

RESEARCH

Open Access



# The role of Wnt/ $\beta$ -catenin-lin28a/let-7 axis in embryo implantation competency and epithelial-mesenchymal transition (EMT)

Qian Li<sup>1,2</sup>, Juanzi Shi<sup>2\*</sup> and Weimin Liu<sup>1\*</sup>

## Abstract

**Background:** The pre-implantation embryo in a competent status and post-implantation fully differentiation of the inner cell mass (ICM) and trophoctoderm (TE) are prerequisites of successful implantation. Type I embryonic epithelial-mesenchymal transition (EMT) involves in these processes. A high level of the mir-let-7 family was found in the dormant mouse embryo of implantation failure in our previous study. Besides, its natural inhibitor lin28a was found to function in maintained stem cell pluripotency and involved in early embryo nucleolus construction. Until now, few studies got involved in the exact molecular mechanism that affects embryo implantation potential. In this study, the possible function of Wnt/ $\beta$ -catenin-lin28a/let-7 pathway in mouse embryo implantation was studied.

**Methods:** ICR mouse, Lin28a/Let-7 g transgenic mice (Lin28a-TG/Let-7 g-TG), and implanting dormant mice models were used for the study.

**Results:** Wnt/ $\beta$ -catenin signaling is essential in embryo implantation, which promotes embryo implantation through directly trigger lin28a expression, thus represses the mir-let-7 family. Lin28a and mir-let-7 both participate in implantation via an inverse function. Lin28a and mir-let-7 participate in embryo implantation through embryonic EMT.

**Conclusions:** Wnt/ $\beta$ -catenin signaling promotes embryo implantation and accompanying embryonic EMT, which is mediated by directly activate lin28a/let-7 axis.

**Keywords:** Wnt/ $\beta$ -catenin signaling, Lin28a protein, Mir-let-7 family, Implantation competency, Embryonic EMT

## Background

Development of the blastocyst to the competent status and orchestrated embryonic EMT that accompany and underlie implantation and embryonic morphogenesis are essential for a successful pregnancy.

The pre-implantation mouse blastocyst is a differentiated, fluid-filled ball that comprises two distinct cell populations, the ICM and surrounded TE [1]. With the exposure of TE cells to the uterine epithelium,

implantation and accompanying embryonic EMT begins. EMT is a biologic process that polarized epithelial cells undergo multiple biochemical changes to gain a mesenchymal cell phenotype. The epithelial-like TE cells transform into invasive trophoblast giant cells, known as the yolk sac placenta, to provide a network of anastomotic channels surrounding the embryo and anchor the placenta. This process is called trophoctoderm EMT [2]. At the same time, ICM segregates into epiblast (EPI) and primitive endoderm (PE) layers. Then gastrulation EMT initiated when the primitive streak begins to form from EPI, the primitive streak generates the mesendoderm, which subsequently separates to form the mesoderm

\* Correspondence: [shijuanzi123@126.com](mailto:shijuanzi123@126.com); [liuwm@ioz.ac.cn](mailto:liuwm@ioz.ac.cn)

<sup>2</sup>Assisted Reproductive Center, Women & Children's Hospital of Northwest China, 73 Hou zai Road, Xi'an, China

<sup>1</sup>Department of Obstetrics and Gynaecology, Laboratory Block, The University of Hong Kong, 21 Sassoon Road, Pokfulam, Hong Kong, SAR, China



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

and the endoderm via an EMT. Those cells remaining in the EPI become ectoderm. Sequential EMTs are needed for the final differentiation of specialized cell types and the formation of the three germ layers that generate all tissue types of the body [3–5]. The exact mechanism that affects embryo implantation competency and EMT is still not clear.

The signaling pathway driven by Wnt/ $\beta$ -catenin seems to be one of the required pathways to achieve blastocyst competency for implantation. Silencing of Wnt/ $\beta$ -catenin signaling in mouse embryo reduced Cdx2, a key transcription factor involved in TE lineage specification, negatively affected the implantation competency of blastocyst [6]. In the post-implantation embryo, Wnt3 deficient mouse embryos have a block in anterior-posterior axis formation, fail to initiate gastrulation, and do not form mesoderm [7]. It was reported active- $\beta$ -catenin detected in different cells of mouse post-implantation embryo related to primitive streak formation [8], emphasizing the importance of this pathway in embryo development.

MicroRNAs constitute a large family of approximately 21-nucleotide-long, noncoding RNAs. They occupy 5% of the human genome but work as key post-transcriptional regulators of more than 70% of genes through mRNA decay and/or translational inhibition [9]. MicroRNA has been found to function in the reproduction system among oogenesis, spermatogenesis, fertilization and early embryonic development, implantation, and placentation. The dysregulation of miRNA may lead to reproductive disease [10–13]. Our previous microarray data suggested that several members of the let-7 family were up-regulated in the mice dormant blastocysts when compared with activated blastocysts, and Integrin-3 $\beta$  works as a down-stream target of mir-let-7 responsible for blastocyst adhesion [14]. Besides, we found mir-let-7a reduces human embryo surrogates' spheroid attachment ability [15].

Lin28a, a natural inhibitor of mir-let-7 family, is a member of a RNA-binding protein (RBP) family conserved in animals [16, 17]. Lin28 family is widely accepted as stem cell maintaining factors that promote stem-cell reprogramming through RNA binding function [18, 19]. Also, lin28a represses the processing of mir-let-7 at both the pri- and pre-miRNA steps, thus affects the translational efficiency of mir-let-7 target genes [20–22]. It has been found that during embryo development, Lin28 enhances the translation of genes important for growth and survival of human embryonic stem cells [23].

In this study, the regulation of mir-let-7 by Wnt/ $\beta$ -catenin-lin28a signaling was investigated, and their function in mouse embryo implantation competency and embryonic EMT were firstly studied.

## Material and method

### Animals and embryo collection

The use of animals was approved by the Committee on Use of Live Animals in Teaching and Research, the University of Hong Kong (Approval no. 3534–4, 4100–16, and 3573–15). Imprinted-coding region (ICR) mice were purchased from Harlan UK Ltd., Bicester, Oxon, UK. We generated a chimeric let-7 g species let-7S21L (let-7 g Stem, mir-21 Loop) in which let-7 g can be induced with doxycycline under the control of the Rosa26 locus (Let-7 g transgenic mice, Let-7 g-TG). Lin28a transgenic (Lin28a-TG) mice was a kind gift from Children's Hospital Boston and Dana Farber Cancer Institute, Boston, which also can be induced expression by doxycycline. Lin28a-TG and Let-7 g-TG mice were housed with food and water under controlled temperature ( $24 \pm 1^\circ\text{C}$ ), humidity ( $55 \pm 2\%$ ), and lighting conditions (daily light period, 0700 to 1900 h). The morning of finding a vaginal plug was designated as Day1 of pregnancy. Embryos at different developmental stages were recovered through flushing the oviduct on the morning of Day 1, 2, or 3 of gestation (1 cell stage embryo, two cell stage embryo, morula) or uterus on Day 4 (blastocyst). Implantation sites were detected on Day 5, Day 6 or Day 7 evening by Chicago blue (1% solution into saline, 2610-05-1, Sigma) via caudal vein injection (20  $\mu\text{l}$ /mice). Transgenic mice were fed with doxycycline hyclate (1 mg/ml, D9891, Sigma) in water from the mating day; for the control group, no doxycycline added. For embryo Wnt activation and inhibition, Wnt activator CHIR99021 (SML1046, Sigma) at 10  $\mu\text{M}$  and Wnt inhibitor mouse DKK (sc-5897-DK) at 1  $\mu\text{g}/\text{ml}$  were used.

### Delayed implantation

The whole surgical procedure was completed before 8:00 am. The Day 4 pregnant female mice were anesthetized by phenobarbitone. The operation area was near the abdominal cavity on two sides of the back. After removing body hair, skin, and muscle incision were made to expose the para-ovarian fatty tissue, which, together with the ovary, were cut between the oviduct and the ovary. The muscles layer and the body skin were sewed up separately. The surgical site was disinfected with 70% alcohol after surgery. Meloxicam (2 mg/kg, 71,125–38-7, Sigma) was fed in water to release their pain. Progesterone (P4, 2 mg/mouse, 5341, Calbiochem) was administered on Day 5 and Day 6 of pregnancy by subcutaneous injection to maintain the delayed implantation status [24]. Estradiol (E2, 25 ng/mouse, E8875, Sigma) was administered on Day 7 to activate dormant embryos.

### Embryo immunostaining

Embryos were fixed in 4% paraformaldehyde at room temperature for 30 min. Then washed three times in

wash buffer (0.01% triton and 0.01% NP40 in PBS), each for 5 min. Permeabilization in 0.4% triton and 0.4% NP40 in PBS for 15 min at room temperature. Incubation with 1% BSA (A3311, Sigma) for 1 h at room temperature. Embryos were then incubated with primary antibodies against active- $\beta$ -catenin (1:200, 19,807 s, CST), lin28a (1:200, 8706 s, CST), total  $\beta$ -catenin (1:200, 610,154, BD) at 4 °C overnight. After 4 rounds of wash in wash buffer, each for 10 min, embryos were incubated with anti-rabbit AF488 (A11070, Life Technology Scientific), anti-rabbit AF568 (A11036, Life Technology Scientific), or anti-mouse AF488 (A21202, Life Technology Scientific) secondary antibody at room temperature for 1 h, washed and incubated with DAPI (1: 500 in PBS, 28718-90-3, Sigma) for 5 min. After washing, fluorescence signals were examined under a Zeiss LSM 700 confocal laser microscope (Jena, Germany).

#### Immunohistochemistry (IHC)

Day 5, Day 6, and Day 7 implantation sites were cut, fixed in 4% PFA, embedded in paraffin, and cut from the transverse surface onto glass slides. Deparaffinized slides in xylene for 2 times. Rehydration the slides in gradient alcohol. 3% H<sub>2</sub>O<sub>2</sub> solution was applied to sections for 30 min to block endogenous peroxidase activity. To unmask the antigenic epitope, slides were boiled in antigen retrieval buffer (S1699, Dako Agilent technologies). Cooled down the slides at room temperature, then rinsed with PBS. Added 1% fetal bovine serum to block the sample at room temperature for 1 h. Applied primary antibody and incubated at 4 °C overnight. Slides were washed with PBS for 4 times. Applied (1:300) biotinylated secondary antibody to the sections and incubated at room temperature for 1 h. Washed with PBS again. Applied ABC buffer (VECTORSTAIN, PK-6100) to the sections and incubated at room temperature for 30 min. Washed slides with PBS. Applied DAB substrate solution (DAKO, K3468) to the sections for color development, Washed slides with PBS. Counterstained slides by Hematoxylin. Dehydrated the tissue slides again through 4 times of alcohol (50, 70, 90 and 100%, 100, 100%), 5 min each. Cleared the tissue slides in three times of xylene and mounting. Antibodies used in this assay were active- $\beta$ -catenin (1:600, 19,807 s, Cell Signaling), lin28a (1:600, 8706 s, Cell Signaling), E-cadherin (1:600, ab76055, Abcam), N-cadherin (1:600, ab18203, Abcam).

#### Embryo qPCR

The High Capacity cDNA RT Kit (4,368,814, Life Technology Scientific) was used to produce cDNA of target miRNAs according to the manufacturer's instructions. The template of miRNAs was obtained by heating the embryos in 6.2  $\mu$ l 5% NP40 (6507, Sigma) at 95 °C for 10

min. 10 embryos were pooled in each sample. The miRNA reverse transcription primers and probes for mature let-7a-5p (000377, Thermo Fisher Scientific), let-7g-5p (002282, Thermo Fisher Scientific), U6snRNA (001973, Thermo Fisher Scientific) were used. The reverse transcription condition was 16 °C for 30 min, 42 °C for 30 min, 4 °C for 5 min. The procedure of qPCR was 50 °C for 2 min, 95 °C for 10 min, 95 °C for 10 s, and 60 °C for 1 min.

#### Mouse primary endometrium (ME) cell isolation

Pregnant Day 4 mouse uteri were cut from mouse abdominal cavity, removed fat tissues, and washed in PBS. Cut longitudinally and digested in 0.5% trypsin (215,240, Becton Dickinson) and DNase I (180,470, Life Technology Scientific) at 4 °C for 90 min and then at 37 °C for 30 min. Tissues were then transferred to complete DMEM/F-12 (11,320,033, Thermo Fisher) and shaken gently for 30 s, three times. The dispersed luminal epithelium and some glandular epithelium in the upper cell suspension were collected and centrifuged at 1500 rpm for 5 min, then washed by centrifugation in complete DMEM-F12 for another 2 times. The recovered epithelial cells were seeded or collected.

#### Chromatin immunoprecipitation (CHIP)

The procedure was according to the manufacturer (185, 913, Abcam). The PCR primers used were as follows: Site 1 forward: GGGGTTGGGTCATTGTCTTT, Reverse: CTCGGTACCAAGCTTGCG; Site 2 forward: GCACTT TCTCTTTTCTAACTGGG, Reverse: GGAGTTCTGG ACCTGGGAA, Site3 forward: GCTTACCTGGGGCT AACTTT, Reverse: ATCTGAATGTCTGCCTCCCA; Site 4 forward: GAGCGCATTTTAGGGTGGTT, Reverse: TTCCCTCCCTCCAAACCTGTG; Site5 forward: CAGTCC TGTTCCCTCCACCAG, Reverse: CATCCACTGCTGAGAGAGGG; Published site 1 forward: GTGCGGGGGA AGATGTAGC, Reverse: CAACTGCTGGTTGGACA CG; Published site 2 forward: GTGGTTTGTGTTTC TGATTGGC, Reverse: GAGTACCCCGGAATTTGA GATC. The PCR procedure was according to Taq 2X Master Mix (M0270, NEB), that was 95 °C for 30 s, 95 °C for 30 s, 45–68 °C for 1 min, 68 °C 1 min per kb, 68 °C for 5 min, 35 cycles from the second step to the fourth step.

#### In vitro embryo attachment and outgrowth

Embryos were retrieved from pregnant Day 4 mouse uteri. The zona pellucida of blastocysts was removed by acid tyrode (CaCl<sub>2</sub> 265 mg, MgCl<sub>2</sub> 0.0469 g, KCl 0.2 g, NaCl 8 g, NaH<sub>2</sub>PO<sub>4</sub> 0.05 g to 1 l and adjust pH to 2). Embryos were then put on to Day 4 ME monolayer. To determine attachment, the plate was shaken at a rate of one rotation/second on an orbital shaker for 20 s after 24 h culture. Blastocysts that remained at the same

location were designated as attached blastocysts. At 48 h, outgrowth was identified as the area covered by trophoblastic monolayer around the embryo.

**Embryo electro-transfection**

Blastocysts were retrieved from pregnant Day 4 mouse uteri. Let-7a inhibitor (AM10050, Thermo Fisher), lin28a siRNA (SC-106990, Santa Cruze), Scramble control siRNA (SC-37007, Santa Cruze) were electro-transfected into blastocysts in a flat electrode (1 mm gap between electrodes) in 20  $\mu$ l of HEPES-buffered saline (150 mM NaCl, 20 mM HEPES) by two sets of 3 electric pulses of 1 ms at 30 V using a 830 Electro Square Porator (BTX Inc., CA). Following electro-transfection, embryos were cultured in fresh M16 medium (M7292, Sigma), and waited for recovery.

**Statistical analysis**

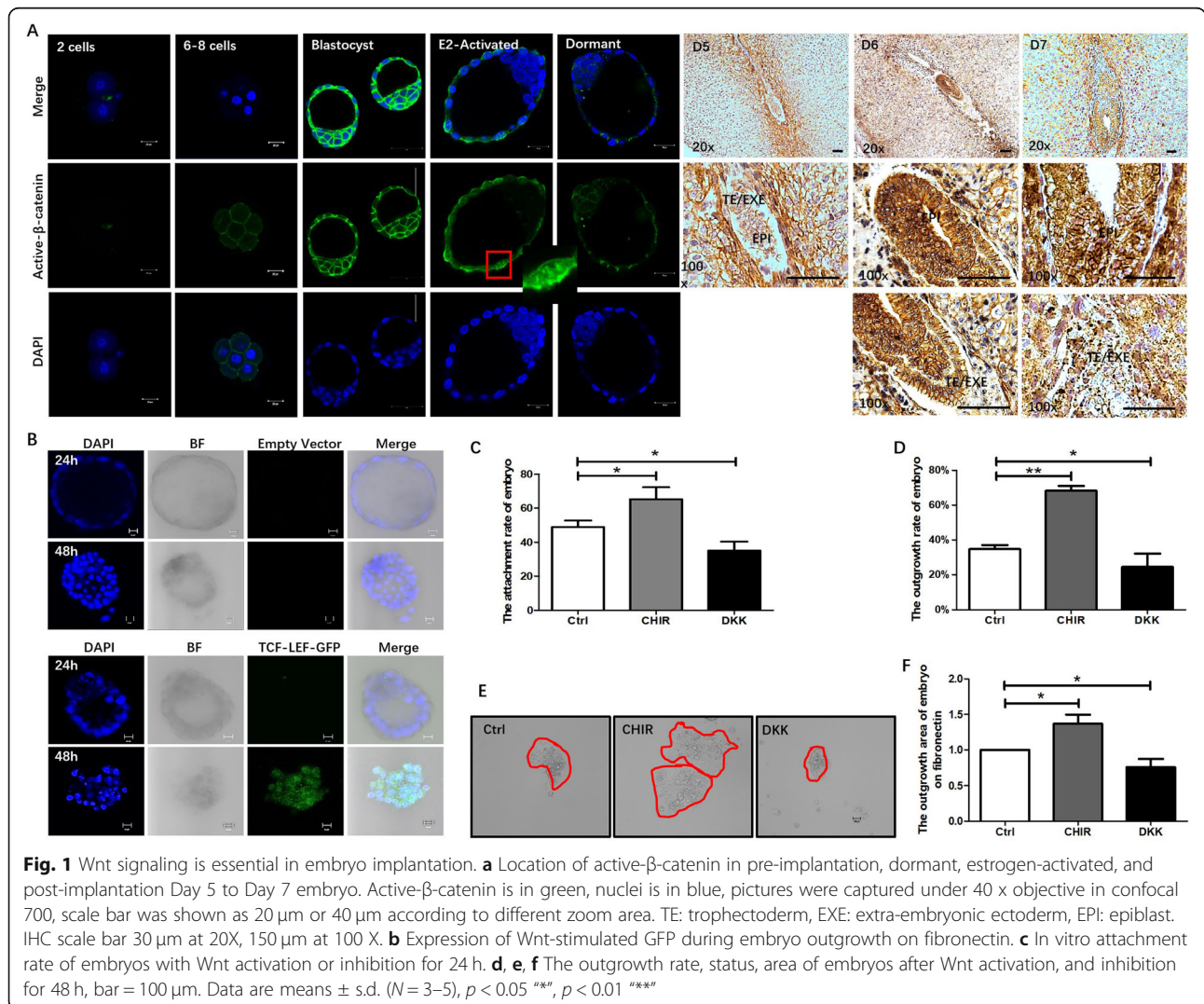
All the images of confocal microscopy were representatives of at least three independent experiments. qPCR

assays were performed in triplicate, and each experiment was repeated for at least three times. Attachment/outgrowth assay has been done at least four times. Data shown were presented as means  $\pm$  sd. Differences were considered statistically significant at  $p < 0.05$  using one way ANOVA.

**Result**

**Wnt signaling is essential in embryo implantation**

Previous researches revealed that Wnt/ $\beta$ -catenin signaling is necessary for embryo implantation competency [6]. Mouse embryo implantation happens at blastocyst stage in Day 4.5. In the pre-implantation embryo, active- $\beta$ -catenin mainly localized on the cellular membrane and increased from 2-cell stage embryos to blastocysts (Fig. 1a). In E2 activated implanting competent embryos, active- $\beta$ -catenin was observed in both membranes, and the nucleus of the trophoctoderm compares with faint immunoactivity only in the membrane of implanting





dormant embryos (Fig. 1a). Compare with pre-implantation embryos, active- $\beta$ -catenin is not only appeared on cellular membrane but also on cytoplasm and nucleus of whole embryo from Day 6 onward, suggesting that Wnt/ $\beta$ -catenin is activated in post-implantation embryos, similar with what happened in E2 activated embryos (Fig. 1a).

To further demonstrate Wnt/ $\beta$ -catenin is necessary for embryo implantation. p-GMT-TCF/LEF-GFP vector, which produces GFP protein upon activation of the canonical Wnt/ $\beta$ -catenin pathway, was electro-transfected into mouse blastocysts. GFP fluorescence was seen automatically expressed in the whole embryo during outgrowth at 48 h, confirming that the pathway was stimulated within this period (Fig. 1b).

The function of Wnt in embryo implantation competency was further studied. Blastocysts were cultured on fibronectin to exclude the effect of the uterine epithelium. Embryos were cultured with Wnt activator CHIR99021 or inhibitor DKK containing M16 medium for 24 h to count attachment and 48 h to observe outgrowth. Wnt stimulation increased embryo attachment (Fig. 1c). At 48 h, more embryos in the Wnt activation group exhibited much evident outgrowth when compared with control, while embryo in the DKK treated group shown decreased outgrowth rate (Fig. 1d). The outgrowth status and outgrowth areas of embryos at 48 h were shown in Fig. 1e and in Fig. 1f, respectively. Similar to the outgrowth rate, Wnt activation enlarged outgrowth area while Wnt inhibition prohibited such expanding ability. Temporal and spatial expression of Wnt ligands in mouse blastocyst has been reported [25, 26], indicating an autocrine manner of embryonic Wnt activation. In addition, the paracrine manner of secrete Wnt ligands from uterus has been testified by detecting Wnt ligands and receptors in peri-implantation mouse uteri and blastocysts in Fig. S1. In summary, Wnt signaling is essential for embryo implantation competency.

#### Repression of let-7a/g by Wnt/ $\beta$ -catenin pathway in the mouse embryo

Several observations suggest that miRNAs are involved in early pre-implantation embryo development and implantation process [14, 27]. Our previous data compare miRNA expression between delay implanting dormant and E2 activated embryos, found a significant decrease in the mir-let-7 family members (including let-7a and let-7g) in the activated embryos [14]. Therefore, the change in let-7 expression in mouse pre-implantation embryos was detected. Mir-let-7a and let-7g were chosen as representative of the family. qPCR detection of the mature let-7a/g shown that the expression decreased more than 50% from the zygote stage to blastocyst, with let-7a decreased at blastocyst stage while let-7

g declined from morula (Fig. 2a), confirming that the mature let-7 expression was suppressed before embryo implantation.

Pathway analysis suggested that Wnt signaling was potentially associated with the differential miRNAs expression between activated and dormant embryos [14]. Wnt activator CHIR99021 and Wnt inhibitor DKK were used to treat blastocysts and cultured for 24 h in M16 medium to determine if the perturbation of Wnt signaling would affect let-7 expression. With Wnt stimulation, active- $\beta$ -catenin goes into the cell nucleus in both ICM and TE cells (Arrow indicated) (Fig. 2b), the corresponding let-7a/g level was decreased (Fig. 2c). DKK treatment decreased active- $\beta$ -catenin in the embryo but up-regulate let-7a/g (Fig. 2b, c). Taken together, the let-7 family member is under negative regulation of Wnt signaling in the embryo.

#### Mir-let-7 family member decreases embryo implantation competency

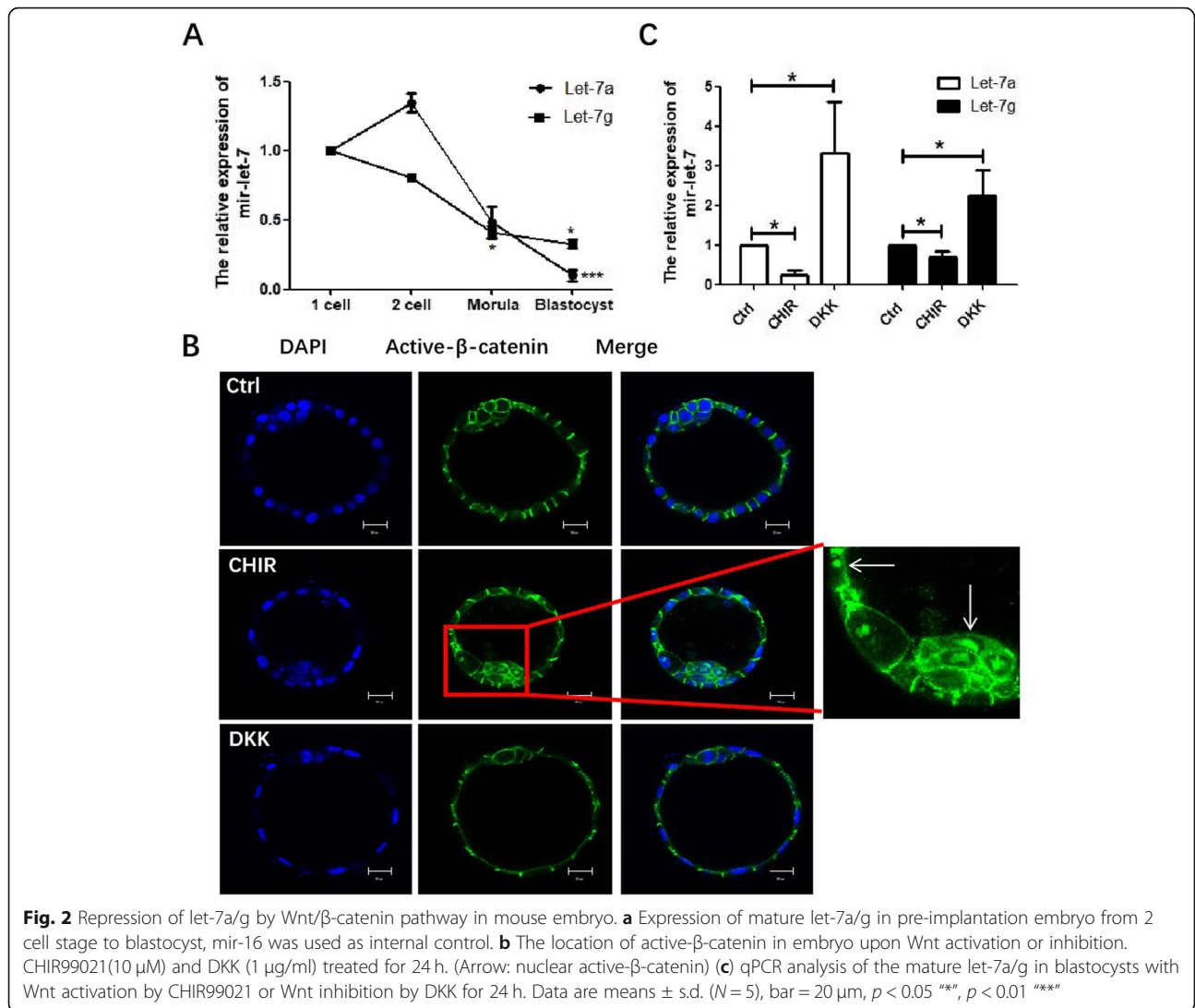
Our lab occasionally owns let-7g-TG mice. As let-7 family members contain the same core sequence, function similarly but location in different chromosomes. Let-7g-TG mice model was used to study the function of let-7 family in embryo implantation. After 4 days of induction, the let-7g level in blastocysts was elevated by more than 15 times relative to the control blastocysts without induction (Fig. 3a). When let-7g overexpressed blastocysts were put onto normal ME in the presence of doxycycline, only 13.33% embryos attached compared with control of 46.66% after 48 h of co-culture (Fig. 3b). At 72 h, let-7g overexpressed embryos had a much lower outgrowth rate (Fig. 3c) and reduced outgrowth area (Fig. 3d, e) than the control without doxycycline treatment.

For the in vivo implantation model, 10–15 Let-7g-TG blastocysts that have been fed with doxycycline for 4 days and control blastocysts that have a normal level of let-7g were transferred into each pseudo-pregnant mice on Day 4. The number of implantation sites was determined on Day 6 of pregnancy (Fig. 3f, g, h). The implantation site in let-7g forced-expressed embryo was evidently decreased when compared with those transferred with normal embryos.

In summary, these results demonstrate that a high level of let-7 family members inhibits embryo implantation competency.

#### Lin28a is directly up-regulated by Wnt/ $\beta$ -catenin and participates in implantation

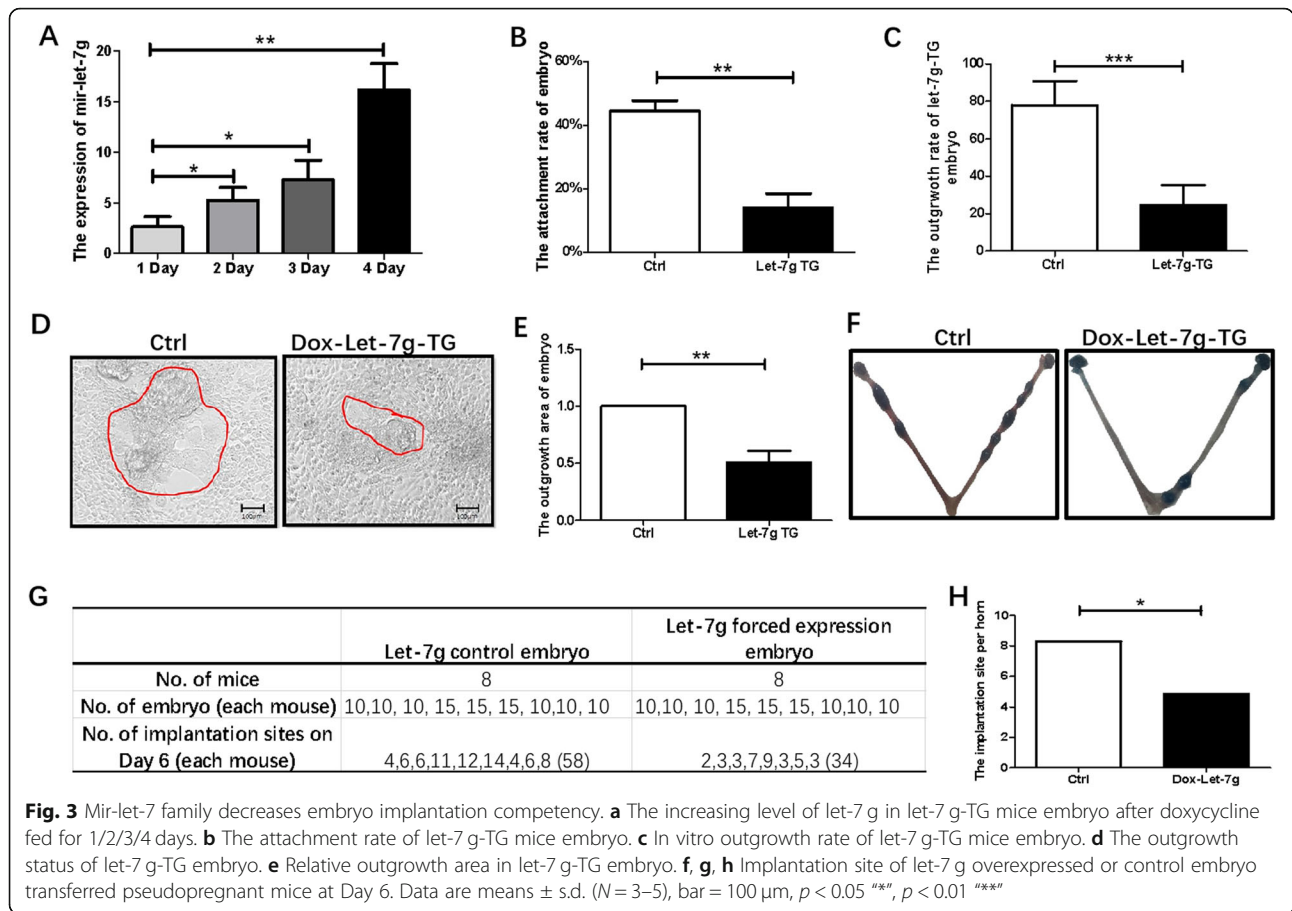
Result in Fig. 2 demonstrated that Wnt signaling modulated let-7 expression. Lin28a is a well-known natural inhibitor of let-7 family [16, 28]. It was hypothesized that lin28a functioned downstream of Wnt signaling to



repress let-7. Therefore, lin28a was determined in the pre-implantation and post-implantation Day 5 to Day 7 embryos. The expression of lin28a was like that of active- $\beta$ -catenin that was up-regulated from 2 cell stage to blastocyst (Fig. 4a). From post-implantation Day 5 onward, lin28a expressed in the cytoplasm and the nucleus in both the TE and the EPI. On Day 5 and Day 6, more lin28a was detected in the EPI than the TE. On Day 7, lin28a expressed in the whole embryo (Fig. 4a).

At the mRNA level, lin28a mRNA increased upon activation and decreased after inhibition of Wnt (Fig. 4b). These results demonstrated that lin28a was positively regulated by Wnt/ $\beta$ -catenin in embryos. To further examine whether  $\beta$ -catenin in canonical Wnt pathway activates lin28a transcription by direct binding to its promoter as a co-activator of LEF/TCF, CHIP assay was performed in mouse trophoblastic stem (mTS) cells, which highly express lin28a as

lin28a is a stem cell maintaining factor [18]. It revealed  $\beta$ -catenin specifically binding to lin28a promoter at five sites in a sequence range upstream of lin28a TSS 1337bp and downstream of TSS 84bp. Besides, two sites that have been published also can be detected in mTS cells (Fig. 4c, d, Pub site1, and 4) [29, 30]. Further, the function of lin28a in embryo implantation competency was detected. The level of lin28a in doxycycline induced Lin28a-TG blastocysts was confirmed by qPCR (Fig. 4e). Lin28a overexpression elevated embryo attachment rate from 41.31 to 53.34% (Fig. 4f). The outgrowth rate increased from 45.67 to 56.31% at 48 h (Fig. 4g). With lin28a overexpression, the embryo outgrowth area was evidently raised (Fig. 4h, i). Collectively, these results demonstrate that lin28a is direct positively regulated by Wnt/ $\beta$ -catenin and important for embryo implantation.



**The function of Wnt in embryo implantation is mediated through lin28a/let-7a axis**

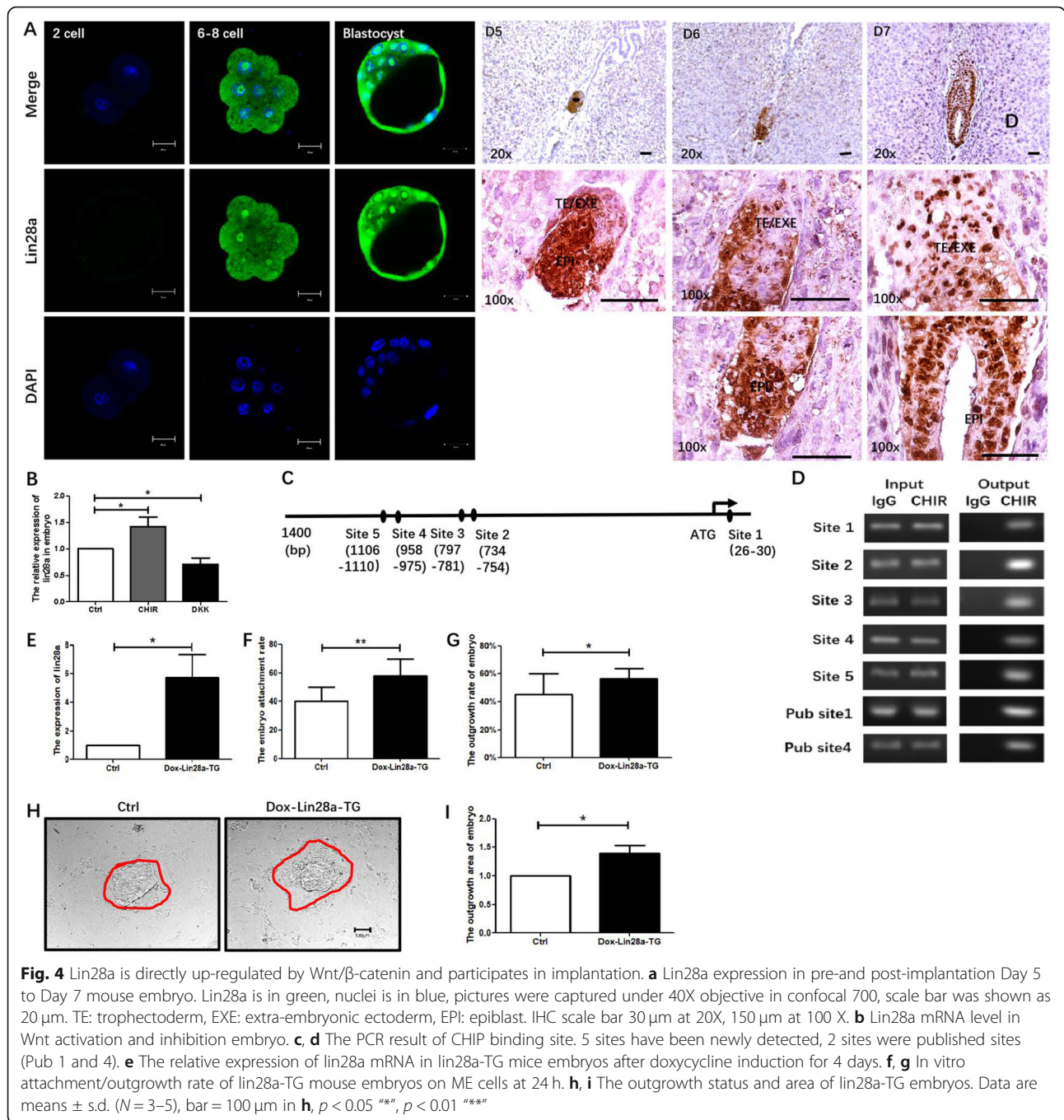
To verify whether the regulation of Wnt/β-catenin on let-7 was through lin28a in mouse embryos. Blastocysts were electro-transfected with scrambled control siRNA and lin28a siRNA respectively and cultured for 24 h in M16 medium. The efficiency of lin28a knockdown was confirmed by immunofluorescence staining and qPCR, respectively (Fig. 5a, b). The ICM in the lin28a-siRNA group is less than control may ascribe to the function of cell proliferation was inhibited by lin28a knockdown [18, 19]. Besides, combined Wnt activation and lin28a knockdown, the level of let-7a restored to that of the control electro-transfected with scrambled siRNA and without Wnt stimulation (Fig. 5c). These outcomes suggest that lin28a mediates the repression action of Wnt on let-7.

It was further hypothesized that Wnt signaling facilitates embryo implantation via lin28a/let-7 axis in the process of implantation. Mouse blastocysts were electro-transfected with let-7a inhibitor or scramble siRNA in the presence or absence of Wnt inhibitor DKK. The treated embryos were cultured on the fibronectin-coated plate in M16 medium for 24 h. DKK alone decreased embryo attachment on fibronectin, while let-7a inhibitor

alone increased embryo attachment. Co-treatment of the embryo with DKK and let-7a inhibitor abolished the suppressive effect of Wnt inhibition on embryo attachment (Fig. 5d), suggesting that let-7a mediated the action of Wnt on embryo implantation. A similar experiment was done with lin28a overexpression in lin28a-TG embryo with DKK treatment (Fig. 5e). With Wnt inhibition, the embryo attachment rate was decreased significantly, while lin28a overexpression increased the attachment rate. When DKK was applied to lin28a overexpressed embryos, the attachment rate rescued to the control level. Taken together, lin28a/let-7a signaling mediates the effects of Wnt on embryo implantation.

**Let-7 family members disturb embryonic EMT**

Embryonic EMT participates in implantation, placentation, and organ development, which is essential for a successful pregnancy [4, 31, 32]. To observe whether the adverse effect of the let-7 family member on embryo implantation was because of disturb EMT process. Blastocysts with or without let-7 g overexpression were transferred to pseudopregnant normal Day 4 female mouse uteri. Implantation sites were detected on Day 6



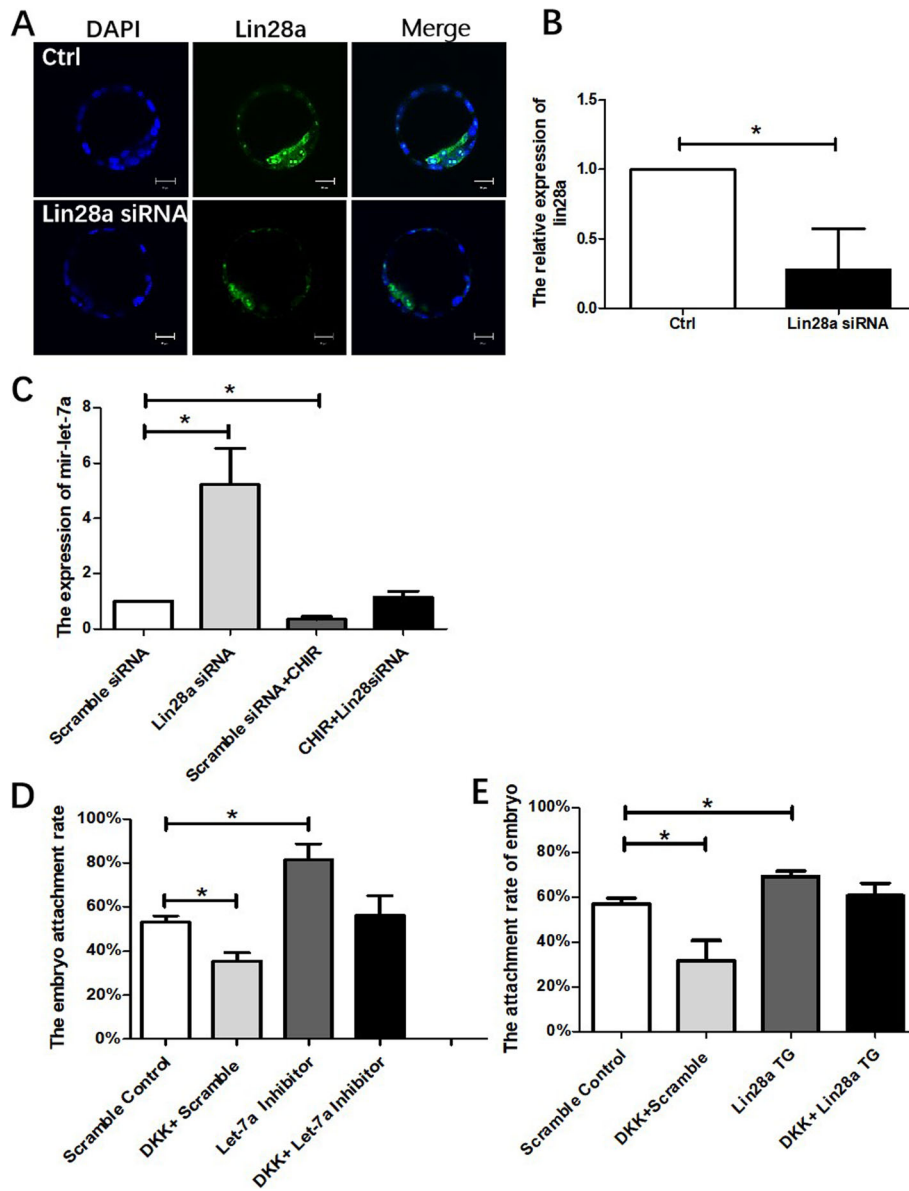
by Chicago blue caudal vein injection, fixed and sectioned for IHC staining. As shown in Fig. 6a, epithelial marker E-cadherin was stronger in let-7 g-TG embryo. In contrast, mesenchymal marker N-cadherin expressed in the whole uterus as the majority of the uterine cells at this stage were mesenchymal cells, which was significantly less in the let-7 g-TG embryos relative to untreated control (Fig. 6b), indicating impairment of EMT. The expression of active- $\beta$ -catenin (Fig. 6c) and lin28a (Fig. 6d) were higher in the control embryos than the

let-7 g overexpressed embryos. These outcomes reveal overexpressed let-7 g in embryo disturb embryo implantation via impairing EMT process. In addition, the in vitro embryonic EMT process was also promoted by lin28a (Fig. S2), while attenuated by the let-7 g (Fig. S3).

### Discussion

Active- $\beta$ -catenin has been reported expressed in the nucleus of mouse embryos from 1 cell to blastocyst [6]. However, nuclear active- $\beta$ -catenin could not be detected

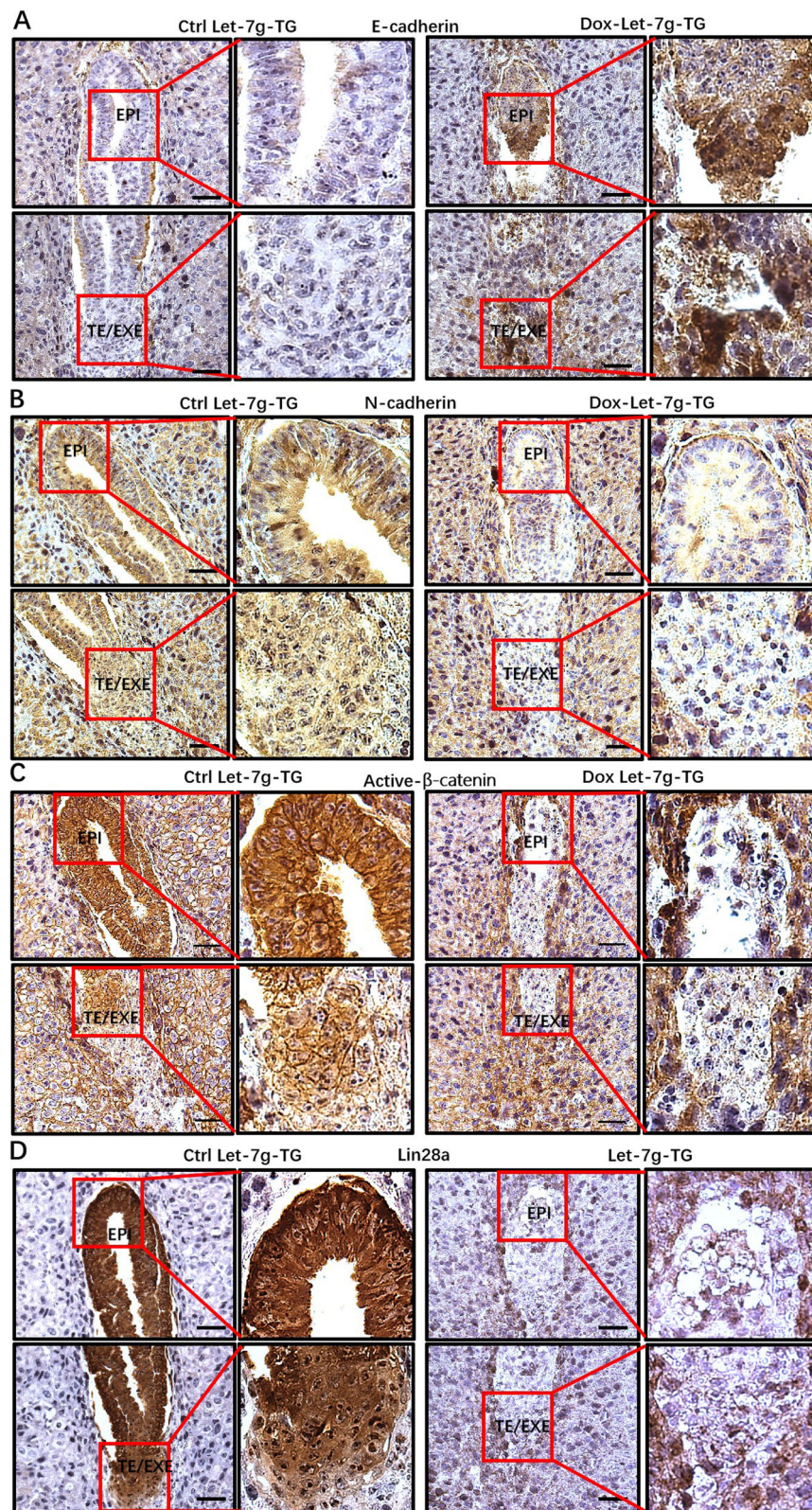




**Fig. 5** The function of Wnt in embryo implantation is mediated through lin28a/let-7a. **a** The efficiency of lin28a siRNA knockdown in blastocysts by immunostaining. **b** The relative lin28a mRNA level in lin28a siRNA knockdown embryo. **c** The relative expression of let-7a in lin28a knockdown but Wnt activation blastocyst. **d** Embryo attachment rate on fibronectin when embryo Wnt was inhibited and/or let7a was knockdown. **e** Embryo attachment rate on fibronectin when embryo Wnt was inhibited by DKK and/or lin28a was overexpressed (lin28a-TG). Data are means  $\pm$  s.d. ( $N = 3-5$ ), bar = 20  $\mu$ m,  $p < 0.05$  "\*",  $p < 0.01$  "\*\*",  $p < 0.001$  "\*\*\*"

in the pre-implantation stages in this research even with the use of the same reported antibody or other antibodies claimed against active- $\beta$ -catenin. Instead, active- $\beta$ -catenin localized to the cellular membrane, despite there was an increasing trend in the pre-implantation stage of pregnancy. From Day 6 onward, active- $\beta$ -catenin was detected in the cell membrane, cytoplasm, and nucleus. The finding is consistent with the current understanding of Wnt signaling in embryo development and implantation, that is, Wnt signaling is not necessary

for blastocyst formation, which is supported by (1) DKK treatment has no apparent adverse effect on embryo develop from 2 cell stage to blastocysts [6, 33, 34]. (2)  $\beta$ -Catenin-null embryos ( $\beta$ -cat  $-/-$ ) can develop normally until Day 6.5 [35]. (3) Truncated  $\beta$ -catenin in the embryo doesn't affect embryo development to blastocyst in vitro [36]. Besides, it is reported that  $\beta$ -catenin forms adhesion junction with E-cadherin soon on the cellular membrane after being synthesized [37]. E-cadherin sequesters  $\beta$ -catenin onto the cellular membrane, without



**Fig. 6** Let-7 family members disturb embryonic EMT. **a** Immunohistochemistry staining of E-cadherin, **b** N-cadherin, **c** Active-β-catenin, **d** Lin28a in control let-7 g and let-7 g overexpressed embryo (Let-7 g-TG) after embryo transfer. EPI: epiblast. TE: trophoctoderm. EXE: extra-embryonic ectoderm. Scale bar: 60 μm



which  $\beta$ -catenin can go into the nucleus [36, 38, 39]. I have detected pre-implantation embryo E-cadherin, it was indeed on the cellular membrane (data not shown). Taken together, firstly, Wnt signaling is not a necessary condition for blastocyst formation; Secondly,  $\beta$ -catenin is sequestered by E-cadherin on cellular membrane. These results together explain the pre-implantation membrane staining of active- $\beta$ -catenin in this finding.

Though active- $\beta$ -catenin mediated canonical Wnt signaling is not required for pre-implantation embryo development, silencing of Wnt/ $\beta$ -catenin signaling in pre-implantation embryo reduces *Cdx2*, a key transcription factor involved in TE lineage specification, and considerably reduced implantation rate upon transfer to pseudo-pregnant recipients, and the Wnt downstream target *c-Myc*, which has been shown to be crucial for pre-implantation embryo development, is also down-regulated in blastocyst TE [6]. Similarly, in porcine parthenogenetic (PA) embryos, inhibition of Wnt signaling doesn't alter the blastocyst formation rate. However, the hatching rate is decreased, and TE development is blocked. In addition, with the activation of Wnt signaling, PA blastocysts hatching improved dramatically [40]. These findings together suggest that Wnt signaling is required for embryo implantation competency.

Embryonic EMT during implantation is designed as type I EMT [3]. In our finding, active- $\beta$ -catenin goes into cytoplasm and nucleus from Day 6.5 onward, stimulation Wnt signaling in embryo improves implantation capacity by strengthening embryo attachment and outgrowth ability. Besides, in *let-7 g* overexpressed, implantation competency weakens embryo, the post-implantation EMT process was disturbed with an impairment of Wnt/ $\beta$ -catenin signaling *in vivo*. *In vitro*, the embryonic EMT process was also promoted by *lin28a* but attenuated by the *let-7 g*. These evidences together provide support for the role of the canonical Wnt/ $\beta$ -catenin pathway in embryo implantation and accompanying EMT.

In the present study, *lin28a* locates in both cytoplasm and nucleus, especially in the nucleolus of mouse embryos (Fig. 4). *Lin28a* has been reported to work in both the cytoplasm and the nucleus; it recruits a TUTase to block the processing of *let-7* precursors by Dicer in the former, while sequesters primary *let-7* transcripts inhibiting their processing by the microprocessor in a TUTase independent manner in the latter [19, 41]. SET7/9-mediated methylation of *lin28a* at lysine 135, a region homologous with the NoLS (Nucleolar Localization Signal) of its homologous family member *lin28b*, increases *lin28a* stability, promotes localization of the molecule in the nucleus and multimerization with *mir-let-7* [42]. Besides, *lin28a* is an essential factor of nucleolar-genesis during early mouse embryonic

development [43]. These evidences combine with the present findings of high expression of *lin28a* in pre and post-implantation embryo and its function in promoting EMT suggest *lin28a* is important for implantation.

In breast cancer, Wnt/ $\beta$ -catenin inhibits breast cancer stem cell expansion through direct binding to *lin28a* promoter and inhibits *let-7a* [30]. Besides, the direct regulation of Wnt to *lin28* was also found in the adult mammalian retina. CHIP assay reveals that  $\beta$ -catenin binding to two distinct sites in the proximal region of *lin28a* and *lin28b* promoters. This binding is critical for *lin28* family transcriptional activation [29]. Interestingly, the two  $\beta$ -catenin binding sites in the two types of research are different, suggesting that distinct  $\beta$ -catenin binding sites may be used to drive *lin28* transcription in different tissues. A similar situation is found in this research: a total of five  $\beta$ -catenin binding sites were detected by CHIP assay in mouse trophoblast stem cells. Besides, the two binding sites that have been reported also can be detected [29]. It is possible that the already published four sites were included in the 5-binding sites in this finding; however, it is undeterminable as there is no detail information for those sites found in their studies.

*Let-7a/g* were decreased suddenly in the pre-implantation embryo. Overexpressed *let-7 g* in *let-7 g*-TG mice embryo decreased embryo implantation ability. Integrin-3 $\beta$  has been proved as a downstream target of *mir-let-7* in our previous study [14]. Integrins are a family of heterodimeric ( $\alpha/\beta$ ) transmembrane molecules that responsible for blastocyst adhesion through activation of phosphoinositide-specific phospholipase C leading to initiation of phosphoinositide signaling [44]. Integrin  $\alpha\beta3$  has been detected on the apical surface of the TE of blastocysts and mediates primary trophoblast adhesion and migration [45]. Thus, decreased integrin-3 $\beta$  in the high *let-7 g* embryo may be a possible reason for impaired embryo implantation ability.

There is only one publication mentioned that the promoter activity of *mir-let-7a2* in the breast cancer cell line could be downregulated by Wnt activator lithium chloride in luciferase assay [46]. It means Wnt could suppress *let-7a2* transcriptional activity through repressing its promoter, but whether this is a direct effect is not known and needs further study.

Wnt signaling and the mesenchymal marker were decreased in *let-7 g* overexpressed embryonic cells. It means *let-7 g* inhibit EMT process in the embryo. It has been reported overexpressed *let-7b* activating epithelial marker E-cadherin and attenuating mesenchymal marker Vimentin, EMT transcriptional factor Twist as well as  $\beta$ -catenin in malignant pleural mesothelioma [47]. In this research, with *let-7 g* overexpression, EMT process in post-implantation embryonic development was blocked.

## Conclusions

In this study, current understanding is Wnt/ $\beta$ -catenin is essential for proper embryo implantation, and EMT, which function via directly binding to lin28a promoter to activate its transcription, thus represses mir-let-7 biogenesis. Lin28a and mir-let-7 family inversely function in embryo implantation competency and EMT under the regulation of Wnt signaling. In conclusion, our studies first shed light on Wnt/ $\beta$ -catenin-lin28a/let-7a pathway in embryo implantation and accompanying EMT.

## Supplementary information

**Supplementary information** accompanies this paper at <https://doi.org/10.1186/s12964-020-00562-5>.

**Additional file 1.**

**Additional file 2.**

**Additional file 3.**

## Abbreviations

ICM: Inner cell mass; TE: Trophoctoderm; EMT: Epithelial-mesenchymal transition; Lin28aTG/Let-7 gTG: Lin28a/Let-7 g transgenic mice; EPI: Epiblast; ICR: Imprinted-coding region; ME: Mouse primary endometrium; mTS: Trophoblastic stem

## Acknowledgments

We are sincerely grateful to all the reviewers for their helpful comments on this article, and those who provide kind help during the whole project.

## Availability of supporting data

All data generated or analyzed during this study are included in this published article (and its supplementary information files).

## Authors' contributions

Conceived and designed the experiments: Weimin LIU. Performed the experiments: Qian LI. Analyzed the data: Qian LI. Wrote the paper: Qian LI. Revise manuscript: Juanzi SHI. All authors read and approved the final manuscript.

## Funding

Not applicable.

## Availability of data and materials

The datasets used and analysed during the current study are available from the corresponding author on reasonable request.

## Ethics approval and consent to participate

The use of animals was approved by the Committee on Use of Live Animals in Teaching and Research, the University of Hong Kong (Approval no. 3534-4, 4100-16, and 3573-15).

## Consent for publication

Not applicable.

## Competing interests

The authors declare that they have no competing interests.

Received: 8 January 2020 Accepted: 23 March 2020

Published online: 11 July 2020

## References

- Lanner F. Lineage specification in the early mouse embryo. *Exp Cell Res*. 2014;321(1):32–9.
- Cross JC. Genetic insights into trophoblast differentiation and placental morphogenesis. *Semin Cell Dev Biol*. 2000;11(2):105–13.
- Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest*. 2009;119(6):1420–8.
- Hay ED. The mesenchymal cell, its role in the embryo, and the remarkable signaling mechanisms that create it. *Dev Dyn*. 2005;233(3):706–20.
- Acloque H, et al. Epithelial-mesenchymal transitions: the importance of changing cell state in development and disease. *J Clin Invest*. 2009;119(6):1438–49.
- Xie H, et al. Inactivation of nuclear Wnt-beta-catenin signaling limits blastocyst competency for implantation. *Development*. 2008;135(4):717–27.
- Liu P, et al. Requirement for Wnt3 in vertebrate axis formation. *Nat Genet*. 1999;22(4):361–5.
- Mohamed OA, Clarke HJ, Dufort D. Beta-catenin signaling marks the prospective site of primitive streak formation in the mouse embryo. *Dev Dyn*. 2004;231(2):416–24.
- Chekulaeva M, Filipowicz W. Mechanisms of miRNA-mediated post-transcriptional regulation in animal cells. *Curr Opin Cell Biol*. 2009;21(3):452–60.
- Gross N, Kropp J, Khatib H. MicroRNA Signaling in Embryo Development. *Biology (Basel)*. 2017;6(3):34.
- Suh N, et al. MicroRNA function is globally suppressed in mouse oocytes and early embryos. *Curr Biol*. 2010;20(3):271–7.
- Bernstein E, et al. Dicer is essential for mouse development. *Nat Genet*. 2003;35(3):215–7.
- Alisch RS, et al. Argonaute2 is essential for mammalian gastrulation and proper mesoderm formation. *PLoS Genet*. 2007;3(12):e227.
- Liu WM, et al. Involvement of microRNA lethal-7a in the regulation of embryo implantation in mice. *PLoS One*. 2012;7(5):e37039.
- Li Q, et al. Regulation of human trophoblast surrogate Jeg-3 spheroids implantation potential by Wnt/beta-catenin pathway and lin28a/let-7a axis. *Exp Cell Res*. 2020;388(1):111718.
- Piskounova E, et al. Lin28A and Lin28B inhibit let-7 microRNA biogenesis by distinct mechanisms. *Cell*. 2011;147(5):1066–79.
- Tu HC, et al. LIN28 cooperates with WNT signaling to drive invasive intestinal and colorectal adenocarcinoma in mice and humans. *Genes Dev*. 2015;29(10):1074–86.
- Yu J, et al. Induced pluripotent stem cell lines derived from human somatic cells. *Science*. 2007;318(5858):1917–20.
- Tsialikas J, Romer-Seibert J. LIN28: roles and regulation in development and beyond. *Development*. 2015;142(14):2397–404.
- King CE, et al. LIN28B fosters colon cancer migration, invasion and transformation through let-7-dependent and -independent mechanisms. *Oncogene*. 2011;30(40):4185–93.
- Wang YC, et al. Lin-28B expression promotes transformation and invasion in human hepatocellular carcinoma. *Carcinogenesis*. 2010;31(9):1516–22.
- Guo Y, et al. Identification and characterization of LIN-28 homolog B (LIN28B) in human hepatocellular carcinoma. *Gene*. 2006;384:51–61.
- Peng S, et al. Genome-wide studies reveal that Lin28 enhances the translation of genes important for growth and survival of human embryonic stem cells. *Stem Cells*. 2011;29(3):496–504.
- Deb K, Reese J, Paria BC. Methodologies to study implantation in mice. *Methods Mol Med*. 2006;121:9–34.
- Mohamed OA, Dufort D, Clarke HJ. Expression and estradiol regulation of Wnt genes in the mouse blastocyst identify a candidate pathway for embryo-maternal signaling at implantation. *Biol Reprod*. 2004;71(2):417–24.
- Hayashi K, et al. Wnt genes in the mouse uterus: potential regulation of implantation. *Biol Reprod*. 2009;80(5):989–1000.
- Melton C, Judson RL, Belloch R. Opposing microRNA families regulate self-renewal in mouse embryonic stem cells. *Nature*. 2010;463(7281):621–6.
- Wang T, et al. Aberrant regulation of the LIN28A/LIN28B and let-7 loop in human malignant tumors and its effects on the hallmarks of cancer. *Mol Cancer*. 2015;14:125.
- Yao K, et al. Wnt regulates proliferation and neurogenic potential of Muller glial cells via a Lin28/let-7 miRNA-dependent pathway in adult mammalian retinas. *Cell Rep*. 2016;17(1):165–78.
- Cai WY, et al. The Wnt-beta-catenin pathway represses let-7 microRNA expression through transactivation of Lin28 to augment breast cancer stem cell expansion. *J Cell Sci*. 2013;126(Pt 13):2877–89.
- Kim DH, et al. Epithelial mesenchymal transition in embryonic development, tissue repair and cancer: a comprehensive overview. *J Clin Med*. 2017;7(1):1.
- Gonzalez DM, Medici D. Signaling mechanisms of the epithelial-mesenchymal transition. *Sci Signal*. 2014;7(344):re8.



33. Meissner A, Jaenisch R. Generation of nuclear transfer-derived pluripotent ES cells from cloned Cdx2-deficient blastocysts. *Nature*. 2006;439(7073):212–5.
34. Niwa H, et al. Interaction between Oct3/4 and Cdx2 determines trophoblast differentiation. *Cell*. 2005;123(5):917–29.
35. Haegel H, et al. Lack of beta-catenin affects mouse development at gastrulation. *Development*. 1995;121(11):3529–37.
36. De Vries WN, et al. Maternal beta-catenin and E-cadherin in mouse development. *Development*. 2004;131(18):4435–45.
37. McCrea PD, Gumbiner BM. Purification of a 92-kDa cytoplasmic protein tightly associated with the cell-cell adhesion molecule E-cadherin (uvomorulin). Characterization and extractability of the protein complex from the cell cytostructure. *J Biol Chem*. 1991;266(7):4514–20.
38. Hendriksen J, et al. Plasma membrane recruitment of dephosphorylated beta-catenin upon activation of the Wnt pathway. *J Cell Sci*. 2008;121(11):1793–802.
39. Du W, et al. From cell membrane to the nucleus: an emerging role of E-cadherin in gene transcriptional regulation. *J Cell Mol Med*. 2014;18(9):1712–9.
40. Huang Y, et al. Moderate expression of Wnt signaling genes is essential for porcine parthenogenetic embryo development. *Cell Signal*. 2013;25(4):778–85.
41. Viswanathan SR, Daley GQ. Lin28: a microRNA regulator with a macro role. *Cell*. 2010;140(4):445–9.
42. Kim SK, et al. SET7/9 methylation of the pluripotency factor LIN28A is a nucleolar localization mechanism that blocks let-7 biogenesis in human ESCs. *Cell Stem Cell*. 2014;15(6):735–49.
43. Vogt EJ, et al. Importance of the pluripotency factor LIN28 in the mammalian nucleolus during early embryonic development. *Development*. 2012;139(24):4514–23.
44. Wang J, Mayernik L, Armant DR. Trophoblast adhesion of the peri-implantation mouse blastocyst is regulated by integrin signaling that targets phospholipase C. *Dev Biol*. 2007;302(1):143–53.
45. Sutherland AE, Calarco PG, Damsky CH. Developmental regulation of integrin expression at the time of implantation in the mouse embryo. *Development*. 1993;119(4):1175–86.
46. Guan H, et al. Characterization and functional analysis of the human microRNA let-7a2 promoter in lung cancer A549 cell lines. *Mol Biol Rep*. 2011;38(8):5327–34.
47. Sohn EJ, et al. Blockage of epithelial to mesenchymal transition and upregulation of let 7b are critically involved in ursolic acid induced apoptosis in malignant mesothelioma cell. *Int J Biol Sci*. 2016;12(11):1279–88.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

