

Draft Genome Sequence of Antarctic Methanogen Enriched from Dry Valley Permafrost

Joy Buongiorno,^a Jordan T. Bird,^a Kirill Krivushin,^{b*} Victoria Oshurkova,^c Victoria Shcherbakova,^c Elizaveta M. Rivkina,^b Karen G. Lloyd,^a Tatiana A. Vishnivetskaya^a

Department of Microbiology, University of Tennessee, Knoxville, Tennessee, USA^a; Institute of Physicochemical and Biological Problems in Soil Science, Russian Academy of Sciences, Pushchino, Russia^b; Skryabin Institute of Biochemistry and Physiology of Microorganisms, Russian Academy of Sciences, Pushchino, Russia^c

* Present address: Kirill Krivushin, University of Alberta, Livestock Gentec, Department of Agricultural, Food and Nutritional Science (AFNS), Edmonton, Alberta, Canada.

A genomic reconstruction belonging to the genus *Methanosarcina* was assembled from metagenomic data from a methane-producing enrichment of Antarctic permafrost. This is the first methanogen genome reported from permafrost of the Dry Valleys and can help shed light on future climate-affected methane dynamics.

Received 11 October 2016 Accepted 14 October 2016 Published 8 December 2016

Citation Buongiorno J, Bird JT, Krivushin K, Oshurkova V, Shcherbakova V, Rivkina EM, Lloyd KG, Vishnivetskaya TA. 2016. Draft genome sequence of Antarctic methanogen enriched from dry valley permafrost. *Genome Announc* 4(6):e01362-16. doi:10.1128/genomeA.01362-16.

Copyright © 2016 Buongiorno et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Karen G. Lloyd, klloyd@utk.edu, or Tatiana A. Vishnivetskaya, tvishniv@utk.edu.

Permafrost currently contributes nearly 25% of all naturally sourced methane (1), a value that is predicted to rise significantly in coming decades (2). However, methane accumulation in permafrost environments is complex and geographically variable, making the trajectory of climate-affected methane dynamics hard to predict. Late Pleistocene permafrost from the Miers Valley (McMurdo Dry Valleys) contained methane in shallow horizons, where isotopic signatures suggested biogenic methane sources (3). In order to determine the biogenicity and timing of methane accumulation, incubation experiments were conducted. Here, we announce a nearly complete genome binned from those methane-producing enrichments.

Anaerobic incubations of permafrost consisted of phosphate-buffered basal medium (4) and gas mixture of H₂/CO₂ (80/20) at 20°C. Methane production was first observed after one year of incubation and is ongoing today (11 years later). After seven years, samples were collected for metagenome sequencing. The total community genomic DNA from the enrichment was extracted using the PowerSoil DNA isolation kit (Mo Bio Laboratories, Inc., Carlsbad, CA, USA), and the DNA library was prepared using the TruSeq DNA sample prep kit version 2 without whole-genome amplification. The Illumina HiSeq 2000 platform was used to acquire paired-end 2 × 100-bp metagenomic reads. Adaptors and low-quality reads were trimmed with the Trimmomatic software (5). VizBin (6) was used to bin together contigs of similar coverage and *k*-mer frequency. Metagenomic binning resulted in recovery of a nearly complete methanogenic genome, determined to be 99.84% complete using the *Euryarchaeota*-specific marker set of housekeeping genes (7), with low contamination (1.41%) and 0% strain heterogeneity.

The genomic reconstruction contained 342 contigs over 1,000 bp in length, with an average coverage of 570× and 38% GC content. RNAmmer (8) identified the 16S rRNA sequence, which BLASTn analysis shows to have 97% nucleotide sequence identity and 100% coverage to *Methanosarcina lacustris*, a psychrotolerant

methanogen isolated from a fen in Moscow (9). Close relatives are *M. subterranea* strain HC-2 and *M. soligelidi* strain DSM 26065, isolated from a deep-subsurface diatomaceous shale formation and Siberian permafrost-affected soil, respectively (10, 11).

Annotation of protein-coding sequences was conducted with Prokka (12). The genome contained 3,593 coding regions, 53 tRNAs, 11 predicted CRISPR regions, and several cytochromes. The entire operon encoding methyl coenzyme M reductase (*Mcr*) and genes for hydrogenotrophic methanogenesis (*fmd*, *ftr*, *mch*, *mta*, *mer*, *mtrABCDEFGHI*, and *hdrABCDE*) were present. Acetoclastic genes encoding carbon monoxide dehydrogenase, acetate kinase, acetyl-coenzyme A synthetase, phosphate acetyltransferase, and the acetyl-CoA decarbonylase/synthase complex provide evidence that this organism is capable of acetoclastic methanogenesis. Methanol metabolism genes encoding the three subunits of methanol—corrinoid protein comethyltransferase—show potential for growth with methanol. The genome contains monomethylamine methyltransferase and dimethylamine corrinoid protein genes, suggesting growth with methylamines. An incomplete formate dehydrogenase operon suggests that growth with formate is not likely.

The genome contains evidence for *de novo* unsaturated diether lipid construction through a functional mevalonate pathway, a signature of adaptation to permanently cold environments (13). DNA double-strand break repair Rad50 ATPase and several heat-shock proteins were detected, indicating that several defense strategies against environmental stress are available to this methanogen.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [MCHG000000000](https://www.ncbi.nlm.nih.gov/assembly/GCA000000000/) for the entire metagenome and [MDTP000000000](https://www.ncbi.nlm.nih.gov/assembly/GCA000000000/) for the reconstructed genome. The versions described in this paper are versions MCHG01000000 and MDTP02000000, respectively.

ACKNOWLEDGMENT

We thank Maria Logacheva from the Laboratory of Evolutionary Genomics of the Moscow State University for help with metagenome sequencing.

FUNDING INFORMATION

This work was funded by National Science Foundation (NSF) (DEB-1442262) for K.G.L., and T.A.V., OCE-1431598 for J.T.B., Center for Dark Energy Biosphere Investigations OCE-0939564 contribution #339 for J.B.), and Russian Foundation for Basic Research (RFBR), 14-14-01115 and 12-04-31416 for K.K. and E.M.R.

REFERENCES

1. Wagner D, Lipski A, Embacher A, Gattinger A. 2005. Methane fluxes in permafrost habitats of the Lena Delta: effects of microbial community structure and organic matter quality. *Environ Microbiol* 7:1582–1592. <http://dx.doi.org/10.1111/j.1462-2920.2005.00849.x>.
2. Anisimov OA. 2007. Potential feedback of thawing permafrost to the global climate system through methane emission. *Environ Res Lett* 2:045016. <http://dx.doi.org/10.1088/1748-9326/2/4/045016>.
3. Gilichinsky DA, Wilson GS, Friedmann EI, McKay CP, Sletten RS, Rivkina EM, Vishnivetskaya TA, Erokhina LG, Ivanushkina NE, Kochkina GA, Shcherbakova VA, Soina VS, Spirina EV, Vorobyova EA, Fyodorov-Davydov DG, Hallet B, Ozerskaya SM, Sorokovikov VA, Laurinavichyus KS, Shatilovich AV. 2007. Microbial populations in Antarctic permafrost: biodiversity, state, age, and implication for astrobiology. *Astrobiology* 7:275–311. <http://dx.doi.org/10.1089/ast.2006.0012>.
4. Kenealy W, Zeikus JG. 1981. Influence of corrinoid antagonists on methanogen metabolism. *J Bacteriol* 146:133–140.
5. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <http://dx.doi.org/10.1093/bioinformatics/btu170>.
6. Laczný CC, Sternal T, Plugaru V, Gawron P, Atashpendar A, Margosian HH, Coronado S, der Maaten Lv, Vlassis N, Wilmes P. 2015. VizBin—an application for reference-independent visualization and human-augmented binning of metagenomic data. *Microbiome* 3:1. <http://dx.doi.org/10.1186/s40168-014-0066-1>.
7. Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <http://dx.doi.org/10.1101/gr.186072.114>.
8. Lagesen K, Hallin P, Rødland EA, Staerfeldt H-H, Rognes T, Ussery DW. 2007. RNAMmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
9. Zhilina T, Zavarzin G. 1991. Low-temperature methane production by pure culture of *Methanosarcina* sp. *Dokl Akad Nauk SSSR* 317:1242–1245.
10. Shimizu S, Ueno A, Naganuma T, Kaneko K. 2015. *Methanosarcina subterranea* sp. nov., a methanogenic archaeon isolated from a deep sub-surface diatomaceous shale formation. *Int J Syst Evol Microbiol* 65: 1167–1171. <http://dx.doi.org/10.1099/ijs.0.000072>.
11. Wagner D, Schirmack J, Ganzert L, Morozova D, Mangelsdorf K. 2013. *Methanosarcina soligelidi* sp. nov., a desiccation- and freeze-thaw-resistant methanogenic archaeon from a Siberian permafrost-affected soil. *Int J Syst Evol Microbiol* 63:2986–2991. <http://dx.doi.org/10.1099/ijs.0.046565-0>.
12. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
13. Nichols DS, Miller MR, Davies NW, Goodchild A, Raftery M, Cavicchioli R. 2004. Cold adaptation in the Antarctic archaeon *Methanococcus burtonii* involves membrane lipid unsaturation. *J Bacteriol* 186: 8508–8515. <http://dx.doi.org/10.1128/JB.186.24.8508-8515.2004>.