# Evaluation of the *CYP1B1* gene as a candidate gene in beagles with primary open-angle glaucoma (POAG)

K. Kato, A. Kamida, N. Sasaki, B.S. Shastry<sup>2</sup>

<sup>1</sup>The laboratory of Veterinary Emergency Medicine and Veterinary Surgery Graduate School of Agricultural and Life Sciences, The University of Tokyo, Tokyo, Japan; <sup>2</sup>Department of Biological Sciences, Oakland University, Rochester, MI

**Purpose:** In humans, primary open-angle glaucoma (POAG) is a complex genetic disorder and is the leading cause of visual impairment. Although all relevant genes were not identified, a small subset of the condition is found to be caused by mutations in the *MYOC* and *CYP1B1* genes. Inherited glaucoma also occurs in several breeds of dogs including beagles. Primary glaucoma in beagles is inherited as an autosomal recessive trait. The purpose of this study is to investigate the role of the *CYP1B1* gene in beagles with POAG.

**Methods:** For the purpose of genetic analysis, total RNAs from the spleen of the canines were isolated and *CYP1B1* cDNA was prepared. Genomic DNA from five affected, two carriers, and 13 randomly selected normal beagles with no sign of glaucoma was amplified by the polymerase chain reaction (PCR) using four pairs of primers. The amplified products were directly sequenced using BigDye terminator cycle sequencing.

**Results:** Genomic DNA analyses have identified a substitution polymorphism (109A→C) in the 5'-untranslated region (UTR) as well as a missense mutation (P93R) in exon 2 of the gene. Three affected, two carriers, and nine normal dogs are heterozygous while two affected and three normal dogs are homozygous for the missense mutation. One normal dog did not show this alteration. Normal dogs also contain the substitution polymorphism in the 5'-UTR. Similar experiments with exon 3 did not identify any additional mutation in the gene.

**Conclusions:** The above results suggest that *CYP1B1* alterations in the coding and UTR are not the primary cause of glaucoma in beagles by possible monogenic association. They may be classified as polymorphisms or they may modify glaucoma phenotype.

Glaucoma in humans is a complex, genetically heterogeneous condition and is the leading cause of blindness worldwide. It is classified into primary open-angle (POAG), primary closed-angle (PCAG) and primary congenital glaucoma (PCG). The disease is frequently associated with elevated intraocular pressure (IOP) and involves defects in the trabecular meshwork (TM) and the anterior chamber of the eye, leading to the obstruction of aqueous outflow, IOP, progressive degeneration of the optic nerve and retinal ganglion cell death causing blindness mostly in adults [1]. Mechanisms independent of IOP are also implicated in glaucomatous degeneration. Several family studies suggest that specific genetic defects may contribute to the pathogenesis of POAG. Although details about the inheritance of the disease remain unclear, genetic regions that are influencing POAG have been identified in large population based studies. Subsequent studies have demonstrated that mutations in myocilin (MYOC) and optineurin (OPTN) genes are associated with POAG in various populations [2]. Mutations in MOYC and OPTN together account for less than 5% of all POAG cases. In addition, mutations in CYP1B1 – a member of the cytochrome P450 superfamily enzymes – have

[3-5]. In children, PCG is an important cause of visual loss and is diagnosed during the neonatal or infantile period [6-8]. Inheritance of PCG is believed to be autosomal recessive [9]. Recently, three PCG loci (2p21, 1p36, and 14q24.3) have been mapped and a spectrum of mutations in *CYP1B1* (GLC 3A) have been implicated in the pathogenesis of the disorder, suggesting that *CYP1B1* is the major PCG causing gene [10-15]. Mutations in *CYP1B1* are associated with wide range of phenotypes and the alteration of this gene could impair the morphogenesis of the outflow angle because it has been proposed that *CYP1B1* participates in iridocorneal angle development [16]. Therefore, it may play an important role in the physiology and development of the eye. Recently it has been suggested that it may be a potential modifier gene in early onset glaucoma [17].

The biological basis of the disease is not fully understood. Investigations in human glaucoma have been hampered

The biological basis of the disease is not fully understood. Investigations in human glaucoma have been hampered because of a lack of suitable inherited POAG animal models of human disease with the exception of inherited glaucoma in beagles. The condition in beagles is transmitted as an autosomal recessive trait and appears when the animals are nine to 18 months old [18]. The pathogenesis, clinical signs, and pharmacological responses of the glaucoma in beagles have been previously investigated and reported [18-20]. For our knowledge, this model system has not been investigated

been implicated in POAG and juvenile open-angle glaucoma

Correspondence to: B.S. Shastry, Department of Biological Sciences, Oakland University, Rochester, MI, 48309; Phone: (248) 370-3577; FAX: (248) 370-4225; email: shastry@oakland.edu

TABLE 1. PRIMERS USED IN CDNA ISOLATION AND MUTATIONAL ANALYSIS.

Region	5'-3' Forward	5'-3' Reverse
5'-UTR	TCCCCTCTCACTGCCTACCA	GGGATGGCGCGTTTTGACTC
Exon 2	CGCGATCTCAGGTTTGCGA	TCATGAGCGCCGTGTGCTT
	TGCGGTTGAGCTGCTCGAA	TCAAAGGCCCCAGCCCTATTACGGA
Nested	AAGCACACGGCGCTCATGACGTT	CACCTCGCAGGTCAGTCTGT
Exon 3	ACACGCATGAAGCACATC	GACTTACGATAGGAAAGAGAGCA

In the table, UTR refers non-translated region (Exon 1).

at the genetic level. Therefore, it is of interest for us to conduct genetic analysis on this only available beagle model of openangle glaucoma. Such studies may provide additional insight about the mechanism of this disease and also complement the human studies.

## **METHODS**

Isolation of canine CYP1B1 cDNA: For the purpose of genetic studies, canine CYP1B1 cDNA was isolated. Three normal beagles were euthanized with an intravenous overdose of pentobarbital sodium (these procedures were approved by the animal care and use committee of the graduate school of agriculture and life sciences, University of Tokyo, Japan) and small pieces of spleen tissues were excised. By using an RN easy mini-kit (Qiagen, Tokyo, Japan) total RNAs were extracted from the tissue. From this total RNAs, single stranded cDNA was synthesized with an oligo(dT)<sub>12-18</sub> primer and the SuperScript III reverse transcriptase (Invitrogen, Tokyo, Japan). The extracted cDNA was stored at -20 °C. For the synthesis of the second strand we have designed the oligonucleotide primers (Table 1) using the sequence information of the Canis familiaris chromosome 17 whole genome shotgun sequence (GenBank). Amplification was carried out according to the procedure supplied by the manufacturer using these primers (Table 1) and Phusion high fidelity DNA polymerase (Finnzymes, Espoo, Finland). Briefly, polymerase chain reaction (PCR) amplification was carried out in a thermocycler (Bio-Rad, Tokyo, Japan) and consisted of an initial denaturation at 98 °C for 30 s, 35 cycles of denaturation at 98 °C for 10 s, annealing at 73 °C for 30 s, and extension at 72 °C for 45 s followed by a final extension at 72 °C for 10 min. The PCR products were electrophoresed on a 2% agarose gel and bands were visualized by ethidium bromide staining. The amplified products were purified by SUPREC-PCR (TaKaRa Bio Inc., Shiga, Japan) or Wizard SV gel and PCR clean up system (Promega, WI).

DNA sequencing and mutational analysis: The purified product was directly sequenced using a BigDye terminator v 1.1 cycle sequencing kit (Applied Biosystems, Foster city, CA) and an ABI PRISM 3130-Avant genetic analyzer (Applied Biosystems). This sequence information was deposited in GenBank (Accession # FJ 824086.1). The sequence data were aligned and compared by using a genetic

information processing software (GENETYX, version 6.1.1.0, Genetyx, Tokyo, Japan) and a sequence scanner software version 1.0 (Applied Biosystems 2005). For mutational analyses, genomic DNA from peripheral blood samples of normal and affected dog was extracted by using an EZ1 – DNA blood kit and the BIOROBOT EZ1 system (Qiagen). Genomic DNA from normal and affected dogs was amplified and subjected to sequence analysis as detailed above by using the same primers (except the nested primer) listed in the Table 1. All three exons were amplified and both strands were sequenced. Sequencing reactions were repeated at least twice using freshly amplified DNA.

## **RESULTS**

In order to understand the genetic basis of POAG in beagles, we have previously shown that the condition is not associated with MYOC [21]. As part of our continued interest in POAG, in this report we have isolated canine CYP1B1 cDNA for mutational analysis. The gene is partially characterized (promoter region and splice sites are not characterized) and contains three exons of which only exons two and three code for the protein while the first exon contains a 5'-untranslated region (UTR) of 354 base pairs. The putative open reading frame begins in the second exon and the gene encodes a 543 amino acid protein. It is approximately 84% homologous to the human gene at the levels of nucleotides and amino acids (Figure 1). For the purpose of genetic analysis, we have analyzed five affected, two carriers, and 13 randomly selected normal beagles with no sign of glaucoma. The results are summarized in Table 2. Our analyses have identified a substitution polymorphism (109A→C) in the 5'-UTR (Figure 2A-D, right-hand side) and it is present in affected, carrier and normal dogs (Figre 2, right-hand side). Additionally, a heterozygous missense mutation (P93R) in exon 2 of the gene was identified in three affected and two carrier dogs (Figure 2C,D, left-hand side) while two affected dogs are homozygous (Figure 2B, left-hand side) for this mutation. Among normal animals, nine of thirteen revealed heterozygosity for the missense mutation (Figure 2A, lefthand side), three of them are homozygous and one normal dog did not contain this alteration. The missense mutation has changed the encoded amino acid proline of the codon 93 to arginine (P93R). Similar experiments with exon 3 did not identify any additional mutation in the gene.

Human	-	orend	MGTSLSPNDP	WPLNPLSIOO	TTLLLLSVL	ATVHVGQRLL	RORRROLRSA	PPGPFAWPLI
			* *** * *	** **	******	* ** ** **	*****	******
Canine	1	:	MATSLGPDAP	LQPSALSAQQ	TTLLLLSVL	AAVHAGQWLL	RQRRRQPGSA	PPGPFAWPLI
	61		GNAAAVGQAA	HI.SFART.ARR	YGDVFOIRLG	SCPIVVLNGE	RAIHOALVOO	GSAFADRPAF
	01		**** * *	******	******	*** *****	*** *****	* ***** *
	61	:					RAIRQALVQQ	
	121						RSRQVLEGHV	
							*** *****	
	121	:	ASFRVVSGGR	SLAFGQYSPR	WKVQRRAAHS	TMRAFSTRQP	RSRRVLEGHV	LAETRELVAL
	181		LVRGSADGAF					
			* **** ***	*******	******	******	******	******
	181	:	LARGSAGGAF	LDPRPLTVVA	VANVMSAVCF	GCRYSHDDAE	FRELLSHNEE	FGRTVGAGSL
	241		VDVMPWLQYF	PNPVRTVFRE	FEQLNRNFSN	FILDKFLRHC	ESLRPGAAPR	DMMDAFILSA
							*** *****	
	241		VDVLPWLQRF	PNPVRTAFRE	FEQLNRNFSN	FVLRKFLRHR	ESLQPGAAPR	DMMDAFILSA
	301		EKKAAGDSHG	GGARLDLENV	PATITDIFGA	SQDTLSTALQ	WLLLLFTRYP	DVQTRVQAEL
			** *	***** * *	*** *****	***** ***	*** *****	** *** **
	301	:	GTEAAEGSGD	GGARLDMEYV	PATVTDIFGA	SQDTLSIALQ	WLLILFTRYP	QVQARVQEEL
	361		DQVVGRDRLP				HATTANTSVL	
			***** ***				**** **	
	361	•	DQVVGRNRLP	CLDDQPNLPY	TMAFLYEGMR	FSSFVPVTIP	HATTTSACVL	GYHIPKDTVV
4	421	:	TO A STATE OF THE PARTY OF THE OWNER OF THE PARTY OF THE				MIFSVGKRRC	
							******	
•	121		FVNQWSVNHD	PAKMANAEDE.	DPARFLDEDG	FIDEDLASSV	MIFSVGKRRC	IGEELSKMQL
4	181	:	FLFISILAHQ					
							*******	
	481	•	FLFISILAHO	CNFKANPDEP	SKMDFNYGLT	IKPKAFSINV	TLRESMELLD	SAVQKLQAEE
,	541	•	TCQ **					
	541	:	DCQ					

Figure 1. A comparative analysis of amino acid sequence between human and canine *CYP1B1* genes. Canine *CYP1B1* is 84% homologous to human gene at the amino acid and nucleotide level. The asterisk indicates identical amino acids.

Table 2. Polymorphic changes identified in CYP1B1 in Beagles.

Analysis of canine	Ex	xon 2	5'-UTR		
samples	Mutational changes	Genotype	Polymorphic changes	Genotype	
1 normal	P93P	homozygote	109 A→C	heterozygote	
9 normal	P93R	heterozygote	ND	_	
3 normal	R93R	homozygote	ND	_	
3 affected	P93R	heterozygote	109 A→C	heterozygote	
2 affected	R93R	homozygote	109 A→C	heterozygote	
2 carriers	P93R	heterozygote	109 A→C	heterozygote	

The affected dogs have POAG. In the table, ND refers to not determined, P refers to proline (normal amino acid), and R refers to arginine.

## **DISCUSSION**

Glaucoma in beagles is inherited as an autosomal recessive trait and the genetic basis of the disorder is not known. In this report, we have isolated and analyzed canine CYP1B1 gene (which is also mutated in human disorder) for mutations. Our analyses have identified a substitution polymorphism in the

5'-UTR and a missense mutation in the exon 2 of the gene. First of all, because normal dogs have the same 5'-UTR substitution (heterozygote) and three of the thirteen normal dogs contain the same missense mutation in both alleles (frequency is about 24%) it is likely that these variations are non-pathogenic. Secondly, these alterations did not segregate with the disease. Hence it is also unlikely that it is pathogenic.

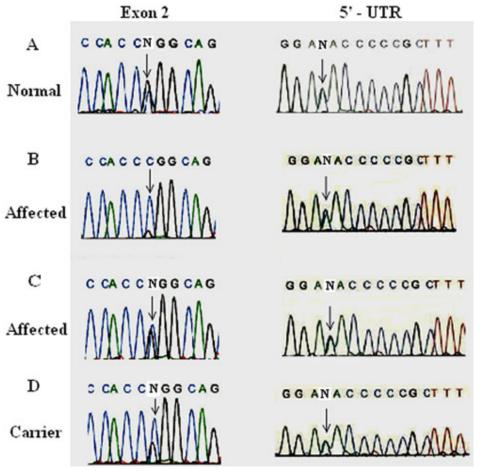


Figure 2. Mutational analysis of CYP1B1. Nucleotide sequence of the mutant parts of the exon 2 (left-hand side) and UTR (right hand-side) of CYP1B1 were shown. The nucleotide sequence change in codon 93 is from CCG to CGG (arrows) that resulted in the amino acid arginine (CGG) instead of normal proline (CCG). This missense mutation is present in the normal (heterozygote, **A**), affected **C**), (heterozygote, carrier (heterozygote, **D**) and also in two affected and three normal dogs (homozygote, **B**). The substitution polymorphism in the 5'-UTR (arrows) is present in all animals tested (righthand side).

The amino acid proline at codon 93 is conserved in normal mouse, rat and humans. It is surprising that its alteration to arginine did not produce the phenotype in homozygote animals (proline is considered to be a helix-breaking residue). Therefore, it is tempting to speculate that this mutation may also not have any functional significance or the mutation is hypomorphic similar to the other reported mutations [22]. It is interesting that many (approximately 72%) normal dogs are heterozygous for the missense mutation in the canine population and there are three classes of normal beagles (no mutation, heterozygotes and homozygotes) at the CYP1B1 locus. Whatever the real explanation for these observed results, glaucoma in beagles does not appear to be a simple recessive trait and it may have a multifactorial etiology. Functional assays as well as mating experiments in the future may clarify the physiological role (if any) of polymorphic missense substitution and mutations. Meanwhile, identification of a putative predisposing gene may substantially improve our knowledge of the pathogenesis of glaucoma in beagles.

## **ACKNOWLEDGEMENTS**

We are thankful for Dr. Kirk N. Gelatt of University of Florida for providing the blood samples from affected and carrier dogs.

#### REFERENCES

- Weinreb RN, Khaw PT. Primary open-angle glaucoma. Lancet 2004; 363:1711-20. [PMID: 15158634]
- Fingert JH, Stone EM, Sheffield VC, Alward WL. Myocilin glaucoma. Surv Ophthalmol 2002; 47:547-61. [PMID: 12504739]
- Acharya M, Mookherjee S, Bhattacharjee A, Bandyaopadhyay AK, Daulat Thakur SK, Bhaduri G, Sen A, Ray K. Primary role of CYP1B1 in Indian juvenile-onset POAG patients. Mol Vis 2006; 12:399-404. [PMID: 16688110]
- Melki R, Colomb E, Lefort N, Brezin AP, Garchon HJ. CYP1B1 mutations in French patients with early-onset primary openangle glaucoma. J Med Genet 2004; 41:647-51. [PMID: 15342693]
- Lopez-garrido MP, Sanchez-Sanchez F, Lopez-Martinez F, Aroca-Aguilar JD, Blanco-Marchite C, Coca-Prados M, Escribano J. Heterozygous CYP1B1 gene mutations in Spanish patients with primary open-angle glaucoma. Mol Vis 2006; 12:748-55. [PMID: 16862072]

- Deluise VP, Anderson DR. Primary infantile glaucoma (congenital glaucoma). Surv Ophthalmol 1983; 28:1-19. [PMID: 6353647]
- Travers JP. The presentation of congenital glaucoma. J Pediatr Ophthalmol Strabismus 1979; 16:241-2. [PMID: 490276]
- 8. Francois J. Congenital glaucoma and its inheritance. Ophthalmologica 1980; 181:61-73. [PMID: 7219964]
- Turacli ME, Aktan SG, Sayli BS, Akarsu N. Therapeutical and genetical aspects of congenital glaucoma. Int Ophthalmol 1992; 16:359-62. [PMID: 1428571]
- Bejjani BA, Lewis RA, Tomey KF, Anderson KL, Dueker DK, Jabak M, Astle WF, Otterud B, Leppert M, Lupski JR. Mutations in CYP1B1, the gene for cytochrome P450 1B1, are the predominant cause of primary congenital glaucoma in Saudi Arabia. Am J Hum Genet 1998; 62:325-33. [PMID: 9463332]
- Bagiyeva S, Marfany G, Gonzalez-Duarte R. Mutational screening of CYP1B1 in Turkish PCG families and functional analyses of newly detected mutations. Mol Vis 2007; 13:1458-68. [PMID: 17893647]
- Dimasi DP, Hewitt AW, Straga T, Pater J, MacKinnon JR, Elder JE, Casey T, Mackey DA, Craig JE. Prevalence of CYP1B1 mutations in Australian patients with primary congenital glaucoma. Clin Genet 2007; 72:255-60. [PMID: 17718864]
- 13. Mashima Y, Suzuki Y, Sergeev Y, Ohtake Y, Tanino T, Kimura I, Miyata H, Aihara M, Tanihara H, Inatani M, Azuma N, Iwata T, Araie M. Novel cytochrome P450 1B1 (CYP1B1) gene mutations in Japanese patients with primary congenital glaucoma. Invest Ophthalmol Vis Sci 2001; 42:2211-6. [PMID: 11527932]
- Stoilov IR, Costa VP, Vasconcellos JP, Melo MB, Betinjane AJ, Carani JC, Oltrogge EV, Sarfarazi M. Molecular genetics of primary congenital glaucoma in Brazil. Invest Ophthalmol Vis Sci 2002; 43:1820-7. [PMID: 12036985]

- Colomb E, Kaplan J, Gaechon HJ. Novel cytochrome P 450 1B1 (CYP1B1) mutations in patients with primary congenital glaucoma in France. Hum Mutat 2003; 22:496. [PMID: 14635112]
- Libby RT, Smith RS, Savinova OV, Zabaleta A, Martin JE, Gonzalez FJ, John SW. Modification of ocular defects in mouse developmental glaucoma models by tyrosinase. Science 2003; 299:1578-81. [PMID: 12624268]
- Vincent AL, Billingsley G, Buys Y, Levin AV, Priston M, Trope G, Williams-Lys D, Heon E. Digenic inheritance of early onset glaucoma: CYP1B1, a potential modifier gene. Am J Hum Genet 2002; 70:448-60. [PMID: 11774072]
- 18. Gelatt KN, Gum GG. Inheritance of primary glaucoma in the beagle. Am J Vet Res 1981; 42:1691-3. [PMID: 7325430]
- Samuelson DA, Gum GG, Gelatt KN. Ultrastructural changes in the aqueous outflow apparatus in beagles with inherited glaucoma. Invest Ophthalmol Vis Sci 1989; 30:550-61.
   [PMID: 2925324]
- Gelatt KN, Gum GG, Gwin RN, Bromberg NM, Merideth RE, Samuelson DA. Inherited primary open-angle glaucoma in the beagles. Am J Pathol 1981; 102:292-5. [PMID: 7468772]
- Kato K, Sasaki N, Gelatt KN, MacKay EO, Shastry BS. Autosomal recessive primary open-angle glaucoma (POAG) in beagles is not associated with mutations in the myocilin (MYOC) gene. Graefes Arch Clin Exp Ophthalmol 2009; 247:1435-6. [PMID: 19225800]
- Campos-Mollo E, Lopez-Garrido MP, Blanco-Marchite C, Grcia-Feijoo J, Peralta J, Belmonte-Martinez J, Ayuso C, Escribano J. CYP1B1 mutations in Spanish patients with primary congenital glaucoma: Phenotypic and functional variability. Mol Vis 2009; 15:417-31. [PMID: 19234632]