III and IV. The highest concentrations of these factors are probably the consequence of a higher mass of cancer cells. This is the further argument for our theory that the analyzed factors can play a prognostic role on the basis of following relationship: the more advanced tumor stage, the higher the concentration of the substance.

In the present study, the diagnostic sensitivity of M-CSF in to-

tal breast cancer patient group (57%) was higher than that of MMP-9 (38%) and TIMP-1 (18%) and slightly lower than that of CA 15-3 (63%). In our previous studies on breast cancer, M-CSF showed a higher sensitivity than CA 15-3 [25, 26]. Concurrently, MMP-9 presented the highest sensitivity at stage I of BC (27%), but still slightly lower than that of CA 15-3 (32%).

Moreover, current findings, similarly to our former works, proved that the combination of the commonly accepted tumor markers and the factors which may be useful in early detection of breast cancer significantly increases the pre-operative diagnostic sensitivity.

In this study, M-CSF had the highest predictive values of all tested factors, similar to that of CA 15-3, at every stage of cancer. The M-CSF diagnostic criteria are the highest for the group with stage IV cancer, which probably resulted from their vital role in the formation of both local and distant metastases [18].

The ROC curve illustrated by a specificity/sensitivity diagram is a crucial value describing all markers. The best cancer markers are characterized by the largest AUCs. In the present study, the AUC of M-CSF was the largest among the analyzed parameters, yet slightly lower than CA 15-3. Additionally, we detected significantly larger AUCs for M-CSF, MMP-9, TIMP-1, and CA 15-3 compared with AUC=0.5. Our data demonstrated that the diagnostic power of the analyzed parameters, in particular M-CSF in total breast cancer patients group, had a higher value than the diagnostic power of M-CSF in cervical cancer [12].

The examined factors are unsuitable for basic screening when considered separately. Their diagnostic usefulness is high in their combined analysis with a comparison tumor marker (in this case with CA 15-3); yet the greatest diagnostic value is observed when they are considered altogether as a new panel of biomarkers.

In conclusion, the obtained results suggest the usefulness of M-CSF, MMP-9, and TIMP-1 in the diagnostics of breast cancer except in stage I-yet only in combination with CA 15-3. Our findings seem to be ambiguous, although they are a promising step forward. The main limitation of the studywas lack of significant differences in concentrations of analyzed factors between patients with stage I and healthy subjects. Therefore, further investigation will be necessary to clarify the utility of this new biomarker panel in clinical management.

Authors' Disclosures of Potential Conflicts Of Interest

No potential conflicts of interest relevant to this article were reported.



REFERENCES

- 1. American Cancer Society. Cancer Facts & Figures 2014. Atlanta, GA: American Cancer Society, 2014:4.
- Trichopoulos D, Adami HO, Ekbom A, Hsieh CC, Lagiou P. Early life events and conditions and breast cancer risk: from epidemiology to etiology. Int J Cancer 2008;122:481-5.
- 3. Harris L, Fritsche H, Mennel R, Norton L, Ravdin P, Taube S, et al. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. J ClinOncol 2007;25: 5287-312.
- 4. Fingleton B. Matrix metalloproteinases: roles in cancer and metastasis. Front Biosci 2006;11:479-91.
- PellikainenJM, Ropponen KM, KatajaVV, KellokoskiJK, EskelinenMJ, KosmaVM. Expression of matrix metalloproteinase (MMP)-2 and MMP-9 in breast cancer with a special reference to activator protein-2, HER2, and prognosis. Clin Cancer Res 2004;10:7621-8.
- Brew K, Dinakarpandian D, Nagase H. Tissue inhibitors of metalloproteinases: evolution, structure and function. BiochimBiophysActa 2000; 1477:267-83.
- Baker AH, Zaltsman AB, George SJ, Newby AC. Divergent effects of tissue inhibitor of metalloproteinase-1,-2, or -3 overexpression on rat vascular smooth muscle cell invasion, proliferation, and death in vitro. TIMP-3 promotes apoptosis. J Clin Invest 1998;101:1478-87.
- Talvensaari-Mattila A andTurpeenniemi-Hujanen T. High preoperative serum TIMP-1 is a prognostic indicator for survival in breast carcinoma. Breast Cancer Res Treat 2005;89:29-34.
- Lin EY, Gouon-Evans V, Nguyen AV, Pollard JW. The macrophage growth factor CSF-1 in mammary gland development and tumor progression. J Mammary Gland BiolNeoplasia 2002;7:147-62.
- 10. Sobin LH, Gospodarowicz MK, et al. eds. TNM classification of malignant tumours. 7th ed. Chichester: John Wiley & Sons, 2011:181-3.
- 11. Jemal A, Murray T, Samuels A, Ghafoor A, Ward E, ThunMJ. Cancer statistics, 2003. CACancer J Clin 2003;53:5-26.
- Ławicki S, Będkowska GE, Gacuta-Szumarska E, Knapp P, Szmitkowski M. Pretreatment plasma levels and diagnostic utility of hematopoietic cytokines in cervical cancer or cervical intraepithelial neoplasia patients. Folia HistochemCytobiol 2012;50:213-9.
- Ławicki S, Będkowska GE, Gacuta-Szumarska E, Czygier M, Szmitkowski M. The plasma levels and diagnostics utility of selected hematopoietic growth factors in endometrial cancer patients and with myoma uteri. Pol MerkurLekarski 2010;28:354-8.
- Ławicki S, Gacuta-Szumarska E, BędkowskaGE, Szmitkowski M. Hematopoietic cytokines as tumor markers in gynecological malignancies. A multivariate analysis in epithelial ovarian cancer patients. Growth Factors 2012;30:357-66.
- 15. Czygier M, Ławicki S, Szmitkowski M. The plasma level of sL-selectin, myeloperoxidase (MPO) and granulocyte-colony stimulating factor (G-

CSF) in breast cancer patients after surgery. PrzeglLek 2009;66:433-6.

- Vasiliades G, Kopanakis N, Vasiloglou M, Zografos G, Margaris H, Masselou K, et al. Role of the hematopoietic cytokines SCF, IL-3, GM-CSF and M-CSF in the diagnosis of pancreatic and ampullary cancer. Int J Biol Markers 2012;27:186-94.
- Bahar B, AcedilAyc lota B, Çoşkun U, Büyükberber S, Benekli M, Yildiz R. Granulocyte colony stimulating factor (G-CSF) and macrophage colony stimulating factor (M-CSF) as potential tumor markers in non small cell lung cancer diagnosis. Asian Pac J Cancer Prev 2010;11:709-12.
- McDermott RS, Deneux L, Mosseri V, Védrenne J, Clough K, Fourquet A, et al. Circulating macrophage colony stimulating factor as a marker of tumour progression. Eur Cytokine Netw2002;13:121-7.
- 19. Kaciński BM. CSF-1 and its receptor in ovarian, endometrial and breast cancer. Ann Med 1995;27:79-85.
- Scholl SM, Lidereau R, de la Rochefordière A, Le-Nir CC, Mosseri V, Noguès C, et al. Circulating levels of the macrophage-colony stimulating factor CSF-1 in primary and metastatic breast cancer patients. A pilot study. Breast Cancer Res Treat 1996;39:275-83.
- Visse R and Nagase H. Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. Circ Res 2003; 92:827-39.
- 22. Giannelli G, Erriquez R, Fransvea E, Daniele A, Trerotoli P, Schittulli F, et al. Proteolytic imbalance is reversed after therapeutic surgery in breast cancer patients. Int J Cancer 2004;109:782–5.
- Jinga DC, Blidaru A, Condrea I, Ardeleanu C, Dragomir C, Szegli G, et al. MMP-9 and MMP-2 gelatinases and TIMP-1 and TIMP-2 inhibitors in breast cancer: correlations with prognostic factors. J Cell Mol Med 2006;10: 499-510.
- 24. Wu ZS, Wu Q, Yang JH, Wang HQ, Ding XD, Yang F, et al. Prognostic significance of MMP-9 and TIMP-1 serum and tissue expression in breast cancer. Int J Cancer 2008;122:2050-6.
- Voorzanger-Rousselot N, Juillet F, Mareau E, Zimmermann J, Kalebic T, Garnero P. Association of 12 serum biochemical markers of angiogenesis, tumour invasion and bone turnover with bone metastases from breast cancer: a crossectional and longitudinal evaluation. Br J Cancer 2006;95:506-14.
- Ławicki S, Bedkowska GE, Szmitkowski M. VEGF, M-CSF and CA 15-3 as a new tumor marker panel in breast malignancies: a multivariate analysis with ROC curve. Growth Factors 2013;31:98-105.
- Ławicki S, Szmitkowski M, Wojtukiewicz M. The pretreatment plasma level and diagnostic utility of M-CSF in benign breast tumor and breast cancer patients. ClinChimActa 2006;371:112-6.
- Iwata H, Kobayashi S, Iwase H, Masaoka A, Fujimoto N, Okada Y. Production of matrix metalloproteinases and tissue inhibitors of metalloproteinases in human breast carcinomas. Jpn J Cancer Res 1996;87:602-11.
- McCarthy K, Maguire T, McGreal G, McDermott E, O'Higgins N, Duffy MJ. High levels of tissue inhibitor of metalloproteinase-1 predict poor outcome in patients with breast cancer. Int J Cancer 1999;84:44-8.