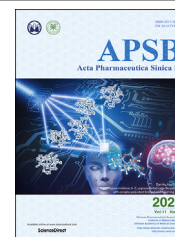




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REVIEW

Pharmacological insights into autophagy modulation in autoimmune diseases



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Autophagy inducer;
Autophagy inhibitor

Abstract As a cellular bulk degradation and survival mechanism, autophagy is implicated in diverse biological processes. Genome-wide association studies have revealed the link between autophagy gene polymorphisms and susceptibility of autoimmune diseases including systemic lupus erythematosus (SLE) and inflammatory bowel disease (IBD), indicating that autophagy dysregulation may be involved in the development of autoimmune diseases. A series of autophagy modulators have displayed protective effects on autoimmune disease models, highlighting the emerging role of autophagy modulators in treating autoimmune diseases. This review explores the roles of autophagy in the autoimmune diseases, with emphasis on four major autoimmune diseases [SLE, rheumatoid arthritis (RA), IBD, and experimental autoimmune encephalomyelitis (EAE)]. More importantly, the therapeutic potentials of small molecular autophagy modulators (including autophagy inducers and inhibitors) on autoimmune diseases are comprehensively analyzed.

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1. Introduction

Autophagy is a vital, conserved, and life-sustaining mechanism by which cytoplasmic cargo is delivered to lysosomes for degradation and by which cellular homeostasis is maintained¹. There are at least three types of autophagy (macroautophagy, microautophagy, and chaperone-mediated autophagy), but macroautophagy (hereafter referred to as autophagy) is the major autophagic degradation form governing organelle quality control and cellular homeostasis in eukaryotic cells. In the process of autophagy, cytosolic components, including toxic protein aggregates and superfluous or damaged organelles, are sequestered into isolated double membrane and delivered to lysosomes for degradation². Dysfunction of autophagy is involved in various kinds of diseases, including cancer, neurodegenerative diseases, infectious diseases, and metabolic diseases³.

Emerging evidence in recent years has revealed that autophagy intrinsically regulates the immune system function⁴. For example, autophagy regulates polarization of macrophages^{5,6}, regulates and is regulated by a wide range of inflammatory cytokines⁷, controls T cell function⁸, and modulates antigen presentation⁹. In the innate immune system, autophagy not only protects cells against invading microbial pathogens¹⁰, but also regulates stress-induced immune cell dysfunction, such as regulation of oxidative stress-induced T cell dysfunction¹¹, modulation of anoxic stress-induced tumor infiltrating lymphocytes activation¹², regulation of endoplasmic reticulum (ER) stress-induced Ig production^{12,13}, and modulation of LPS-triggered inflammatory responses¹⁴. Autophagy-related genes have been implicated in tissue-destructive inflammation and in the pathogenesis of several autoimmune and inflammatory disorders, including systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), inflammatory bowel disease (IBD), pancreatitis, and cancer^{15–18}. Interestingly, SLE, RA, and IBD all belong to autoimmune diseases, indicating that autophagy may play critical role in autoimmune regulation. In fact, a number of autophagy activators and inhibitors have been applied for modulation of autoimmune diseases, making pharmacological autophagy modulation a potentially potent therapeutic strategy for treating these diseases¹⁹.

However, there remain many unanswered questions. For example, whether autophagy regulates autoimmune response through a universal regulatory mechanism? Whether autophagy functions differently in different autoimmune diseases? Whether autophagy modulation is a promising approach for autoimmune diseases treatment? These questions are discussed in this review.

2. Molecular mechanism of autophagy

Autophagy proceeds in five main steps: (1) autophagy initiation, (2) phagophore formation, (3) phagophore expansion, (4) fusion with lysosome, and (5) degradation by lysosome (Fig. 1). Firstly, the complex including Unc-51 like autophagy activating kinase 1 (ULK1), FAK family kinase-interacting protein of 200 kDa (FIP200), autophagy-related protein 13 (ATG13), and autophagy-related protein 101 (ATG101), is the major player to regulate the autophagy initiation²⁰. ULK1 kinase can be activated by AMP-activated protein kinase (AMPK) under glucose starvation condition²¹ and by inhibition of mammalian target of rapamycin complex 1 (mTOR1) under amino acid deficiency condition to induce autophagy²². After autophagy induction, ATG9-containing vesicles are recruited by the ULK1 complex to the autophagosome to facilitate membrane delivery²³. Subsequently, the class III phosphatidylinositol 3-kinase (PI3KC3) complex [vacuolar protein sorting 34 (VPS34), vacuolar protein sorting 15 (VPS15), Beclin-1 (BECN1), autophagy-related protein 14 (ATG14), and nuclear receptor binding factor 2 (NRBF2)] is activated to generate phosphatidylinositol 3-phosphate [PI(3)P] on the phagophore. The PI(3)P effectors, such as FYVE-containing protein 1 (DFCP1) and WD-repeat protein that interact with PtdIns (WIPI) proteins, are recruited to phagophore to induce autophagosome formation²⁴. Then, two ubiquitin-like systems, the ATG12–ATG5–autophagy-related 16 like 1 (ATG16L1) complex and the microtubule-associated protein light chain 3 (LC3) complex, are recruited to drive autophagosome membrane elongation and vesicle expansion²⁵. Besides autophagosome membrane elongation, LC3-II is also involved in autophagosome closure. Finally, the autophagosomes fuses with the lysosome *via* soluble *N*-ethylmaleimide-sensitive-factor attachment protein receptor (SNARE) for degradation²⁶.

3. Overview of autoimmune diseases

Autoimmunity is defined as immune responses that attack an organism's own cells or tissues^{27,28}. In the development of autoimmune diseases, genetic predisposition or environmental stimulus break immune tolerance, resulting in autoantibodies and self-reactive lymphocytes formation which ultimately cause tissue damage²⁸. Autoimmunity-derived disease is one of the major health problems in the world, and autoimmunity has been viewed as an ever endless world²⁹. There are nearly 100 distinct autoimmune diseases based on different syndromes; typical examples

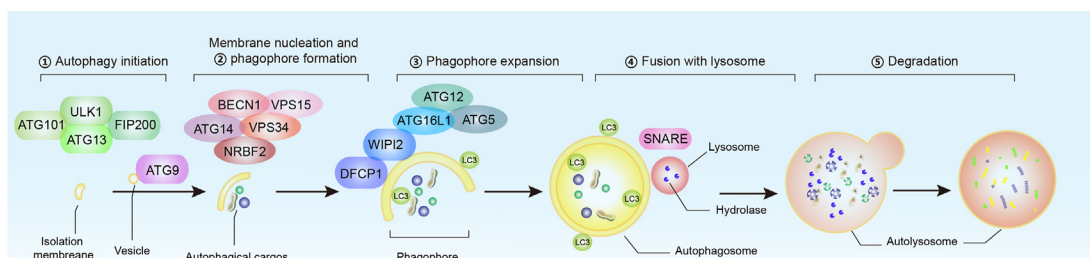


Figure 1 Process of autophagy. Autophagy is initiated and proceeds by diverse autophagy-related proteins to deliver different cargos to lysosomes for degradation.

of autoimmune diseases include multiple sclerosis (MS), type 1 diabetes (T1D), RA, and SLE.

Multiple genes have been reported to be associated with autoimmune diseases. Monogenic mutations of *AIRE*, *TNFRSF6*, *FOXP3*, and *CD25* have been found to induce autoimmune diseases. These genes are mostly associated with lymphocyte development or the functional regulation of lymphocytes, impairing T cell selection in the thymus and periphery tolerance^{30–33}. In fact, most autoimmune diseases involve multiple genetic factors. The major histocompatibility complex (MHC) is the strongest gene complex associated with human autoimmunity^{34,35}. For example, genetic variants *HLA-II-DQ2* and *HLA-II-DQ8* have been found in T1D and celiac disease (CeD), *HLA-II-DR3*, *HLA-II-DR2*, and *HLA-II-DR8* have been found in SLE, while *HLA-II-DR4* and *HLA-III-TNF* have been identified in RA³⁵. Other genes such as protein tyrosine phosphatase non-receptor type 22 (*PTPN22*), *IRF5-TNPO3*, and *BACH2* have all been found to be associated with autoimmunity²⁸. Environmental influence is a risk factor for the induction of autoimmune diseases. Nutrition, microbiota, invading pathogens, pharmacological agents, hormones, ultraviolet light, silica solvents, heavy metals, vaccines, and collagens implants are all potential stimuli that can elicit autoimmune responses^{28,36,37}. For example, Epstein–Barr virus (EBV) has been identified as a cofactor in numerous autoimmune diseases, such as SLE, RA, and MS^{38,39}. Excess intake of dietary iodine increases the incidence of autoimmune thyroid diseases (AITD)⁴⁰, and smoking is a well-recognized risk factor for RA and SLE^{41,42}.

As the precise etiology of autoimmune diseases is poorly understood, treatment is a challenge for clinicians. Currently, the basic therapeutic approach is to block the inflammatory pathway, such as by administration of agents to inhibit tumor necrosis factor (TNF- α), interleukin 6 (IL-6), or IL-12^{43–45}. Other immunotherapies, such as stem cell therapies, have been introduced⁴². However, none of these methods can cure the autoimmune diseases, and not all have been proven effective in humans. Modifying host immune system and identification of novel means of effective modulation are crucial for improving therapy of autoimmune diseases.

4. Autophagy in autoimmune diseases

Evidence from genome-wide association studies (GWAS) study and autophagy-defective animal models reveals autophagy dysregulation is associated with a number of autoimmune diseases, such as SLE, Crohn's disease (CD), RA, MS, and T1D¹⁹. Clarifying the roles of autophagy in these different autoimmune diseases can consolidate our understanding and perhaps suggest promising pathway for future research (Fig. 2).

4.1. SLE

SLE is an autoimmune disease characterized by production of large amounts of antibodies that act against self-antigens, especially double-stranded DNA (dsDNA), phospholipids, and small nuclear RNA-binding proteins. Various organs, such as joints, skin, kidneys, blood cells, brain, heart, and lungs can be affected^{46–48}. Polymorphisms of several autophagy-related genes have been associated with SLE. Five SLE-related single-nucleotide polymorphisms (SNPs) have been found near to or in the *ATG5* gene locus^{49,50}. In 2010, phosphatidylinositol 3-kinase catalytic subunit type 3 (*PIK3C3*) promoter variant

(rs3813065 C) was reported to be strongly associated with the presence of SLE in African–American patients⁵¹. In the following years, several autophagic proteins, including BECN1, ATG5, and ATG7 were observed to be required in LC3-associated phagocytosis (LAP) to facilitate apoptotic cell clearance which inhibits the development of autoimmunity⁵². Precise analysis revealed that deficiency of LAP-related proteins, including BECN1, ATG7, ATG3, ATG5, ATG12, ATG16L1, and Run domain BECN1-interacting and cysteine-rich domain-containing protein (RUBICON), accelerated the development of SLE-like disease condition in mice. However, mice lacking ULK1 and FIP200 that are not required for LAP in myeloid cells, did not show SLE-like disease¹⁸. Therefore, whether LAP is a critical factor to connect autophagy and SLE remains a topic worth intensive studying. In the clinical study, LAP levels were found to be elevated in blood and liver monocytes from patients with fibrosis and cirrhosis, two conditions often accompanying autoimmune diseases, because SLE-induced chronic liver damage may eventually develop into liver cirrhosis. It appears possible that LAP activation might protect against chronic inflammation-induced liver damage⁵³.

Mitophagy, a selective form of autophagy to degrade damaged mitochondria, is also an essential factor in SLE pathogenesis. Mitochondrial homeostasis is fundamental to maintaining immune system balance. In the innate immune system, diverse damage-associated molecular patterns (DAMPs), such as phospholipid cardiolipin, *N*-formyl peptides, ATP, and mitochondrial DNA (mtDNA) which released from damaged mitochondria, were threaten to activate the immune response⁵⁴ and induce autoimmune diseases^{55,56}. In addition, mitochondrial reactive oxygen species (ROS) is able to drive spontaneous neutrophil extracellular traps (NETs) formation, which is implicated in the SLE pathogenesis⁵⁷. Mitophagy serves as a protective mechanism to keep mitochondria homeostasis and is proposed to be involved in autoimmune diseases, such as SLE. In fact, mitophagy levels were found to be suppressed in T cells in SLE patients⁵⁸. In addition, in mitophagy-deficient mice lacking PTEN-induced kinase 1 (*PINK1*) or Parkin RBR E3 ubiquitin protein ligase (*PRKN*) after exhaustive exercise, released mtDNA was able to activate the stimulator of interferon genes protein (STING)-mediated DNA sensing pathway, resulting in higher levels of type I interferon (IFN), a potential enhancer of autoimmunity, compared to wild-type mice⁵⁹. Interestingly, deletion of immune-related GTPase family M protein (IRGM), a protein linking autoimmunity to autophagy, displayed defective mitophagy flux and resulted in overproduced DAMPs. These DAMPs stimulated DNA/RNA sensing-signaling to activate IFN expression and its responses⁶⁰. Thus, promoting mitophagy emerges as a possible way to inhibit autoimmune diseases, although the mechanism remains unclear⁶¹. Collectively, LAP and mitophagy are two vital cellular processes to limit auto-antigen production.

Innate inflammatory pathways that are regulated by autophagy may aggravate SLE. Retinoic acid-inducible gene 1 (RIG-1)-mitochondrial antiviral signaling protein (MAVS) RNA-sensing and cyclic GMP-AMP (cGAS)—STING DNA-sensing pathways are two innate immune responses to defend against pathogenic viral genomes or endogenous damaged DNA⁶². In the activated state, sensors stimulate multiple signaling cascades for production of type I IFNs and proinflammatory cytokines to eliminate threats. Notably, excessive activation of cGAS—STING pathway is implicated in auto-inflammatory and autoimmune diseases, such as Aicardi–Goutières syndrome and SLE^{63–65}. Accumulating evidence suggested that RIG-1 or the STING sensing pathway-induced

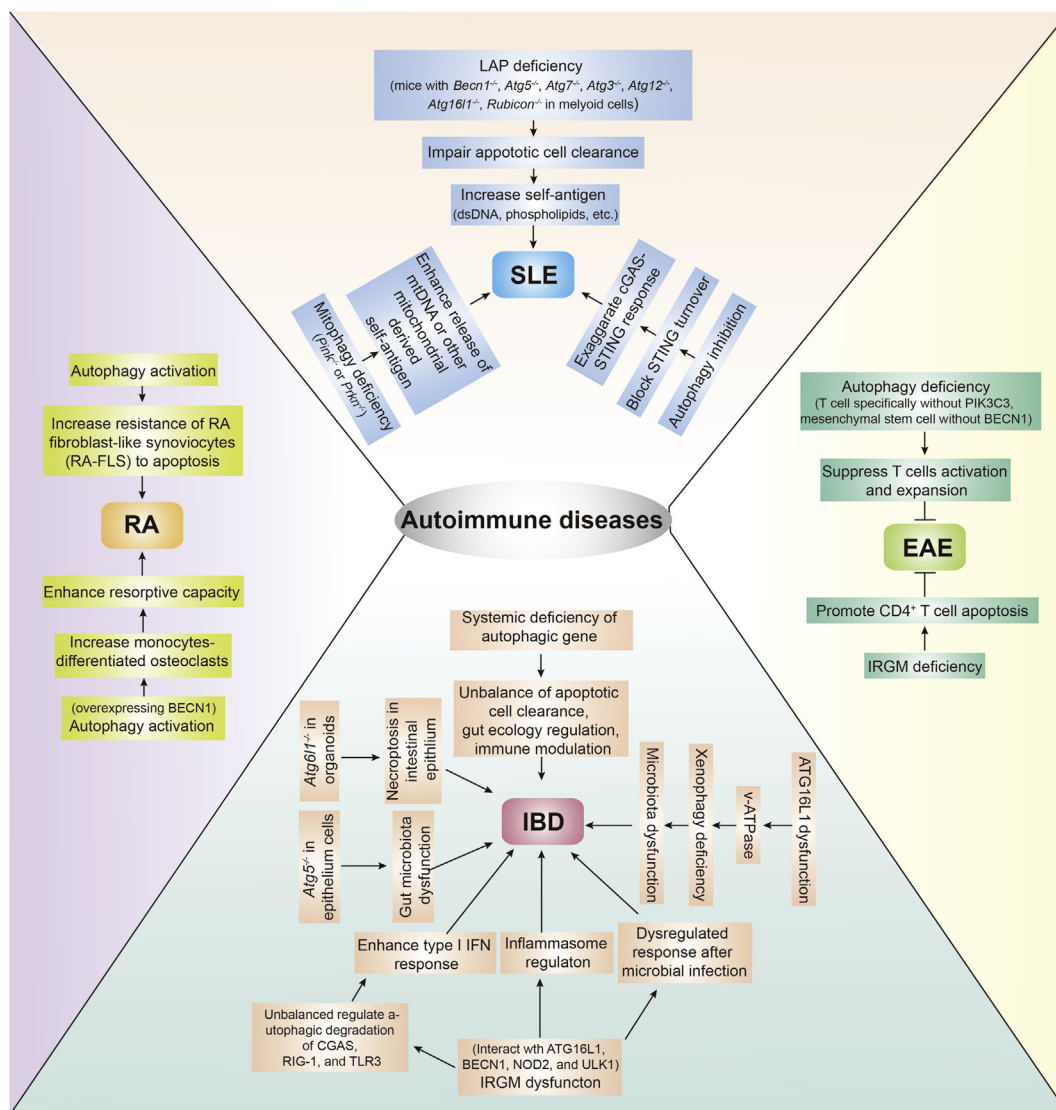


Figure 2 Autophagy regulation in autoimmune diseases. Autophagy displays differently in different autoimmune diseases. In SLE or IBD, autophagy enhancement contributes to ameliorating syndrome *via* reducing self-antigens production, cell-specific function regulation, and inflammatory pathway regulation. However, autophagy activation can aggravate RA disease syndrome by inhibiting RA-FLS apoptosis and increasing osteoclast differentiation. Inhibition of autophagy attenuates EAE by prohibiting T cell proliferation and activation.

sustained IFN response is detrimental to autoimmune diseases therapy^{66,67}. Interestingly, autophagy is able to regulate STING turnover *via* phosphorylating p62 by TANK-binding kinase 1 (TBK1) and attenuate cGAS–STING signaling⁶⁸. Evidence indicates that STING can be degraded *via* autophagy pathway. Considering STING is closely associated with ER⁶⁹, and it's a primordial function of cGAS–STING pathway to induce autophagy in a manner independent of TBK1 or IFN induction⁷⁰. It is highly possible that autophagy regulates autoimmune response through the STING pathway by promoting STING turnover.

4.2. IBD

IBD is a complicated idiopathic inflammatory disease that can be triggered by genetic or environmental stimuli. It is classified into CD and ulcerative colitis (UC) based on the region of the gastrointestinal (GI) tract that is inflamed. UC is confined to the inflammation occurring at the mucosal surface, while CD involves

transmural inflammation from mouth to anus⁷¹. It is widely accepted that the pathogenesis of IBD results from the loss of immune tolerance to intestinal antigens due to genetic or environmental factors. Several studies also defined IBD as auto-inflammatory syndrome of the gastrointestinal tract considering the limited observations of self-antigens⁷².

Since genetic polymorphisms in ATG16L1 have been identified as a strong risk for developing CD⁷³, roles of autophagy in the development of IBD have been extensively studied. Different autophagy-related genes such as *ULK1*⁷⁴, *ATG4B*⁷³, *PIK3C3*⁷⁵, and *NRBF2*⁷⁶, have been revealed to be implicated in IBD. Studies have determined that autophagy is essential for intestinal homeostasis maintenance, gut ecology regulation, immune modulation, and anti-microbial protection⁷⁷. However, the questions of what are the critical factors associating autophagy and IBD have not been answered.

It should be noted that in zebrafish, *pik3c3* mutants showed IBD-like features with high inflammatory response, and cell

junctions in intestinal epithelial cells were disrupted⁷⁵. However, gut microorganisms seem unnecessary for these IBD-like features⁷⁵, indicating that gut microbiome dysfunction may not be a critical pathogenic element. In our recent study, NRBF2, as a component of the PI3KC3 complex, has been revealed to be involved in the IBD regulation *via* modulating apoptotic cell clearance⁷⁶, initiating a discussion as to whether apoptotic cell-derived factors are vital targets for IBD therapy and whether PI3KC3-regulated apoptotic cell clearance is one of the critically affected factors involved in autoimmune diseases.

Autophagy-related inflammatory signaling regulation is another vital process implicated in IBD. Myeloid deficiency in *ULK1*, a LAP-unnecessary autophagic gene, is not necessary in SLE¹⁸, but is involved in IBD⁷⁸. These results suggest that ULK1 perhaps regulates a LAP-independent immune response, which contributes to the IBD pathogenesis. Interestingly, IRGM was able to interact and stabilize autophagic proteins, such as ATG16L1, BECN1, nucleotide-binding oligomerization domain-containing protein 2 (NOD2), and ULK1, to govern the autophagy-dependent microbial infection defense and regulate inflammatory response. Interestingly, all the genes that encode the above mentioned proteins were identified in IBD genetic polymorphism study^{73,75,78}. These findings suggested that autophagy can be involved in IBD *via* the IRGM-associated inflammatory pathway. IRGM is closely associated with auto-inflammatory or autoimmune diseases. IRGM deficiency has been found to be involved in ankylosing spondylitis⁷⁹, AITD⁸⁰, CD⁸¹, experimental autoimmune encephalomyelitis (EAE)⁸², and hepatic steatosis⁸³. Genetically *Irgm* knocked-out mice show hallmarks of Sjogren's syndrome, a kind of autoimmune diseases⁸⁴. Although the core mechanism by which IRGM acts in innate immune systems remains undetermined, IRGM has been involved in distinct inflammatory responses. For instance, IRGM is involved in the autophagic degradation of inflammasomes⁸⁵. Recently, a study has found that IRGM may mediate autoimmune diseases *via* regulating autophagic degradation of cGAS, RIG-1, and toll-like receptor 3 (TLR3), resulting in the regulation of type I IFN response⁶⁰. This is the beginning of uncovering the precise mechanism by which IRGM modulates the autoimmune system and autophagy.

Xenophagy and autophagy-regulated epithelium cell function are two factors assumed to be involved in IBD pathogenesis. Autophagy induced by invading pathogens is termed xenophagy⁸⁶. As microbiota dysfunction is a critical part of the pathogenesis of IBD, xenophagy is regarded as an essential connection between autophagy and IBD. Studies have found that ATG16L1 is linked with V-ATPase and critically mediates xenophagy⁸⁷, and ULK1 kinase also directly phosphorylates ATG16L1 to drive xenophagy⁸⁸. Therefore, xenophagy is a potential mechanism to regulate IBD pathogenesis. However, there is still very limited evidence that directly supports the roles of xenophagy in IBD. Intestinal epithelium function is another hotspot in recent studies of IBD pathogenesis. Epithelium not only acts as a physical barrier to pathogens, but also communicates with other immune cells for immune regulation⁸⁶. Previous studies have found that *Atg5* deficiency in epithelium cells alters the composition of the gut microbiota. Transcriptome analysis has showed that two IBD-associated factors, nuclear receptor ROR-gamma (RORC) and T-box transcription factor 21 (TBX21), were upregulated in mice with impaired autophagy in intestinal epithelial cells⁸⁹. In addition, *pik3c3* mutation in zebrafish displayed disrupted cell-junctions in intestinal epithelial cells⁷⁵. *Atg4b*^{-/-} mice presented abnormal Paneth cells⁹⁰ and intestinal organoids lacking

ATG16L1 elevated necroptosis in intestinal epithelium⁷⁷. The current evidence revealed that defects of xenophagy or autophagy in epithelial cells may contribute to the pathogenesis of IBD.

4.3. RA

RA is a chronic autoimmune disease characterized by synovial inflammation and bone loss. Dysfunction of immune cells and RA fibroblast-like synoviocytes (RA-FLS) have been identified as players in RA⁹¹. CD4⁺ T cells, B cells, macrophages, and RA-FLS infiltrate the synovium, resulting in destroyed articular structure and hyperplasia of the intimal lining⁹¹.

In the pathogenesis of RA, RA-FLS is a key player since it is a tissue-specific cell and is capable of producing local inflammatory cytokines and enzymes. Inhibition of aggressive RA-FLS formation or induction of RA-FLS apoptosis is one of the therapeutic approaches for treating RA. Interestingly, previous studies have elucidated that autophagy is able to regulate RA-FLS survival. Autophagy activation enhanced the resistance of RA-FLS to apoptosis *via* releasing ER stress and attenuating mitochondria dysfunction^{92,93}.

Another vital factor affecting development of RA is the differentiation of osteoclasts. Accumulation of osteoclast precursors and mature osteoclasts at inflammatory sites contributes to articular erosion and systemic osteoporosis⁹⁴. In the previous study, autophagy is also able to mediate osteoclast-mediated bone destruction in RA. Autophagy activation by overexpressing BECN1 increased monocyte-differentiated osteoclasts and enhanced resorptive capacity. In contrast, genetic and pharmacologic inhibition of autophagy reduced bone resorption and protected mice from TNF- α -induced bone erosion, proteoglycan loss and chondrocyte death¹⁷.

Unlike beneficial roles of autophagy in immune cell regulation in LAP and IBD, autophagy regulation in RA seems to be controversial. Autophagy activation drives RA development, which elicits discussions about the roles of autophagy in different cell types and auto-immune diseases.

4.4. EAE

EAE is a widely used mouse model for multiple sclerosis study. EAE is typically triggered by activated T cells migration into the central nervous system (CNS), and displays robust inflammatory response after immunization of susceptible mice with myelin-associated proteins⁹⁵. Although multiple innate immune cell types are involved during EAE, autophagy has been mostly studied in T cell differentiation and surviving in the EAE models. *Atg5*-deficient T lymphocytes dramatically increased dead CD8⁺ T cells in periphery. In addition, CD4⁺ and CD8⁺ T cells failed to undergo efficient proliferation while T cell receptor (TCR) activation after knocking out *Atg5*⁹⁶. These results suggested that autophagy is necessary in the adaptive immune response. One successful therapeutic strategy for EAE is silencing over-activated T cells by either inducing T cell apoptosis or blocking T cell activation, indicating autophagy inhibition is beneficial for EAE.

One study has revealed that *Pik3c3* deficiency contributes to EAE resistance in a mouse model⁹⁷. The study established *Pik3c3*-deficient T cells, and found that these cells failed to differentiate into T helper 1 cells, and the mice without *Pik3c3* in T cells cannot mount autoreactive T cell response in experimental autoimmune EAE. Further, they found that mice with *Pik3c3*-deficient T cells exhibited impaired metabolism and lower levels

of active mitochondria. Researchers have also found *Becn1*-deficiency and pharmacological inhibition of autophagy in mesenchymal stem cells (MSC) suppressed CD4⁺ T cells activation and expansion in EAE⁹⁸. Moreover, deficiency of *Irgm* was reported to suppress EAE by promoting apoptosis of activated CD4⁺ T cells⁸². These results indicate that inhibiting autophagy is beneficial for therapy of EAE, whose pathogenesis is guided by T cell dysfunction. There is less evidence confirming whether overreacted autophagy can exaggerate EAE in these studies; however, autophagy inducers, such as rapamycin and spermidine, have been proven to inhibit EAE development^{99,100}.

Taken together, in the innate immune system, autophagy may exert a general inhibitory effect of immune response to ensure the accurate control of immune responses, at least partially by regulating inflammasome and DNA sensing pathway *via* degrading the functional proteins. Thus, the autoimmune diseases with overactivated inflammatory response can possibly be regulated by autophagy induction. However, autophagy as a cytoprotective mechanism enhances cellular resistance to cell death, by which to prohibit the death of over reacted lymphocytes or RA-FLS in EAE and RA, may result in enhanced autoimmune responses. Moreover, autophagy functions quite differently in various cell types and results in distinct influences in autoimmune diseases. For examples, mitophagy, xenophagy, and LAP are selective autophagic ways to degrade potential antigens that may induce autoimmune response in IBD and SLE. While mitophagy and xenophagy are present in all the cell types, LAP is limited to phagocytes. Therefore, precise understanding of autoimmune disease pathogenesis is important for utilization of autophagy regulator in therapeutic strategy. Tissue- or cell-specific autophagy function may be a pivotal topic for discussion in the future work for autoimmune study.

5. Therapeutic potential of autophagy regulators in autoimmune diseases

Although novel therapeutic strategies such as the use of anti-TNF- α and cytokines inhibitors are being developed, application of immunosuppressors to suppress the systemic immune system is still the most common approach for autoimmune diseases treatment¹⁰¹. However, the adverse effects of immunosuppressors limit their long-time administration. There is an urgent need for safe drugs that can be used long-term. Exploring novel therapeutic strategies is an important avenue for developing these drugs. Autophagy dysregulation, as a common biological event in the development of autoimmune diseases, has triggered interests in testing small molecule autophagy regulators in the treatment of autoimmune diseases. A wide spectrum of small molecule autophagy regulators have been identified and tested in various disease models. Several autophagy inducers and inhibitors have already been applied for autoimmune disease therapies in multiple models (Fig. 3 and Table 1^{2,100,102–135}). Autophagy inducers, such as rapamycin, spermidine, and vitamin D, and autophagy inhibitors, such as chloroquine, have been used for the regulation of diverse autoimmune diseases. In addition, certain drugs used for autoimmune therapy have also been found to be associated with autophagy. For example, the anti-inflammatory drug, glucocorticoids, has been found to initiate autophagy¹³⁶.

5.1. Autophagy inducers

Among the positive autophagy regulators used for autoimmune modulation, two categories are noteworthy. One is mTOR

pathway inhibitors; the other one is histone deacetylase inhibitors.

mTOR is a master regulator of cell growth and proliferation¹³⁷. Genetic or pharmacologic inhibition of mTOR has been demonstrated for autophagy induction^{138,139}. Inhibition of mTORC1 can directly activate ULK1 activity to induce autophagy¹⁴⁰, and mTORC2 inhibition indirectly allows forkhead box O3a (FOXO3A) nuclear translocation to accelerate autophagic vesicles formation¹⁴¹. The mTOR pathway also plays regulatory roles in the immune system, especially in mTOR-modulated T cell development and differentiation¹⁴². Therefore, mTOR is a critical regulator both in autophagy and the immune system. In fact, rapamycin and its analog, as notable mTORC1 pathway inhibitors, are accepted as promising immunosuppressants because of their anti-proliferative effects on immune cells and their ability to induce Treg differentiation^{143–146}. In fact, these drugs have been utilized in different autoimmune diseases, such as allergic encephalomyelitis, adjuvant arthritis, and the humoral (IgE) immune response. Extensively, rapamycin administration blocks T cell activation in SLE patients and show obvious therapeutic efficacy^{147–149}. Rapamycin-mediated mTORC1 pathway inhibition regulates lineage specification in the T cell compartment, which has been regarded as target for autoimmune diseases treatment^{150,151}. As autophagy and immune system regulation can be regulated in parallel by mTOR, whether the rapamycin-induced immunosuppression effects are, to some extent, dependent on autophagy is still unclear. Assessing therapeutic effects of rapamycin on autoimmune disease models in autophagy deficiency condition may help to answer the question.

Histone deacetylase inhibitors (HDACIs) are repeatedly reported to activate autophagy in different cellular and animal models^{152–154}. At present, the molecular mechanism underlying HDACIs-induced autophagy is unclear, but several studies suggested it is potentially associated with the mTORC1 pathway and FOXO-dependent pathway^{152,155,156}. In addition, many HDACIs have been observed to modulate the autoimmune response and been applied in treating different autoimmune diseases. Trichostatin A (TSA) was suggested to ameliorate EAE and suppress collagen antibody-induced arthritis in animal models^{122,123}. Valproic acid treatment attenuated inflammation in experimental autoimmune neuritis (EAN) and EAE^{126,127}. Vorinostat treatment ameliorated EAE¹²⁴. These studies explain that the HDACIs were able to upregulate Treg cells, suppress dendritic functions and diminish lymphoproliferation. However, since molecular functions of histone modification are complicated, it's difficult to dissect how HDACIs regulate autoimmune diseases. Whether autophagy functions as a strong mediator in the connections between HDACIs and autoimmune diseases can be examined in advanced studies.

Other autophagy inducers, such as spermidine and vitamin D, have also been implicated in the regulations of autoimmune diseases. Spermidine is a kind of polyamines that exists in all mammalian cells. Administration of spermidine has been demonstrated to extend the lifespan in diverse animals and cells *via* inducing autophagy¹⁵⁷. In aging yeast, spermidine administration triggered deacetylation of histone H3, resulting in autophagy induction^{157,158}. In recent years, people have found that spermidine can affect autoimmune diseases^{100,159,160}. Spermidine administration alleviated severity of EAE *via* inducing inhibitory macrophages^{100,160}. Most recently, a novel study found that spermidine can also elicit metabolic fitness of dendritic cells (DC) and mediate the autoimmune response¹⁵⁹. Vitamin D (VD) is a precursor of a multifunctional hormone, the major types of which are

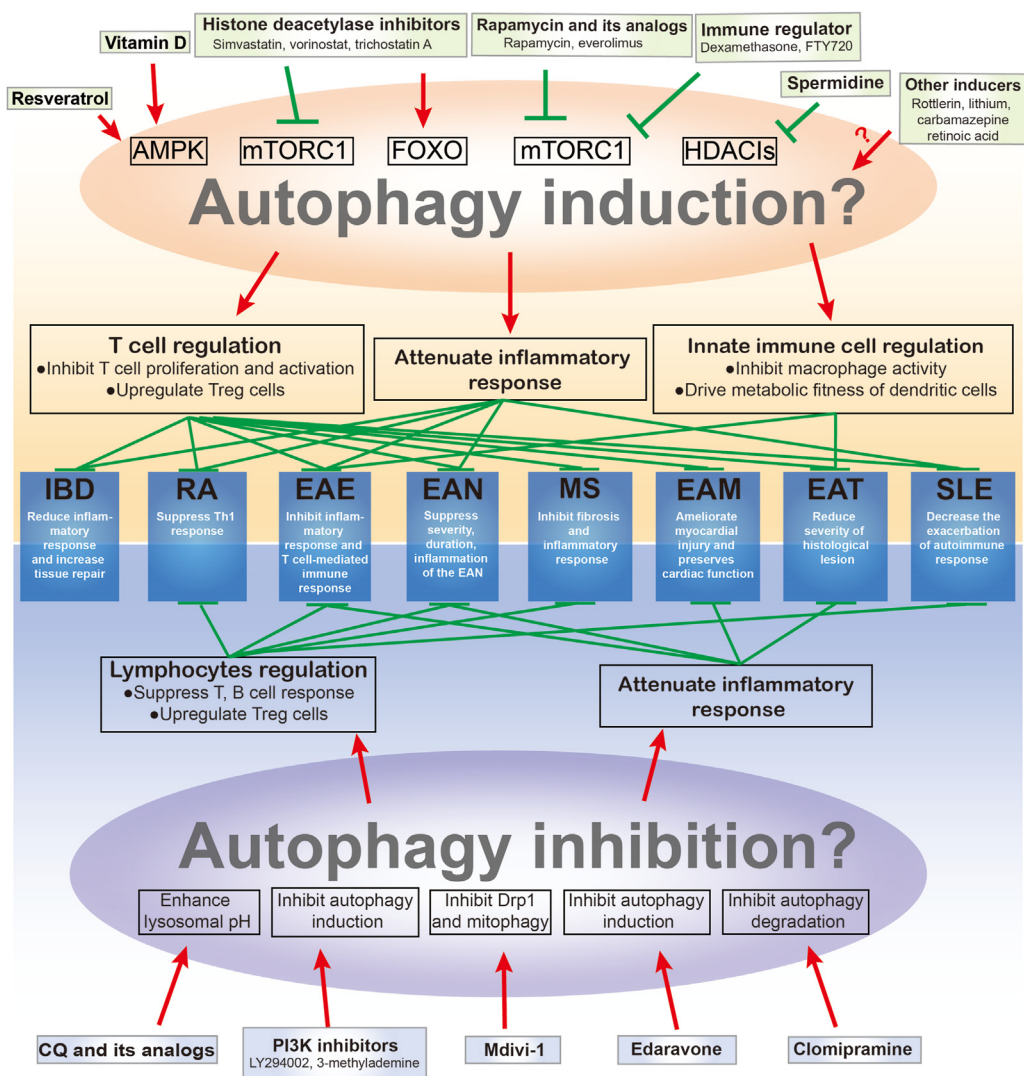


Figure 3 Potential autophagic regulators of autoimmune diseases. Typical autophagy regulators that have been utilized to treat autoimmune diseases in diverse animal models are shown.

ergocalciferol (vitamin D₂) and cholecalciferol (vitamin D₃)¹⁶¹. Once VD diffuses into cells, it binds with VD receptor (VDR) to dimerize with the retinoid X receptor and regulate multiple gene expressions¹⁶². Low VD status and VDR polymorphism have been regarded as environmental risk factors in autoimmune diseases¹⁶³. A deficiency of VD has been found in multiple sclerosis, SLE, RA, thyroiditis, and autoimmune gastritis^{164–167}. However, it's still not clear whether the low level of VD in patients with autoimmune diseases is the cause or the consequence. Some animal experiments found that VD administration reduced the incidence and severity of EAE symptoms, suggesting that VD or VDR potentially play protective roles in autoimmune diseases^{168,169}. Interestingly, accumulating evidence demonstrates that VD treatment can trigger autophagy in different cell types including cancer cells and monocytes *via* CAMKK- β - and AMPK-dependent cascades^{170–172}. Another study uncovered that VD improved autophagy in M2 macrophages¹⁷². It would be interesting to check whether VD enhances LAP, and in this way, regulates autoimmune diseases. Spermidine and VD are important molecules which display multiple biological functions in different cell types.

Whether autophagy regulation is an important mediator for their activity in anti-autoimmune diseases should be further investigated.

5.2. Autophagy inhibitors

Not only autophagy inducers, but also autophagy inhibitors have been revealed to exert protective effects on different autoimmune diseases. Blocking autophagy disturbs lymphocyte proliferation or activation, resulting in reduced autoimmune response in several autoimmune disease models.

Chloroquine (CQ), an anti-malarial drug and autophagy inhibitor that works by increasing lysosomal pH, has been proven to have anti-inflammatory effects and to act on RA, especially when used with other immunosuppressive drugs¹⁷³. Numerous studies have revealed that CQ played diverse roles in antigen presentation, inflammatory pathway and Treg cell proliferation^{129,174}. However, CQ is a strong lysosome activity inhibitor which also has autophagy-independent functions. To what extent the autophagy inhibition is involved in the anti-inflammatory effects of CQ need further investigated.

Table 1 Pharmacological studies of autophagy regulators on autoimmune diseases therapy.

Compound	Effect on autophagy	Animal model	Dose	Administration	Effect	Ref.
Rapamycin	Activator	Myelin oligodendrocyte glycoprotein (MOG)-induced EAE	1 mg/kg/day	ip or oral gavage for 15 days, and followed 3 days once for another 45 or 80 days	Inhibit EAE	102
		Murine autoimmune lymphoproliferative syndrome (ALPS)	5 mg/kg/day	Oral gavage for 5 days a week	Attenuate ALPS	103
		Experimental autoimmune myositis (EAM)	1 or 3 mg/kg/day	Oral gavage for about 10 days	Relieve symptoms of EAM	104
		Experimental autoimmune myocarditis	2 mg/kg/day	Oral gavage for about 17 days	Ameliorate myocardial injury and preserves cardiac function	105
		Experimental autoimmune uveoretinitis (EAU)	0.1 mg/kg/day	Intravenous infusion by a mini osmotic pump for about 14 days	Reduce number of cells in the immunization sites, lower lymphocyte proliferative response	106
Resveratrol	Activator	EAM	50 mg/kg/day	Intraperitoneal injection for 2 weeks	Ameliorate myocardial injury and preserves cardiac function	107
		EAE and Theiler's murine encephalomyelitis virus-induced demyelinating disease (TMEV-IDD)	0.04% in the chow (approximately 20 mg/kg/day)	Resveratrol containing food for about 2 months	Exacerbate demyelination and inflammation	108
		MOG-induced EAE	10, 25, and 50 mg/kg/day	ip injection for 20 days	Ameliorate the clinical severity of MS	109
Retinoic acid	Activator	Lupus nephritis in NZB/WF ₁ mice	0.5 mg each time	ip injection three times per week for 5–7 months	Alleviate autoimmune renal disorder and prolongs survivals	110
		EAE	75 mg/kg/day	Oral gavage from Days 6–11	Suppressive activity on T cell-mediated immune response	111
		EAU	0.2 mg/mouse/day	ip injection every other day for 21 days	Ameliorate severity of EAU and reduces the Th1/Th17 responses	112
Carbamazepine	Activator	EAE	N/A	Carbamazepine-supplemented chow for 28 days	Improve the clinical course of the disease	113
Everolimus/ RAD001	Activator	Immunization with hIPRBp161–180-induced autoimmune uveoretinitis	5 mg/kg/day	Oral gavage daily for 21 days	Reduce the histopathological uveitis score	114
		P0 peptide 180–199-induced autoimmune neuritis	1 mg/kg/day	Oral gavage daily for 16 consecutive days post-immunization	Protect mice from the symptoms of EAN	115
Lithium	Activator	EAE	0.2% lithium carbonate in pelleted food	Lithium-containing food and two injections of LiCl on the 1st and 2nd days	Markedly suppress the clinical symptoms of EAE	116
FTY720	Activator	Experimental autoimmune neuritis (EAN)	1 mg/kg FTY720 in 1 mL PBS	ip injection once daily for 30 days	Greatly reduce the severity and duration of EAN	117
		Experimental autoimmune myasthenia gravis (EAMG)	1 mg/kg	Oral administration (three days a week) for 5 weeks	Suppress the production of both anti-torpedo californica AChR antibody and anti-mouse AChR antibody	2
Spermidine	Activator	EAE	30 mmol/L in drinking water	Administration by drinking water for about one month	Alleviate the severities of EAE, particularly of optic neuritis	100

(continued on next page)

Table 1 (continued)

Compound	Effect on autophagy	Animal model	Dose	Administration	Effect	Ref.
Vitamin D	Activator	EAE	0.1 µg	Every other day for 15 days	The association of MOG with VD was able to control EAE development	118
		Experimental autoimmune thyroiditis (EAT)	0.1 or 0.2 µg/kg/day	ip injection for 21 days	Administration alone did not affect the incidence of thyroiditis and reduced by up to 26% the severity of histological lesion	119
Dexamethasone	Activator	MRL-lpr female mice that developed an aggressive autoimmune nephritis	0.4 mg/kg per day	Administered by drinking water	Nephritis was ameliorated without alteration of TNF-α and ICAM-1 gene transcription	120
Rottlerin	Activator	Silica-exacerbated systemic autoimmune disease in New Zealand mixed mice	10 µg/institution once a week	Instillation for 14 weeks	Decrease the exacerbation of autoimmunity by silica exposure	121
Trichostatin A	Activator	Collagen-induced rheumatoid arthritis (CIA)	2 mg/kg/day	ip for 7 days	Suppress Th1 response and exert protective effects on CIA	122
		EAE	7.5 mg/kg/dose/day	ip for 40 days	Histone deacetylase (HDAC) inhibition by trichostatin A ameliorate EAE	123
Vorinostat	Activator	EAE	100 mg/kg/day	Oral gavage for 1 month	Suppress DCs and DCs-mediated Th1 and Th17 cell functions in EAE	124
Simvastatin	Activator	<i>Fas^{tgld/gld} apoE^{-/-}</i> mice that lack functional Fas ligand and apolipoprotein E and exhibit accelerated atherosclerosis and aggravated lupus-like features	0.125 mg/kg/day	ip for 12 weeks	Atherosclerosis degree and inflammatory response were reduced.	125
Valproic acid	Activator	EAE	250 or 500 mg/kg/day	Oral gavage daily from Days 7–18 or from Days 9–19	Suppress systemic and local inflammation to improve EAE	126
		EAN	300 mg/kg/day	Days 10–21	Effectively suppress inflammation in EAN	127
Niclosamide	Activator or Inhibitor	HOCl-induced systemic sclerosis	10 mg/kg every other day	ip injection for 6 weeks	Reverse fibrosis and inhibit immune response	128
Chloroquine	Inhibitor	EAE	3, 5, and 10 mg/kg/day	ip injection for 5 days	Suppress inflammation and increase Treg cells	129
LY294002	Inhibitor	EAM	40 µmol/L, 10 µL volume per day	ip injection for about 1 week	Inhibit cardiac injury	130
Mdivi-1	Inhibitor	EAE	25 mg/kg/day	ip injection for 27 days	Modulate the balance between Th1/Th17 and regulatory T cells	131
Edaravone	Inhibitor	EAM	1–10 mg/kg/day	ip for 3 weeks	Protect against acute EAM by scavenging hydroxyl free radicals	132
		EAE	6 mg/kg/day	From Days 5–19	iNOS was reduced	133
3-Methyladenine (3-MA)	Inhibitor	EAE	24 mg/kg/day	ip on Days 5, 10, 15, and 20	Attenuate IL-17-induced aggravated EAE	134
Clomipramine	Inhibitor	EAN	20 mg/kg/day	ip for 28 days	Suppress clinical signs of EAN	135

Phosphoinositide 3-kinase (PI3K) inhibitors, such as 3-methyladenine (3-MA), wortmannin, and LY294002, are widely used autophagy inhibitors based on their inhibitory effects on autophagy induction. Both 3-MA and LY294002 have been utilized in different autoimmune animal models, in which, the autoimmune diseases syndrome was ameliorated. 3-MA administration ameliorated the neurologic severity of EAN¹⁷⁵ and inhibited IL-17-induced aggravated myocarditis severity¹³⁴. LY294002 significantly alleviated experimental autoimmune myositis (EAM) injury in mice¹³⁰. As PI3K activation is essential for lymphocyte proliferation, the reduction of activated lymphocytes may also be attributed to the effects of 3-MA or LY294002 on autoimmune diseases.

Many other autophagy inhibitors with unsettled understanding in autophagy are used in autoimmune diseases. Mitochondrial division inhibitor 1 (Mdivi-1) is a selective dynamic-related protein 1 (Drp1) inhibitor. It has been found to act as a mitophagy inhibitor¹⁷⁶. In 2019, Li et al.¹³¹ reported that Mdivi-1 reduced EAE severity. Th1 and Th17 cells were reduced, and regulatory T cells were promoted after Mdivi-1 administration. Edaravone is an oxygen-free radical scavenger and showed effects of inhibiting autophagy after oxygen-glucose deprivation or recovery injury¹⁷⁷. Accumulating evidence from different animal models shows that edaravone is a promising compound to be utilized for autoimmune therapy. Edaravone significantly ameliorated EAE¹³³, acute autoimmune myocarditis¹³², experimental autoimmune thyroiditis¹⁷⁸, and fibrosis in skin models of systemic sclerosis¹⁷⁹. Clomipramine is an anti-depressant, which also inhibits autophagic degradation^{180,181}. In 1998, clomipramine was found to suppress clinical signs of EAN and inhibit adaptive immune responses¹³⁵. In addition, clomipramine has been systematically screened out for progressive sclerosis treatment as it has displayed the ability to inhibit T cell proliferation and B lymphocytes activity¹⁸². Again, though these compounds displayed both anti-autoimmune effects and autophagy inhibition activity, more work still need to confirm that autophagy inhibition plays a clear role in their pharmacological effects on autoimmune diseases.

6. Conclusions and discussion

Autoimmune disease is a category covering a wide range of diseases, all associated with an abnormally activated self-immune response, and involving diverse immune cells. In innate immune disorders, dysfunction of autophagy has been repeatedly reported. For example, phagocytosis deficiency, xenophagy, or mitophagy dysfunction limits the clearance of apoptotic cells, pathogens, and damaged mitochondria, inducing over-productions of self-antigens. In addition, over-active innate immune responses (dysregulated DNA and RNA sense pathways) and dysregulated proliferation or activation in lymphocytes can also aggravate self-immune responses. In this complicated autoimmune scenario, autophagy seems to play quite complicated roles in regulating different autoimmune diseases.

A large number of studies have found that autophagy or autophagy-related proteins have protective roles in autoimmune diseases. Many autophagy-related proteins have been found to regulate LAP and enhance apoptotic cell clearance. As such, LAP becomes a link to connect autophagy and autoimmune diseases, especially SLE. Innate immune signaling, including inflammatory and DNA or RNA sensing pathways, are essential targets to be regulated by autophagy. The autoimmune diseases-related gene

IRGM has recently been found to regulate autophagic degradation of cGAS, RIG-1, and TLR3, and by this way to limit IFN production. However, it is not known whether IRGM is a key mediator connecting autophagy function and immune responses; research into this question and into the exact regulating mechanism involved is ongoing. Mitophagy, as a selective form of autophagy, is another key event to be considered in autoimmune diseases. Many DAMPs can be released from damaged mitochondria, and mitophagy is an essential way to inhibit the immune response by removing damage mitochondria, but the specific modulation mechanism is still largely unknown. Knowing more about the mitophagy and identifying specific modulation factors would help to establish the link between mitophagy modulation and autoimmune diseases therapy.

However, we found that autophagy was not always beneficial in all autoimmune conditions. For example, it was reported that, in mice, inhibition of autophagy promotes resistance of mice to EAE, inhibits T cell differentiation, and reduces T cell responses. In RA model, apoptosis of over-activated RA-FLS is restricted. Therefore, autophagy appears to play different roles in different autoimmune diseases, possibly depending on cell type. We also noticed that unlike SLE¹⁸ and IBD⁷⁵, EAE and RA symptoms do not spontaneously appeared in autophagy-deficient conditions, suggesting a fact that EAE and RA may be less sensitive to autophagy inhibition, compared with SLE and IBD.

Pharmacological regulators of autophagy have been used for autoimmune disease therapy. It triggers concerns because autophagy regulators have displayed different roles in pharmacological studies. Both autophagy activators and inhibitors can be beneficial in some autoimmune disease models. Although more and more autophagy regulators have been identified, most of them do not specifically target autophagy. For example, rapamycin, spermidine, VD, and CQ are all compounds that regulate autophagy and autoimmune diseases, but there is limited evidence showing that the effects on autoimmune diseases are based on autophagy regulation; other cellular pathways could be involved. Lack of specific autophagy regulators makes it difficult to evaluate the effects of autophagy regulation in treating autoimmune diseases. Though rapamycin, 3-MA, and CQ are largely used as autophagy inducers or inhibitors in research articles, autophagy is not the solely affected pathway. Identification of highly specific autophagy regulators would be of great importance for future therapeutic application.

The complex pathogenesis of autoimmune diseases has been depicted as a mosaic of genetic predisposition, hormonal effects, and environmental factors. Autophagy appears to play various roles in different autoimmune diseases *via* distinct regulation pathway. Taken together, knowing more about autophagy regulation in the autoimmune system and identifying specific pharmacological autophagy regulators or pathways of regulation are vital for applying autophagy regulators in autoimmune diseases therapy.

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Author contributions

Jia-Hong Lu initiated this review and designed the fame. Ming-Yue Wu wrote the manuscript. Er-Jin Wang, Du Feng, Min Li, and Richard Ye helped revise the manuscript.

Conflicts of interest

The authors declare no conflicts of interest.

References

- Ravikumar B, Sarkar S, Davies JE, Futter M, Garcia-Arencibia M, Green-Thompson ZW, et al. Regulation of mammalian autophagy in physiology and pathophysiology. *Physiol Rev* 2010;**90**:1383–435.
- Kohno T, Tsuji T, Hirayama K, Iwatsuki R, Hirose M, Watabe K, et al. A novel immunomodulator, FTY720, prevents development of experimental autoimmune myasthenia gravis in C57BL/6 mice. *Biol Pharm Bull* 2005;**28**:736–9.
- Jiang P, Mizushima N. Autophagy and human diseases. *Cell Res* 2014;**24**:69–79.
- Cui J. *Autophagy regulation of innate immunity*, vol. 1209. Singapore: Springer Nature Singapore Pte Ltd.; 2019.
- Chang CP, Su YC, Lee PH, Lei HY. Targeting NFKB by autophagy to polarize hepatoma-associated macrophage differentiation. *Autophagy* 2013;**9**:619–21.
- Chang CP, Su YC, Hu CW, Lei HY. TLR2-dependent selective autophagy regulates NF- κ B lysosomal degradation in hepatoma-derived M2 macrophage differentiation. *Cell Death Differ* 2013;**20**:515–23.
- Ge Y, Huang M, Yao YM. Autophagy and proinflammatory cytokines: interactions and clinical implications. *Cytokine Growth Factor Rev* 2018;**43**:38–46.
- Botbol Y, Guerrero-Ros I, Macian F. Key roles of autophagy in regulating T-cell function. *Eur J Immunol* 2016;**46**:1326–34.
- Crotzer VL, Blum JS. Autophagy and its role in MHC-mediated antigen presentation. *J Immunol* 2009;**182**:3335–41.
- Watson RO, Manzanillo PS, Cox JS. Extracellular *M. tuberculosis* DNA targets bacteria for autophagy by activating the host DNA-sensing pathway. *Cell* 2012;**150**:803–15.
- He MX, McLeod IX, Jia W, He YW. Macroautophagy in T lymphocyte development and function. *Front Immunol* 2012;**3**:22.
- Ko A, Kanehisa A, Martins I, Senovilla L, Chargari C, Dugue D, et al. Autophagy inhibition radiosensitizes *in vitro*, yet reduces radioresponses *in vivo* due to deficient immunogenic signalling. *Cell Death Differ* 2014;**21**:92–9.
- Bhattacharya A, Eissa NT. Autophagy as a stress response pathway in the immune system. *Int Rev Immunol* 2015;**34**:382–402.
- Wang J, Wu MY, Su H, Lu J, Chen X, Tan J, et al. iNOS interacts with autophagy receptor p62 and is degraded by autophagy in macrophages. *Cells* 2019;**8**:1255.
- Gukovsky I, Li N, Todoric J, Gukovskaya A, Karin M. Inflammation, autophagy, and obesity: common features in the pathogenesis of pancreatitis and pancreatic cancer. *Gastroenterology* 2013;**144**:1199–209.
- Keller CW, Sina C, Kotur MB, Ramelli G, Mundt S, Quast I, et al. ATG-dependent phagocytosis in dendritic cells drives myelin-specific CD4⁺ T cell pathogenicity during CNS inflammation. *Proc Natl Acad Sci U S A* 2017;**114**:E11228–37.
- Lin NY, Beyer C, Giefl A, Kireva T, Scholtysek C, Uderhardt S, et al. Autophagy regulates TNF α -mediated joint destruction in experimental arthritis. *Ann Rheum Dis* 2013;**72**:761–8.
- Martinez J, Cunha LD, Park S, Yang M, Lu Q, Orchard R, et al. Noncanonical autophagy inhibits the autoinflammatory, lupus-like response to dying cells. *Nature* 2016;**533**:115–9.
- Bonam SR, Wang F, Muller S. Autophagy: a new concept in autoimmunity regulation and a novel therapeutic option. *J Autoimmun* 2018;**94**:16–32.
- Zachari M, Ganley IG. The mammalian ULK1 complex and autophagy initiation. *Essays Biochem* 2017;**61**:585–96.
- Egan DF, Shackelford DB, Mihaylova MM, Gelino S, Kohnz RA, Mair W, et al. Phosphorylation of ULK1 (hATG1) by AMP-activated protein kinase connects energy sensing to mitophagy. *Science* 2011;**331**:456–61.
- Lamb CA, Yoshimori T, Tooze SA. The autophagosome: origins unknown, biogenesis complex. *Nat Rev Mol Cell Biol* 2013;**14**:759–74.
- Noda T. Autophagy in the context of the cellular membrane-trafficking system: the enigma of Atg9 vesicles. *Biochem Soc Trans* 2017;**45**:1323–31.
- Kishi-Itakura C, Koyama-Honda I, Itakura E, Mizushima N. Ultrastructural analysis of autophagosome organization using mammalian autophagy-deficient cells. *J Cell Sci* 2014;**127**:4984.
- Itakura E, Mizushima N. Characterization of autophagosome formation site by a hierarchical analysis of mammalian Atg proteins. *Autophagy* 2010;**6**:764–76.
- Nakatogawa H, Ichimura Y, Ohsumi Y. Atg8, a ubiquitin-like protein required for autophagosome formation, mediates membrane tethering and hemifusion. *Cell* 2007;**130**:165–78.
- Stetson DB. Editorial overview: autoimmunity: a new frontier awaits. *Curr Opin Immunol* 2018;**55**:iii–iv.
- Wang L, Wang FS, Gershwin ME. Human autoimmune diseases: a comprehensive update. *J Intern Med* 2015;**278**:369–95.
- Toubi E. Editorial: autoimmunity—the ever endless world. *Immunol Res* 2018;**66**:633–6.
- Nagamine K, Peterson P, Scott HS, Kudoh J, Minoshima S, Heino M, et al. Postional cloning of the *APECED* gene. *Nat Genet* 1997;**17**:393–8.
- Sakaguchi S. Naturally arising Foxp3-expressing CD25⁺ CD4⁺ regulatory T cells in immunological tolerance to self and non-self. *Nat Immunol* 2005;**6**:345–52.
- Gambineri E, Torgerson TR, Ochs HD. Immune dysregulation, polyendocrinopathy, enteropathy, and X-linked inheritance (IPEX), a syndrome of systemic autoimmunity caused by mutations of FOXP3, a critical regulator of T-cell homeostasis. *Curr Opin Rheumatol* 2003;**15**:430–5.
- Shah S, Wu E, Rao VK, Tarrant TK. Autoimmune lymphoproliferative syndrome: an update and review of the literature. *Curr Allergy Asthma Rep* 2014;**14**:10–4.
- Cui Y, Sheng Y, Zhang X. Genetic susceptibility to SLE: recent progress from GWAS. *J Autoimmun* 2013;**41**:25–33.
- Sollid LM, Pos W, Wucherpfennig KW. Molecular mechanisms for contribution of MHC molecules to autoimmune diseases. *Curr Opin Immunol* 2014;**31**:24–30.
- Colafrancesco S, Perricone C, Priori R, Valesini G, Shoenfeld Y. Sjögren's syndrome: another facet of the autoimmune/inflammatory syndrome induced by adjuvants (ASIA). *J Autoimmun* 2014;**51**:10–6.
- Peng J, Narasimhan S, Marchesi JR, Benson A, Wong FS, Wen L. Long term effect of gut microbiota transfer on diabetes development. *J Autoimmun* 2014;**53**:85–94.
- Draborg AH, Duus K, Houen G. Epstein-barr virus in systemic autoimmune diseases. *Clin Dev Immunol* 2013;**2013**:535738.
- Zivadnov R, Zorzon M, Weinstock-Guttman B, Serafin M, Bosco A, Bratina A, et al. Epstein-Barr virus is associated with grey matter atrophy in multiple sclerosis. *J Neurol Neurosurg Psychiatry* 2009;**80**:620–5.
- Rose NR, Bonita R, Burek CL. Iodine: an environmental trigger of thyroiditis. *Autoimmun Rev* 2002;**1**:97–103.
- Eklblom-Kullberg S, Kautiainen H, Alha P, Leirisalo-Repo M, Miettinen A, Julkunen H. Smoking, disease activity, permanent damage and dsDNA autoantibody production in patients with systemic lupus erythematosus. *Rheumatol Int* 2014;**34**:341–5.
- Afridi HI, Kazi TG, Talpur FN, Naher S, Brabazon D. Relationship between toxic metals exposure via cigarette smoking and rheumatoid arthritis. *Clin Lab* 2014;**60**:1735–45.

43. Zhao M, Liu S, Luo S, Wu H, Tang M, Cheng W, et al. DNA methylation and mRNA and microRNA expression of SLE CD4⁺ T cells correlate with disease phenotype. *J Autoimmun* 2014;**54**:127–36.
44. Song SNJ, Yoshizaki K. Tocilizumab for treating rheumatoid arthritis: an evaluation of pharmacokinetics/pharmacodynamics and clinical efficacy. *Expert Opin Drug Metabol Toxicol* 2015;**11**:307–16.
45. Meng Y, Dongmei L, Yanbin P, Jinju F, Meile T, Binzhu L, et al. Systematic review and meta-analysis of ustekinumab for moderate to severe psoriasis. *Clin Exp Dermatol* 2014;**39**:696–707.
46. Harley JB, Kelly JA, Kaufman KM. Unraveling the genetics of systemic lupus erythematosus. *Springer Semin Immunopathol* 2006;**28**:119–30.
47. Goulielmos GN, Zervou MI, Vazgiourakis VM, Ghodke-Puranik Y, Garyfallos A, Niewold TB. The genetics and molecular pathogenesis of systemic lupus erythematosus (SLE) in populations of different ancestry. *Gene* 2018;**668**:59–72.
48. Muñoz LE, Lauber K, Schiller M, Manfredi AA, Herrmann M. The role of defective clearance of apoptotic cells in systemic autoimmunity. *Nat Rev Rheumatol* 2010;**6**:280–9.
49. Zhou X, Lu X, Lv J, Yang H, Qin L, Zhao M, et al. Genetic association of PRDM1–ATG5 intergenic region and autophagy with systemic lupus erythematosus in a Chinese population. *Ann Rheum Dis* 2011;**70**:1330–7.
50. International Consortium for Systemic Lupus Erythematosus Genetics (SLEGEN), Harley JB, Alarcón-riquelme ME, Criswell LA, Jacob CO, Kimberly RP, et al. Genome-wide association scan in women with systemic lupus erythematosus identifies susceptibility variants in ITGAM, PXX1, KIAA1542 and other loci. *Nat Genet* 2008;**40**:204–10.
51. Kariuki SN, Franek BS, Mikolaitis RA, et al. Promoter variant of PIK3C3 is associated with autoimmunity against Ro and Sm epitopes in African-American lupus patients. *J Biomed Biotechnol* 2010;**2010**.
52. Martínez J, Almendinger J, Oberst A, Ness R, Dillon CP, Fitzgerald P, et al. Microtubule-associated protein 1 light chain 3 alpha (LC3)-associated phagocytosis is required for the efficient clearance of dead cells. *Proc Natl Acad Sci U S A* 2011;**108**:17396–401.
53. Wan JH, Weiss E, Mkaddem S Ben, Mabire M, Choinier PM, Picq O, et al. LC3-associated phagocytosis protects against inflammation and liver fibrosis via immunoreceptor inhibitory signaling. *Sci Transl Med* 2020;**12**:eaaw8523.
54. Banoth B, Cassel SL. Mitochondria in innate immune signaling. *Transl Res* 2018;**202**:52–68.
55. Hajizadeh S, DeGroot J, TeKoppele JM, Tarkowski A, Collins LV. Extracellular mitochondrial DNA and oxidatively damaged DNA in synovial fluid of patients with rheumatoid arthritis. *Arthritis Res Ther* 2003;**5**:234–40.
56. Caielli S, Athale S, Domic B, Murat E, Chandra M, Banchereau R, et al. Oxidized mitochondrial nucleoids released by neutrophils drive type I interferon production in human lupus. *J Exp Med* 2016;**213**:697–713.
57. Fazeli G, Stetter M, Lisack JN, Wehman AM. *C. elegans* blastomeres clear the corpse of the second polar body by LC3-associated phagocytosis. *Cell Rep* 2018;**23**:2070–82.
58. Caza TN, Fernandez DR, Talaber G, Oaks Z, Haas M, Madaio MP, et al. HRES-1/Rab4-mediated depletion of Drp1 impairs mitochondrial homeostasis and represents a target for treatment in SLE. *Ann Rheum Dis* 2014;**73**:1888–97.
59. Sliter DA, Martínez J, Hao L, Chen X, Sun N, Fischer TD, et al. Parkin and PINK1 mitigate STING-induced inflammation. *Nature* 2018;**561**:258–62.
60. Jena KK, Mehto S, Nath P, Chauhan NR, Sahu R, Dhar K, et al. Autoimmunity gene IRGM suppresses cGAS-STING and RIG-I-MAVS signaling to control interferon response. *EMBO Rep* 2020;**21**:e50051.
61. Xu Y, Shen J, Ran Z. Emerging views of mitophagy in immunity and autoimmune diseases. *Autophagy* 2020;**16**:3–17.
62. Zevini A, Olganier D, Hiscott J. Crosstalk between cytoplasmic RIG-I and STING sensing pathways. *Trends Immunol* 2017;**38**:194–205.
63. Crow YJ, Manel N. Aicardi-Goutières syndrome and the type I interferonopathies. *Nat Rev Immunol* 2015;**15**:429–40.
64. Gall A, Treuting P, Elkon KB, Loo YM, Gale M, Barber GN, et al. Autoimmunity initiates in nonhematopoietic cells and progresses via lymphocytes in an interferon-dependent autoimmune disease. *Immunity* 2012;**36**:120–31.
65. Gao D, Li T, Li XD, Chen X, Li QZ, Wight-Carter M, et al. Activation of cyclic GMP-AMP synthase by self-DNA causes autoimmune diseases. *Proc Natl Acad Sci U S A* 2015;**112**:E5699–705.
66. Black CM, Silman AJ, Herrick AI, Denton CP, Wilson H, Newman J, et al. Interferon- σ does not improve outcome at one year in patients with diffuse cutaneous scleroderma: results of a randomized, double-blind, placebo-controlled trial. *Arthritis Rheum* 1999;**42**:299–305.
67. Ronnblom LE, Alm GV, Oberg KE. Autoimmunity after alpha-interferon therapy for malignant carcinoid tumors. *Ann Intern Med* 1991;**115**:178–83.
68. Prabakaran T, Bodda C, Krapp C, Zhang B, Christensen MH, Sun C, et al. Attenuation of cGAS-STING signaling is mediated by a p62/SQSTM1-dependent autophagy pathway activated by TBK1. *EMBO J* 2018;**37**:e97858.
69. Wang Q, Liu X, Cui Y, Tang Y, Chen W, Li S, et al. The E3 Ubiquitin ligase AMFR and INSIG1 bridge the activation of TBK1 kinase by modifying the adaptor STING. *Immunity* 2014;**41**:919–33.
70. Gui X, Yang H, Li T, Tan X, Shi P, Li M, et al. Autophagy induction via STING trafficking is a primordial function of the cGAS pathway. *Nature* 2019;**567**:262–6.
71. Kaistha A, Levine J. Inflammatory bowel disease: the classic gastrointestinal autoimmune disease. *Curr Probl Pediatr Adolesc Health Care* 2014;**44**:328–34.
72. Yang Z, Goronzy JJ, Weyand CM. Autophagy in autoimmune disease. *J Mol Med (Berl)* 2015;**93**:707–17.
73. Liu JZ, Van Sommeren S, Huang H, Ng SC, Alberts R. Europe PMC Funders Group Association analyses identify 38 susceptibility loci for inflammatory bowel disease and highlight shared genetic risk across populations. *Nat Genet* 2015;**47**:979–86.
74. Singh SB, Ornatowski W, Vergne I, Naylor J, Delgado M, Roberts E, et al. Human IRGM regulates autophagy and cell-autonomous immunity functions through mitochondria. *Nat Cell Biol* 2010;**12**:1154–65.
75. Zhao S, Xia J, Wu X, Zhang L, Wang P, Wang H, et al. Deficiency in class III PI3-kinase confers postnatal lethality with IBD-like features in zebrafish. *Nat Commun* 2018;**9**:2639.
76. Wu MY, Liu L, Wang EJ, Xiao HT, Cai CZ, Wang J, et al. PI3KC3 complex subunit NRBF2 is required for apoptotic cell clearance to restrict intestinal inflammation. *Autophagy* 2021;**17**:1096–111.
77. Larabi A, Barnich N, Nguyen HTT. New insights into the interplay between autophagy, gut microbiota and inflammatory responses in IBD. *Autophagy* 2020;**16**:38–51.
78. Henckaerts L, Cleynen I, Brinar M, John JM, Van Steen K, Rutgeerts P, et al. Genetic variation in the autophagy gene ULK1 and risk of Crohn's disease. *Inflamm Bowel Dis* 2011;**17**:1392–7.
79. Xia Q, Wang M, Yang X, Li X, Zhang X, Xu S, et al. Autophagy-related IRGM genes confer susceptibility to ankylosing spondylitis in a Chinese female population: a case-control study. *Gene Immun* 2017;**18**:42–7.
80. Yao QM, Zhu YF, Wang W, Song ZY, Shao XQ, Li L, et al. Polymorphisms in autophagy-related gene IRGM are associated with susceptibility to autoimmune thyroid diseases. *BioMed Res Int* 2018;**2018**:7959707.
81. Parkes M, Barrett JC, Prescott NJ, Tremelling M, Anderson CA, Fisher SA, et al. Sequence variants in the autophagy gene IRGM and multiple other replicating loci contribute to Crohn's disease susceptibility. *Nat Genet* 2007;**39**:830–2.

82. Xu H, Wu Z, Fang F, Guo L, Chen D, Chen JX, et al. Genetic deficiency of Irgm1 (LRG-47) suppresses induction of experimental autoimmune encephalomyelitis by promoting apoptosis of activated CD4⁺ T cells. *FASEB J* 2010;**24**:1583–92.
83. Bellini G, Miraglia del Giudice E, Nobili V, Rossi F. The IRGM rs10065172 variant increases the risk for steatosis but not for liver damage progression in Italian obese children. *J Hepatol* 2017;**67**:653–5.
84. Azzam KM, Madenspacher JH, Cain DW, Lai L, Gowdy KM, Rai P, et al. Irgm1 coordinately regulates autoimmunity and host defense at select mucosal surfaces. *JCI Insight* 2017;**2**:e91914.
85. Mehto S, Jena KK, Nath P, Chauhan S, Kolapalli SP, Das SK, et al. The Crohn's disease risk factor IRGM limits NLRP3 inflammasome activation by impeding its assembly and by mediating its selective autophagy. *Mol Cell* 2019;**73**:429–45.e7.
86. Bauckman KA, Owusu-Boaitey N, Mysorekar IU. Selective autophagy: xenophagy. *Methods* 2015;**75**:120–7.
87. Xu Y, Zhou P, Cheng S, Lu Q, Nowak K, Hopp AK, et al. A bacterial effector reveals the V-ATPase–ATG16L1 axis that initiates xenophagy. *Cell* 2019;**178**:552–66.e20.
88. Alsaadi RM, Losier TT, Tian W, Jackson A, Guo Z, Rubinsztein DC, et al. ULK1-mediated phosphorylation of ATG16L1 promotes xenophagy, but destabilizes the ATG16L1 Crohn's mutant. *EMBO Rep* 2019;**20**:e46885.
89. Yang L, Liu C, Zhao W, He C, Ding J, Dai R, et al. Impaired autophagy in intestinal epithelial cells alters gut microbiota and host immune responses. *Appl Environ Microbiol* 2018;**84**: e00880-18.
90. Cabrera S, Fernández ÁF, Mariño G, Aguirre A, Suárez MF, Español Y, et al. ATG4B/autophagin-1 regulates intestinal homeostasis and protects mice from experimental colitis. *Autophagy* 2013;**9**:1188–200.
91. Firestein GS. Evolving concepts of rheumatoid arthritis. *Nature* 2003;**423**:356–61.
92. Kim EK, Kwon JE, Lee SY, Lee EJ, Kim DS, Moon SJ, et al. IL-17-mediated mitochondrial dysfunction impairs apoptosis in rheumatoid arthritis synovial fibroblasts through activation of autophagy. *Cell Death Dis* 2017;**8**:e2565.
93. Kato M, Ospelt C, Gay RE, Gay S, Klein K. Dual role of autophagy in stress-induced cell death in rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum* 2014;**66**:40–8.
94. Gravallese EM, Harada Y, Wang JT, Gorn AH, Thornhill TS, Goldring SR. Identification of cell types responsible for bone resorption in rheumatoid arthritis and juvenile rheumatoid arthritis. *Am J Pathol* 1998;**152**:943–51.
95. Bernard CC. Experimental autoimmune encephalomyelitis in mice: genetic control of susceptibility. *J Immunogenet* 1976;**3**:263–74.
96. Pua HH, Dzhalgalov I, Chuck M, Mizushima N, He YW. A critical role for the autophagy gene *Atg5* in T cell survival and proliferation. *J Exp Med* 2007;**204**:25–31.
97. Yang G, Song W, Postoak JL, Chen J, Martinez J, Zhang J, et al. Autophagy-related protein PIK3C3/VPS34 controls T cell metabolism and function: PIK3C3/VPS34 in T cell metabolism and function. *Autophagy* 2021;**17**:1193–204.
98. Dang S, Xu H, Xu C, Cai W, Li Q, Cheng Y, et al. Autophagy regulates the therapeutic potential of mesenchymal stem cells in experimental autoimmune encephalomyelitis. *Autophagy* 2014;**10**:1301–15.
99. Hou H, Miao J, Cao R, Han M, Sun Y, Liu X, et al. Rapamycin ameliorates experimental autoimmune encephalomyelitis by suppressing the mTOR–STAT3 pathway. *Neurochem Res* 2017;**42**:2831–40.
100. Guo X, Harada C, Namekata K, Kimura A, Mitamura Y, Yoshida H, et al. Spermidine alleviates severity of murine experimental autoimmune encephalomyelitis. *Investig Ophthalmol Vis Sci* 2011;**52**:2696–703.
101. Rosman Z, Shoenfeld Y, Zandman-Goddard G. Biologic therapy for autoimmune diseases: an update. *BMC Med* 2013;**11**:88.
102. Yang Q, Wang R, Zhu L. *Autophagy: biology and diseases*, vol. 1206. Singapore: Springer Nature Singapore Pte Ltd.; 2019.
103. Esposito M, Ruffini F, Bellone M, Gagliani N, Battaglia M, Martino G, et al. Rapamycin inhibits relapsing experimental autoimmune encephalomyelitis by both effector and regulatory T cells modulation. *J Neuroimmunol* 2010;**220**:52–63.
104. Prevel N, Allenbach Y, Klatzmann D, Salomon B, Benveniste O. Beneficial role of rapamycin in experimental autoimmune myositis. *PLoS One* 2013;**8**:e74450.
105. Maeda K, Shioi T, Kosugi R, Yoshida Y, Takahashi K, Machida Y, et al. Rapamycin ameliorates experimental autoimmune myocarditis. *Int Heart J* 2005;**46**:513–30.
106. Roberge FG, Xu D, Chan CC, de Smet MD, Nussenblatt RB, Chen H. Treatment of autoimmune uveoretinitis in the rat with rapamycin, an inhibitor of lymphocyte growth factor signal transduction. *Curr Eye Res* 1993;**12**:197–203.
107. Yoshida Y, Shioi T, Izumi T. Resveratrol ameliorates experimental autoimmune myocarditis. *Circ J* 2007;**71**:397–404.
108. Sato F, Martinez NE, Shahid M, Rose JW, Carlson NG, Tsunoda I. Resveratrol exacerbates both autoimmune and viral models of multiple sclerosis. *Am J Pathol* 2013;**183**:1390–6.
109. Wang D, Li SP, Fu JS, Zhang S, Bai L, Guo L. Resveratrol defends blood–brain barrier integrity in experimental autoimmune encephalomyelitis mice. *J Neurophysiol* 2016;**116**:2173–9.
110. Kinoshita K, Yoo B-S, Nozaki Y, Sugiyama M, Ikoma S, Ohno M, et al. Retinoic acid reduces autoimmune renal injury and increases survival in NZB/W F₁ mice. *J Immunol* 2003;**170**:5793–8.
111. Massacesi L, Castigli E, Vergelli M, Olivetto J, Abbamondi AL, Sarlo F, et al. Immunosuppressive activity of 13-*cis*-retinoic acid and prevention of experimental autoimmune encephalomyelitis in rats. *J Clin Invest* 1991;**88**:1331–7.
112. Keino H, Watanabe T, Sato Y, Okada AA. Anti-inflammatory effect of retinoic acid on experimental autoimmune uveoretinitis. *Br J Ophthalmol* 2010;**94**:802–7.
113. Black JA, Liu S, Carrithers M, Carrithers LM, Waxman SG. Exacerbation of experimental autoimmune encephalomyelitis after withdrawal of phenytoin and carbamazepine. *Ann Neurol* 2007;**62**:21–33.
114. Hennig M, Bauer D, Wasmuth S, Busch M, Walscheid K, Thanos S, et al. Everolimus improves experimental autoimmune uveoretinitis. *Exp Eye Res* 2012;**105**:43–52.
115. Han R, Gao J, Zhai H, Xiao J, Ding Y, Hao J. RAD001 (everolimus) attenuates experimental autoimmune neuritis by inhibiting the mTOR pathway, elevating Akt activity and polarizing M2 macrophages. *Exp Neurol* 2016;**280**:106–14.
116. De Sarno P, Axtell RC, Raman C, Roth KA, Alessi DR, Jope RS. Lithium prevents and ameliorates experimental autoimmune encephalomyelitis. *J Immunol* 2008;**181**:338–45.
117. Zhang Z, Zhang ZY, Fauser U, Schluesener HJ. FTY720 ameliorates experimental autoimmune neuritis by inhibition of lymphocyte and monocyte infiltration into peripheral nerves. *Exp Neurol* 2008;**210**:681–90.
118. Chiuso-Minicucci F, Ishikawa LL, Mimura LA, Fraga-Silva TF, França TG, Zorzella-Pezavento SF, et al. Treatment with vitamin D/MOG association suppresses experimental autoimmune encephalomyelitis. *PLoS One* 2015;**10**:e0125836.
119. Fournier C, Gepner P, Sadouk M, Charreire J. *In vivo* beneficial effects of cyclosporin A and 1,25-dihydroxyvitamin D3 on the induction of experimental autoimmune thyroiditis. *Clin Immunol Immunopathol* 1990;**54**:53–63.
120. Jevnikar AM, Singer GG, Brennan DC, Xu HW, Kelley VR. Dexamethasone prevents autoimmune nephritis and reduces renal expression of Ia but not costimulatory signals. *Am J Pathol* 1992;**141**:743–51.
121. Brown JM, Schwanke CM, Pershouse MA, Pfau JC, Holian A. Effects of rottlerin on silica-exacerbated systemic autoimmune disease in New Zealand mixed mice. *Am J Physiol Lung Cell Mol Physiol* 2005;**289**:L990–8.
122. Zhou X, Hua X, Ding X, Bian Y, Wang X. Trichostatin differentially regulates Th1 and Th2 responses and alleviates rheumatoid arthritis in mice. *J Clin Immunol* 2011;**31**:395–405.

123. Camelo S, Iglesias AH, Hwang D, Due B, Ryu H, Smith K, et al. Transcriptional therapy with the histone deacetylase inhibitor trichostatin A ameliorates experimental autoimmune encephalomyelitis. *J Neuroimmunol* 2005;**164**:10–21.
124. Ge Z, Da Y, Xue Z, Zhang K, Zhuang H, Peng M, et al. Vorinostat, a histone deacetylase inhibitor, suppresses dendritic cell function and ameliorates experimental autoimmune encephalomyelitis. *Exp Neurol* 2013;**241**:56–66.
125. Aprahamian T, Bonegio R, Rizzo J, Perlman H, Lefer DJ, Rifkin IR, et al. Simvastatin treatment ameliorates autoimmune diseases associated with accelerated atherosclerosis in a murine lupus model. *J Immunol* 2006;**177**:3028–34.
126. Zhang Z, Zhang ZY, Wu Y, Schluesener HJ. Valproic acid ameliorates inflammation in experimental autoimmune encephalomyelitis rats. *Neuroscience* 2012;**221**:140–50.
127. Zhang Z, Zhang ZY, Fauser U, Schluesener HJ. Valproic acid attenuates inflammation in experimental autoimmune neuritis. *Cell Mol Life Sci* 2008;**65**:4055–65.
128. Morin F, Kavian N, Nicco C, Cerles O, Chéreau C, Batteux F. Niclosamide prevents systemic sclerosis in a reactive oxygen species-induced mouse model. *J Immunol* 2016;**197**:3018–28.
129. Thomé R, Moraes AS, Bombeiro AL, Farias A, dos S, Francelin C, da Costa TA, et al. Chloroquine treatment enhances regulatory T cells and reduces the severity of experimental autoimmune encephalomyelitis. *PLoS One* 2013;**8**:e65913.
130. Liu HS, Zhang J, Guo JL, Lin CY, Wang ZW. Phosphoinositide 3-kinase inhibitor LY294002 ameliorates the severity of myosin-induced myocarditis in mice. *Curr Res Transl Med* 2016;**64**:21–7.
131. Li YH, Xu F, Thome R, Guo MF, Sun ML, Song G Bin, et al. Mdivi-1, a mitochondrial fission inhibitor, modulates T helper cells and suppresses the development of experimental autoimmune encephalomyelitis. *J Neuroinflammation* 2019;**16**:149.
132. Nimata M, Okabe TA, Hattori M, Yuan Z, Shioji K, Kishimoto C. MCI-186 (edaravone), a novel free radical scavenger, protects against acute autoimmune myocarditis in rats. *Am J Physiol Heart Circ Physiol* 2005;**289**:2514–8.
133. Moriya M, Nakatsuji Y, Miyamoto K, Okuno T, Kinoshita M, Kumanogoh A, et al. Edaravone, a free radical scavenger, ameliorates experimental autoimmune encephalomyelitis. *Neurosci Lett* 2008;**440**:323–6.
134. Yuan J, Yu M, Li HH, Long Q, Liang W, Wen S, et al. Autophagy contributes to IL-17-induced plasma cell differentiation in experimental autoimmune myocarditis. *Int Immunopharm* 2014;**18**:98–105.
135. Zhu J, Bengtsson BO, Mix E, Ekerling L, Thorell LH, Olsson T, et al. Clomipramine and imipramine suppress clinical signs and T and B cell response to myelin proteins in experimental autoimmune neuritis in Lewis rats. *J Autoimmun* 1998;**11**:319–27.
136. Grandér D, Kharaziha P. Autophagy as the main means of cytotoxicity by glucocorticoids in hematological malignancies. *Autophagy* 2009;**5**:1198–200.
137. Kim YC, Guan KL. mTOR: a pharmacologic target for autophagy regulation. *J Clin Invest* 2015;**125**:25–32.
138. Vargas JNS, Wang C, Bunker E, Hao L, Maric D, Schiavo G, et al. Spatiotemporal control of ULK1 activation by NDP52 and TBK1 during selective autophagy. *Mol Cell* 2019;**74**:347–62.e6.
139. Scott RC, Schuldiner O, Neufeld TP. Role and regulation of starvation-induced autophagy in the *Drosophila* fat body. *Dev Cell* 2004;**7**:167–78.
140. Mokas S, Mills JR, Garreau C, Fournier MJ, Robert F, Arya P, et al. Uncoupling stress granule assembly and translation initiation inhibition. *Mol Biol Cell* 2009;**20**:2673–83.
141. Mammucari C, Milan G, Romanello V, Masiero E, Rudolf R, Del Piccolo P, et al. FoxO3 controls autophagy in skeletal muscle *in vivo*. *Cell Metabol* 2007;**6**:458–71.
142. Delgoffe GM, Kole TP, Zheng Y, Zarek PE, Matthews KL, Xiao B, et al. The mTOR kinase differentially regulates effector and regulatory T cell lineage commitment. *Immunity* 2009;**30**:832–44.
143. Faivre S, Kroemer G, Raymond E. Current development of mTOR inhibitors as anticancer agents. *Nat Rev Drug Discov* 2006;**5**:671–88.
144. Martee RR, Klicius J, Galet S. Inhibition of the immune response by rapamycin, a new antifungal antibiotic. *Can J Physiol Pharmacol* 1977;**55**:48–51.
145. Gao W, Lu Y, El Essawy B, Oukka M, Kuchroo VK, Strom TB. Contrasting effects of cyclosporine and rapamycin in *de novo* generation of alloantigen-specific regulatory T cells. *Am J Transplant* 2007;**23**:1722–32.
146. Yang K, Shrestha S, Zeng H, Karmaus PWF, Neale G, Guertin DA, et al. T cell exit from quiescence and differentiation into Th2 cells depend on Raptor-mTORC1-mediated metabolic programming. *Immunity* 2014;**39**:1043–56.
147. Fernandez DR, Telarico T, Bonilla E, Li Q, Banerjee S, Middleton FA, et al. Activation of mTOR controls the loss of TCR in lupus T cells through HRES-1/Rab4-regulated lysosomal degradation. *J Immunol* 2009;**182**:2063–73.
148. Lai ZW, Borsuk R, Shadakshari A, Yu J, Dawood M, Garcia R, et al. Mechanistic target of rapamycin activation triggers IL-4 production and necrotic death of double-negative T cells in patients with systemic lupus erythematosus. *J Immunol* 2013;**191**:2236–46.
149. Fermande D, Bonilla E, Mirza N, Niland B, Perl A. Rapamycin reduces disease activity and normalizes T cell activation-induced calcium fluxing in patients with systemic lupus erythematosus. *Arthritis Rheum* 2006;**54**:2983–8.
150. Delgoffe GM, Pollizzi KN, Waickman AT, Heikamp E, David J, Horton MR, et al. The mammalian target of rapamycin (mTOR) regulates T helper cell differentiation through the selective activation of mTORC1 and mTORC2 signaling. *Nat Immunol* 2011;**12**:295–303.
151. Piranavan P, Bhamra M, Perl A. Metabolic Targets for treatment of autoimmune diseases. *Immunometabolism* 2020;**2**:e200012.
152. Gammoh N, Lam D, Puente C, Ganley I, Marks PA, Jiang X. Role of autophagy in histone deacetylase inhibitor-induced apoptotic and nonapoptotic cell death. *Proc Natl Acad Sci U S A* 2012;**109**:6561–5.
153. Shao A, Wang Z, Wu H, Dong X, Li Y, Tu S, et al. Enhancement of autophagy by histone deacetylase inhibitor trichostatin A ameliorates neuronal apoptosis after subarachnoid hemorrhage in rats. *Mol Neurobiol* 2016;**53**:18–27.
154. Mrakovcic M, Kleinheinz J, Fröhlich LF. Histone deacetylase inhibitor-induced autophagy in tumor cells: implications for p53. *Int J Mol Sci* 2017;**18**:1883.
155. Gupta M, Ansell SM, Novak AJ, Kumar S, Kaufmann SH, Witzig TE. Inhibition of histone deacetylase overcomes rapamycin-mediated resistance in diffuse large B-cell lymphoma by inhibiting Akt signaling through mTORC2. *Blood* 2009;**114**:2926–35.
156. Zhang J, Ng S, Wang J, Zhou J, Tan SH, Yang N, et al. Histone deacetylase inhibitors induce autophagy through FOXO1-dependent pathways. *Autophagy* 2015;**11**:629–42.
157. Eisenberg T, Knauer H, Schauer A, Büttner S, Ruckenstein C, Carmona-Gutierrez D, et al. Induction of autophagy by spermidine promotes longevity. *Nat Cell Biol* 2009;**11**:1305–14.
158. Pietrocola F, Lachkar S, Enot DP, Niso-Santano M, Bravo-San Pedro JM, Sica V, et al. Spermidine induces autophagy by inhibiting the acetyltransferase EP300. *Cell Death Differ* 2015;**22**:509–16.
159. Li G, Ding H, Yu X, Meng Y, Li J, Guo Q, et al. Spermidine suppresses inflammatory DC function by activating the FOXO3 pathway and counteracts autoimmunity. *iScience* 2020;**23**:100807.
160. Yang Q, Zheng C, Cao J, Cao G, Shou P, Lin L, et al. Spermidine alleviates experimental autoimmune encephalomyelitis through inducing inhibitory macrophages. *Cell Death Differ* 2016;**23**:1850–61.
161. Bikle DD. Vitamin D: newly discovered actions require reconsideration of physiologic requirements. *Trends Endocrinol Metabol* 2010;**21**:375–84.

162. Kamen DL, Tangpricha V. Vitamin D and molecular actions on the immune system: modulation of innate and autoimmunity. *J Mol Med* 2010;**88**:441–50.
163. Bizzaro G, Antico A, Fortunato A, Bizzaro N. Vitamin D and autoimmune diseases: is vitamin D receptor (VDR) polymorphism the culprit? *Isr Med Assoc J* 2017;**19**:438–43.
164. Antico A, Tozzoli R, Giavarina D, Tonutti E, Bizzaro N. Hypovitaminosis D as predisposing factor for atrophic type a gastritis: a case-control study and review of the literature on the interaction of vitamin D with the immune system. *Clin Rev Allergy Immunol* 2012;**42**:355–64.
165. Amital H, Szekanez Z, Szücs G, Dankó K, Nagy E, Csépanyi T, et al. Serum concentrations of 25-OH vitamin D in patients with systemic lupus erythematosus (SLE) are inversely related to disease activity: is it time to routinely supplement patients with SLE with vitamin D?. *Ann Rheum Dis* 2010;**69**:1155–7.
166. Lin J, Liu J, Davies ML, Chen W. Serum Vitamin D level and rheumatoid arthritis disease activity: review and meta-analysis. *PLoS One* 2016;**11**:e0146351.
167. Agmon-Levin N, Theodor E, Segal RM, Shoenfeld Y. Vitamin D in systemic and organ-specific autoimmune diseases. *Clin Rev Allergy Immunol* 2013;**45**:256–66.
168. Ahangar-Parvin R, Mohammadi-Kordkhalili M, Azizi SV, Nemati M, Khorramdelazad H, Taghipour Z, et al. The modulatory effects of vitamin D on the expression of IL-12 and TGF- β in the spinal cord and serum of mice with experimental autoimmune encephalomyelitis. *Iran J Pathol* 2018;**13**:10–22.
169. Haghmorad D, Yazdanpanah E, Jadid Tavaf M, Zargarani S, Soltanmohammadi A, Mahmoudi MB, et al. Prevention and treatment of experimental autoimmune encephalomyelitis induced mice with 1,25-dihydroxyvitamin D₃. *Neurol Res* 2019;**41**:943–57.
170. Høyer-Hansen M, Bastholm L, Mathiasen IS, Elling F, Jäättelä M. Vitamin D analog EB1089 triggers dramatic lysosomal changes and Beclin 1-mediated autophagic cell death. *Cell Death Differ* 2005;**12**:1297–309.
171. Yuk JM, Shin DM, Lee HM, Yang CS, Jin HS, Kim KK, et al. Vitamin D₃ induces autophagy in human monocytes/macrophages via cathelicidin. *Cell Host Microbe* 2009;**6**:231–43.
172. Das LM, Binko AM, Traylor ZP, Peng H, Lu KQ. Vitamin D improves sunburns by increasing autophagy in M2 macrophages. *Autophagy* 2019;**15**:813–26.
173. Rintelen B, Andel I, Sautner J, Leeb BF. Leflunomide/chloroquin combination therapy in rheumatoid arthritis: a pilot study. *Clin Rheumatol* 2006;**25**:557–9.
174. Taherian E, Rao A, Malemud CJ, Askari AD. The biological and clinical activity of anti-malarial drugs in autoimmune disorders. *Curr Rheumatol Rev* 2013;**9**:45–62.
175. Zhou S, Chen X, Xue R, Zhou Q, Hu P, Ouyang X, et al. Autophagy is involved in the pathogenesis of experimental autoimmune neuritis in rats. *Neuroreport* 2016;**27**:337–44.
176. Luo X, Liu R, Zhang Z, Chen Z, He J, Liu Y. Mitochondrial division inhibitor 1 attenuates mitophagy in a rat model of acute lung injury. *BioMed Res Int* 2019;**2019**:2193706.
177. Yin J, Zhou Z, Chen J, Wang Q, Tang P, Ding Q, et al. Edaravone inhibits autophagy after neuronal oxygen-glucose deprivation/recovery injury. *Int J Neurosci* 2019;**129**:501–10.
178. Li H, Min J, Mao X, Wang X, Yang Y, Chen Y. Edaravone ameliorates experimental autoimmune thyroiditis in rats through HO-1-dependent STAT3/PI3K/AKT pathway. *Am J Transl Res* 2018;**10**:2037–46.
179. Yoshizaki A, Yanaba K, Ogawa A, Iwata Y, Ogawa F, Takenaka M, et al. The specific free radical scavenger edaravone suppresses fibrosis in the bleomycin-induced and tight skin mouse models of systemic sclerosis. *Arthritis Rheum* 2011;**63**:3086–97.
180. Rossi M, Munarriz ER, Bartesaghi S, Milanese M, Dinsdale D, Guerra-Martin MA, et al. Desmethylclomipramine induces the accumulation of autophagy markers by blocking autophagic flux. *J Cell Sci* 2009;**122**:3330–9.
181. Rossi M, Rotblat B, Ansell K, Amelio I, Caraglia M, Misso G, et al. High throughput screening for inhibitors of the HECT ubiquitin E3 ligase ITCH identifies antidepressant drugs as regulators of autophagy. *Cell Death Dis* 2014;**5**:e1203.
182. Faissner S, Mishra M, Kaushik DK, Wang J, Fan Y, Silva C, et al. Systematic screening of generic drugs for progressive multiple sclerosis identifies clomipramine as a promising therapeutic. *Nat Commun* 2017;**8**:1990.