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Perspective

## PERSPECTIVE

# Unraveling the potential of non-conventional yeasts in biotechnology

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One sentence summary: This article highlights the importance of NCY in research and industry and offers some perspectives to accelerate the application of NCY in biotechnology.

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## **ABSTRACT**

Cost-effective microbial conversion processes of renewable feedstock into biofuels and biochemicals are of utmost importance for the establishment of a robust bioeconomy. Conventional baker's yeast Saccharomyces cerevisiae, widely employed in biotechnology for decades, lacks many of the desired traits for such bioprocesses like utilization of complex carbon sources or low tolerance towards challenging conditions. Many non-conventional yeasts (NCY) present these capabilities, and they are therefore forecasted to play key roles in future biotechnological production processes. For successful implementation of NCY in biotechnology, several challenges including generation of alternative carbon sources, development of tailored NCY and optimization of the fermentation conditions are crucial for maximizing bioproduct yields and titers. Addressing these challenges requires a multidisciplinary approach that is facilitated through the 'YEAST4BIO' COST action. YEAST4BIO fosters integrative investigations aimed at filling knowledge gaps and excelling research and innovation, which can improve biotechnological conversion processes from renewable resources to mitigate climate change and boost transition towards a circular bioeconomy. In this perspective, the main challenges and research efforts within YEAST4BIO are discussed, highlighting the importance of collaboration and knowledge exchange for progression in this research field.

Keywords: lignocellulosic sugars; volatile fatty acids; non-conventional yeasts; biochemicals; biofuels

## INTEREST IN NON-CONVENTIONAL YEASTS

Yeast biotechnology has been applied for decades for production of foods and beverages, biopharmaceuticals, industrial biocatalysts and bioethanol. So far, the bioconversion processes have mainly relied on strains of conventional baker's yeast Saccharomyces cerevisiae as cell factories. In next generation production

processes, renewable and low-cost carbon sources including forestry, agricultural and industrial waste- and side-streams are converted to various bioproducts including biofuels, bulk and fine biochemicals and biomaterials (Mattanovich, Sauer and Gasser 2014; Kavšček *et al.* 2015). Such processes require a range of different cell factories tailored for the specific purposes, and

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S. cerevisiae lacks many of the traits needed for efficient conversion of these alternative substrates to products of interest. For example, wild type S. cerevisiae strains utilize primarily hexose sugars such as glucose and do not readily use multiple carbon sources simultaneously. This hampers the yeast's usefulness in fermentations of mixed-sugar substrates such as lignocellulosic hydrolysates, which apart from hexoses contain high concentrations of pentoses (Katahira et al. 2008). Saccharomyces cerevisiae is a good producer of alcohols, esters and organic acids but does not naturally accumulate high amounts of intracellular lipids and consume other residual carbon sources like short chain fatty acids, which limits its applications as cell factory (Zhang, Nielsen and Liu 2021). Thus, either extensive S. cerevisiae strain development is needed, or, alternatively, the biotech industry needs to turn to other microorganisms that are naturally more suited for intended tasks. In this respect, other yeast species, collectively referred to as non-conventional yeasts (NCY), are progressively gaining more attention as new potential workhorses for biotechnological applications (Navarrete and Martínez 2020).

To date, a few thousand different yeast species have been identified and described to various degrees. Only a small number of these species have been characterized in relatively large detail and are frequently used as cell factories, including but not limited to Yarrowia lipolytica, Komagataella phaffii (Pichia pastoris), Hansenula/Ogataea polymorpha, Kluyveromyces lactis, Kluyveromyces marxianus, Zygosaccharomyces bailii, Rhodotorula toruloides and Phaffia rhodozyma (Radecka et al. 2015; Rebello et al. 2018; Wen et al. 2020; Binati et al. 2021). Besides, a few hundred NCY species have now been genome sequenced and growth phenotyped (Shen et al. 2018), and the results show interesting traits in many of these emerging yeasts. It is projected that the vast majority of the existing yeast species have still not been isolated and identified (Lachance 2006), and thus yeast biodiversity can be considered as a largely untapped resource for biotechnological applications.

NCY is a hugely diverse group of organisms, where the phylogenetic span of just the hemiascomycetous yeasts is as big as the whole phylum of chordates (Dujon et al. 2004). This means that collectively, the NCY's are far more multifaceted than S. cerevisiae alone. Many NCY can grow and ferment at wide ranges of process conditions and can utilize, apart from glucose, a large variety of carbon sources present in renewable feedstock such as monosaccharides (xylose, galactose, fructose, arabinose), disaccharides (lactose, cellobiose, xylobiose) and even polysaccharides (xylan and cellulose) (Shen et al. 2018; Ravn et al. 2021). Some NCY species are highly tolerant to bioprocessinduced stresses including low pH, elevated temperatures and high osmolarities as well as inhibitory compounds formed during plant biomass pretreatment (Deparis et al. 2017; Mukherjee et al. 2017; Palma et al. 2017; Navarrete et al. 2021). On the product side, some NCY are superior to S. cerevisiae when it comes to accumulation of metabolites and synthesis and secretion of recombinant proteins (Corchero et al. 2013). Additionally, some oleaginous yeasts such as Rhodotorula glutinis and Lipomyces starkeyi can accumulate lipids up to 60%-70% of their biomass. Thus, the ability of non-conventional oleaginous yeasts to accumulate high quantities of lipids offers the commercial potential for production of lipids or 'single-cell oils', advanced biofuels generation and accumulation of lipid-soluble fine chemicals of high value, such as carotenoids and surfactants (Bharathiraja et al. 2017).

In the last decade, great progress has been made in NCY utilization for biotechnological applications, as well as in proposing solutions to improve their efficiency (Wagner and Alper 2016; Wang et al. 2020). Nonetheless, due to the many potential applications of these microorganisms that remain unexplored, there are numerous challenges that need to be addressed to fully exploit the advantages of NCY in biotechnology. First of all, to be able to use new strains and species in industrial production processes, extensive characterization and testing of the microorganism(s) of interest is needed. Second, when feeding NCY with alternative low-cost substrates, a detailed understanding of the substrate composition and its interaction with the microorganism of choice is required. Third, process development including optimization of the NCY cultivation parameters and modes is usually needed to boost production titers, yields and productivity. Fourth, strain development using adaptive laboratory evolution (ALE) is often essential to improve/enhance the interesting traits. Metabolic engineering was for long restricted to model organisms such as S. cerevisiae. Nowadays, it is an emerging technology in NCY research, much thanks to the recent technology revolution in terms of synthetic biology, genome sequencing and CRISPR/Cas9. However, for most NCY, metabolic engineering tools are still lacking or in the process of being developed, and great progress in this field is expected within the coming years (Raschmanová et al. 2018).

## YEAST4BIO: DRIVING RESEARCH AND INNOVATION OF NCY-BASED BIOPROCESSES FORWARD THROUGH CROSS-DISCIPLINARY COLLABORATION

To address the many challenges with developing novel NCY cell factories and bioprocesses, collaboration, sharing of knowledge and cross-disciplinary research approaches are highly needed. Lately, this has been greatly facilitated through the 'YEAST4BIO' network funded by COST (European Cooperation in Science and Technology) (https://yeast4bio.eu/). The YEAST4BIO COST Action brings together yeast research communities from both industry and academia in Europe and beyond, to foster integrative investigations aimed at filling knowledge gaps and excelling research and innovation in the field. The relevance of YEAST4BIO is clear as there is an urgent need to improve biotechnological conversion processes from renewable resources to mitigate climate change and transition towards a stable and circular bioeconomy.

Meeting the challenges in NCY utilization for bioproduct generation requires a multidisciplinary approach. Therefore, YEAST4BIO has gathered researchers with a combination of skills and experience in biomass fractionation and hydrolysis, yeast cultivation and fermentation and NCY strain characterization and development through the means of molecular biology, genetics and physiology. Besides facilitation of networking, collaboration and knowledge exchange, YEAST4BIO aims to provide its members and in extension the global scientific community with: (i) a compilation of existing scientific knowledge of NCY and applications at the European level and beyond; (ii) standardized experimental techniques and procedures for production of carbon sources from low-cost alternative substrates to be used as platform molecules for NCY; (iii) access to genetic and systems/synthetic biology tools already available to be applied on NCY and (iv) a portfolio of bioproducts to be produced from both

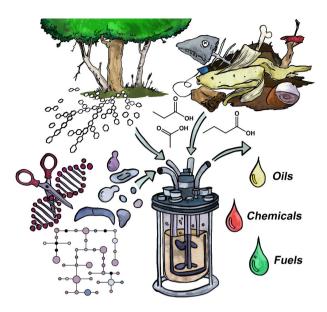


Figure 1. Overview of the main challenges addressed by YEAST4BIO, where renewable and low-cost carbon sources are generated and used as starting materials for generation of bioproducts using NCY, which can be characterized and developed using systems and synthetic biology techniques.

carboxylic and sugar platforms by means of NCYs. Altogether, these deliverables will help facilitating NCY-based conversion of renewable raw materials into biofuels and other important bioproducts in a zero-waste circular economy context.

#### YEAST4BIO FOCUS AREAS

YEAST4BIO has identified three main challenges of utmost importance for exploring and exploiting the full potential of NCY in biotechnology. These challenges, namely renewable and lowcost substrate generation, efficient NCY-based bioproduct formation and molecular strain characterization and development, are described in more detail below and depicted in Fig. 1.

## Renewable and low-cost carbon source generation for NCY

Glucose and sucrose are classical carbon sources for industrial yeast processes, however, the price for these sugars can make up as much as 60%-80% of the overall production cost (Koutinas et al. 2014). Thus, the successful expansion of industrial biotechnology towards a sustainable economy drives research to the utilization of alternative low-cost carbon sources.

With the aim of converting renewable raw materials into fermentable carbon sources for NCY, two main routes can be considered: (i) enzymatic hydrolysis of biomass-contained carbohydrates, and (ii) production of carboxylic acids (i.e. volatile fatty acids, VFAs) from organic residues by anaerobic fermentation. Recent biotechnological advances in biomass deconstruction and anaerobic fermentation have facilitated utilization of both lignocellulosic sugars and carboxylates for the biotechnological production of biofuels and (fine) biochemicals. However, each carbon source generation route faces specific challenges that are assessed and studied within YEAST4BIO.

## The Sugar Platform

The production of sugars as platform molecules is one of the most attractive biotechnological ways to valorize organic wastes or lignocellulosic substrates. To date, there have been several initiatives to optimize the sugar release by means of pretreatment and enzymatic catalysts from biomass. Enzymatic hydrolysis is a relatively new concept and during the last two decades, significant advances have been made in the development of more efficient and cheaper enzymatic catalysts (Cannella and Jørgensen 2014). Notwithstanding, significant progress is still needed to increase performance and productivity of cellulosic enzymes to reduce the costs for production of sugars.

The organic residues are composed of different types of polysaccharides whose enzymatic hydrolysis ultimately releases a set of monosaccharides including glucose, galacturonic acid, arabinose, galactose, rhamnose, xylose and mannose, which can be transformed in bioproducts. For processes applying NCY that can utilize a wide range of sugars, optimization of the release of all kinds of sugars is of utmost importance. Hence, YEAST4BIO-associated researchers aim to boost the profitability of the bioconversion processes by maximizing the sugar production with a minimum requirement of enzyme loading. Furthermore, newly identified and characterized enzymes and recombinant enzymes are currently tested on lignocellulose (Agrawal, Tsang and Chadha 2021), and new enzymatic cocktails enriched in accessory enzymes are being formulated (Singhania et al.

#### The Carboxylic Acids Platform

Besides being interesting bioproducts, carboxylic acids can also be used as renewable carbon sources for NCY-based biosynthesis (Llamas et al. 2020b). Given the great heterogeneity of food wastes, agricultural residues or even lignocellulosic feedstock, the anaerobic fermentation is considered a very technologically favorable process for obtaining an assimilable carbon source for the yeasts. Furthermore, the anaerobic fermentation process is almost completely independent of the composition of the substrate. When limiting the anaerobic digestion to the first three stages to avoid the production of biogas, the organic matter contained in the waste results in a mixture of VFAs (Magdalena and González-Fernández 2019). The VFAs can have different chain lengths (between C2 and C6), and their prevalence depends on the composition of the substrate and on the conditions of the anaerobic reactor (pH, temperature, organic load, hydraulic time of residence etc.) (Magdalena, Greses and González-Fernández 2019; Llamas et al. 2021). VFA production is nowadays considered as a novel technology for the development of future biotechnology industries based on these alternative carbon sources.

To date, research efforts addressed towards the implementation of a stable carboxylate platform are scarce compared to the numerous studies on the sugar platform. Thus, YEAST4BIO research activities aim to advance the understanding on how feedstock composition may affect carbon source output and VFA profiles for robust anaerobic fermentation technologies.

## **Bioproduct Formation using NCY**

Numerous NCY are more efficient than S. cerevisiae in converting carbon sources from alternative feedstock such as lignocellulosic wastes and organic residues into bioproducts. These microorganisms will surely play a key role in production of biobased chemicals from both sugar and carboxylate platforms in the near future.

## Bioproducts using NCY and the sugar platform

The most typical bioproduct associated with yeast is ethanol. In contrast to wild type S. cerevisiae that most readily ferments hexose sugars to ethanol, multiple NCY including Scheffersomyces

stipitis and Spathaspora passalidarum can also produce ethanol from xylose, the second to glucose most abundant carbon source in plant biomass (Veras, Parachin and Almeida 2017). Other NCY bioproducts from the sugar platform include lipids, 2phenylethanol, flavin coenzyme, weak acids, industrially relevant enzymes, bioplastics, etc. Some relatively well characterized yeasts that are used as production hosts include K. marxianus for production bioethanol, aroma compounds and biosurfactants (Karim, Gerliani and Aïder 2020), K. phaffi (Pichia pastoris) for recombinant protein and enzyme production (Heistinger, Gasser and Mattanovich 2020) and Y. lipolytica for production of biosurfactants, carotenoids and lipids (Ledesma-Amaro and Nicaud 2016). A number of less well-characterized yeasts are also emerging as potential cell factories, including, Z. bailii for organic acids and both Candida intermedia and Candida boidinii for sugar alcohols (Bedő et al. 2021; Kuanyshev et al. 2021). However, as most NCY still await identification and/or detailed characterization, the scientific community and biotechnological industries are most likely only beginning to realize the vast range of other bioproducts that NCY can generate through the use of the sugar platform.

With the goal of developing economically-viable production processes, large research efforts within the YEAST4BIO community are directed to push product titers, yield and productivity toward theoretic limits. To reach this goal, more knowledge on NCY metabolism, genetics and physiology is essential. There is also a need to carefully tailor the cultivation parameters, growth medium composition and yeast species and strain of choice for each specific raw material and targeted bioproduct. The unique nature of each biotechnology process adds to the complexity and difficulty of scale-up, which is also being addressed within YEAST4BIO.

#### Bioproducts using NCY and the carboxylic acids platform

Some NCY such as Y. lipolytica, Cutaneotrichosporon oleaginosus (previously known as Trichosporon oleaginosus, Cryptococcus curvatus, Apiotrichum curvatum or Candida curvata) R. toruloides, Lipomyces lipofer and Williopsis saturnus have demonstrated their ability to utilize VFAs as platform molecules for production of biochemicals (Llamas et al. 2020a). However, high concentrations of VFAs can exert inhibitory effects on yeast affecting final product titers and yields (Ma et al. 2019; Llamas, Tomás-Pejó and González-Fernández 2020). The maximum concentrations of the different VFAs that can be used without compromising process yields remain to be elucidated. Thus, exploiting the intrinsic tolerance to acid stress present in some NCY species is of utmost importance when using these carbon sources. Remarkably, the metabolic pathways involved in the conversion of VFAs to targeted products by NCY are still unknown or only partially elucidated, and many research efforts are directed to unravel the metabolism behind VFAs utilization in NCY (Park et al. 2021).

For an efficient implementation of the carboxylate platform in NCY-based industrial processes, it is crucial to determine the capacity of NCY to convert these acids into products of interest. In this sense, YEAST4BIO aims to shed light both on the NCY metabolic pathways involved in VFAs utilization and the process conditions favoring VFAs conversion into targeted products.

## NCY potential and application of systems/synthetic biology techniques

The implementation of novel system and synthetic biology tools for NCY is highly required to facilitate the utilization of these microorganisms in biotechnological applications. However, there are still some challenges that should be addressed to make the application of these new techniques for NCY a reality. The limited -omics (genomics, proteomics, transcriptomics and metabolomics) studies so far on NCY and the lack of knowledge on their wide variety of metabolic pathways and production capacity are among the main challenges to progress in the application of NCY. Systems biology approaches, using proteomic, transcriptomic, metabolomic, and fluxomic analyses can be potentially applied to NCY for the identification and characterization of DNA parts as well as for understanding the metabolic networks and regulation. Furthermore, -omic information such as genomics and transcriptomics can be used to create reliable genome-scale metabolic models, which can guide strain design strategies. These genome scale metabolic models can also integrate further -omic results such as proteomics, metabolomics and fluxomics to more reliably simulate metabolic fluxes in silico. Some NCY have curated genome-scale metabolic models that are available to be used. Examples of these are Y. lipolytica (Kerkhoven et al. 2016), K. phaffii (Cankorur-Cetinkaya, Dikicioglu and Oliver 2017), O. polymorpha (Liebal et al. 2021), K. lactis (Dias et al. 2014), K. marxianus (Marcišauskas, Ji and Nielsen 2019), R. toruloides (Dinh et al. 2019; Tiukova et al. 2019).

In addition, identified and characterized DNA parts such as promoters and terminators can be used to generate novel synthetic biology tools tailored for each NCY. Some of these tools being developed include Golden Gate libraries of parts and CRISPR technologies for genome engineering and transcriptional regulation. Developing novel NCY strains requires a multidisciplinary approach involving metabolic engineering for new pathways and enzymesu6 to expand the substrate usage by the microorganism; engineering microorganisms for better stress response to industrial conditions and for higher productivity, as well as biochemical engineering to apply these organisms under optimal process configurations. Different NCY have different degrees of development regarding systems and synthetic biology tools, being some of the most advanced Y. lipolytica (Larroude et al. 2018), K. marxianus (Rajkumar and Morrissey 2020), K. phaffii (Gao, Jiang and Lian 2021) and D. hansenii (Strucko et al. 2021). In any case, the developments of these tools and their efficiency are still behind those of S. cerevisiae, and there is still much that need to be done to domesticate these organisms. YEAST4BIO will explore these limitations and will bring together researchers working with different NCY to address current challenges by sharing learnings and brainstorming.

## YEAST4BIO THEMATIC ISSUE COVERING **DIFFERENT ASPECTS OF NCY IN BIOTECHNOLOGY**

Some of the challenges mentioned in the previous sections are covered in the different review articles included on this Thematic Issue. These review articles show both the breadth and the depth of research within the Action and gather different exper-

In this Thematic Issue, different systems and synthetic biology tools and their application for NCY are reviewed. Godinho and co-authors (2021) present N.C. Yeastract and CommunityYeastract databases to study gene and genomic transcription regulation in non-conventional yeasts. Binati and colleagues (2021) discuss the applications of NCY in food and food additive production. Ergün and collaborators (2021) describe the interplay between metabolic model and transcriptional control of K. phaffii and their use in the production of proteins. In addition, Ata and colleagues (2021) review the features of K. phaffii that makes it non-conventional and discuss its potential in industrial applications. In another work, Solieri and collaborators (2021) digest and discuss the current knowledge about life cycle and cell identity regulatory circuits with special focus on the genetic improvement of NCY. Park and co-workers (2021) offer their perspective on the bioproducts generated by NCY using carboxylic acids as substrates with Y. lipolytica while Karaalioğlu and Yuceer (2021) review how NCY can produce volatile compounds. Altogether, these collaborative works exemplify the importance of NCY in research and industry, identify current challenges and future perspectives and clearly imply one step forward towards the application of NCY in biotechnology.

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#### **REFERENCES**

- Agrawal D, Tsang A, Chadha BS. Economizing the lignocellulosic hydrolysis process using heterologously expressed auxiliary enzymes feruloyl esterase D (CE1) and  $\beta$ -xylosidase (GH43) derived from thermophilic fungi Scytalidium thermophilum. Bioresour Technol 2021;339.
- Ata O, Ergün BC, Fickers P et al. What makes komagataella phaffii non-conventional?. FEMS Yeast Res 2021. In press.
- Bedő S, Fehér A, Khunnonkwao P et al. Optimized bioconversion of xylose derived from pre-treated crop residues into xylitolby using Candida boidini i. Agronomy 2021;11:79.
- Bharathiraja B, Sridharan S, Sowmya V et al. Microbial oil a plausible alternate resource for food and fuel application. Bioresour Technol 2017;233:423-32
- Binati RL, Salvetti E, Bzducha-Wróbel A et al. Non-conventional yeasts for food and additives production in a circular economy perspective. FEMS Yeast Res 2021;21:foab052.
- Cankorur-Cetinkaya A, Dikicioglu D, Oliver SG. Metabolic modeling to identify engineering targets for Komagataella phaffii: the effect of biomass composition on gene target identification. Biotechnol Bioeng 2017;114:2605-15.
- Cannella D, Jørgensen H. Do new cellulolytic enzyme preparations affect the industrial strategies for high solids lignocellulosic ethanol production? Biotechnol Bioeng 2014;111: 59-68.
- Corchero JL, Gasser B, Resina D et al. Unconventional microbial systems for the cost-efficient production of high-quality protein therapeutics. Biotechnol Adv 2013;31:140-53.
- Deparis Q, Claes A, Foulquié-Moreno MR et al. Engineering tolerance to industrially relevant stress factors in yeast cell factories. FEMS Yeast Res 2017;17.
- Dias O, Pereira R, Gombert AK et al. iOD907, the first genomescale metabolic model for the milk yeast Kluyveromyces lactis. Biotechnol J 2014;9:776-90.
- Dinh H V., Suthers PF, Chan SHJ et al. A comprehensive genomescale model for Rhodosporidium toruloides IFO0880 accounting for functional genomics and phenotypic data. Metab Eng Commun 2019;9:e00101.

- Dujon B, Sherman D, Fischer G et al. Genome evolution in yeasts. Nature 2004: 430:35-44.
- Ergün BG, Berrios J, Binay B et al. Recombinant protein production in Pichia pastoris: from transcriptionally redesigned strains to bioprocess optimization and metabolic modelling. FEMS Yeast Res 2021; foab057.
- Gao J, Jiang L, Lian J. Development of synthetic biology tools to engineer Pichia pastoris as a chassis for the production of natural products. Synth Syst Biotechnol 2021;6:110-9.
- Godinho CP, Palma M, Oliveira J et al. The N.C. Yeastract and communityyeastract databases to study gene and genomic transcription regulation in non-conventional yeasts. FEMS Yeast Res 2021;21:foab045.
- Heistinger L, Gasser B, Mattanovich D. Microbe profile: komagataella phaffii: a methanol devouring biotech yeast formerly known as pichia pastoris. Microbiol (United Kingdom) 2020;166:614-6.
- Karaalioğlu O, Yuceer Y. Non-Conventional yeasts to produce aroma compounds by using agri-food waste materials. FEMS Yeast Res 2021 In Press.
- Karim A, Gerliani N, Aïder M. Kluyveromyces marxianus: an emerging yeast cell factory for applications in food and biotechnology. Int J Food Microbiol 2020;333:108818.
- Katahira S, Ito M, Takema H et al. Improvement of ethanol productivity during xylose and glucose co-fermentation by xylose-assimilating s. cerevisiae via expression of glucose transporter sut1. Enzyme Microb Technol 2008;43:115-9.
- Kavšček M, Stražar M, Curk T et al. Yeast as a cell factory: current state and perspectives. Microb Cell Fact 2015;14:94.
- Kerkhoven EJ, Pomraning KR, Baker SE et al. Regulation of amino-acid metabolism controls flux to lipid accumulation in yarrowia lipolytica. npj Syst Biol Appl 2016;2:1-7.
- Koutinas AA, Chatzifragkou A, Kopsahelis N et al. Design and techno-economic evaluation of microbial oil production as a renewable resource for biodiesel and oleochemical production. Fuel 2014;**116**:566–77.
- Kuanyshev N, Rao C V., Dien B et al. Domesticating a food spoilage yeast into an organic acid-tolerant metabolic engineering host: lactic acid production by engineered zygosaccharomyces bailii. Biotechnol Bioeng 2021;118:372-82.
- Lachance M-A. Yeast biodiversity: how many and how much?In: Biodiversity and Ecophysiology of Yeasts. Berlin: Springer-Verlag, 2006, 1-9.
- Larroude M, Rossignol T, Nicaud JM et al. Synthetic biology tools for engineering yarrowia lipolytica. Biotechnol Adv 2018;36:2150-64.
- Ledesma-Amaro R, Nicaud JM. Yarrowia lipolytica as a biotechnological chassis to produce usual and unusual fatty acids. Prog Lipid Res 2016;61:40-50.
- Liebal UW, Fabry BA, Ravikrishnan A et al. Genome-scale model reconstruction of the methylotrophic yeast ogataea polymorpha. BMC Biotechnol 2021;21:23.
- Llamas M, Dourou M, González-Fernández C et al. Screening of oleaginous yeasts for lipid production using volatile fatty acids as substrate. Biomass Bioenergy 2020a;138: 105553.
- Llamas M, Greses S, Tomás-Pejó E et al. Tuning microbial community in non-conventional two-stage anaerobic bioprocess for microalgae biomass valorization into targeted bioproducts. Bioresour Technol 2021;337:125387.
- Llamas M, Magdalena JA, González-Fernández C et al. Volatile fatty acids as novel building blocks for oil-based chemistry via oleaginous yeast fermentation. Biotechnol Bioeng 2020b;117:238-50.

- Llamas M, Tomás-Pejó E, González-Fernández C. VOLATILE FATTY ACIDS FROM ORGANIC WASTES AS NOVEL LOW-COST CARBON SOURCE FOR yarrowia lipolytica. N Biotechnol 2020;56:123-9.
- Ma X, Gao Z, Gao M et al. Microbial lipid production from food waste saccharified liquid under two-stage process. Bioresour Technol 2019;289: 121626.
- Magdalena JA, González-Fernández C. Microalgae biomass as a potential feedstock for the carboxylate platform. Molecules 2019:24:4404.
- Magdalena JA, Greses S, González-Fernández C. Impact of organic loading rate in volatile fatty acids production and population dynamics using microalgae biomass as substrate. Sci Rep 2019;9:1-11.
- Marcišauskas S, Ji B, Nielsen J. Reconstruction and analysis of a kluyveromyces marxianus genome-scale metabolic model. BMC Bioinformatics 2019;20:1-9.
- Mattanovich D, Sauer M, Gasser B. Yeast biotechnology: teaching the old dog new tricks. Microb Cell Fact 2014;13:1-5.
- Mukherjee V, Radecka D, Aerts G et al. Phenotypic landscape of non-conventional yeast species for different stress tolerance traits desirable in bioethanol fermentation. Biotechnol Biofuels 2017;10:216.
- Navarrete C, Frost AT, Ramos-Moreno L et al. A physiological characterization in controlled bioreactors reveals a novel survival strategy for debaryomyces hansenii at high salinity. Yeast 2021;38:302-15.
- Navarrete C, Martínez J L. Non-conventional yeasts as superior production platforms for sustainable fermentation based bio-manufacturing processes. AIMS Bioeng 2020;7: 289-305.
- Palma M, Münsterkötter M, Peça J et al. Genome sequence of the highly weak-acid-tolerant zygosaccharomyces bailii IST302, amenable to genetic manipulations and physiological studies. FEMS Yeast Res 2017;17: fox025.
- Park YK, González-Fernández C, Robles-Iglesias R et al. Bioproducts generation from carboxylate platforms by the non-conventional yeast yarrowia lipolytica. FEMS Yeast Res 2021;21:foab047.
- Radecka D, Mukherjee V, Mateo RQ et al. Looking beyond saccharomyces:the potential of non-conventional yeast species fordesirable traits in bioethanol fermentation. FEMS Yeast Res 2015;15:fov053.

- Rajkumar A S, Morrissey J P. Rational engineering of kluyveromyces marxianus to create a chassis for the production of aromatic products. Microb Cell Fact 2020;19:207.
- Raschmanová H, Weninger A, Glieder A et al. Implementing CRISPR-Cas technologies in conventional and nonconventional yeasts: current state and future prospects. Biotechnol Adv 2018;36:641-65.
- Ravn JL, Engqvist MKM, Larsbrink J et al. CAZyme prediction in ascomycetous yeast genomes guides discovery of novel xylanolytic species with diverse capacities for hemicellulose hydrolysis. Biotechnol Biofuels 2021;14:150.
- Rebello S, Abraham A, Madhavan A et al. Non-conventional yeast cell factories for sustainable bioprocesses. FEMS Microbiol Lett 2018;365:fny222.
- Shen XX, Opulente DA, Kominek J et al. Tempo and mode of genome evolution in the budding yeast subphylum. Cell 2018;175:1533-45.
- Singhania R R, Dixit P, Patel KumarA et al. Role and significance of lytic polysaccharide monooxygenases (LPMOs) in lignocellulose deconstruction. Bioresour Technol 2021;335:125261.
- Solieri L, Cassanelli S, Huff F et al. Insights on life cycle and cellidentity regulatory circuits for unlocking genetic improvement in zygosaccharomyces and kluyveromyces yeasts. FEM-SYeast Res 2021;foab058
- Strucko T, Andersen NL, Mahler MR et al. A CRISPR/Cas9 method facilitates efficient oligo-mediated gene editing in debaryomyces hansenii. Synth Biol 2021;6:ysab031
- Tiukova IA, Prigent S, Nielsen J et al. Genome-scale model of rhodotorula toruloides metabolism. Biotechnol Bioeng 2019;116:3396-408.
- Veras HCT, Parachin NS, Almeida JRM. Comparative assessment of fermentative capacity of different xylose-consuming yeasts. Microb Cell Fact 2017;16:1-8.
- Wagner JM, Alper HS. Synthetic biology and molecular genetics in non-conventional yeasts: current tools and future advances. Fungal Genet Biol 2016;89:126-36.
- Wang J, Ledesma-Amaro R, Wei Y et al. Metabolic engineering for increased lipid accumulation in yarrowia lipolytica – a review. Bioresour Technol 2020;313:123707.
- Wen Z, Zhang S, Odoh CK et al. Rhodosporidium toruloides A potential red yeast chassis for lipids and beyond. FEMS Yeast Res 2020;20:foaa038.
- Zhang Y, Nielsen J, Liu Z. Yeast based biorefineries for oleochemical production. Curr Opin Biotechnol 2021;67:26-34.