Check for updates

# 

**Citation:** Tanabe MB, Prochaska J, Morales ML, Lopez M, Baca-Turpo B, Arque E, et al. (2022) Geospatial analysis of the associations between environmental contamination with livestock feces and children with chronic fascioliasis in the Anta province of Cusco, Peru. PLoS Negl Trop Dis 16(6): e0010499. https://doi.org/10.1371/journal. pntd.0010499

Editor: Uwem Friday Ekpo, Federal University of Agriculture Abeokuta, NIGERIA

Received: November 14, 2021

Accepted: May 14, 2022

Published: June 16, 2022

**Peer Review History:** PLOS recognizes the benefits of transparency in the peer review process; therefore, we enable the publication of all of the content of peer review and author responses alongside final, published articles. The editorial history of this article is available here: https://doi.org/10.1371/journal.pntd.0010499

**Copyright:** © 2022 Tanabe et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability Statement:** The raw data used for this manuscript is available at the Harvard

RESEARCH ARTICLE

# Geospatial analysis of the associations between environmental contamination with livestock feces and children with chronic fascioliasis in the Anta province of Cusco, Peru

Melinda Barbara Tanabe<sup>1</sup>, John Prochaska<sup>2</sup>, Maria Luisa Morales<sup>3,4</sup>, Martha Lopez<sup>3,4</sup>, Benicia Baca-Turpo<sup>3,4</sup>, Eulogia Arque<sup>3,4</sup>, Miguel Mauricio Cabada<sup>1,3,4</sup>\*

 Division of Infectious Diseases, Department of Medicine, University of Texas Medical Branch, Galveston, Texas, United States of America, 2 Graduate School of Biomedical Sciences, University of Texas Medical Branch, Galveston, Texas, United States of America, 3 Alexander von Humboldt Tropical Medicine Institute, Department of Medicine, Universidad Peruana Cayetano Heredia, San Martin de Porres, Lima, Perú,
UPCH–UTMB Collaborative Research Center—Cusco, Universidad Peruana Cayetano Heredia, Wanchaq, Cusco, Peru

\* micabada@utmb.edu

# Abstract

Fasciola hepatica is a neglected parasitic infection with significant human health and livestock industry impact. The Andean Altiplano harbors an estimated 50% of the Fasciola's world infection burden. There is scarce data regarding the spatial associations between different Fasciola hosts. In this project, we aimed to determine the geospatial relationships between Fasciola eggs passed in feces of different livestock species and the risk of infection among each household as a unit. We used data from a cross-sectional study evaluating children and livestock feces for Fasciola infection around households in three districts of Anta province, in the Cusco region of Peru. Each sample was geographically tagged and evaluated for fascioliasis using microscopy methods. A total of 2070 households were included, the median age was 9.1 years (6.7-11.8), 49.5% were female, and 7.2% of the households had at least one infected child. A total of 2420 livestock feces samples were evaluated. The infection rate in livestock samples was 30.9%. The highest infection rate was found in sheep with 40.8%, followed by cattle (33.8%), and swine (26.4%). The median distance between a household with an infected child to a positive animal sample was 44.6 meters (IQR 14.7-112.8) and the distance between a household with no infected children to a positive animal sample was 62.2 meters (IQR 18.3–158.6) (p = 0.025). The multivariable logistic regression adjusted by presence of poor sanitation, unsafe water consumption, altitude, and presence of multiple infected children per household demonstrated an association between household infection and any cattle feces at a 50 meters radius (Uninfected: OR 1.42 (95%CI 1.07-1.89), p = 0.017. Infected: OR 1.89 (95%CI 1.31–2.73), p = 0.001), positive cattle feces at a 100 meters radius (OR 1.35 (95% Cl 1.08–1.69), p = 0.008), and negative cattle feces at a 200 meters radius (OR 1.08 (95% Cl 1.01–1.15), p = 0.022). We identified potential hot and cold spots for fascioliasis in the Anta province. An association between environmental contamination with feces from different livestock species and infected children in rural

Dataverse public repository (https://dataverse. harvard.edu). Full data cannot be shared publicly as it violates human subject research compliance as it provides identifiable information, therefore all data but geographic location coordinates are available. The database without identifiable information can be requested via this repository. https://doi.org/10. 7910/DVN/YHEQXU.

**Funding:** M.M.C. work was supported by the National Institute for Allergy and Infectious Diseases, National Institutes of Health grant no. 1R01AI104820 (https://www.niaid.nih.gov). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

households was found in our study. Local health authorities may apply this strategy to estimate the risk of infection in human populations and apply targeted interventions.

#### Author summary

*Fasciola hepatica* is the parasite that causes fascioliasis in the Andes region. The factors associated with transmission of the disease among humans have not been well described in the area studied. In this study, we evaluate the spatial relationships between infected households and the environmental burden of *Fasciola* eggs passed by different livestock in the Cusco region of Peru. We geographically marked and tested for fascioliasis feces provided by children and livestock. Infected and uninfected cattle as well as sheep feces were more likely to be in proximity to human households with infected children than to households without infected children. We were able to identify and map the areas of higher and lower infections rates. This information and strategy may be used in the control of the disease.

## Introduction

*Fasciola hepatica* is the causative agent of fascioliasis in the Andes region. It is considered an emergent infectious disease, with reports suggesting the establishment of new endemic areas in locations where sporadic transmission occurred in the past [1-3]. Climate change and man-made environmental modifications are probably associated with the increased prevalence and geographic distribution of Fasciola [4]. Fascioliasis causes significant financial burden in the livestock industry around the world. Over 90% of the human burden is located in low-resource areas in small farming communities [5,6]. School age children are disproportionally vulnerable to the infection [7,8].

*Fasciola hepatica* has a complex life cycle that includes snail intermediate hosts and a variety of mammalian definite hosts (Fig 1). Animals and humans with fascioliasis often have overlapping endemic areas. A Venezuelan study reported that animal and human fascioliasis was associated with snail presence and consumption of untreated water [9]. Mas-Coma et al. reported a prevalence of fascioliasis between 0–65% in cattle and 0–68% in humans with geographic overlap in different host endemic areas in the Bolivian Altiplano [10–11]. In Brazil, the low report of fascioliasis in humans over 60 years differed from the high reports of animal infection in the Southern states, specifically cattle [12–13]. In part, this could be explained by lack of surveillance and diagnosis in humans, as the disease seems to be spread in an area with overlapping human and livestock disease, however epidemiology is far from being elucidated [12]. The inter-host dynamics in *Fasciola* infection are not fully understood and may explain the inconsistencies between studies as well as differences in transmission patterns. Although no direct fecal-oral transmission of fascioliasis is possible, the role of *Fasciola's* environmental load caused by infected livestock in human transmission needs further scrutiny.

In this study, we aimed to determine the geospatial association between *Fasciola* eggs passed in feces of different livestock species and the risk of infection among each household in the Cusco region of Peru.

#### Methods

#### **Ethics statement**

The Institutional Ethics Committee of Universidad Peruana Cayetano Heredia and the University of Texas Medical Branch Institutional Review Boards approved the study protocol. The



https://doi.org/10.1371/journal.pntd.0010499.g001

consent process was performed in Quechua or Spanish language according to subject's preference. A written and verbal informed consent was obtained from the children's guardians and assent was obtained from children older than 6 years before any study procedure. Children with fascioliasis were referred to the Ministry of Health for treatment with triclabendazole.

We performed a cross-sectional study among children in 26 communities of the Anta Province in the Cusco region of Peru [15]. Children between 3 and 16 years of age enrolled in preschools and schools of the districts of Ancahuasi, Anta, and Zurite, were invited to participate between August of 2013 and August of 2018. Containers and instructions to collect only their own feces sample were provided to each child at pre-school and schools. Freshly produced feces samples were collected the next morning in consecutive days. Children that failed to produce a feces sample were given instructions again to collect a sample the next day. Each child's household was visited within 2 weeks of enrolment. Field workers visited each household to collect demographic, socio-economic, and epidemiological data. The Simple Poverty Scorecard (Microfinance Risk Management LLC) was used to evaluate the probability of the child's household to be under the poverty lines [16]. The entrance to the main living space of the house was geographically tagged at the time of visit. For the present study, we selected all children that provided at least one fecal sample for testing and had Global Position System (GPS) coordinates for the house where they resided regardless of infection status. Households were considered the analysis unit and houses with one or multiple children infected were defined as *Fasciola* positive households. The analysis was adjusted for the presence of multiple infected children in one household. If no infected children were found in a house, the household was defined as *Fasciola* negative household. The field workers collected up to four fresh fecal samples passed by different livestock species in the immediate vicinity of each house. Livestock samples were considered fresh if they were still moist at the time of collection. The livestock samples closest to the household were selected, geographically tagged, and classified according to the species. For the present study we focused on feces passed by cattle, swine, and sheep and selected one sample per species in each household. We were unable to ascertain if feces samples were from livestock owned by a specific household, but the proximity between the living space and the samples made ownership likely. If no GPS data was available for a livestock sample, the house coordinates were used as a proxy only for the radius analysis but not for distance analysis.

All fecal samples were collected in individual clean containers, maintained at 4-8°C, and transported as soon as possible to the laboratory at the Cusco Branch–Tropical Medicine Institute of Universidad Peruana Cayetano Heredia in Cusco city. Children's fecal samples were preserved fresh and in 10% formalin. Fresh samples were tested by the Kato–Katz method within 24–48 hours of collection. The samples preserved in 10% formalin were tested by the Lumbreras rapid sedimentation method within a week. [17] Chronic fascioliasis in children was defined as having *F. hepatica* eggs in any of the samples collected. Livestock feces were analyzed by the Lumbreras rapid sedimentation method for the presence of *Fasciola* eggs. Internal and external quality control procedures for data collection, data management, and laboratory procedures were used in the study. Known positive and negative stool samples were introduced in the laboratory routine to evaluate technician proficiency. Positive microscopy results were confirmed by a second observer.

The statistical analysis was performed using the SAS Statistical Package v. 9.4 (SAS Institute Inc., 2013). We used ArcGIS v. 10.7.1 (ESRI, 2019) for the analysis of infection distribution and inter-host associations. We calculated frequencies, means  $\pm$  standard deviation ( $\pm$ SD), and medians with interquartile ranges (IQR) to describe the distribution of the variables. We calculated the median distances from the different households to the closest infected or noninfected livestock feces. Livestock feces without GPS data were excluded from the radius analysis. The student-t test, Fisher/X<sup>2</sup> test, or Mann-Whitney-U test were used accordingly to compare the households with and without Fasciola infection. A backwards logistic regression analysis was performed using Fasciola positive household as the dependent variable to evaluate the spatial association between infection status of feces samples from different livestock species. We delineated areas around each household at 50, 100, and 200-meter radius and counted the number of positive and negative livestock feces samples within each radius area. In each radius model, we included the covariates animal feces count inside each radius area per species and infection status (sheep positive, sheep negative, cattle positive, cattle negative, pig positive, and pig negative). All models were adjusted by reported poor sanitation, unsafe water intake, and presence of multiple children infected in the household. Poor sanitation was defined as defecating in the fields or using superficial pits. Unsafe water intake was defined as not consuming water from the chlorinated municipal supply. Poverty likelihood based on the Simple Poverty Scorecard was not included in the regression model due to missing values in 17.6% of the sample. We used a p value < 0.2 for variable retention in the regression model.

The hot spots, cold spots, and spatial outliers analysis was performed in two different ways, one including all animals feces and another one including animals feces and household

infection status. Identification of hot and cold spots of *Fasciola* infected households was performed via Getis-Ord G statistics [18]. The statistic takes into account the interaction of the zone with itself plus the relationships with its surroundings and allows for a statistical significance evaluation of clustering via p-value as compared to a random distribution [19–20]. Other spatial patterns of the infection (outliers and clusters) in each household were analyzed by using Anselin Local Moran's I [18]. This index provides data regarding each location's risk based on statistically significant clustering patterns (high-high, low-low) and outliers (high-low, lowhigh) via a Z score [19]. The Z score indicates the presence of apparent similarity (clusters) or dissimilarity (outliers) as compared to that expected from random distribution. A positive Z score indicated clustering of high or low values (positive or negative cases) while a negative Z score indicated outliers. A high-low outlier represents a statistically significant high/positive value surrounded by low/negative values, while a low-high outlier represents the opposite. All maps were created under the WGS 1984 UTM Zone 18S coordinate system and Transverse Mercator as the elected projected system. A p < 0.05 was considered statistically significant.

# Results

Our study population consisted of 3000 children from 2122 households. After excluding 42 households lacking GPS information and 10 households where no livestock feces samples were collected, we included 2070 individual households in the analysis (Table 1). The median age was 9.1 years (6.7–11.8) and 1025 (49.5%) were female. Most children resided in the Anta district (50.8%). The median years of education was 6 (IQR 3–11) for the mothers and 9 (IQR 6–11) for the fathers. The mean poverty score was 39.6 ( $\pm$ 10.7) indicating that about half of the study households had a 34.4% or higher likelihood of living under a USD 3.75/day poverty line. [16] The median elevation of the households was 3382 meters (IQR 3352–3479). Overall, 7.2% of the households had at least one *Fasciola* infected child. From the 611 households

Table 1.	Children demographic characteristics in I	Fasciola positive and negative households.
----------	---	--

		Total (N = 2070)	Fasciola Positive (n = 148)	Fasciola Negative (n = 1922)	p-value
		Me	dian (IQR)*	·	
Education of the father (years)		9 (6-11)	7 (6-11)	9 (6-11)	0.007
Education of the mother (years)		6 (3-11)	6 (2-6.2)	6 (4-11)	<0.001
Altitude of the house (meters)		3382 (3352–3479)	3427 (3356–3610)	3381 (3351–3477)	<0.001
		Me	an (+/- SD)*	·	-
Poverty score		39.6 (±10.7)	34.9 (±9.4)	40.0 (±10.7)	<0.001
		·	N (%)*	·	
Sex	Male	1045 (50.5)	74 (7.1)	971 (92.9)	0.902
	Female	1025 (49.5)	74 (7.2)	951 (92.8)	
Location	Anta	1052 (50.8)	71 (6.7)	981 (93.3)	0.085
	Ancahuasi	745 (35.9)	64 (8.6)	681 (91.4)	
	Zurite	273 (13.1)	13 (4.8)	260 (95.2)	
Unsafe water consumption <sup>#</sup>	Yes No	90 (4.3) 1980 (95.7)	5 (5.6) 143 (7.2)	85 (94.4) 1837 (92.8)	0.5484
Poor sanitation <sup>\$</sup>	Yes No	578 (27.9) 1492 (72.1)	56 (9.7) 92 (6.2)	522 (90.3) 1400 (93.8)	0.0053

<sup>+</sup> P value based on Mann-Whitney-U,  $X^2$  as appropriate. Bolded values = significant to p< 0.05

\*Median and IQR values provided if variables failed to have a normal distribution by Shapiro Wilk test

# Defined as main water source not coming from municipal supply

\$ Defined as defecation in the opened field or in shallow pits.

https://doi.org/10.1371/journal.pntd.0010499.t001

where more than one child was tested for *Fasciola* infection, only 12 had multiple infected children. There were significant differences between fascioliasis positive and negative households in maternal and paternal years of education, poverty score, altitude of the household, and presence of unsafe sanitation (Table 1).

We collected 2648 samples of livestock feces, but only 2420 had GPS information and were included for distance analysis (Table 2). Cattle was the most frequent livestock species included (40.8%), followed by swine (32.8%), and sheep (26.4%). The overall *Fasciola* infection rate in livestock samples was 30.9%. A similar number of feces samples were collected from Ancahuasi and Anta with no significant differences in overall prevalence (Table 2). The highest infection prevalence was found in sheep feces (258/640, 40.3%) with Ancahuasi (129/292, 44.1%) as the place with the highest prevalence followed by Anta (109/290, 37.5%) and Zurite (20/58, 34.4%). Most cattle positive feces (328/987, 33.2%) were found in Anta (153/420, 36.4%) followed by Zurite (46/131, 35.1%) and Ancahuasi (129/436, 29.6%). Swine positive feces (163/793, 20.5%) were more often found in Ancahuasi (87/353, 24.6%) followed by Anta (66/363, 18.2%) and Zurite (10/77, 12.9%) (Table 2).

The median distance from a *Fasciola* positive household to the closest infected livestock feces sample was 44.6 meters (IQR = 14.7-112.8), while the median distance from a negative household to the closest infected livestock feces sample was 62.2 meters (IQR = 18.3-158.6) (p = 0.025) (Table 3). There was no significant difference between the distances from a *Fasciola* positive or negative household to a non-infected livestock feces sample (18.7 meters (IQR = 9.1–47.4) versus 22.4 meters (IQR = 9.1–66.5), p = 0.219). The median distance from a positive household to the closest infected cattle feces sample was 85.9 meters (IQR = 32-220.8) while the distance from a negative household to the closest infected cattle feces sample was 137.4 meters (IQR = 44.5-344.1) (p = 0.006). The distance of a positive household to infected swine feces (207.3 meters (IQR = 66.6-444.2)) was significantly different to the median distance from a negative household (236.9 meters (IQR = 89-610.5)) (p = 0.044). The distances from Fasciola positive or negative households to positive sheep feces samples were not significantly different. No differences were observed between Fasciola positive or negative households and negative feces samples from any livestock species (Table 3). The logistic regression models analyzing the effect of the distance to the closest animal feces on the household infection status adjusted by multiple infections in the household and altitude of the house demonstrated an OR 0.997 (95% CI 0.995–1.000, p = 0.0174). The effect modification on the model by distance to the closest feces by livestock species demonstrated no significant relationship to the household infection status (p = 0.7053). Similarly, the effect of the distance to the closest feces by animal status was not statistically significant (p = 0.9384).

The adjusted multivariable logistic regression using a 50-meter radius around households demonstrated a higher likelihood of finding cattle feces around infected households

		Total (N = 2420) n (%)	Fasciola Positive (N = 749) n (%)	Fasciola Negative (N = 1671) n (%)	p-value*
Location*	Ancahuasi	1081 (44.7)	345 (31.9)	736 (68.1)	< 0.535
	Anta	1073 (44.3)	328 (30.6)	745 (69.4)	
	Zurite	266 (11.0)	76 (28.6)	190 (71.4)	
Species	Cattle	987 (40.8)	328 (33.2)	659 (66.8)	< 0.001
	Swine	793 (32.8)	163 (20.6)	630 (79.4)	
	Sheep	640 (26.4)	258 (40.3)	382 (59.7)	

Table 2. "Distribution of the livestock samples and comparison between Fasciola positive and negative samples ".

\*P value based on two-sided X<sup>2</sup> tests.

Bolded values = significant to p < 0.05

https://doi.org/10.1371/journal.pntd.0010499.t002

	Positive Household (median, IQR)	Negative Household (median, IQR)	p-value *		
Distances to the closest livestock feces sample (meters)					
Positive livestock feces	44.6 (14.7–112.8)	62.2 (18.3–158.6)	0.025		
Negative livestock feces	18.7 (9.12–47.4)	22.4 (9.1–66.5)	0.219		
Distances to the closest fe	stances to the closest feces sample according to livestock species (meters)**				
Positive cow feces	85.9 (32-220.8)	137.4 (44.5–344.1)	0.006		
Positive pig feces	207.3 (66.6–444.2)	236.9 (89-610.5)	0.044		
Negative sheep feces	82 (26.1–179.4)	99.2 (33–199.1)	0.235		
Negative pig feces	49.1 (17–116.7)	57.5 (19.6–147.6)	0.290		
Negative cow feces	48.7 (16.8–157.3)	56 (18.2–165)	0.578		
Positive sheep feces	162.4 (82.7–317.2)	147.9 (57.3–373.6)	0.675		

Table 3. "Median distances from *Fasciola* positive or negative households to closest positive or negative livestock feces sample".

\* P value calculated for Mann-Whitney-U statistics.

\*\* Unable to report subgroup sizes, as feces might be close to more than one household.

Bolded values = significant to p< 0.05

https://doi.org/10.1371/journal.pntd.0010499.t003

independently of their infection status. However, infected households had a higher likelihood of being close to *Fasciola* infected cattle feces (OR = 1.89 (95%CI 1.31–2.73), p = 0.007) than to *Fasciola* negative cattle feces (OR = 1.42 (95%CI 1.06–1.89), P = 0.017). Using a 50-meter radius around households, *Fasciola* positive sheep feces were associated with a lower likelihood of the household being infected with *Fasciola* (OR = 0.52 (95%CI 0.32–0.84), p = 0.007). *Fasciola* negative sheep feces (p = 0.939) and swine feces regardless of their infection status (positive p = 0.7711, negative p = 0.7968) were not associated with infection in the households (Tables 4 and S1). Using a 100-meter radius in the adjusted logistic regression model, infected households still had a higher likelihood of having *Fasciola* positive sheep feces (DR = 1.35 (95%CI 1.08–1.69), p = 0.049) but had a decreased likelihood of having *Fasciola* positive sheep feces (OR 0.77 (95%CI 0.60–0.99), p = 0.042). Any swine feces and *Fasciola* negative cattle or sheep feces showed no

Table 4. "Multivariable logistic regression modeling factors assessing the likelihood of household positivity status by proximity to different types of livestock feces under different distance radius".

Radius length *	Odds Ratio	95% Confidence Interval	p-value***
50 meters **			
Cattle negative	1.42	1.07-1.89	0.017
Sheep positive	0.52	0.32-0.84	0.007
Cattle positive	1.89	1.31-2.73	0.001
100 meters **			
Sheep positive	0.77	0.60-0.99	0.049
Cattle positive	1.35	1.08–1.69	0.008
200 meters **			
Cattle negative	1.08	1.01-1.15	0.022

\* The radius calculations included all animal feces, the house GPS was used in lieu of the feces sample GPS data when the information was missing.

\*\* models adjusted by presence of poor sanitation, unsafe water, multiple household Fasciola infections, altitude of the household.

\*\*\* Only statistically significant values to p< 0.05 are shown

https://doi.org/10.1371/journal.pntd.0010499.t004

association with household infection status (Tables 4 and S2). Using a 200-meter radius, infection in the household was only associated with *Fasciola* negative cattle feces in the adjusted analysis (OR = 1.08 (95%CI: 1.01-1.15), p = 0.022). Any swine or sheep feces, and *Fasciola* positive cattle feces showed no association with household infection status (Tables 4 and S3). All models were adjusted by presence of poor sanitation, unsafe water intake, and presence of multiple infectious within a household.

When assessing for effect of altitude in the household infection status, the models were analyzed by altitude strata at each radius using median altitude as the cutoff (S4 Table). Using a model adjusted for sanitation, safe water intake, and multiple household infections at 50, 100, and 200 meters radius, there were no variables associated with the household infection status in the lower altitude strata. At the higher altitude strata and using a 50 meter radius, the presence of cattle positive feces (OR = 2.22 (95% CI = 1.28-3.84), p = 0.0046), cattle negative feces (OR = 1.68 (95%CI = 1.16-2.43) p = 0.0058), sheep positive feces (OR = 0.59 (95%CI = 0.35-0.97), p = 0.0389) were associated with *Fasciola* positive household status. At the higher altitudes and using a 100 meters radius, cattle positive feces (OR = 1.51 (95%CI = 1.02-2.24), p = 0.0370), cattle negative feces (OR = 1.35 (95%CI = 1.07-1.70), p = 0.0100), and sheep positive feces (OR = 0.71 (95%CI = 0.52-0.99), p = 0.0446) remained associated with Fasciola infected households (S5 Table).

For the identification of outlier and cluster analysis of all livestock and household fascioliasis, we localized 76 high-high clusters, 726 low-low clusters, 62 high-low outliers, and 136 lowhigh outliers (Fig 2A). When only livestock feces were analyzed, we localized 141 high-high clusters, 338 low-low clusters, 65 high-low outliers, and 134 low-high outliers (Fig 2B). For the identification of hot/cold spots based on the confidence intervals, when livestock and household fascioliasis were considered, we identified 16 hot and 3 cold spots zones with over > 90% confidence (Fig 3A). We identified 4 hot and 3 cold spot zones when all livestock feces were considered at > 90% confidence (Fig 3B). When comparing data from households in hot and cold spots areas, hot spot households were located at higher median altitude (3574 meters (IQR = 3427.5–3764) versus 3473 meters (IQR = 3370–3497), p = 0.001), had lower socioeconomic scores (34 (IQR = 28.5–41) versus 43 (IQR = 36–49), p < 0.001), had less median years of education for the fathers (6 (IQR 5–10) versus 11 (IQR = 6–11), p < 0.001) and the mothers (4.5 (IQR = 2–6) versus 6 (IQR = 4–11), p < 0.001) than cold spot households.

## Discussion

Geographic coordinate systems and computational spatial analysis are important tools to study infectious diseases epidemiology [22]. Geographic information systems have been most useful for creating dynamic scenarios of infectious disease epidemics, forecasting transmission patterns of epidemic outbreaks, and assessing the effectiveness of infection control interventions [23]. Given its complex lifecycle, multiple reservoirs, patchy distribution, and emerging character, *Fasciola hepatica* is an ideal parasite to model using quantitative spatiotemporal analysis [24]. Using these tools, we found a positive association between environmental contamination with cattle feces and households where children with chronic fascioliasis reside. The presence of any cattle feces was associated with *Fasciola* infection in the household with increasing infection likelihood with decreasing distance. Cattle can pass between 60–106 pounds of manure daily which could explain a higher burden of environmental contamination compared to other livestock species [25]. Parkinson et al. found a significant correlation (R 0.769, p = 0.02) between human and cattle infection in 12 rural communities of the Bolivian Altiplano [26]. Mas Coma et al. in the same area, reported a high prevalence of cattle fasciolia-sis and, based on metacercaria production and infectivity, suggested that this livestock species



Fig 2. "Outlier and Cluster Analysis map based on Anselin Local Moran's I statistic of all livestock and human Fascioliasis in the Anta province of Cusco, Peru" [21]. (A) Map based on infected and uninfected livestock and household infection status. (B) Map based on infected and uninfected livestock

feces (cattle, sheep, swine). (C) Inset showing the location of the study area (maroon dot) in the Cusco region of Peru. This map was created using ArcGIS v. 10.7.1 (ESRI, 2019). https://www.arcgis.com/home/item.html?id=67372ff42cd145319639a99152b15bc3.

https://doi.org/10.1371/journal.pntd.0010499.g002

plays an important role in maintaining transmission to human [27]. Other authors have suggested that consumption of contaminated water from natural sources or uncontrolled manmade canals is associated with fascioliasis transmission in humans and livestock [11,28]. Open defecation due to lack of latrines have been associated with Fasciola infection in observational studies in Bolivian Altiplano [11]. No direct transmission of fascioliasis between livestock and human is possible and the interactions between definitive hosts and snails are in need of further scrutiny to understand parasite transmission dynamics.

In our study, the presence of any cattle feces around the house, in particular Fasciola infected cattle feces, increased the likelihood of finding infection in the household and finding Fasciola infected sheep feces was associated with a decreased likelihood. These associations were maintained only at the higher altitude strata in the altitude stratified analysis. Higher altitude has been correlated with higher rates of fascioliasis likely due to increased survival of the intermediate host, longer cercarial shedding periods, and higher cercarial production [29]. Studies in South America have suggested an important role of sheep as Fasciola reservoirs at different altitudes based on infection prevalence [30]. Similarly, in our study, the prevalence of infection in sheep samples was the highest, but overall the proportion of sheep feces collected was the smallest compared to cattle and swine. It is unclear if this could have introduced sampling bias affecting our results. In the Anta province, sheep owners with the largest number of animals tend to keep herds close to pastures and away from their households especially at lower altitudes which may have also affected the evaluation of the environmental burden of sheep feces containing Fasciola eggs. Another potential explanation is a higher socioeconomic status among sheep owners which has been demonstrated to decrease the risk of infection in children [15]. The unexpected inverse association between household fascioliasis and sheep positive feces emphasize the complexities of Fasciola circulation between hosts and the need for further research on transmission.

Infected animal feces were significantly closer to infected households as compared to noninfected feces, especially cattle and swine feces. Experimental studies involving Fasciola eggs of sheep, cattle, and pigs have shown no differences in infection intensity or infectivity rates [30-31]. The particular association with cattle could be partially explained by the high number of this livestock species in the area probably constituting the main reservoir of fascioliasis in the Anta province and having a major role in transmission [11]. Using spatial analysis, we identified areas with high and low transmission of disease [32,33]. The large number of low-low outliers reflects the confidence on the locations of low risk of the disease. The small number of high-high outliers as compared to large number of "hot spots" when human data was added to livestock data, may suggest a discrepancy between human and animal risk of infection. In our study, the rates of human fascioliasis were considerably lower compared to the rates of animal fascioliasis. Areas of low-high and high-low transmission, remained similar when more data was added suggesting that in some locations of the Anta province infection risk is not welldefined [33]. In studies in Bolivia, the areas with higher prevalence of animal and human disease were located near water sources [10]. There is a lack of spatial analysis data about fascioliasis in South America and extrapolation of data from European studies might not be appropriate given differences in epidemiological behaviors. For example, spatial analysis done on Danish cattle herds showed that the presence of streams, wetlands, and pastures had a significant association with the presence of livestock infection [34]. While a study based in Switzerland, demonstrated that cattle were most likely infected in streams as they correspond to



Fig 3. "Significant hot/cold spot analysis of livestock and human fascioliasis in the Anta province of Cusco, Peru" [18]. (A) Map based on infected and uninfected livestock feces and household infection status. (B) Map based on infected and uninfected livestock feces (cattle, sheep, swine). (C) Inset showing the location of the study area (maroon dot) in the Cusco region of Peru. This map was created using ArcGIS v. 10.7.1 (ESRI, 2019). https://www.arcgis.com/home/item.html?id=f33a34de3a294590ab48f246e99958c9.

https://doi.org/10.1371/journal.pntd.0010499.g003

areas where the snail prevalence is higher [35]. In this study, we did not account for distance to large water bodies, as this information was not available. In Figs 2B or 3B, we can locate a large water body (northeast of the border) where the small number of children sampled failed to show a higher infection rate. This could indicate a prominent presence of snails in smaller bodies of water influencing the infection prevalence in the area [36]. The hot spots were more likely to be in locations where poverty likelihood and altitude were higher and parent's education was lower compared to cold spots. Previous studies by our group, have associated lower maternal education status and lower socioeconomic status with increased *Fasciola* infection rates [15]. In the future, comparing the outliers with the differences in their corresponding environmental conditions could be analyzed to define other risk factors for fascioliasis.

There are several limitations in our study that need to be considered. We did not have data regarding distance to different waterbodies (small or large) which may have limited our analysis model. In our study, we were aiming to understand the rarely explored relationship between livestock and human fascioliasis. Livestock feces found around the households was collected and no samples were collected from individual animals. This precluded the calculation of fascioliasis prevalence in the different livestock species. Due to missing data, we were unable to use the socioeconomic score in the predictive model selection. However, elevation of the household correlated well with the socioeconomic score and we used it as a surrogate measure of poverty. Despite the relatively large sample of children tested for fascioliasis, the number of infected subjects was low limiting our statistical power. The lack of specific topographic variables for the predictive model could have decreased the accuracy of our risk estimates.

#### Conclusion

In the Anta province of Cusco, the spatial distribution of *Fasciola hepatica* eggs in the environment was associated with the distribution of human fascioliasis. There was a different risk depending on livestock species with egg positive cattle and swine feces being independently associated with fascioliasis in the household. Further research to characterize areas where environmental egg contamination clusters with human cases is need to understand the dynamics of parasite transmission and identify potential strategies to optimize surveillance. The complexities of *Fasciola* infection among human and livestock emphasize the need for a one-health approach to research and control.

### Supporting information

**S1** Table. Number of livestock feces inside the 50 m buffer as per household status. (DOCX)

S2 Table. Number of livestock feces inside the 100 m buffer as per household status. (DOCX)

S3 Table. Number of livestock feces inside the 200 m buffer as per household status. (DOCX)

S4 Table. Adjusted univariate regression model for sheep positivity feces on household positivity status stratified by altitude of the household. (DOCX)

S5 Table. Adjusted Multivariate Logistic regression analysis stratified by higher or lower altitudes above the sea level associated with amount of livestock feces around different radiuses around the household.

(DOCX)

## Acknowledgments

We thank the Cusco Regional Health Directorate of the Peruvian Ministry of Health. The contents of this manuscript are solely the responsibility of the authors and do not necessarily represent the official views of the National Institute for Allergy and Infectious Disease.

## **Author Contributions**

Conceptualization: Melinda Barbara Tanabe, John Prochaska, Miguel Mauricio Cabada.

**Data curation:** Melinda Barbara Tanabe, Maria Luisa Morales, Martha Lopez, Benicia Baca-Turpo, Eulogia Arque, Miguel Mauricio Cabada.

Formal analysis: Melinda Barbara Tanabe, John Prochaska.

Funding acquisition: Miguel Mauricio Cabada.

Investigation: Martha Lopez, Benicia Baca-Turpo, Eulogia Arque, Miguel Mauricio Cabada.

Methodology: Melinda Barbara Tanabe, Maria Luisa Morales, Miguel Mauricio Cabada.

Project administration: Maria Luisa Morales, Martha Lopez, Miguel Mauricio Cabada.

Resources: Martha Lopez.

Software: John Prochaska.

Supervision: John Prochaska, Maria Luisa Morales, Martha Lopez, Miguel Mauricio Cabada.

Visualization: Miguel Mauricio Cabada.

Writing - original draft: Melinda Barbara Tanabe.

Writing – review & editing: Melinda Barbara Tanabe, John Prochaska, Benicia Baca-Turpo, Eulogia Arque, Miguel Mauricio Cabada.

#### References

- Sah R, Khadka S, Khadka M, Gurubacharya D, Sherchand JB, Parajuli K, et al. Human fascioliasis by Fasciola hepatica: the first case report in Nepal. BMC Res Notes. 2017; 10(1):439. https://doi.org/10. 1186/s13104-017-2761-z PMID: 28870243
- Qureshi AW, Zeb A, Mansoor A, Hayat A, Mas-Coma S. *Fasciola hepatica* infection in children actively detected in a survey in rural areas of Mardan district, Khyber Pakhtunkhawa province, northern Pakistan. *Parasitol Int.* 2019; 69:39–46. https://doi.org/10.1016/j.parint.2018.11.003 PMID: 30445198
- Hatami H, Asmar M, Masoud J, Mansouri F, Namdaritabar H, Ramazankhani A. The First Epidemic and New-emerging Human Fascioliasis in Kermanshah (Western Iran) and a Ten-year Follow Up, 1998– 2008. Int J Prev Med. 2012; 3(4):266–72. PMID: 22624083
- Fuentes MV, Valero MA, Bargues MD, Esteban JG, Angles R, Mas-Coma S. Analysis of climatic data and forecast indices for human fascioliasis at very high altitude. *Ann Trop Med Parasitol.* 1999; 93 (8):835–50. PMID: 10715678
- Mehmood K, Zhang H, Sabir AJ, Abbas RZ, Ijaz M, Durrani AZ, et al. A review on epidemiology, global prevalence and economical losses of fasciolosis in ruminants. *Microb Pathog.* 2017; 109:253–62. https://doi.org/10.1016/j.micpath.2017.06.006 PMID: 28602837

- Nyindo M, Lukambagire AH. Fascioliasis: An Ongoing Zoonotic Trematode Infection. *Biomed Res Int.* 2015; 2015:786195. https://doi.org/10.1155/2015/786195 PMID: 26417603
- Marcos L, Maco V, Samalvides F, Terashima A, Espinoza JR, Gotuzzo E. Risk factors for *Fasciola hepatica* infection in children: a case-control study. *Trans R Soc Trop Med Hyg.* 2006; 100(2):158–66. https://doi.org/10.1016/j.trstmh.2005.05.016 PMID: 16181652
- Lopez M, White AC Jr., Cabada MM. Burden of *Fasciola hepatica* Infection among children from Paucartambo in Cusco, Peru. *Am J Trop Med Hyg.* 2012; 86(3):481–5. https://doi.org/10.4269/ajtmh.2012. 11-0448 PMID: 22403322
- Corrales Y, Ferrer E, Fernandez J, Gauta J, Garcia M, Aguirre A, et al. Diagnosis and Risk Factors of Bovine and Human Fascioliasis in Cattle Farms from a Venezuelan Andean Rural Area. *Acta Parasitol.* 2021. https://doi.org/10.1007/s11686-021-00341-3 PMID: 33547991
- Mas-Coma S, Angles R, Esteban JG, Bargues MD, Buchon P, Franken M, et al. The Northern Bolivian Altiplano: a region highly endemic for human fascioliasis. *Trop Med Int Health*. 1999; 4(6):454–67. https://doi.org/10.1046/j.1365-3156.1999.00418.x PMID: 10444322
- Angles R, Buchon P, Valero MA, Bargues MD, Mas-Coma S. One Health Action against Human Fascioliasis in the Bolivian Altiplano: Food, Water, Housing, Behavioural Traditions, Social Aspects, and Livestock Management Linked to Disease Transmission and Infection Sources. *Int. J. Environ. Res. Public Health.* 2022; 19(3):1120. https://doi.org/10.3390/ijerph19031120 PMID: 35162146
- Schwantes JB, Quevedo P, D'Ávila MF, Molento MB, Graichen DAS. Fasciola hepatica in Brazil: genetic diversity provides insights into its origin and geographic dispersion. *J Helminthol.* 2019; 94:e83. https://doi.org/10.1017/S0022149X19000774 PMID: 31495340
- Pritsch I, Molento M. Recount of reported cases of human fascioliasis in Brazil over the last 60 years. J. Trop. Pathol. 2018; 47(2)
- 14. Center for Disease Control and Prevention. Fasciola- Biology 2018 [Available from: https://www.cdc. gov/parasites/fasciola/biology.html].
- Cabada MM, Morales ML, Webb CM, Yang L, Bravenec CA, Lopez M, et al. Socioeconomic Factors Associated with *Fasciola hepatica* Infection Among Children from 26 Communities of the Cusco Region of Peru. *Am J Trop Med Hyg.* 2018; 99(5):1180–5. https://doi.org/10.4269/ajtmh.18-0372 PMID: 30226136
- 16. Schreiner M. A Simple Poverty Scorecard for Peru. 2012 [Available from: https://www. simplepovertyscorecard.com/PER\_2010\_ENG.pdf]
- Lopez M, Morales ML, Konana M, Hoyer P, Pineda-Reyes R, White AC, Jr., et al. Kato-Katz and Lumbreras rapid sedimentation test to evaluate helminth prevalence in the setting of a school-based deworming program. *Pathog Glob Health*. 2016; 110(3):130–4. <u>https://doi.org/10.1080/20477724</u>. 2016.1187361 PMID: 27376503
- 18. Anselin L. Local Indicators of Spatial Association-LISA. Geographical Analysis. 2010; 27(2).
- Abdulhafedh A. Identifying Vehicular Crash High Risk Locations along Highways via Spatial Autocorrelation Indices and Kernel Density Estimation. World J. Eng. 2017; 5:198–215.
- **20.** Ord JK, Getis A. Testing for local spatial autocorrelation in the presence of global autocorrelation. *J. Reg. Sci.* 2001; 41(3), 411–432
- Esri H, Garmin, Intermap, increment P Corp., GEBCO, USGS, FAO, NPS, NRCAN, GeoBase, IGN, Kadaster NL, Ordnance Survey, Esri Japan, METI, Esri China (Hong Kong), National Geographic, UNEP-WCMC, NASA, NOAA, OpenStreetMap contributors and the GIS User community., cartographer Map Sources: ArcMAP v. 10.7.1; 2019.
- 22. Chowell G, Rothenberg R. Spatial infectious disease epidemiology: on the cusp. *BMC Med.* 2018; 16 (1):192. https://doi.org/10.1186/s12916-018-1184-6 PMID: 30333024
- Nyirenda SS, Sakala M, Moonde L, Kayesa E, Fandamu P, Banda F, et al. Prevalence of bovine fascioliasis and economic impact associated with liver condemnation in abattoirs in Mongu district of Zambia. BMC Vet Res. 2019; 15(1):33. https://doi.org/10.1186/s12917-019-1777-0 PMID: 30665406
- Webb CM, Cabada MM. Recent developments in the epidemiology, diagnosis, and treatment of Fasciola infection. Curr Opin Infect Dis. 2018; 31(5):409–14. <u>https://doi.org/10.1097/QCO.</u> 00000000000482 PMID: 30113327
- Amherst. UoM. Umass extension crops, dairy, livestock and equine program: Center of agriculture, food and the environment.; 2020 [Available from: https://ag.umass.edu/crops-dairy-livestock-equine/ fact-sheets/manure-inventory
- Parkinson M O'Neill S, Dalton J. Endemic human fasciolosis in the Bolivian Altiplano. *Epidemiol Infect*. 2007; 135(4):669–74. https://doi.org/10.1017/S095026880600728X PMID: 17064455

- 27. Mas-Coma S, Buchon P, Funatsu IR, Angles R, Artigas P, Valero MA, et al. Sheep and Cattle Reservoirs in the Highest Human Fascioliasis Hyperendemic Area: Experimental Transmission Capacity, Field Epidemiology, and Control Within a One Health Initiative in Bolivia. *Front. Vet. Sci.* 2020; 7.
- Esteban JG, González C, Bargues MD, Angles R, Sánchez C, Náquira C, Mas-Coma S. High fascioliasis infection in children linked to a man-made irrigation zone in Peru. *Trop Med Int Health*. 2002 Apr; 7 (4):339–48. https://doi.org/10.1046/j.1365-3156.2002.00870.x PMID: 11952950
- Mas-coma S, Funatsu I, Bargues M. Fasciola hepatica and lymnaeid snails occurring at very high altitude in South America. *Parasitology*. 2001; 123(7), 115–127. <u>https://doi.org/10.1017/s0031182001008034</u> PMID: 11769277
- Mas-Coma S, Buchon P, Funatsu IR, Angles R, Artigas P, Valero MA, Bargues MD. Sheep and Cattle Reservoirs in the Highest Human Fascioliasis Hyperendemic Area: Experimental Transmission Capacity, Field Epidemiology, and Control Within a One Health Initiative in Bolivia. *Front Vet Sci.* 2020 Oct 27; 7:583204. https://doi.org/10.3389/fvets.2020.583204 PMID: 33195605
- Mas-Coma S, Funatsu IR, Angles R, Buchon P, Mas-Bargues C, Artigas P, et al. Domestic pig prioritized in one health action against fascioliasis in human endemic areas: Experimental assessment of transmission capacity and epidemiological evaluation of reservoir role. *One Health*. 2021; 13:100249. https://doi.org/10.1016/j.onehlt.2021.100249 PMID: 33997234
- Purwanto P, Utaya S, Handoyo B, Bachri S, Astuti IS, Utomo KSB, et al. Spatiotemporal Analysis of COVID-19 Spread with Emerging Hotspot Analysis and Space–Time Cube Models in East Java, Indonesia. *ISPRS Int. J. Geoinf.* 2021; 10(3):133.
- McGinnis ML. A Spatiotemporal Analysis of Environmental Risk Factors of Lyme Disease in the Northeastern United States: University of Southern California; 2018.
- Olsen A, Frankena K, Bodker R, Toft N, Thamsborg SM, Enemark HL, et al. Prevalence, risk factors and spatial analysis of liver fluke infections in Danish cattle herds. *Parasit Vectors*. 2015; 8:160. <u>https:// doi.org/10.1186/s13071-015-0773-x</u> PMID: 25888827
- Knubben-Schweizer G, Torgerson PR. Bovine fasciolosis: control strategies based on the location of Galba truncatula habitats on farms. *Vet Parasitol.* 2015; 208(1–2):77–83. <u>https://doi.org/10.1016/j.vetpar.2014.12.019</u> PMID: 25596803
- 36. Charlier J., Soenen K., De Roeck E. et al. Longitudinal study on the temporal and micro-spatial distribution of Galba truncatula in four farms in Belgium as a base for small-scale risk mapping of Fasciola hepatica. *Parasit Vectors*. 2014: 7, 528 https://doi.org/10.1186/s13071-014-0528-0 PMID: 25425397