

miR-10b as a Clinical Marker and a Therapeutic Target for Metastatic Breast Cancer

Technology in Cancer Research & Treatment
Volume 24: 1-7
© The Author(s) 2025
Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/15330338251339256
journals.sagepub.com/home/tct



Alan Halim, PhD¹ , Bryan Kim, BS^{1,2} , Elizabeth Kenyon, MS^{1,3}, and Anna Moore, PhD^{1,3}

Abstract

Despite advances in cancer detection and treatment, metastatic breast cancer continues to carry a poor prognosis due to the lack of diagnostic and therapeutic resources that are specific to the metastatic process. MicroRNA-10b (miR-10b) is a small, non-coding RNA that is the focus of many studies due to its unique role as a driver of metastasis. The pathways it is involved in and the properties it confers have been reviewed previously and, collectively, are suggestive of the potential of miR-10b as a clinical marker and as a therapeutic target specific to metastatic disease. With the goal of application of our understanding of miR-10b to the clinic, in this mini-review, we highlight the studies that support the utility of miR-10b for these translational purposes.

Keywords

breast cancer, metastasis, microRNA, biomarker, cancer therapy

Received: 2 December 2024; revised: 25 March 2025; accepted: 14 April 2025

Introduction

MicroRNAs (miRNAs) are small, noncoding RNAs composed of 19–25 nucleotides that function as gene silencers.¹ They have been linked to the regulation of developmental processes since the discovery of the first miRNA in 1993,² and their role in pathological processes was demonstrated in 2002, when miR-15 and miR-16-1 were identified as tumor suppressors downregulated in the majority of chronic lymphocytic leukemia cases.³ Since then, studies have continued to find numerous miRNAs that function as oncogenes or tumor suppressors.

Uniquely, miR-10b was one of the first microRNAs to be implicated specifically in cancer metastasis, with early studies demonstrating that its expression confers properties of increased migration, invasion, and metastatic potential onto breast cancer cells.⁴ It has since been associated with other roles in breast cancer models, such as chemoresistance,⁵ stemness,^{6,7} and viability,^{8–10} and its roles appear to be conserved across at least 17 other types of cancer.¹¹ These studies, as well as mechanistic insights, have been reviewed previously.^{11,12} Altogether, these features make miR-10b a potentially valuable clinical marker and promising therapeutic target for the treatment of cancer, particularly metastatic disease, which comprises up to 90% of cancer-related

deaths.^{13,14} With five-year survival rates for metastatic breast cancer being a dismal 30%,¹⁵ here, we review the utility of miR-10b as a clinical marker for breast cancer and the therapeutic effects observed with its inhibition in *in vivo* breast cancer models using clinically viable methods.

miR-10b as a Clinical Marker in Breast Cancer

In patient primary tumors, miR-10b has been correlated with greater tumor diameter, worse histological and clinical grades, and greater vascularization.^{16–18} Additionally, miR-10b appears to positively correlate with HER2 status^{16,19} and negatively correlate with both estrogen and progesterone receptors.¹⁶ This variability among breast cancer subtypes, as well as intratumoral

¹Precision Health Program, Michigan State University, East Lansing, MI, USA

²Department of Radiology, College of Human Medicine, Michigan State University, East Lansing, MI, USA

³Department of Radiology, College of Human Medicine, Michigan State University, East Lansing, MI, USA

Corresponding Author:

Anna Moore, PhD, 766 Service Rd., Rm. 2022, East Lansing, MI 48824, USA.
Email: moorea57@msu.edu



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and

distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access page (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

heterogeneity in miR-10b expression,²⁰ may explain why studies comparing miR-10b expression in breast primary tumors in general (ie, mixed subtypes and at ratios that vary across studies) versus normal tissues have yielded inconsistent results.^{4,16,21,22} Nevertheless, because of the reproducible correlations between miR-10b expression and tumor grade and the phenotypes miR-10b confers in laboratory settings, many studies have begun investigating the potential of miR-10b as a diagnostic or prognostic indicator of breast cancer and metastatic disease. Circulating tumor cells²³ and the secretion of miR-10b by breast cancer cells^{24,25} also provide a rationale for serum-based analyses.

Multiple studies suggest that serum miR-10b can be used to discern patients with primary breast cancer from those without,^{26,27} with one study reporting a sensitivity of 83.30% and specificity of 100%.²⁶ This far exceeds the sensitivity of breast cancer markers already in use in the clinic, such as CA15-3 and CEA,²⁸ and demonstrates the potential of miR-10b as a diagnostic marker. As a prognostic marker, miR-10b expression has been correlated with disease relapse,^{22,29} which may be specific to breast cancer, as this correlation was not observed in a pan-cancer meta-analysis.³⁰ Furthermore, miR-10b expression has been linked to worse overall survival through meta-analysis of three breast cancer studies.³⁰ MiR-10b may also be useful as a lab value for disease monitoring, as miR-10b expression in patient serum significantly decreased post-operation relative to matched pre-operation levels and further decreased post-radiotherapy.²⁷ More recently, one group proposed to include miR-10b among a panel of miRNAs for predicting response to treatment,³¹ demonstrating another possible clinical application of miR-10b. In support of this, elevated miR-10b in patient serum has been shown to be predictive of anemia in response to chemoradiotherapy.³²

Given its association with pro-metastatic features, it is possible that miR-10b has the greatest utility as a marker in the context of metastasis. Indeed, one consistent finding is the upregulation of miR-10b in metastatic breast primary tumors relative to non-metastatic tumors or healthy tissue.^{4,16–18} Similarly, several studies suggest that miR-10b expression in serum could be used as a marker for breast cancer spread to lymph nodes, finding significant upregulation in patients with metastatic lymph nodes relative to those without.^{27,33,34} In one study, the median miR-10b expression of patients with spread to lymph nodes ($n=35$) was 4.44-fold that of patients without spread to lymph nodes ($n=25$; $p<0.01$), and the odds ratio between miR-10b expression and lymph node spread was calculated to be 2.19. When used as a diagnostic test, a threshold expression value identified patients with spread to lymph nodes with a sensitivity of 71% and a specificity of 72%.³³ These findings are particularly notable given that miR-10b is upregulated in lymph node metastasis samples relative to their matched primary tumors, as reported in a study that included 43 pairs of samples across four different types of cancer,³⁵ as it implicates increased miR-10b at every stage of the metastatic process, from primary tumor to circulation to metastases. These findings also align with the expression patterns of miR-10b relative to HER2, ER, and PR status described earlier,

as HER2-overexpressing tumors have increased rates of metastasis relative to hormone receptor-positive tumors.

miR-10b as a Therapeutic Target

The use or inhibition of miRNAs in therapeutic settings has been extensively reviewed in recent publications.^{36–38} MiRNAs make for promising therapeutic targets due to their regulation of hundreds of genes that affect several, often-related pathways or processes.^{39,40} Given that they are believed to have relatively minor effects on each individual gene,⁴⁰ modulation of one miRNA is less likely to perturb normal biological function the way that other therapeutics do. This is evident in studies investigating therapeutic inhibition of miR-10b. Though believed to be an important contributor to early development, genetic knockout of miR-10b in mice is nonlethal, with no significant effect on body weight, overall survival, fertility, or complete blood count.⁴¹ Moreover, the expression of miR-10b in normal tissues is low compared to primary breast tumors.⁴² These findings collectively suggest that inhibition of miR-10b in patients would have little or no off-target effects while still potentially having profound effects on cancers that depend on miR-10b activity. Here, we highlight the different approaches that have been used across cancer models to specifically silence miR-10b *in vivo*, their results, and their clinical promise.

At the core of most of the methods used to inhibit miR-10b is the delivery of an anti-miR-10b antisense oligonucleotide (ASO), an RNA molecule with base complementarity to miR-10b. By binding miR-10b, the ASO renders miR-10b non-functional and triggers its degradation. The challenges to delivery of ASOs include nucleases, a short blood half-life, and charge-charge repulsion at the cell membrane.^{36,37,43} As such, modifications to the ASO or conjugation to a vehicle are generally required. One of the more basic strategies toward therapeutic miR-10b inhibition utilized an ASO with a phosphorothioate backbone, 2'-O-methylation of ribose sugars, and a cholesterol moiety at the 3' end, all contributing toward stability and delivery.⁴⁴ In mice implanted with the spontaneously metastatic 4T1 mouse breast cancer cell line, intravenous administration of the ASO (twice per week, 50 mg ASO/kg bodyweight) resulted in no effect on primary tumor size while decreasing pulmonary metastases by 86%, relative to mice treated with a control oligonucleotide of a similar sequence but with mismatches or with PBS. Important for this study and other methods of therapy using ASOs, this inhibition was shown to be specific to miR-10b and the findings were reproduced when miR-10b was inhibited in the cancer cells prior to implantation, demonstrating that the effects were due to miR-10b suppression in cancer cells and not in other cells (eg, the tumor microenvironment). Moreover, treated mice displayed no behavioral changes, and blood and histologic analyses revealed minimal signs of toxicity.

Although the described modifications to the ASO appear to have overcome some of the limitations to delivery *in vivo*, a major criticism is the high dosage that was required for

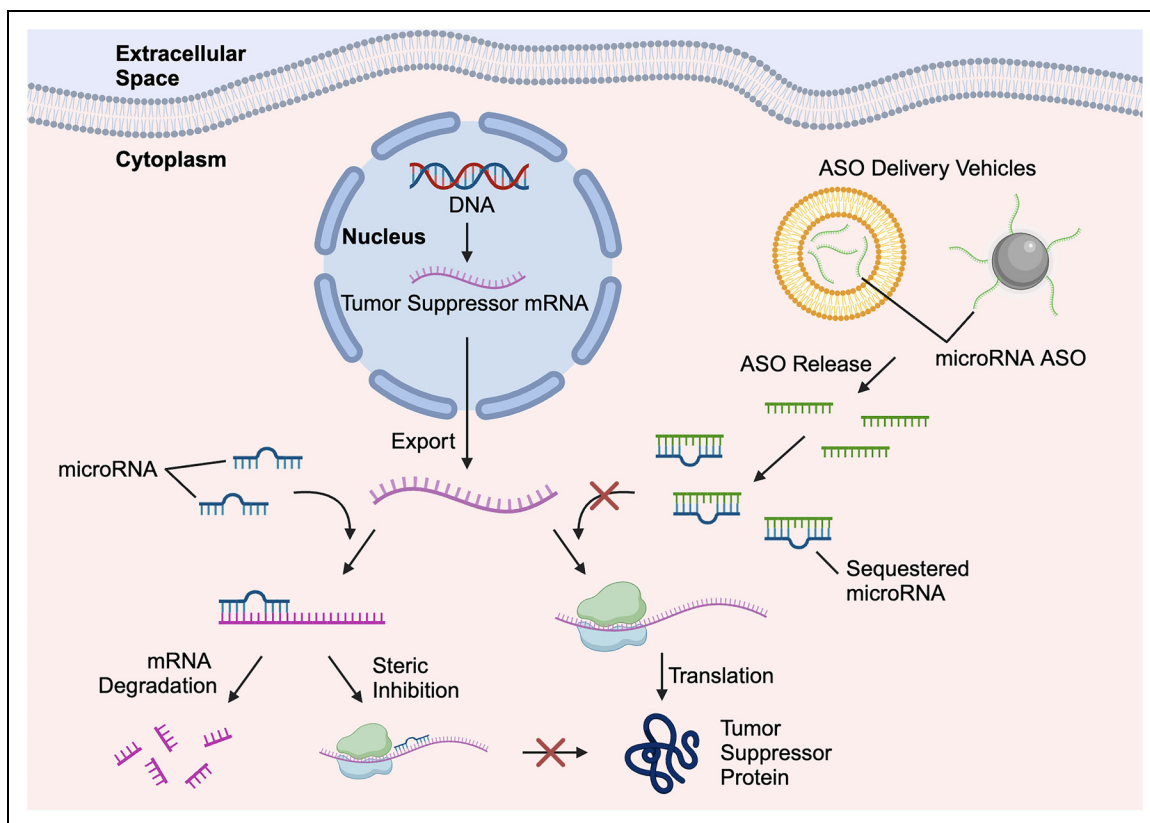


Figure 1. Anti-microRNA Oligonucleotide Delivery. Nanoparticles are at the Forefront of Anti-microRNA Oligonucleotide Delivery. Once Nanoparticles Carrying Anti-microRNA Oligonucleotides Enter the Cell, the Payload is Released from the Nanoparticle Construct, Resulting in Sequestration of Aberrantly Expressed microRNAs. Consequently, Translation of Tumor Suppressor Proteins is Restored, Leading to Reduced Tumorigenesis. Created in BioRender by Kim, B. (2025).

therapeutic effect, which the authors acknowledge may be the reason for liver and spleen enlargement in both the anti-miR-10b and control oligonucleotide treatment mice relative to PBS treatment. To further optimize the use of ASOs as therapeutics, extensive research has gone into the formulation of vehicles for ASO delivery. Various delivery strategies have been employed to increase the delivery efficiency of ASOs to cancers such as conjugation to cell-penetrating peptides, antibodies, and vitamins, such as folate.⁴⁵ While these constructs indeed increased the circulation half-life of ASOs and delivery to tumors, one disadvantage of these approaches is the difficulty in tracking accumulation of these constructs in the clinic. At the forefront of delivery methods for miR-10b ASOs are nanoparticles, which have been widely used as vehicles of various payloads, including nucleic acids, to increase delivery to the tumor (Figure 1). Furthermore, nanoparticles are remarkably versatile, allowing them to be engineered for unique and specific properties with precise surface modifications, tailored sizes, and targeting moieties, making them ideal platforms for drug delivery. Due to the modular nature of nanoparticles, a wide variety of pre-clinically and clinically relevant imaging modalities, such as fluorescence imaging and magnetic resonance imaging, respectively, can be conferred by surface modification or nanoparticle core design. In early studies, polylysine nanoparticles

carrying anti-miR-10b ASOs demonstrated their ability to inhibit invasion and migration; however these particles were not carried forward into *in vivo* testing.⁴⁶

To date, one of the more promising treatment approaches involves the conjugation of anti-miR-10b ASOs to an iron oxide magnetic nanoparticle (MN) core. This compound, termed MN-anti-miR10b, was first administered intravenously to mice bearing implanted MDA-MB-231 breast cancer cells.⁹ Similar to the study using an unconjugated ASO, weekly MN-anti-miR10b (10 mg Fe/kg bodyweight) had no effect on primary tumor size; however, its administration inhibited the formation of lymph node metastases and suppressed the growth of established metastatic tumors, as visualized by bioluminescence imaging (BLI). Histological analyses of lymph node metastases revealed a significant reduction in cell proliferation but not an increase in apoptosis. In a subsequent study, MN-anti-miR10b was used with adjuvant doxorubicin in mice with established spontaneous metastases, and the combination induced stable regression of the metastases by the fourth and final week of treatment.⁸ At sacrifice at week 12, there was no evidence of metastasis in the lymph nodes by histology. Importantly, the therapeutic study was initially performed in an immunocompromised model of breast cancer metastases (MDA-MB-231 cells) and was then reproduced in an

immunocompetent model (4T1 cells),⁴⁷ and its activity was demonstrated in a case report in a feline model of breast cancer.⁴⁸ In all these studies, treatment with MN-anti-miR10b did not appear to cause adverse effects.^{8,9,48}

In 2023, MN-anti-miR10b, under the name TTX-MC138 (TransCode Therapeutics), completed an early Phase I clinical trial for its use in “advanced solid tumors”,⁴⁹ and a multicenter Phase I/II study began in 2024.⁵⁰ To date, two cohorts of patients have been administered TTX-MC138 with no reports of significant safety concerns or dose-limiting toxicities.⁵¹ An increased dosage of TTX-MC138 administered to cohort two resulted in greater activity while remaining safe for patients. A third cohort is currently in recruitment for further dosage escalation studies.

Building on the mounting evidence implicating other miRNAs in multiple cancers, especially metastatic breast cancer, recent studies have explored targeting other miRNAs in addition to miR-10b. Devulapally et al investigated the anti-tumor efficacy of PLGA-b-PEG nanoparticles coloaded with anti-miR-10b and anti-miR-21 (another miRNA implicated in breast cancer) ASOs and decorated with uPA-peptide for tumor cell targeting.⁵² These PLGA-b-PEG nanoparticles decreased tumor growth in a subcutaneous, triple-negative breast cancer model after systemic administration. Interestingly, breast cancer cells treated with co-loaded anti-miR-10b and anti-miR-21 PLGA-b-PEG nanoparticles prior to intravenous injection formed tumors significantly slower than untreated cells. In another study, mesoporous silica nanoparticles loaded with anti-miR-10b ASOs and miR-34 mimic was shown to decrease tumor growth in both *ex ovo* and orthotopic *in vivo* models of human triple-negative breast cancer.⁵³

An important consideration when using nanoparticles as a delivery vehicle for anti-miR-10b oligonucleotides is the accumulation of these therapeutic nanoparticles in off-target sites. Systemically injected nanoparticles are commonly subject to accumulation in the reticuloendothelial system organs, such as the liver and spleen. Importantly, systemic inhibition of miR-10b does not seem to be associated with adverse effects, as reported by the various studies discussed above.^{8,9,41,48} Targeted nanoparticles or biomimetic carriers, such as leukosomes,⁵⁴ could improve delivery efficiency of anti-miR-10b oligonucleotides in future studies to improve therapeutic outcomes.

Beyond ASOs, another approach to miR-10b inhibition is the repurposing of existing compounds. One group used a luciferase reporter to screen 450 small molecule inhibitors for miR-10b suppression, identifying thirteen compounds.⁵⁵ Of these, the vascular endothelial and platelet derived growth factors inhibitor linifanib was determined to be the only molecule that suppressed miR-10b specifically, finding no effect on five other clinically relevant miRNAs, including miR-10a. Administration of once-daily oral linifanib to mice implanted with bioluminescent MDA-MB-231 breast cancer cells led to tumor bioluminescence, volume, and weight comparable to that of mice treated with liposome-delivered anti-miR-10b ASO, both of which showed decreases in all three parameters relative to vehicle and scrambled ASO controls.⁵⁵ As it has

already demonstrated therapeutic potential in clinical trials for various cancers, including colorectal,⁵⁶ lung,⁵⁷ and hematologic,⁵⁸ linifanib has unique potential as a readily available miR-10b inhibitor; however, adverse effects may ultimately limit its use.⁵⁵ Another non-ASO-based approach toward miR-10b inhibition is the use of anti-miR-10b CRISPR plasmid constructs delivered via lipid-polymer nanoparticles. In this system, payload delivery to the tumor was increased by destruction of the nanoparticle by focused ultrasound.⁵⁹ Consistent with other approaches, these nanoparticles failed to reduce primary tumor volume but reduced the number of metastatic lung nodules, further demonstrating the cruciality of miR-10b in breast cancer metastasis development.

While extensively studied in breast cancer, anti-miR-10b nanoparticle therapy has been studied in other cancers including colorectal cancer and glioblastoma. Wang et al investigated the therapeutic efficacy of a novel EGFR-targeted quantum dot nanoparticle to co-deliver the chemotherapy 5-fluorouracil and anti-miR-10b ASO in human colorectal cancer xenografts, resulting in decreased growth of primary tumors and reduced development of metastases in established human cell line xenograft models.⁶⁰ Multiple nanoparticle constructs have been developed to target miR-10b in glioblastoma. Teplyuk et al reported on a novel lipid nanoparticle loaded with anti-miR-10b ASOs which was continuously delivered intracranially into the tumor via osmotic pump leading to reduction in orthotopic glioblastoma growth.⁶¹ However, it was insufficient for complete elimination of the tumor. Other groups have explored the co-delivery of anti-miR-10b and anti-miR-21 ASOs as a therapeutic strategy against glioblastoma using an cRGD-targeted PLGA nanoparticle alongside temozolomide (TMZ).^{62,63} Chen et al demonstrated the therapeutic potential of MN-anti-miR10b and cytotoxic synergy with TMZ in glioblastoma cell lines *in vitro*,⁶⁴ resembling the apparent synergistic effect seen with MN-anti-miR10b and doxorubicin. Interestingly, sequential treatment with MN-anti-miR10b followed by TMZ increased cell killing, suggesting that anti-miR-10b therapy may sensitize cancer cells to chemotherapy. These findings could potentially be translated into the clinic to bolster the effects of traditional chemotherapies and perhaps lower the dose of chemotherapy needed for effective treatment.

Conclusion

In summary, many studies convincingly show that miR-10b overexpression in breast cancer correlates with worse prognosis and disease state. Based on these observations, strong cases have been made that miR-10b is both a robust diagnostic biomarker and specific therapeutic target for metastatic breast cancer. In fact, elevated levels of miR-10b in serum was found to be predictive of worse prognosis in melanoma, non-small cell lung cancer, and pancreatic adenocarcinoma, further supporting the utility of miR-10b as a diagnostic marker.^{65–67} Low miR-10b expression in normal tissues, along with the evidence that miR-10b knock-out mice develop normally, indicate that systemic administration of anti-miR-10b therapeutics could be an avenue of precision therapy for metastatic breast cancers.

Moreover, miR-10b has been implicated in the tumorigenesis of many other cancers of metastatic and non-metastatic nature, such as pancreatic cancer, colorectal cancer, and glioblastoma, and has been shown to be effective targets in these cancers. Importantly, in some studies, anti-miR-10b therapy synergized with chemotherapy to improve therapeutic efficacy, suggesting that there is a potential to use it as an adjuvant treatment. Taken together, the critical role of miR-10b in cancer progression in breast cancer as well as other cancers makes it a promising target for novel cancer therapeutics and a potential arm in combination therapy.

Acknowledgments

Portions of this manuscript are derived from the introduction chapter of the dissertation of Dr Alan Halim. The chapter had not previously been peer reviewed. The text is reproduced here with permission from the copyright holder, and the complete dissertation is publicly available.

ORCID iDs

Alan Halim  <https://orcid.org/0000-0001-9419-4552>

Bryan Kim  <https://orcid.org/0000-0001-8282-3366>

Anna Moore  <https://orcid.org/0000-0002-5218-7204>

Ethics Statement

N/A for mini-review as we report on other people's findings.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was supported in part by the National Institute of Health [R01CA221771, R01CA261691].

Declaration of Conflicting Interests

The author(s) declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: A.M. is a co-founder and shareholder of TransCode Therapeutics Inc. Other authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

References

- Kim VN. MicroRNA biogenesis: Coordinated cropping and dicing. *Nat Rev Mol Cell Biol.* 2005;6(5):376-385.
- Lee RC, Feinbaum RL, Ambros V. The *C. elegans* heterochronic gene *lin-4* encodes small RNAs with antisense complementarity to *lin-14*. *Cell.* 1993;75(5):843-854.
- Calin GA, Dumitru CD, Shimizu M, et al. Frequent deletions and down-regulation of microRNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A.* 2002;99(24):15524-15529.
- Ma L, Teruya-Feldstein J, Weinberg RA. Tumour invasion and metastasis initiated by microRNA-10b in breast cancer. *Nature.* 2007;449(7163):682-688.
- Ahmad A, Ginnebaugh KR, Yin S, Bollig-Fischer A, Reddy KB, Sarkar FH. Functional role of miR-10b in tamoxifen resistance of ER-positive breast cancer cells through down-regulation of HDAC4. *BMC Cancer.* 2015;15:540.
- Bahena-Ocampo I, Espinosa M, Ceballos-Cancino G, et al. miR-10b expression in breast cancer stem cells supports self-renewal through negative PTEN regulation and sustained AKT activation. *EMBO Rep.* 2016;17(5):648-658.
- Gastelum-Lopez MLA, Aguilar-Medina M, Garcia Mata C, et al. Organotypic 3D cell-architecture impacts the expression pattern of miRNAs-mRNAs network in breast cancer SKBR3 cells. *Noncoding RNA.* 2023;9(6):66.
- Yoo B, Kavishwar A, Ross A, et al. Combining miR-10b-targeted nanotherapy with low-dose doxorubicin elicits durable regressions of metastatic breast cancer. *Cancer Res.* 2015;75(20):4407-4415.
- Yigit MV, Ghosh SK, Kumar M, et al. Context-dependent differences in miR-10b breast oncogenesis can be targeted for the prevention and arrest of lymph node metastasis. *Oncogene.* 2013;32(12):1530-1538.
- Yoo B, Greninger P, Stein GT, et al. Potent and selective effect of the mir-10b inhibitor MN-anti-mir10b in human cancer cells of diverse primary disease origin. *PLoS One.* 2018;13(7):e0201046.
- Sheedy P, Medarova Z. The fundamental role of miR-10b in metastatic cancer. *Am J Cancer Res.* 2018;8(9):1674-1688.
- Ma L. Role of miR-10b in breast cancer metastasis. *Breast Cancer Res.* 2010;12(5):210.
- Dillekas H, Rogers MS, Straume O. Are 90% of deaths from cancer caused by metastases? *Cancer Med.* 2019;8(12):5574-5576.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell.* 2000;100(1):57-70.
- American Cancer Society: *Breast Cancer Facts & Figures 2022-2024*. American Cancer Society, Inc; 2022.
- Liu Y, Zhao J, Zhang PY, et al. MicroRNA-10b targets E-cadherin and modulates breast cancer metastasis. *Med Sci Monit.* 2012;18(8):BR299-BR308.
- Zhang J, Yang J, Zhang X, Xu J, Sun Y, Zhang P. MicroRNA-10b expression in breast cancer and its clinical association. *PLoS One.* 2018;13(2):e0192509.
- Liu X, Guan Y, Wang L, Niu Y. MicroRNA-10b expression in node-negative breast cancer-correlation with metastasis and angiogenesis. *Oncol Lett.* 2017;14(5):5845-5852.
- Bertoli G, Cava C, Castiglioni I. MicroRNAs: New biomarkers for diagnosis, prognosis, therapy prediction and therapeutic tools for breast cancer. *Theranostics.* 2015;5(10):1122-1143.
- Raychaudhuri M, Schuster T, Buchner T, et al. Intratumoral heterogeneity of microRNA expression in breast cancer. *J Mol Diagn.* 2012;14(4):376-384.
- Iorio MV, Ferracin M, Liu CG, et al. MicroRNA gene expression deregulation in human breast cancer. *Cancer Res.* 2005;65(16):7065-7070.
- Markou A, Yousef GM, Stathopoulos E, Georgoulas V, Lianidou E. Prognostic significance of metastasis-related microRNAs in early breast cancer patients with a long follow-up. *Clin Chem.* 2014;60(1):197-205.
- Gasch C, Plummer PN, Jovanovic L, et al. Heterogeneity of miR-10b expression in circulating tumor cells. *Sci Rep.* 2015;5:15980.

24. Glynn CL, Khan S, Kerin MJ, Dwyer RM. Isolation of secreted microRNAs (miRNAs) from cell-conditioned media. *Microna*. 2013;2(1):14-19.
25. Singh R, Pochampally R, Watabe K, Lu Z, Mo YY. Exosome-mediated transfer of miR-10b promotes cell invasion in breast cancer. *Mol Cancer*. 2014;13:256.
26. Mar-Aguilar F, Mendoza-Ramirez JA, Malagon-Santiago I, et al. Serum circulating microRNA profiling for identification of potential breast cancer biomarkers. *Dis Markers*. 2013;34(3):163-169.
27. Khalighfard S, Alizadeh AM, Irani S, Omranipour R. Plasma miR-21, miR-155, miR-10b, and Let-7a as the potential biomarkers for the monitoring of breast cancer patients. *Sci Rep*. 2018; 8(1):17981.
28. Hou MF, Chen YL, Tseng TF, et al. Evaluation of serum CA27.29, CA15-3 and CEA in patients with breast cancer. *Kaohsiung J Med Sci*. 1999;15(9):520-528.
29. Eissa S, Matboli M, Shehata HH, Essawy NO. MicroRNA-10b and minichromosome maintenance complex component 5 gene as prognostic biomarkers in breast cancer. *Tumour Biol*. 2015; 36(6):4487-4494.
30. Huang Q, Song Q, Zhong W, Chen Y, Liang L. MicroRNA-10b and the clinical outcomes of various cancers: A systematic review and meta-analysis. *Clin Chim Acta*. 2017;474:14-22.
31. Xu SB, Fan RH, Qin X, Han RM. microRNA prognostic signature for postoperative success of metastatic orthopedic cancers: Implications for precision microsurgery. *Front Cell Dev Biol*. 2021;9:704505.
32. Davey MG, Abbas R, Kerin EP, et al. Circulating microRNAs can predict chemotherapy-induced toxicities in patients being treated for primary breast cancer. *Breast Cancer Res Treat*. 2023;202(1): 73-81.
33. Chen W, Cai F, Zhang B, Barekati Z, Zhong XY. The level of circulating miRNA-10b and miRNA-373 in detecting lymph node metastasis of breast cancer: Potential biomarkers. *Tumour Biol*. 2013;34(1):455-462.
34. Mangolini A, Ferracin M, Zanzi MV, et al. Diagnostic and prognostic microRNAs in the serum of breast cancer patients measured by droplet digital PCR. *Biomark Res*. 2015;3(1):12.
35. Baffa R, Fassan M, Volinia S, et al. MicroRNA expression profiling of human metastatic cancers identifies cancer gene targets. *J Pathol*. 2009;219(2):214-221.
36. Diener C, Keller A, Meese E. Emerging concepts of miRNA therapeutics: From cells to clinic. *Trends Genet*. 2022;38(6): 613-626.
37. Rupaimoole R, Slack FJ. MicroRNA therapeutics: Towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov*. 2017;16(3):203-222.
38. Kim T, Croce CM. MicroRNA: Trends in clinical trials of cancer diagnosis and therapy strategies. *Exp Mol Med*. 2023;55(7): 1314-1321.
39. Lam JK, Chow MY, Zhang Y, Leung SW. siRNA versus miRNA as therapeutics for gene silencing. *Mol Ther Nucleic Acids*. 2015; 4(9):e252.
40. Selbach M, Schwanhauser B, Thierfelder N, Fang Z, Khanin R, Rajewsky N. Widespread changes in protein synthesis induced by microRNAs. *Nature*. 2008;455(7209):58-63.
41. Kim J, Siverly AN, Chen D, et al. Ablation of miR-10b suppresses oncogene-induced mammary tumorigenesis and metastasis and reactivates tumor-suppressive pathways. *Cancer Res*. 2016;76(21):6424-6435.
42. Han X, Yan S, Weijie Z, et al. Critical role of miR-10b in transforming growth factor-beta1-induced epithelial-mesenchymal transition in breast cancer. *Cancer Gene Ther*. 2014;21(2):60-67.
43. Yigit MV, Moore A, Medarova Z. Magnetic nanoparticles for cancer diagnosis and therapy. *Pharm Res*. 2012;29(5): 1180-1188.
44. Ma L, Reinhardt F, Pan E, et al. Therapeutic silencing of miR-10b inhibits metastasis in a mouse mammary tumor model. *Nat Biotechnol*. 2010;28(4):341-347.
45. Yu B, Zhao X, Lee LJ, Lee RJ. Targeted delivery systems for oligonucleotide therapeutics. *AAPS J*. 2009;11(1):195-203.
46. Jin H, Yu Y, Chrisler WB, Xiong Y, Hu D, Lei C. Delivery of MicroRNA-10b with polylysine nanoparticles for inhibition of breast cancer cell wound healing. *Breast Cancer (Auckl)*. 2012; 6:9-19.
47. Yoo B, Kavishwar A, Wang P, et al. Therapy targeted to the metastatic niche is effective in a model of stage IV breast cancer. *Sci Rep*. 2017;7:45060.
48. Savan NA, Saavedra PV, Halim A, et al. Case report: MicroRNA-10b as a therapeutic target in feline metastatic mammary carcinoma and its implications for human clinical trials. *Front Oncol*. 2022;12:959630.
49. An Open-Label, Single-Center, Phase 0, Microdose Study to Demonstrate Delivery of TTX-MC138-NODAGA-Cu64 to Radiographically Confirmed Metastases in Subjects With Advanced Solid Tumors [article online], 2023. Available from <https://clinicaltrials.gov/study/NCT05908773>.
50. A Phase 1/2 Multicenter, Open-Label, Dose-escalation and Expansion Study of TTX-MC138 in Subjects With Advanced Solid Tumors [article online], 2023. Available from <https://clinicaltrials.gov/study/NCT06260774>.
51. TransCode Therapeutics Announces Completion of Cohort 3 Initial Dosing with Lead Candidate in Phase 1 Clinical Trial [article online], 2025. Available from <https://ir.transcodetherapeutics.com/news-releases/news-release-details/transcode-therapeutics-announces-completion-cohort-3-initial>.
52. Devulapally R, Sekar NM, Sekar TV, et al. Polymer nanoparticles mediated codelivery of anti-miR-10b and anti-miR-21 for achieving triple negative breast cancer therapy. *ACS Nano*. 2015;9(3): 2290-2302.
53. Ahir M, Upadhyay P, Ghosh A, et al. Delivery of dual miRNA through CD44-targeted mesoporous silica nanoparticles for enhanced and effective triple-negative breast cancer therapy. *Biomater Sci*. 2020;8(10):2939-2954.
54. Shadbad MA, Safaei S, Brunetti O, et al. A systematic review on the therapeutic potentiality of PD-L1-inhibiting MicroRNAs for triple-negative breast cancer: Toward single-cell sequencing-guided biomimetic delivery. *Genes (Basel)*. 2021;12(8):1206.
55. Monroig-Bosque PDC, Shah MY, Fu X, et al. OncomiR-10b hijacks the small molecule inhibitor linifanib in human cancers. *Sci Rep*. 2018;8(1):13106.

56. Chan E, Goff LW, Cardin DB, et al. Phase II study of the Multikinase inhibitor of angiogenesis, Linifanib, in patients with metastatic and refractory colorectal cancer expressing mutated KRAS. *Invest New Drugs*. 2017;35(4):491-498.
57. Ramalingam SS, Shtivelband M, Soo RA, et al. Randomized phase II study of carboplatin and paclitaxel with either linifanib or placebo for advanced nonsquamous non-small-cell lung cancer. *J Clin Oncol*. 2015;33(5):433-441.
58. Wang ES, Yee K, Koh LP, et al. Phase 1 trial of linifanib (ABT-869) in patients with refractory or relapsed acute myeloid leukemia. *Leuk Lymphoma*. 2012;53(8):1543-1551.
59. Li Y, Wu P, Zhu M, et al. High-Performance delivery of a CRISPR interference system via lipid-polymer hybrid nanoparticles combined with ultrasound-mediated microbubble destruction for tumor-specific gene repression. *Adv Healthc Mater*. 2023;12(10):e2203082.
60. Wang D, Wang L, Zheng L, et al. Enhancing the management of metastatic tumors by robust co-delivery of 5-fluorouracil/MicroRNA-10b inhibitor using EGFR-targeted nanovehicles. *Adv Healthc Mater*. 2023;12(15):e2202989.
61. Teplyuk NM, Uhlmann EJ, Gabriely G, et al. Therapeutic potential of targeting microRNA-10b in established intracranial glioblastoma: First steps toward the clinic. *EMBO Mol Med*. 2016;8(3):268-287.
62. Malhotra M, Sekar TV, Ananta JS, et al. Targeted nanoparticle delivery of therapeutic antisense microRNAs presensitizes glioblastoma cells to lower effective doses of temozolomide in vitro and in a mouse model. *Oncotarget*. 2018;9(30):21478-21494.
63. Ananta JS, Paulmurugan R, Massoud TF. Tailored nanoparticle codelivery of antimiR-21 and antimiR-10b augments glioblastoma cell kill by temozolomide: Toward a “personalized” anti-microRNA therapy. *Mol Pharm*. 2016;13(9):3164-3175.
64. Chen M, Kim B, Robertson N, Mondal SK, Medarova Z, Moore A. Co-administration of temozolomide (TMZ) and the experimental therapeutic targeting miR-10b, profoundly affects the tumorigenic phenotype of human glioblastoma cells. *Front Mol Biosci*. 2023;10:1179343.
65. Bai M, Zhang H, Si L, Yu N, Zeng A, Zhao R. Upregulation of Serum miR-10b is associated with poor prognosis in patients with melanoma. *J Cancer*. 2017;8(13):2487-2491.
66. Muratore S, Zeng X, Korc M, et al. Metastatic pancreatic adenocarcinoma after total pancreatectomy islet autotransplantation for chronic pancreatitis. *Am J Transplant*. 2016;16(9):2747-2752.
67. Setiawan L, Setiabudy R, Kresno SB, et al. Circulating miR-10b, soluble urokinase-type plasminogen activator receptor, and plasminogen activator inhibitor-1 as predictors of non-small cell lung cancer progression and treatment response. *Cancer Biomark*. 2024;39(2):137-153.