



Anti-Apoptotic Role of Sanhuang Xiexin Decoction and Anisodamine in Endotoxemia

Zixuan Liu^{1,2†}, Wenxiang Wang^{1,2†}, Jie Luo^{1,2}, Yingrui Zhang^{1,2}, Yunsen Zhang², Zhiqiang Gan^{1,2}, Xiaofei Shen³, Yi Zhang^{1,2*} and Xianli Meng^{4*}

¹Ethnic Medicine Academic Heritage Innovation Research Center, Chengdu University of Traditional Chinese Medicine, Chengdu, China, ²School of Ethnic Medicine, Chengdu University of Traditional Chinese Medicine, Chengdu, China, ³TCM Regulating Metabolic Diseases Key Laboratory of Sichuan Province, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, China, ⁴Innovative Institutes of Chinese Medicine and Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu, China

OPEN ACCESS

Edited by:

Hong Zhang, Shanghai University of Traditional Chinese Medicine, China

Reviewed by:

Jwu-Lai Yeh, Kaohsiung Medical University, Taiwan Ina Yosifova Aneva, Bulgarian Academy of Sciences, Bulgaria

*Correspondence:

Yi Zhang zhangyi@cdutcm.edu.cn Xianli Meng xlm999@cdutcm.edu.cn

⁺These authors have contributed equally to this work

Specialty section:

This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

Received: 20 February 2020 Accepted: 24 March 2021 Published: 21 April 2021

Citation:

Liu Z, Wang W, Luo J, Zhang Y, Zhang Y, Gan Z, Shen X, Zhang Y and Meng X (2021) Anti-Apoptotic Role of Sanhuang Xiexin Decoction and Anisodamine in Endotoxemia. Front. Pharmacol. 12:531325. doi: 10.3389/fphar.2021.531325 Endotoxemia is characterized by initial uncontrollable inflammation, terminal immune paralysis, significant cell apoptosis and tissue injury, which can aggravate or induce multiple diseases and become one of the complications of many diseases. Therefore, anti-inflammatory and anti-apoptotic therapy is a valuable strategy for the treatment of endotoxemia-induced tissue injury. Traditional Chinese medicine exhibits great advantages in the treatment of endotoxemia. In this review, we have analyzed and summarized the active ingredients and their metabolites of Sanhuang Xiexin Decoction, a famous formula in endotoxemia therapy. We then have summarized the mechanisms of Sanhuang Xiexin Decoction against endotoxemia and its mediated tissue injury. Furthermore, silico strategy was used to evaluate the anti-apoptotic mechanism of anisodamine, a well-known natural product that widely used to improve survival in patients with septic shock. Finally, we also have summarized other anti-apoptotic natural products as well as their therapeutic effects on endotoxemia and its mediated tissue injury.

Keywords: endotoxemia, anti-apoptosis, immunosuppression, herbal medicine, sanhuang xiexin decoction, anisodamine

INTRODUCTION

Possible sources of plasma endotoxin are bacteria from infected local tissues, blood, respiratory tract, digestive tract, food, or other ingested matter (Munford, 2016). The intestinal and other epithelial cells, such as those of the skin or lungs, are the first line of defense against endotoxins entering the bloodstream. Bacteria release bacterial lipopolysaccharide (LPS) or endotoxins mainly through lymphatic channels (e.g., small intestinal lymphatic system) and thoracic duct into the circulation from infected parts. Furthermore,

1

Abbreviations APAF1, Apoptotic protease activating factor-1; BAK1, B-Cell leukemia-2 antagonist killer-1; BAX, Bcl2 associated X-protein; Bcl2, B-Cell leukemia-2; BID, BH3 interacting death domain; BIM, B-Cell leukemia-2 interacting protein; CLSM, Confocal laser scanning microscopy; ECP, Eos cationic protein; EDN, Eos-derived neurotoxin; ETX, endotoxin; FADD, Fas-associated via death domain; FasL, Fas ligand; FM, Fluorescence microscopy; ICAD, Inhibitor of caspase-activated dNase; INOS, inducible nitric oxide synthase; LPS, lipopolysaccharide; MBP, Major base proteins of eosinophils; MODS, organ dysfunction syndrome; MOMP, Mitochondrial outer membrane permeabilization; NO, nitric oxide; OM, Optical microscopy; PTEN, phosphatase and tensin homolog deleted on chromosome-10; PUMA, p53-upregulated modulator of apoptosis; p53AIP1, p53-regulated apoptosis-inducing protein-1; RIP, Receptor-interacting protein; ROS, reactive oxygen species; SIRS, systemic inflammatory response syndrome; TNFR1, TNF receptor 1; TLM, Transmission electron microscopy; TRADD, Tumor necrosis factor receptor-1-associated death domain



LPS can also be absorbed from the gastrointestinal mucosa, especially the damaged intestinal mucosa (Brenchley, 2006) into the portal circulation. Erridge (Erridge, 2011) and Deitch (Deitch, 2012) showed that the largest source of LPS detected in the peripheral venous blood was from the small intestine. In addition, blood-borne bacteria can release LPS directly into the bloodstream to cause diseases, such as N. meningitidis (Ovstebo et al., 2005; Coureuil et al., 2014) and E. coli (Simpson et al., 2000). However, when the amount of endotoxin is overloaded, the host defense fails to eliminate and control the infection, leading to endotoxemia or even sepsis (Piwowar, 2014). In addition to endotoxemia/sepsis, excessive amounts of LPS are also implicated in the pathogenesis of a range of diseases, such as atherosclerosis, alcoholic liver disease, autoimmunity, metabolic syndrome, renal injury, multiple organ failure, depression, and chronic fatigue. Interestingly, these diseases both share a common pathophysiological basis: the imbalance between induction of apoptosis and resistance to apoptosis (Manco et al., 2010).

Apoptosis is a conserved process of programmed cell death characterized by chromatin fragmentation and condensation, membrane blebbing, and nuclear collapse. Apoptosis also plays important roles in a variety of physiological processes, including embryogenesis, tissue remodeling, immune response, and carcinogenesis (Kerr et al., 1972; Williams, 1991). In this process, cells that are no longer needed or that will be detrimental to an organism or tissue are disposed of in a highly ordered manner. This process can effectively block the development of inflammatory responses and tumorigenesis. **Figure 1** illustrates how endotoxemia occurs, with six apoptosis hallmarks and related assays to distinguish apoptosis cells from normal healthy ones.

It has been shown that various herbal ingredients exhibit potent anti-apoptotic effects and improve the survival rate in experimental endotoxemia models. In the last decade, an increasing number of studies have emphasized that the reduced incidence of various endotoxin-related pathologies is closely related to herbal medicine, which is a good source of beneficial natural anti-inflammatory, anti-oxidative stress, and anti-coagulant bioactive compounds. Moreover, the active ingredients of many herbs have been used to treat endotoxemia and its complications with impressive efficacy. For example, the administration of anisodamine significantly improved the survival rate of septic shock. Considering the complexity of the apoptosis-related endotoxemia mechanisms and the fact that the modulatory effects of herbs are difficult to be systematically presented in various experimental studies. Therefore, we wrote this review to provide a general and descriptive overview of the potential role and underlying mechanisms of apoptosis-related progression in endotoxemia, as well as biomarkers and assays for endotoxemia, and the bidirectional regulation of apoptosis in endotoxemia by herbal components and their metabolism (especially Sanhuang Xiexin decoction and Anisodamine).

This review explores these issues by taking a novel perspective on apoptosis regulation in the treatment of endotoxemia. It begins with the herbal ingredients that may affect the inflammatory response, visceral protection, and apoptosis of endotoxemia *in vitro* and *in vivo*. How the major herbal ingredients are metabolized will then be discussed, followed by a summary of these metabolites and their effects. The review concludes by considering the influencing factors that determine whether herbal ingredients regulating apoptosis are advantageous in therapeutic endotoxemia.

ENDOTOXEMIA IS ASSOCIATED WITH VARIOUS DISEASES

Numerous variables can be activated in endotoxemia, including circulating glucocorticoid, cytotoxic T lymphocytes, oxygen free



radicals, nitric oxide, heat shock proteins, FAS ligand, and proinflammatory cytokines (e.g., TNF- α , IL-1, and IL-6); all of these factors may initiate apoptosis (Harjai et al., 2013). Recently, regulation of the apoptotic process has been considered as a new therapeutic strategy for the treatment of endotoxemia and its complications (Eduardo and Fresno, 2013). Therefore, an indepth understanding of the molecular regulation of endotoxininduced apoptosis could provide a scientific basis for the intervention and treatment of the various diseases mentioned in **Figure 1**.

ETX INDUCES APOPTOSIS IN VITRO AND IN VIVO

ETX receptors could be divided into the following categories: scavenger receptor (SR), lipopolysaccharide-binding protein (LBP), cluster of differentiation antigen 14 (CD14) (Ulevitch and Tobias, 1995), β 2-integrins (such as CD11/CD18), and Toll-like receptors (TLRs). They are extensively distributed on the surface of immune cells such as monocytes, macrophages, and

neutrophils and play an important role in the recognition and elimination of ETX in the host. The binding of LPS to LBP activates transmembrane receptors such as SR, \u03b32-integrins, and TLRs to transmit signals into the cell. These signals further promote the activation of transcription factors and induce the expression of pro-inflammatory genes. The expression of these overproduced pro-inflammatory products can further cause the induction of apoptosis. Apoptosis is induced by the intrinsic (mitochondrial) pathway and the extrinsic (death receptormediated) pathway (Danial and Korsmeyer, 2004). The intrinsic pathway involves mitochondria and depends mainly on Bcl2 family proteins and caspase 9/7/6/3 (Hengartner, 2000; Leist and Jäättelä, 2001). The extrinsic pathway is involved in death signaling, such as TNF-a with TNF receptor 1 (TNFR1). Then, the activation of death signaling further activates caspase-8, which cleaves procaspase-3 into its active form and activates various genes such as P53, Fas, Bcl2, and NFκB alone (Micheau and Jürg, 2003; Peter and Krammer, 2003). In contrast, the NF-κB signaling cascade and p53 activate proapoptotic Bcl2 family proteins and inhibit anti-apoptotic Bcl2 family proteins and inhibitors of apoptotic proteins (cIAPs) via transcription, as well as several other p53 targets such as BAX, Noxa (Latin for damage, a BH3 protein involved only in regulating cell death decisions), p53-upregulated apoptosis regulator (PUMA) and BID to trigger apoptosis. Furthermore, p53 also transactivates several other genes that may contribute to apoptosis, including phosphatase and tensin homolog deleted on chromosome-10 (PTEN), APAF1, Perp, and genes that cause elevated levels of reactive oxygen species (ROS). The overproduction of ROS leads to extensive oxidative damage to all components of mitochondria. Mitochondrial DNA damage induced by ROS further disrupts the oxidative phosphorylation of mitochondria, which leads to a range of human diseases. Herbal ingredients are often involved in the treatment of endotoxemia with anti-inflammatory, antioxidant, and anti-apoptosis effects, which are interrelated and affect each other. Interestingly, however, there is no one-to-one correlation between anti-inflammatory/antioxidant and antiapoptotic/pro-apoptotic.

Endotoxin has been reported to significantly induce apoptosis in vitro and in vivo (O'Brien and Abraham, 2004; Neff et al., 2006; Li et al., 2018). ETX can directly stimulate the production of cytokines by monocytes-macrophages to activate the body's local or overall immune system against bacterial infections. On the other hand, if the infection is severe, large amounts of ETX in the body come into contact with mononuclear-macrophages and release vast inflammatory cytokines such as TNF-a leading to an uncontrolled inflammatory response. In addition, ETX stimulates the production of large amounts of nitric oxide (NO) by inducing non-calcium-dependent inducible nitric oxide synthase (INOS) in vascular cells. ETX also activates monocytes, macrophages, and neutrophils, and further promotes the expression of apoptosis-related receptors, such as Fas, TLR-2, and CD14, which can also induce apoptosis and tissue damage (Hotchkiss et al., 2002). Apoptosis of parenchymal cells can lead to dysfunction in multiple organs, while apoptosis of immune cells manifests as impaired host immune tolerance and immunosuppression (Hotchkiss et al., 2005), ultimately leading to septic shock, systemic inflammatory response syndrome (SIRS), organ dysfunction syndrome (MODS), and death. Interestingly, in sepsis, both pro- and anti-apoptotic members of the apoptosis-related pathway are upregulated, but the ratio is more favorable to the pro-apoptosis. In addition, lymphocytes are susceptible to endotoxin-induced apoptosis, with increased apoptosis of CD4⁺ T cells (Jäättelä and Tschopp, 2003), B cells, and follicular dendritic cells in the spleen of patients, while oxidative stress can lead to apoptosis of thymocytes (Fehsel et al., 1995). Although there is evidence that granulocytes can limit inflammatory damage through apoptosis, this is macrophage-dependent because the phagocytosis of apoptotic cells by macrophages effectively prevents the release of toxic content from dead cells. If a large number of macrophages undergo apoptosis, the apoptotic granulocytes will eventually lyse and release toxic molecules such as elastase and myeloperoxidase from neutrophils, major



FIGURE 2 | Metabolic pathways of main components of Sanhuang Xiexin Decoction.



base proteins (MBP) from eosinophils, Eos cationic protein (ECP), and Eos-derived neurotoxin (EDN), leading to further uncontrolled inflammatory responses. However, the nonimmune cells are mainly parenchymal cells of the liver, lung, and intestine (Suzuki et al., 2011; Liu H. et al., 2015; Guo et al., 2019), while cardiomyocytes (Liu et al., 2013; Zhang et al., 2015; Tian et al., 2019) and nerve cells (Peña et al., 2011; Reichardt et al., 2017) of apoptosis have also been reported.

ANTI-APOPTOSIS MECHANISM OF HERBAL MEDICINE IN ENDOTOXEMIA

It has been observed that various herbal ingredients can improve the healing of endotoxemia, not only through anti-inflammatory effects, but also through their regulation of apoptosis in many types of immune cells (including lymphocytes and macrophages), endothelial cells, and parenchymal cells in different solid organs, especially heart, liver, and lung (Cheng et al., 2014). Although apoptosis has been extensively studied in various diseases such as cancer, neurodegenerative diseases, and HIV infection, its role in sepsis and its regulation as a novel therapeutic approach in the treatment of endotoxemia has attracted attention in recent years.

Sanhuang Xiexin Decoction and Its Metabolites

Sanhuang Xiexin decoction was firstly described in the *Synopsis of Golden Chamber* by Zhongjing Zhang of the Eastern Han Dynasty and is a classical formula widely used in Chinese medicine to treat fire and detoxify. It is composed of Rhizoma Rhei (*Rheum palmatum* L.), Rhizoma Coptidis (*Coptis chinensis* Franch), and Radix Scutellaria (*Scutellaria baicalensis* Georgi). Previous studies have shown that Sanhuang Xiexin Decoction has effective therapeutic effects on endotoxemia through anti-inflammation (Lo et al., 2005a; Lo et al., 2005b). Other recent studies have shown that it also possesses potent anti-apoptotic activity. The main anti-apoptotic ingredients are aloe-emodin,



coptisine, and baicalin. Figure 2 depicts its formula and the metabolic steps and products of coptisine and baicalin in vivo. For example, aloe-emodin can attenuate myocardial infarction and cardiomyocyte apoptosis (Yu et al., 2019). The metabolites are rhein, aloe-emodin sulfates/glucuronides, and rhein sulfates/ glucuronides (Yu et al., 2016). Among them, rhein exhibited significant anti-apoptosis in endotoxemia and showed significant protective effects against endotoxin-induced kidney injury (Yu et al., 2015). Coptisine, a major compound from Rhizoma Coptidis that clears heat and dampness, purging fire for removing the toxin has been reported to exhibit protective effects on HaCaT keratinocytes by blocking the mitochondriadependent apoptotic pathway (Choi, 2019). Furthermore, baicalin is the most reported anti-apoptotic component of Sanhuang Xiexin Decoction, showing anti-apoptotic effects in a variety of cell types, including cardiomyocytes (Liou et al., 2011; Liou et al., 2012), C2C12 myoblast (Pan et al., 2019), arterial endothelial cells (Shou et al., 2017), neuronal cells (Zheng, et al., 2015), hepatic stellate cells (Wu et al., 2018a; Wu et al., 2018b), endplate chondrocytes (Pan et al., 2017), renal cells (Zhu et al., 2016) and colonic epithelial cells (Yao et al., 2016). Baicalein 6-O- β -D-glucopyranuronoside (Akao et al., 2013) and baicalein (Wang et al., 2015) are the main metabolites of baicalin that inhibit the apoptosis of intestinal epithelial cells, thus reducing the chance of endotoxemia in cirrhosis (Liu Y. et al., 2015) and cardiomyocyte injury (Zhu et al., 2019).

Molecular Docking Prediction of Anisodamine for Anti-Apoptotic Treatment of Endotoxemia

Anisodamine is a well-known natural product that is widely used to improve the survival of patients with septic shock. The binding capacity of anisodamine and its 23 intracorporal metabolites (Chen et al., 2005) to endothelial cell apoptosis-related receptors was evaluated by molecular docking technology. As shown in **Figure 3**, anisodamine and its metabolites bind mainly to TNFR1 and APO3 with high affinity. Other individual metabolites, such as N-demethyl-6 beta hydroxytropine and 1-sulfate conjugated hydroxyanisodamine also bind to FAS and APO2 with strong affinity. Thus, the therapeutic effect of anisodamine on endotoxemia involves molecular mechanisms associated with apoptosis. However, further evaluation regarding the molecular mechanisms of anisodamine *in vitro* and *in vivo* is necessary.

Other Anti-Apoptosis Ingredients Derived From Herbal Medicines

As seen in **Figure 4**, immune paralysis and organ damage are the main culprits for the poor prognosis of endotoxemia. Many herbal ingredients are effective in the treatment of endotoxemia by inhibiting apoptosis. Their effects, target organs/cells, dosage, relevant targets, and indications have been summarized in **Table 1**. The activation of the TLR4/NF- κ B pathway leads to exacerbation of inflammation and apoptosis, which in turn leads to worsening endotoxemia. Therefore, inhibition of inflammation, oxidative stress, and apoptosis is a new research focus in the treatment of endotoxemia (Guan et al., 2019; Li et al., 2019; Zhao et al., 2020).

Meng et al. confirmed that sanguinarine attenuates ETXinduced inflammation and apoptosis by blocking the TLR4/NF- κ B pathway (Meng et al., 2018). However, in general, the mechanism of herbal ingredients based on this pathway for the treatment of endotoxemia is less studied, and it remains to be verified whether the inhibition of this pathway signal can be used to evaluate and screen potential herbal ingredients. Furthermore, leukocyte adhesion molecules have been suggested as new targets

in	nti-apoptosis, nephroprotection, anti-			
Thaliporphine A	flammation, anti-oxidant	J: Serum Cr, NGAL, KIM-1, IL-18, TNF-α, IL-6; MDA, NO, MPO, iNOS. Bax/Bcl2, caspase-3/-8/-9	Sepsis complication of acute kidney injury	Fouad et al. (2016)
	nti-inflammation, anti-apoptosis; adioprotection	J: p38/NF-κB pathway; TNF-a, caspase 3, serum cTnl, LDH, Reverse steeper EDPVR, Gentler ESPVR	Cardiac depression	Lee et al. (2010)
	lepatocyte protection, anti-apoptosis, nti-oxidation	 ↑: PI3K/Akt/mTOR pathway ↓: Hydroperoxides, MDA, SOD, NADPH, POX ↑: CAT, SOD, G-POX 	Endotoxemia	Ben et al. (2000)
	nti-apoptosis, anti-oxidation, anti- flammation, immunoregulation	1: O/H, OOE, GT O/K ↓: SOD, CAT, GSH, MDA, NO, DPPH; TNF-α, IL-1β, IL-6 ↑: AST, ALT	Acute liver injury	Latha et al. (2017
Soybean oil, olive oil A	nti-inflammation, immunoregulation	 J: Mitochondrial pathway, Apoptosis of lymphocyte, Caspase-8, Bax, Bcl- xl, caspase-3, TNF-a, IL-6 †: PI3K/Akt phosphorylation 	Sepsis complications of acute lung injury and acute respiratory distress syndrome	Bi et al. (2010)
	nti-apoptosis, anti-inflammation, nmunoregulation	\downarrow : NF-κB, TNF-α, AST, NO, AST, caspase-3/-6, PARP \uparrow : Phosphorylation of MAPK (ERK1/2), SOD, IL-10	Sepsisshock complication of hepaticdysfunction	Chen and Wu. (2014)
in	nti-inflammation, pro-apoptosis, nmunoregulation, anti-platelet, anti- ypertension, antioxidation	↓: TNF-a, IL-6 ↑: IL-10, sTRAIL/Apo-2L	Preeclampsia	Makris et al. (2005)
Salvia miltiorrhizae In in ap	nprove microcirculation, anti- flammation, antioxidation, anti- poptosis, mucosal protection, nacrophage protection	J: p65NF-κB, PLA2, Bax	Severe acute pancreatitis; obstructive jaundice	Zhang et al. (2009)
Sophocarpine A	nti-inflammation, anti-apoptosis, ntioxidation	J: p38/JNK, NF-κB, PI3K/AKT, Bcl-xl, H ₂ O ₂ , O ₂ , NO, CYP2E/NRF2, CYPE2, ROS, ALT, ALT, ALP, AST, IL-1β, TNF- α, IL-6, Cyto-C, Apaf1, caspase-9/-3	Septicliver injury	Zheng et al. (2018)
0	Cytoprotection, anti-apoptosis, nmunoregulation	 ↑: SOD, CAT, GSH, SOD1, Nrf2 ↓: ROS, caspase-3, p53, mitochondrial membrane depolarization A Pale 	Endotoxemia	Wu et al. (2019)
0	ntioxidation, anti-apoptosis, anti- iflammation, antianaphylaxis	1: Bcl2 J: NF-κB, MAPK, TLR4, JNK pathways; eotaxin-1, ROS, caspase- 3, NADPH, PLCγ1-PKCβ2-NADPH	Allergic airway diseases; peribronchial eosinophilia	Cho et al. (2014)
	nti-inflammation, antioxidation, anti- brosis, antiviral	 ↑: Akt, ERK ↓: NF-κB, MIP-2, MPO, MDA, TNF- α, IL-6, IL-8, ALT, AST, LDH, ALP, NO, sICAM-1, caspase-3 	Acute liver injury, Hepaticischemia	Zhang et al. (2011)
	nti-apoptosis, geno protective, ntioxidation	J: AST, ALT, TB, ALB, NO ↑: TAC	Liver damage	Allam et al. (2017
•	nti-apoptosis	↓: MABP, iNOS, PI, IL-6, IL-18, ET- 1, NO	Septic shock	Pei et al. (2013)
12-O-Tetradecanoyl-phorbol Ai 13-Acetate (TPA)	nti-apoptosis	↓: NF-κΒ ↑: C/EBP, p38 MAPK	Endotoxemia	Sunil et al. (2002
	nti-inflammation, antioxidation, anti- poptosis	\downarrow : NF-κB, NO, iNOS, CAT, SOD, ROS, Caspase 3, PiκBa, TNF-α, IL-1β, IL-6	Human dermalfibroblast	Gasparrin et al. (2018)
Betulinicacidderivative BA5 In	nmunoregulation	†: pAMPK, SIRT1463, PGC1α, GPX, GR, GST, IL-10, Nrf2-AMPK ↓: NF-κB, TNF-α, IL-2, IL-4, IL-6, IL- 17A, IFN-γ, NO, Activated LY, CD4 ⁺	Inflammationliver injury Delayed hypersensitivity	Meira et al. (2017
	nti-inflammation, antioxidation, anti- poptosis	Tcellcalcineurin activity 1: nitrate/nitrite	Intestinaldamage	Chiang et al. (2006)
	nti-inflammation	J: NF-κΒ, NO, IL-8, IL-6, MCP-1, P-p65, P-ΙκΒα J: PI3K/AKT, TAK1 + TAB2	Septic shock	(2008) Wang et al. (2018)

Herbal medicine	Effects	Related targets	Endotoxemia related indications	References
Cinnamaldehyde (CA) Linalool (LIN)	Anti-inflammation	J: NF-κB, TLR4/MD2, MyD88, NLRP3, ASC, caspase-1, TNF-α, IL- 1β, IL-18, nitrate/nitrite, IFN-γ, HMGB-1	Endotoxemia	Lee et al. (2018)
Gryllusbimaculatus	Anti-apoptosis, anti-inflammation,	↓: ROS, NO, IL-6, TNF-α, TG, TLR4,	Alcoholic liver diseases	Hwang et al.
Extracts (GBE)	antioxidation	P-JNK, P-p38, OHdG, p-MLCK, p-ROCK, p-srcFK	Intestinal inflammation	(2019)
Scytonemin	Antiproliferative	$ \downarrow: cdc25C, p.cdc25C, Cell Endotoxemia cycleregulatory kinases, GST-PLK, GST-Tie2, Protein kinase A, Protein kinase C\beta2, CDK1/cyclin B, GST-Myt1, GST-checkpoint kinase 1, GST-polo-like kinase 1$		Stevensonet al. (2002)

	TABLE 1 (Continue	d) Effect of herbal ingredients	on anti-apoptosis related pathway	v molecular (excepted those	mentioned in the text).
--	---------------------	---------------------------------	-----------------------------------	-----------------------------	-------------------------

for the development of anti-inflammatory agents (Mukaida et al., 1996), which is mainly attributed to the enhancement of TNFα-induced apoptosis by β2 integrins (CD11/CD18) at 4 and 8 h after ETX stimulation (Walzog et al., 1997). Moreover, free total rhubarb anthraquinones (FTRAs) attenuated intestinal injury and enhanced intestinal barrier function by regulating intestinal immune function in rats with endotoxin-induced acute pancreatitis (Xiong, et al., 2018). In addition, endotoxemia can also induce apoptosis through oxidative stress. For example, piperlongumine exhibits prominent anti-inflammatory effects by inhibiting the ETX-induced p65 NFκB signaling cascade and markedly suppresses cytokine storm production, which is closely associated with ROS-mediated induction of late apoptosis and reduced expression of antiapoptotic Bcl2 protein (Thatikonda et al., 2020). To sum up, Figure 4 reveals how the recognition signals of endotoxin, inflammation, and oxidative stress responses induce apoptosis directly or indirectly. These herb ingredients show significant therapeutic effects on endotoxin, inhibiting endotoxin-induced elevation in Ca2+ and adhesion factors, and activation of signaling pathways such as TLR4/NF-KB, MAPK, PI3K-AKT, and mTOR/STAT3.

FUTURE PROSPECTS

In conclusion, numerous studies have shown that the antiapoptotic treatment of endotoxemia can improve immunosuppression and protect parenchymal organ function. A range of bacterial components such as ETX can induce a cellular stress response and induce apoptosis. However, ETX, as a caspase inhibitor, can inhibit apoptosis by engaging its O-antigen (O Ag) moiety to directly bind caspases, contributing to the successful colonization of bacteria to support their intracellular propagation (Günther et al., 2019), but to the detriment of humans. In addition, it is interesting to note that many anti-apoptotic herbal ingredients in endotoxemia exhibit pro-apoptotic activity in cancer. For example, salidroside increases survival in endotoxemic mice (Guan et al., 2011) and has been reported to inhibit apoptosis in bone marrow mesenchymal stem cells (Wei et al., 2013), H9c2 cardiomyocytes

(Sun et al., 2018), pulmonary arterial smooth muscle cells (Gui et al., 2017), pheochromocytoma cells (Hu et al., 2016), neural stem cells (Yan et al., 2018), cardiomyocytes (Zhu et al., 2015) and umbilical vein endothelial cells (Chen and Wu, 2014), while inducing apoptosis in colorectal cancer cells (Fan et al., 2016) gastric cancer AGS cells (Rong et al., 2020). It is thus clear that anti-apoptosis is not a single effective approach for the treatment of endotoxemia with herbal medicine, but must be combined with anti-inflammation, antioxidative stress, and anticoagulation to improve the prognosis of patients with endotoxemia. In addition, it should be noted that the anti-apoptotic activity of herb ingredients is also affected by the pathological state of the host. In any case, anti-apoptotic therapy plays an important role in endotoxemia but has long been neglected, and a considerable number of experimental studies on endotoxemia do not involve the detection of apoptotic-related indicators. This suggests that the impact of apoptosis on the pathophysiological process of endotoxemia should be emphasized and that antiapoptosis is a potential new approach for the treatment of endotoxemia.

AUTHOR CONTRIBUTIONS

ZL conceived the study. WW, JL, YinZ, YuZ, and GQ reviewed and summarized the literature. ZL wrote the manuscript and created all the figures. XM and YiZ paid all fees for the publication of this manuscript. YiZ, XM, and XS supervised the study and gave final approval for the published version. The final version of the manuscript was read and approved by all authors.

FUNDING

This work was supported by the National Natural Science Foundation of China (81773974), the National Key R&D program of China (2017YFC1703904), the Science and Technology Department of Sichuan Province (19SYXHZ0095, 2018JY0467).

REFERENCES

- Akao, T., Sato, K., He, J.-X., Ma, C.-M., and Hattori, M. (2013). Baicalein 6-O-β-d-Glucopyranuronoside is a main metabolite in the plasma after oral administration of baicalin, a flavone glucuronide of scutellariae Radix, to rats. *Biol. Pharm. Bull.* 36 (5), 748–753. doi:10.1248/bpb.b12-00850
- Allam, A. A., Gabr, S., Ajarem, J., Alghadir, A., Sekar, R., and Chow, B. K. (2017). Geno protective and anti-apoptotic effect of green tea against perinatal lipopolysaccharide-exposure induced liver toxicity in rat newborns. *Ajtcam* 14 (2), 166–176. doi:10.21010/ajtcam.v14i2.18
- Ben, S. V., Lomnitski, L., Nyka, A., Carbonatto, M., Peano, S., Zurovsky, Y., et al. (2000). Effect of natural antioxsidants and apocynin on LPS-induced endotoxemia in rabbit. *Hum. Exp. Toxicol.* 19 (11), 604–614. doi:10.1191/ 096032700666138364
- Bi, M. H., Ott, J., Fischer, T., Hecker, M., Dietrich, H., Schaefer, M. B., et al. (2010). Induction of lymphocyte apoptosis in a murine model of acute lung injurymodulation by lipid emulsions. *Shock* 33 (2), 179–188. doi:10.1097/shk. 0b013e3181ac4b3b
- Brenchley, J. M. (2006). Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Retrovirology* 3 (1 Suppl. ment), S98. doi:10.1186/1742-4690-3-s1-s98
- Chen, H. X., Wang, H., Chen, Y., and Zhang, H. S. (2005). Liquid chromatographytandem mass spectrometry analysis of anisodamine and its phase I and II metabolites in rat urine. J. Chromatogr. B Analyt. Technol. Biomed. Life Sci. 824, 21–29. doi:10.1016/j.jchromb.2005.07.036
- Chen, Z., and Wu, X. (2014). Salidroside attenuates high glucose-induced apoptosis in human umbilical vein endothelial cells via activating the Ca(2)+/CaM/ CAMKIIð/eNOS pathway. *Zhonghua Xin Xue Guan Bing Za Zhi* 42 (4), 327–333. doi:10.3760/cma.j.issn.0253-3758.2014.04.013
- Cheng, Y.-J., Cheng, S.-M., Teng, Y.-H., Shyu, W.-C., Chen, H.-L., and Lee, S.-D. (2014). Cordyceps sinensisPrevents apoptosis in mouse liver with D-galactosamine/lipopolysaccharide-induced fulminant hepatic failure. Am. J. Chin. Med. 42 (2), 427–441. doi:10.1142/s0192415x14500281
- Chiang, Y.-H., Jen, L.-N., Su, H.-Y., Lii, C.-K., Sheen, L.-Y., and Liu, C.-T. (2006). Effects of garlic oil and two of its major organosulfur compounds, diallyl disulfide and diallyl trisulfide, on intestinal damage in rats injected with endotoxin. *Toxicol. Appl. Pharmacol.* 213 (1), 46–54. doi:10.1016/j.taap.2005. 08.008
- Cho, I. H., Gong, J. H., Kang, M. K., Lee, E. J., Park, J. H., Park, S. J., et al. (2014). Astragalin inhibits airway eotaxin-1 induction and epithelial apoptosis through modulating oxidative stress-responsive MAPK signaling. *BMC Pulm. Med.* 14, 122. doi:10.1186/1471-2466-14-122
- Choi, Y. H. (2019). Activation of the Nrf2/HO-1 signaling pathway contributes to the protective effects of coptisine against oxidative stress-induced DNA damage and apoptosis in HaCaT keratinocytes. *gpb* 38 (4), 281–294. doi:10.4149/ gpb_2019014
- Coureuil, M., Bourdoulous, S., Marullo, S., and Nassif, X. (2014). Invasive meningococcal disease: a disease of the endothelial cells. *Trends Mol. Med.* 20 (10), 571–578. doi:10.1016/j.molmed.2014.08.002
- Danial, N. N., and Korsmeyer, S. J. (2004). Cell death. Cell 116 (2), 205–219. doi:10. 1016/s0092-8674(04)00046-7
- Deitch, E. A. (2012). Gut-origin sepsis: evolution of a concept. *The Surgeon* 10 (6), 350–356. doi:10.1016/j.surge.2012.03.003
- Eduardo, L. C., and Fresno, C. D. (2013). Pathophysiology of endotoxin tolerance: mechanisms and clinical consequences. *Crit. Care* 17 (6), 242. doi:10.1186/ cc13110
- Erridge, C. (2011). Diet, commensals and the intestine as sources of pathogenassociated molecular patterns in atherosclerosis, type 2 diabetes and nonalcoholic fatty liver disease. *Atherosclerosis* 216 (1), 1–6. doi:10.1016/j. atherosclerosis.2011.02.043
- Fan, X.-J., Wang, Y., Wang, L., and Zhu, M. (2016). Salidroside induces apoptosis and autophagy in human colorectal cancer cells through inhibition of PI3K/ Akt/mTOR pathway. Oncol. Rep. 36 (6), 3559–3567. doi:10.3892/or.2016. 5138
- Fehsel, K., Kröncke, K. D., Meyer, K. L., Huber, H., Wahn, V., and Kolb-Bachofen, V. (1995). Nitric oxide induces apoptosis in mouse thymocytes. *J. Immunol.* 155 (6), 2858–2865.

- Fouad, A. A., Qutub, H. O., and Al-Melhim, W. N. (2016). Nephroprotection of punicalagin in rat model of endotoxemic acute kidney injury. *Toxicol. Mech. Methods* 26 (7), 538–543. doi:10.1080/15376516.2016.1211207
- Gasparrini, M., Giampieri, F., Forbes-Hernandez, T. Y., Afrin, S., Cianciosi, D., Reboredo-Rodriguez, P., et al. (2018). Strawberry extracts efficiently counteract inflammatory stress induced by the endotoxin lipopolysaccharide in human dermal fibroblast. *Food Chem. Toxicol.* 114, 128–140. doi:10.1016/j.fct.2018. 02.038
- Guan, F., Zhou, X., Li, P., Wang, Y., Liu, M., Li, F., et al. (2019). MG53 attenuates lipopolysaccharide-induced neurotoxicity and neuroinflammation via inhibiting TLR4/NF-κB pathway *in vitro* and *in vivo*. Prog. Neuro-Psychopharmacol. Biol. Psychiatry 95, 109684. doi:10.1016/j.pnpbp.2019. 109684
- Guan, S., Feng, H., Song, B., Guo, W., Xiong, Y., Huang, G., et al. (2011). Salidroside attenuates LPS-induced pro-inflammatory cytokine responses and improves survival in murine endotoxemia. *Int. Immunopharmacol.* 11 (12), 2194–2199. doi:10.1016/j.intimp.2011.09.018
- Gui, D., Cui, Z. M., Zhang, L., Yu, C., Yao, D., Xu, M., et al. (2017). Salidroside attenuates hypoxia-induced pulmonary arterial smooth muscle cell proliferation and apoptosis resistance by upregulating autophagy through the AMPK-mTOR-ULK1 pathway. *BMC Pulm. Med.* 17 (1), 191. doi:10. 1186/s12890-017-0477-4
- Günther, S. D., Fritsch, M., Seeger, J. M., Schiffmann, L. M., Snipas, S. J., Coutelle, M., et al. (2019). Cytosolic Gram-negative bacteria prevent apoptosis by inhibition of effector caspases through lipopolysaccharide. *Nat. Microbiol.* 5 (2), 354–367. doi:10.1038/s41564-019-0620-5
- Guo, J. M., Liu, Z., Zhang, D. f., Chen, Y. Y., Qin, H. R., Liu, T. T., et al. (2019). TLR4 agonist monophosphoryl lipid A alleviated radiation-induced intestinal injury. J. Immunol. Res. 2009, 2121095. doi:10.1155/2019/2121095
- Harjai, M., Bogra, J., Kohli, M., and Pant, A. B. (2013). Is suppression of apoptosis a new therapeutic target in sepsis?. *Anaesth. Intensive Care* 41 (2), 175–183. doi:10.1177/0310057x1304100207
- Hengartner, M. O. (2000). The biochemistry of apoptosis. Nature 407 (6805), 770-776. doi:10.1038/35037710
- Hotchkiss, R. S., Coopersmith, C. M., and Karl, I. E. (2005). Prevention of lymphocyte apoptosis—a potential treatment of sepsis?. *Clin. Infect. Dis.* 41, 465–469. doi:10.1086/431998
- Hotchkiss, R. S., Tinsley, K. W., Swanson, P. E., and Karl, I. E. (2002). Endothelial cell apoptosis in sepsis. *Crit. Care Med.* 30, 225–228. doi:10.1097/00003246-200205001-00009
- Hu, Y., Lv, X., Zhang, J., and Meng, X. (2016). Comparative study on the protective effects of salidroside and hypoxic preconditioning for attenuating anoxiainduced apoptosis in pheochromocytoma (PC12) cells. *Med. Sci. Monit.* 22, 4082–4091. doi:10.12659/msm.897640
- Hwang, B. B., Chang, M. H., Lee, J. H., Heo, W., Kim, J. K., Pan, J. H., et al. (2019). The edible insect *Gryllus bimaculatus* protects against gut-derived inflammatory responses and liver damage in mice after acute alcohol exposure. *Nutrients* 11 (4), E857. doi:10.3390/nu11040857
- Jäättelä, M., and Tschopp, J. (2003). Caspase-independent cell death in T lymphocytes. *Nat. Immunol.* 4 (5), 416–423. doi:10.1038/ni0503-416
- Kerr, J. F. R., Wyllie, A. H., and Currie, A. R. (1972). Apoptosis: a basic biological phenomenon with wideranging implications in tissue kinetics. *Br. J. Cancer* 26 (4), 239–257. doi:10.1038/bjc.1972.33
- Latha, S., Chaudhary, S., and Ray, R. S. (2017). Hydroalcoholic extract of Stevia rebaudiana bert. leaves and stevioside ameliorates lipopolysaccharide induced acute liver injury in rats. *Biomed. Pharmacother*. 95, 1040–1050. doi:10.1016/j. biopha.2017.08.082
- Lee, A. S., Chen, W. P., Kuo, Y. L., Ho, Y. J., Lee, S. S., and Su, M. J. (2010). Thaliporphine preserves cardiac function of endotoxemic rabbits by both directly and indirectly attenuating NFκB signaling pathway. *Plos One* 7 (6), e39174. doi:10.1371/journal.pone.0039174
- Lee, S.-C., Wang, S.-Y., Li, C.-C., and Liu, C.-T. (2018). Anti-inflammatory effect of cinnamaldehyde and linalool from the leaf essential oil of Cinnamomum osmophloeum Kanehira in endotoxin-induced mice. J. Food Drug Anal. 26 (1), 211–220. doi:10.1016/j.jfda.2017.03.006
- Leist, M., and Jäättelä, M. (2001). Four deaths and a funeral: from caspases to alternative mechanisms. Nat. Rev. Mol. Cel Biol. 2 (8), 589–598. doi:10.1038/ 35085008

- Li, C., Li, L., Chen, K., Wang, Y., Yang, F. X., and Wang, G. (2019). UFL1 alleviates lipopolysaccharide-induced cell damage and inflammation via regulation of the TLR4/NF-κB pathway in bovine mammary epithelial cells. Oxid. Med. Cel Longev. 2019, 6505373. doi:10.1155/2019/6505373
- Li, L., Wan, G., Han, B., and Zhang, Z. (2018). Echinacoside alleviated LPS-induced cell apoptosis and inflammation in rat intestine epithelial cells by inhibiting the mTOR/STAT3 pathway. *Biomed. Pharmacother*. 104, 622–628. doi:10.1016/j. biopha.2018.05.072
- Liou, S.-F., Hsu, J.-H., Liang, J.-C., Ke, H.-J., Chen, I.-J., Wu, J.-R., et al. (2012). San-Huang-Xie-Xin-Tang protects cardiomyocytes against hypoxia/reoxygenation injury via inhibition of oxidative stress-induced apoptosis. *J. Nat. Med.* 66 (2), 311–320. doi:10.1007/s11418-011-0592-0
- Liou, S.-F., Ke, H.-J., Hsu, J.-H., Liang, J.-C., Lin, H.-H., Chen, I.-J., et al. (2011). San-huang-xie-xin-tang prevents rat hearts from ischemia/reperfusion-induced apoptosis through eNOS and MAPK pathways. *Evidence-Based Complement*. *Altern. Med.* 2011, 1. doi:10.1093/ecam/neq061
- Liu, H., Yu, X., Yu, S., and Kou, J. (2015). Molecular mechanisms in lipopolysaccharide-induced pulmonary endothelial barrier dysfunction. *Int. Immunopharmacol.* 29 (2), 937–946. doi:10.1016/j.intimp.2015.10.010
- Liu, L., Wang, P., Liang, C., He, D., Yu, Y., and Liu, X. (2013). Distinct effects of Nampt inhibition on mild and severe models of lipopolysaccharide-induced myocardial impairment. *Int. Immunopharmacol.* 17 (2), 342–349. doi:10.1016/ j.intimp.2013.06.017
- Liu, Y., Ye, F., Zou, W. J., Sun, Y., Wang, R., Han, P. P., et al. (2015). Baicalein reduces the occurrence of cirrhotic endotoxemia by reducing intestinal mucosal apoptosis. *BMC Complement. Altern. Med.* 15, 161. doi:10.1186/s12906-015-0682-8
- Lo, Y. C., Lin, Y. L., Yu, K. L., Lai, Y. H., Wu, Y. C., Ann, L. M., et al. (2005a). San-Huang-Xie-Xin-Tang attenuates inflammatory responses in lipopolysaccharide-exposed rat lungs. J. Ethnopharmacol. 101 (1-3), 68–74. doi:10.1016/j.jep.2005.03.015
- Lo, Y. C., Tsai, P. L., Huang, Y. B., Shen, K. P., Tsai, Y. H., Wu, Y. C., et al. (2005b). San-Huang-Xie-Xin-Tang reduces lipopolysaccharides-induced hypotension and inflammatory mediators. *J. Ethnopharmacol.* 96 (1-2), 99–106. doi:10. 1016/j.jep.2004.09.023
- Makris, A., Thornton, C. E., Xu, B., and Hennessy, A. (2005). Garlic increases IL-10 and inhibits TNFa and IL-6 production in endotoxin-stimulated human placental explants. *Placenta* 26 (10), 828–834. doi:10.1016/j.placenta.2004. 10.019
- Manco, M., Putignani, L., and Bottazzo, G. F. (2010). Gut microbiota, lipopolysaccharides, and innate immunity in the pathogenesis of obesity and cardiovascular risk. *Endocr. Rev.* 31 (6), 817–844. doi:10.1210/er.2009-0030
- Meira, C. S., Espírito Santo, R. F. d., dos Santos, T. B., Orge, I. D., Silva, D. K. C., Guimarães, E. T., et al. (2017). Betulinic acid derivative BA5, a dual NF-kB/ calcineurin inhibitor, alleviates experimental shock and delayed hypersensitivity. *Eur. J. Pharmacol.* 815, 156–165. doi:10.1016/j.ejphar.2017.09.008
- Meng, Y.-y., Liu, Y., Hu, Z.-f., Zhang, Y., Ni, J., Ma, Z.-g., et al. (2018). Sanguinarine attenuates lipopolysaccharide-induced inflammation and apoptosis by inhibiting the TLR4/NF-κB pathway in H9c2 cardiomyocytes. *Curr. Med. Sci.* 38 (2), 204–211. doi:10.1007/s11596-018-1867-4
- Micheau, O., and Jürg, T. (2003). Induction of tnf receptor i-mediated apoptosis via two sequential signaling complexes. *Cell* 114 (2), 0–190. doi:10.1016/s0092-8674(03)00521-x
- Mukaida, N., Ishikawa, Y., Lkeda, N., Fujioka, N., Watanabe, S.-i., Kuno, K., et al. (1996). Novel insight into molecular mechanism of endotoxin shock: biochemical analysis of LPS receptor signaling in a cell-free system targeting NF-κB and regulation of cytokine production/action through β2 integrin *in vivo. J. Leukoc. Biol.* 59 (2), 145–151. doi:10.1002/jlb.59.2.145
- Munford, R. S. (2016). Endotoxemia-menace, marker, or mistake?. J. Leukoc. Biol. 100 (4), 687–698. doi:10.1189/jlb.3ru0316-151r
- Neff, S. B., Z'graggen, B. R., Neff, T. A., Jamnicki-Abegg, M., Suter, D., Schimmer, R. C., et al. (2006). Inflammatory response of tracheobronchial epithelial cells to endotoxin. Am. J. Physiol. Lung Cell Mol. Physiol. 290 (1), 86–96. doi:10.1152/ ajplung.00391.2004
- O'Brien, J. M., and Abraham, E. (2004). Human models of endotoxemia and recombinant human activated protein C. *Crit. Care Med.* 32, S202–S208. doi:10. 1097/01.ccm.0000126123.34119.98

- Ovstebo, R., Brandtzaeg, P., Brusletto, B., Bente Foss Haug, K., Lande, K., Hoiby, E. A., et al. (2005). Use of robotized DNA isolation and real-time PCR to quantify and identify close correlation between levels of Neisseria meningitidis DNA and lipopolysaccharides in plasma and cerebrospinal fluid from patients with systemic meningococcal disease. J. Clin. Microbiol. 43 (1), 532. doi:10.1128/ jcm.43.1.532.2005
- Pan, Y., Song, D., Zhou, W., Lu, X., Wang, H., and Li, Z. (2019). Baicalin inhibits C2C12 myoblast apoptosis and prevents against skeletal muscle injury. *Mol. Med. Rep.* 20 (1), 709–718. doi:10.3892/mmr.2019.10298
- Pan, Y., Chen, D., Lu, Q., Liu, L., Li, X., and Li, Z. (2017). Baicalin prevents the apoptosis of endplate chondrocytes by inhibiting the oxidative stress induced by H₂O₂. *Mol. Med. Rep.* 16 (3), 2985–2991. doi:10.3892/mmr.2017.6904
- Pei, H., Zhang, Y., Wu, H., Ren, L., Jia, X., Zhang, Y., et al. (2013). Relationship between iNOS expression and apoptosis in cerebral tissue, and the effect of sini injection in endotoxin shock rats. *J. Traditional Chin. Med.* 33 (4), 486–491. doi:10.1016/s0254-6272(13)60153-3
- Peña, G., Cai, B., Ramos, L., Vida, G., Deitch, E. A., and Ulloa, L. (2011). Cholinergic regulatory lymphocytes re-establish neuromodulation of innate immune responses in sepsis. J. Immunol. 187 (2), 718–725. doi:10.4049/ jimmunol.1100013
- Peter, M. E., and Krammer, P. H. (2003). The CD95(APO-1/Fas) DISC and beyond. Cell Death Differ. 10 (1), 26–35. doi:10.1038/sj.cdd.4401186
- Piwowar, A. (2014). The advanced oxidation protein products as potential diagnostic and prognostic factor in diseases of the indicated participation of oxidative stress. *Postepy. Hig. Med. Dosw.* 68, 446–458. doi:10.5604/17322693. 1101545
- Reichardt, F., Chassaing, B., Nezami, B. G., Li, G., Tabatabavakili, S., Mwangi, S., et al. (2017). Western diet induces colonic nitrergic myenteric neuropathy and dysmotility in mice via saturated fatty acid- and lipopolysaccharide-induced TLR4 signalling. J. Physiol. 595 (5), 1831–1846. doi:10.1113/jp273269
- Rong, L., Li, Z., Leng, X., Li, H., Ma, Y., Chen, Y., et al. (2020). Salidroside induces apoptosis and protective autophagy in human gastric cancer AGS cells through the PI3K/Akt/mTOR pathway. *Biomed. Pharmacother.* 122, 109726. doi:10. 1016/j.biopha.2019.109726
- Shou, X., Wang, B., Zhou, R., Wang, L., Ren, A., Xin, S., et al. (2017). Baicalin suppresses hypoxia-reoxygenation-induced arterial endothelial cell apoptosis via suppressing pkcδ/p53 signaling. *Med. Sci. Monit.* 23, 6057–6063. doi:10. 12659/msm.907989
- Simpson, A. J. H., Opal, S. M., Angus, B. J., Prins, J. M., Palardy, J. E., Parejo, N. A., et al. (2000). Differential antibiotic-induced endotoxin release in severe melioidosis. J. Infect. Dis. 181 (3), 1014–1019. doi:10.1086/315306
- Stevenson, C. S., Capper, E. A., Roshak, A. K., Marquez, B., Eichman, C., Jackson, J. R., et al. (2002). The identification and characterization of the marine natural product scytonemin as a novel antiproliferative pharmacophore. *J. Pharmacol. Exp. Ther.* 303 (2), 858–866. doi:10.1124/jpet.102.036350
- Sun, M. Y., Ma, D. S., Zhao, S., Wang, L., Ma, C. Y., and Bai, Y. (2018). Salidroside mitigates hypoxia/reoxygenation injury by alleviating endoplasmic reticulum stress—induced apoptosis in H9c2 cardiomyocytes. *Mol. Med. Rep.* 18 (4), 3760–3768. doi:10.3892/mmr.2018.9403
- Sunil, V. R., Connor, A. J., Lavnikova, N., Gardner, C. R., Laskin, J. D., and Laskin, D. L. (2002). Acute endotoxemia prolongs the survival of rat lung neutrophils in response to 12-O-tetradecanoyl-phorbol 13-acetate. *J. Cel. Physiol.* 190 (3), 382–389. doi:10.1002/jcp.10074
- Suzuki, K., Murakami, T., Kuwahara-Arai, K., Tamura, H., Hiramatsu, K., and Nagaoka, I. (2011). Human anti-microbial cathelicidin peptide LL-37 suppresses the LPS-induced apoptosis of endothelial cells. *Int. Immunol.* 23 (3), 185–193. doi:10.1093/intimm/dxq471
- Thatikonda, S., Pooladanda, V., Sigalapalli, D. K., and Godugu, C. (2020). Piperlongumine regulates epigenetic modulation and alleviates psoriasis-like skin inflammation via inhibition of hyperproliferation and inflammation. *Cell Death Dis.* 11 (1), 21. doi:10.1038/s41419-019-2212-y
- Tian, W., Guo, H.-S., Li, C.-Y., Cao, W., Wang, X.-Y., Mo, D., et al. (2019). PFKFB3 promotes endotoxemia-induced myocardial dysfunction through inflammatory signaling and apoptotic induction. *Toxicol. Appl. Pharmacol.* 368, 26–36. doi:10.1016/j.taap.2019.02.007
- Ulevitch, R. J., and Tobias, P. S. (1995). Receptor-dependent mechanisms of cell stimulation by bacterial endotoxin. *Annu. Rev. Immunol.* 13, 437–457. doi:10. 1146/annurev.iy.13.040195.002253

- Walzog, B., Jeblonski, F., Zakrzewicz, A., and Gaehtgens, P. (1997). β2 integrins (CD11/CD18) promote apoptosis of human neutrophils. FASEB J. 11 (13), 1177–1186. doi:10.1096/fasebj.11.13.9367353
- Wang, C.-Z., Zhang, C.-F., Chen, L., Anderson, S., Lu, F., and Yuan, C.-S. (2015). Colon cancer chemopreventive effects of baicalein, an active enteric microbiome metabolite from baicalin. *Int. J. Oncol.* 47 (5), 1749–1758. doi:10.3892/ijo.2015.3173
- Wang, Z., Zhao, S., Song, L., Pu, Y., Tan, N., Liu, X., et al. (2018). Natural cyclopeptide RA-V inhibits the NF-κB signaling pathway by targeting TAK1. *Cell Death Dis* 9 (7), 715. doi:10.1038/s41419-018-0743-2
- Wei, Y. P., Bai, H., Sun, Y. Q., Bao, S., Xi, R., and Liu, L. (2013). Effect of salidroside on apoptosis of bone marrow mesenchymal stem cells induced by ara-C. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 21 (6), 1572–1577. doi:10.7534/j.issn. 1009-2137.2013.06.039
- Williams, G. T. (1991). Programmed cell death: apoptosis and oncogenesis. *Cell* 65 (7), 1097–1098. doi:10.1016/0092-8674(91)90002-g
- Wu, J., Luo, Y., Jiang, Q., Li, S., Huang, W., Xiang, L., et al. (2019). Coptisine from Coptis chinensis blocks NLRP3 inflammasome activation by inhibiting caspase-1. *Pharmacol. Res.* 147, 104348. doi:10.1016/j.phrs.2019.104348
- Wu, X., Zhi, F., Lun, W., Deng, Q., and Zhang, W. (2018a). Baicalin inhibits PDGF-BB-induced hepatic stellate cell proliferation, apoptosis, invasion, migration and activation via the miR-3595/ACSL4 axis. *Int. J. Mol. Med.* 41 (4), 1992–2002. doi:10.3892/ijmm.2018.3427
- Wu, X., Zhou, W., Wei, Q., Chen, P., and Li, Y. (2018b). Cytoprotective effects of the medicinal herb Astragalus membranaceus on lipopolysaccharide—exposed cells. Mol. Med. Rep. 18 (5), 4321–4327. doi:10.3892/mmr.2018.9483
- Xiong, Y. X., Chen, L., Fan, L., Wang, L. L., Zhou, Y. J., Qin, D. L., et al. (2018). Free total Rhubarb Anthraquinones protect intestinal injury via regulation of the intestinal immune response in a rat model of severe acute pancreatitis. *Front. Pharmacol.* 9, 75. doi:10.3389/fphar.2018.00075
- Yan, R., Xu, H., and Fu, X. (2018). Salidroside protects hypoxia-induced injury by up-regulation of miR-210 in rat neural stem cells. *Biomed. Pharmacother*. 103, 1490–1497. doi:10.1016/j.biopha.2018.04.184
- Yao, J., Cao, X., Zhang, R., Li, Y. X., Xu, Z. L., Zhang, D. G., et al. (2016). Protective Effect of baicalin against experimental colitis via suppression of oxidant stress and apoptosis. *Pharmacogn. Mag.* 12 (47), 225–234. doi:10.4103/0973-1296. 186342
- Yu, C.-P., Shia, C.-S., Lin, H.-J., Hsieh, Y.-W., Lin, S.-P., and Hou, Y.-C. (2016). Analysis of the pharmacokinetics and metabolism of aloe-emodin following intravenous and oral administrations in rats. *Biomed. Chromatogr.* 30 (10), 1641–1647. doi:10.1002/bmc.3735
- Yu, C., Qi, D., Sun, J. F., Li, P., and Fan, H. Y. (2015). Rhein prevents endotoxininduced acute kidney injury by inhibiting NF-κB activities. *Sci. Rep.* 5, 11822. doi:10.1038/srep11822
- Yu, Y., Liu, H., Yang, D., He, F., Yuan, Y., Guo, J., et al. (2019). Aloe-emodin attenuates myocardial infarction and apoptosis via up-regulating miR-

133 expression. Pharmacol. Res. 146, 104315. doi:10.1016/j.phrs.2019. 104315

- Zhang, B., Liu, Y., Zhang, J.-S., Zhang, X.-H., Chen, W.-J., Yin, X.-H., et al. (2015). Cortistatin protects myocardium from endoplasmic reticulum stress induced apoptosis during sepsis. *Mol. Cell Endocrinol.* 406, 40–48. doi:10.1016/j.mce. 2015.02.016
- Zhang, F., Wang, X., Tong, L., Qiao, H., Li, X., You, L., et al. (2011). Matrine attenuates endotoxin-induced acute liver injury after hepatic ischemia/reperfusion in rats. *Surg. Today* 41 (8), 1075–1084. doi:10.1007/s00595-010-4423-9
- Zhang, X. P., Li, C. Y., Zhang, J., Ruan, Y. F., and Ma, M. L. (2009). Protection of salvia miltiorrhizae to the spleen and thymus of rats with severe acute pancreatitis or obstructive jaundice. *Mediators Inflamm.* 2009 (2), 186136. doi:10.1155/2009/186136
- Zhao, J., Yan, S., Zhu, X., Bai, W., Li, J., and Liang, C. (2020). PTPRO exaggerates inflammation in ulcerative colitis through TLR4/NF-κB pathway. *J. Cell Biochem.* 121 (2), 1061–1071. doi:10.1002/jcb.29343
- Zheng, W.-x., Cao, X.-l., Wang, F., Wang, J., Ying, T.-z., Xiao, W., et al. (2015). Baicalin inhibiting cerebral ischemia/hypoxia-induced neuronal apoptosis via MRTF-A-mediated transactivity. *Eur. J. Pharmacol.* 767, 201–210. doi:10.1016/ j.ejphar.2015.10.027
- Zheng, Y. J., Yan, M., Lu, L. B., Chang, L. W., Jin, J. B., and Xiao, M. D. (2018). Sophocarpine attenuates LPS-induced liver injury and improves survival of mice through suppressing oxidative stress, inflammation, and apoptosis. *Mediators Inflamm.* 2018, 5871431. doi:10.1155/2018/5871431
- Zhu, J., Wang, X. J., and Li, J. (2019). Pharmacological mechanism and apoptosis effect of baicalein in protecting myocardial ischemia reperfusion injury in rats. *Pak. J. Pharm. Sci.* 32, 407–412.
- Zhu, L., Wei, T., Gao, J., Chang, X., He, H., Luo, F., et al. (2015). The cardioprotective effect of salidroside against myocardial ischemia reperfusion injury in rats by inhibiting apoptosis and inflammation. *Apoptosis* 20 (11), 1433–1443. doi:10.1007/s10495-015-1174-5
- Zhu, Y., Fu, Y., and Lin, H. (2016). Baicalin inhibits renal cell apoptosis and protects against acute kidney injury in pediatric sepsis. *Med. Sci. Monit.* 22, 5109–5115. doi:10.12659/msm.899061

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Liu, Wang, Luo, Zhang, Zhang, Gan, Shen, Zhang and Meng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.